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United States Department of Agriculture

Agricultural Research Service

Index-Catalogue of Medical and Veterinary Zoology

COMMONWEALTH INSTITUTE OF PARASITOLOGY,
(FORMERLY COMMONWEALTH INSTITUTE OF HELMINTHOLOGY),
395A, HATFIELD ROAD,
ST. ALBANS,
HERTS. AL4 0XU
ENGLAND

Supplement 23, Part 2

Parasite-Subject Catalogue
Parasites: Protozoa

United States Department of Agriculture

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Parasite-Subject Catalogue Parasites: Protozoa

By
Shirley J. Edwards, In Charge
Martha W. Hood, Zoologist
Judith H. Shaw, Zoologist
Jane D. Rayburn, Technical Information Specialist
Margie D. Kirby, Technical Information Specialist
Deborah T. Hanfman, Technical Information Specialist
Judith A. Zidar, Technical Information Specialist

Preface

The Index-Catalogue of Medical and Veterinary Zoology is an index to the world's literature on animal parasites of animals, including man. The Catalogue is distributed to qualified individuals and libraries throughout the world without charge. It has been maintained in cumulative files since 1892. Only the Author Catalogue has been published in its entirety. A revision of the Author Catalogue of the Index-Catalogue of Medical and Veterinary Zoology, consisting of Parts 1 to 18, was published during the period 1932-52. Beginning in 1953, a series of supplements designed to publish the backlog was initiated. This was completed with Supplement 6, published in 1956. From 1956 to 1964, supplements covering authors A to Z were issued on an annual basis.

Beginning with Supplement 15, the Parasite-Subject Catalogues, containing indices to the author references, have been issued. The Author Catalogues of Supplements 15-21 continued the format of previous supplements. Users should note that for each reference in the Author Catalogues of these supplements the author(s) plus the date and letter (e.g., Smith, J.; and Doe, L., 1978 b) are the key to all items in the Parasite-Subject Catalogues derived from that reference. In other words, when using the Parasite-Subject Catalogues of Supplements 15-21, it is necessary to consult the Author Catalogue of the corresponding supplement for complete bibliographic information.

Commencing with Supplement 22, basic bibliographic information is included with each entry in Parts 2-7. It should be emphasized, however, that it will still be useful to consult the Author Catalogue for a variety of other information that may be found there: Title of the reference, translated title, language of text and summaries, issue date, library from which the original may be obtained, published corrections, related references by the same author, and other miscellaneous information.

Each supplement consists of the following parts:

Part 1, Authors: A-Z
Part 2, Parasite-Subject Catalogue: Parasites: Protozoa
Part 3, Parasite-Subject Catalogue: Parasites: Trematoda and Cestoda
Part 4, Parasite-Subject Catalogue: Parasites: Nematoda and Acanthocephala
Part 5, Parasite-Subject Catalogue: Parasites: Arthropoda and Miscellaneous Phyla
Part 6, Parasite-Subject Catalogue: Subject Headings and Treatment
Part 7, Parasite-Subject Catalogue: Hosts

Users should bear in mind that this is an Index-Catalogue, not a treatise, and should not expect to find reasons for any given entry. Nor does citing of synonymy mean that it is necessarily correct. The same statement holds for hosts, locations, localities, authorship of taxa, designation of new taxa, etc. These items are cited as given by the author(s) of the publication being indexed.

The information included in any given supplement represents only the publications that have been indexed in that supplement; and therefore, exclusion of, or limited entries for, any given author or parasite has no significance. No pretension is made for completeness, and assistance in correcting errors or obtaining additional information is appreciated. Reprints of papers on parasitology are requested.

Explanatory Note

Author Catalogue

The Author Catalogue (Part 1 of each supplement) contains full bibliographic information for each publication indexed during the compilation of that supplement. A symbol for the library from which the original publication may be obtained is given at the end of each entry, e.g., Wa, Wm, Wc, etc. A key to these library symbols may be found in Supplements 10 and 20. A list of serial abbreviations new to our files is published at the beginning of each Author Catalogue.

Parasite Catalogues

The Parasite Catalogues (Parts 2-5 of each supplement) are divided by parasite phyla (Protozoa, Trematoda, etc.). They are arranged alphabetically by genera, parasitic diseases, and higher taxa and then alphabetically by species within genera. Entries under each heading are in turn arranged alphabetically by authors and then chronologically for each author. Each entry consists of the name of the parasite or parasitic disease, the author(s) of the publication, date, abbreviated title of the publication, volume, number, inclusive pages, and a subheading. Illustrations of parasites are indicated by the word illus. following the name of the parasite.

Fasciola hepatica, illus.
  Smith, J.; and Doe, L., 1978, J. Parasitol., v. 64 (1), 30-38
    Fasciola hepatica, white mice, successful vaccination with culture incubate antigens and antigens from sonic disruption of immature worms

    Subheading

  Bibliographic information (author line(s))

Entry term

A variety of information is found indented beneath the author line(s) of each entry: Classification, hosts, synonymy, keys, treatment, etc. Subheadings are guides to the subject matter of the publication.

(1) **Classification:** In entries based on systematic articles, the subheading may give the higher taxa in which the taxon has been placed or it may list the lower taxa included in a higher taxon.

(2) **Hosts:** The only hosts recorded are those that pertain directly to the author's own work. Scientific host names are used unless the author gives only common names, in which case the host names are given exactly as in the original publication.

However, when host common names are in Cyrillic alphabet languages, host Latin names are assigned and listed instead of the common name; these are in square brackets [].

Locations of parasites in or on hosts are given in parentheses (). Where a host-parasite relationship is well known, a host may be given under a parasite name and not in the Host Catalogue; this applies particularly to parasites of medical and veterinary importance and of worldwide distribution. A + before the host name on the parasite entry means that no host entry was made for this particular reference.

(3) **Synonymy:** Usually only those synonyms which the author indicates as new, or which are new to the files of the Index-Catalogue of Medical and Veterinary Zoology, are given.

(4) **Keys:** The subheading "key" indicates that the name is included in a taxonomic key.

(5) **Treatment:** When there are several antiparasitic agents mentioned in a publication, a general term is used in the subheading, e.g., anthelmintics, insecticides, protozoacides. However, in the Treatment Catalogue, all agents tested by the investigator(s) are listed.

(6) **Geographic Distribution:** When there are multiple hosts and geographic localities, the appropriate locality is recorded after each host name; when the hosts of a parasite are all from one locality, they are recorded as "all from" this locality.

(7) **Other Subject Matter:** Phrases indicate other subject matter discussed (e.g., immunity, metabolism, morphology, etc.).

Subject Headings Catalogue

The Subject Headings Catalogue (the first section of Part 6 of each supplement) is an alphabetic arrangement of entry terms from a controlled list of subject headings. Each entry consists of the subject heading, bibliographic information, and a subheading reflecting the information contained in the paper. Subject headings with numerous entries are separated into alphabetized subdivisions, e.g.,

Immunity
Immunity, Agglutination
Immunity, Allergy

Treatment Catalogue

In the Treatment Catalogue (a section of Part 6 of each supplement), all entries referring to one antiparasitic agent are grouped under one heading (regardless of the name used by the investigator) and are then listed alphabetically by author. Other names for the same agent are cross-referenced to the name used for filing. When generic and chemical names are available, preference is given to those names as headings rather than to trade names or code numbers and letters. Code number designations for compounds are entered in the Number Index in numerical order and cross-referenced to the name under which they are listed in the alphabetical section. Salts of a compound are usually grouped together, e.g., piperazine adipate, piperazine citrate, etc., are all listed under Piperazine. Sometimes verifying synonymy of drug names is impossible; consequently, groupings and cross-references are not always authenticated although as many as possible have been checked with reliable sources. In some instances, the cross-references are based entirely on information in papers indexed and verification was not possible. Foreign language terminology has been anglicized where feasible. Chemosterilants, Molluscicides, and Repellents are entered under these three collective headings and not under the individual chemical. The format is the same as the parasite entries: Entry term (in this case, drug name), bibliographic information, and subheading.

Host Catalogue

The Host Catalogue (Part 7 of each supplement) is arranged alphabetically by genera, common names, and higher taxa and then alphabetically by species within genera. Nominate subspecies are interfiled with the species. Entries under each heading are in turn arranged alphabetically by author(s) and then chronologically for each author. The format is the same as in the other Catalogues, i.e., entry term (in this case, host name), bibliographic information, and subheading. Indented beneath the author line(s) of each host entry are all the parasites of a particular phylum that were reported from this host in the paper in question. Body locations of these parasites will be found in parentheses () either in the subheading or with the host name. Experimental infection is reported as such. When there are multiple parasites and geographic localities, the appropriate locality is recorded after each parasite name; when the parasites from this host are all from one locality, they are recorded as ''all from'' this locality. When authors use only common names of hosts, scientific names are cautiously supplied from authoritative sources after careful consideration. Cross-references from the common name used by the author to the scientific name supplied by the Index-Catalogue are filed among the host entries. Such supplied names are given in square brackets []. If a scientific name cannot be supplied, English common names are used. Scientific names or English common names are always supplied for common names in Cyrillic alphabet languages, and no cross-references are made. Surveys of parasites of humans and domestic animals are often indexed under geographic headings and entered in Part 6, Subject Headings, in addition to appearing in the Host Catalogue. In this case, all parasite phyla are grouped under the same host entry, and individual parasite entries are not included in the Parasite Catalogue.

Visitors are welcome to come to the Animal Parasitology Institute to use the cumulative files. Arrangements should be made in advance for lengthy visits.

All correspondence should be addressed to:

Index-Catalogue of Medical and Veterinary Zoology
Animal Parasitology Institute
USDA, ARS, BARC-East, Building 1180
Beltsville, Maryland 20705 U.S.A.

It is hoped that these Catalogues will serve as a useful tool to workers in the field of parasitology. Users are requested to preserve the Catalogues, since they are not designed for general distribution and the edition is limited.

The compilers thank the staffs of the National Agricultural Library, the National Library of Medicine, and all other libraries who have aided us invaluably by making publications available to us.

Trade names are used in this publication solely for the purpose of providing specific information. Mention of a trade name does not constitute a guarantee or warranty of the product by the U.S. Department of Agriculture or an endorsement by the Department over other products not mentioned.

Acanthamoeba (Hartmannella), illus.
Ashton, N.; and Stamm, W., 1975, Tr. Ophth. Soc. United Kingdom, v. 95 (2), 214-220
acanthamoebic infections of human eyes, histologic features, diagnostic problems, clinical report

Acanthamoeba
Botto, C.; and Rodriguez, N., 1975, Rev. Uruguaya Patol. Clin. y Microbiol., v. 13 (1), 3-16
human amoebic meningoencephalitis, etiology, epidemiology, pathology, diagnosis, therapy, review

Acanthamoeba
Dutta, G. P., 1979, J. Scient. and Indust. Research, India, v. 38 (4), 199-218
pathogenic and nonpathogenic aerobic free-living amoebae, characterization of cytoplasmic inclusions, cytochemistry and ultrastructure, review

Acanthamoeba
Feldman, R. E., 1979, Acta Bioquim. Clin. Latinoam., v. 13 (3), 309-311
pathogenic free-living amoebae, brief review

Acanthamoeba
Gottlieb, B.; and Reyes, H., 1977, Rev. Med. Chile, v. 105 (7), 467-471
Naegleria, Acanthamoeba, Hartmannella, free-living amoeba now thought to be agents responsible for human meningoencephalitis, diagnosis, pathology, therapy, review

Acanthamoeba
Hoffman, E. O.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (1, pt. 1), 29-38
primary amebic meningoencephalitis, human (brain, pancreas), clinical findings, postmortem studies, electron microscopy, immunohistologic studies, evidence slightly more indicative of Acanthamoeba than Naegleria infection: Louisiana

Acanthamoeba, illus.
Lastovica, A. J., 1977, Proc. Electron Microsc. Soc. South. Africa, v. 7, 145-146
pathogenic Acanthamoeba trophozoites, transmission and scanning electron microscopy

Acanthamoeba, illus.
Martinez, A. J.; and Janitschke, K., 1979, Immun. u. Infekt., v. 7 (2), 57-64
Naegleria fowleri, Acanthamoeba, human, histopathology, protozoology and taxonomy, epidemiology, treatment, animal models, review

Acanthamoeba
Reyes Armijo, E.; and Carrillo Roman, A., 1975, Rev. Med. Hosp. Gen., Mexico, v. 38 (4), 217-225
human amoebic cerebral infections, case reports, pathology, medical and surgical management: Mexico

Acanthamoeba
Rivera, F.; et al., 1979, Tr. Am. Micr. Soc., v. 98 (3), 465-469
protozoa polluting tap water, concentration and identification in culture: Federal District, Mexico City

Acanthamoeba
Scholten, T., 1979, Canad. Med. Ass. J., v. 120 (3), 264, 267-268 [Letter]
soil amoebae potentially pathogenic to man, existence in Canada, results of survey from various areas of Ontario

Acanthamoeba
Simitzis, A. M.; Le Goff, F.; and L'Azou, M. T., 1979, Ann. Parasitol., v. 54 (2), 121-127
free-living amebae, 9 strains isolated from nasal mucosa of 1039 healthy humans sampled, pathogenicity for mice and in cell culture tested

Acanthamoeba-Hartmanella group amoeba, illus.
Stibbs, H. H.; et al., 1979, J. Invert. Path., v. 33 (2), 159-170
Hartmanella-Acanthamoeba group amoeba isolated from Biomphalaria glabrata (Schistosoma mansoni susceptible and resistant strains) (pericardium, mantle), in vitro killing of S. mansoni sporocysts by amoebae; whether amoebae contribute to snail resistance is unknown

Acanthamoeba
Willaert, E., 1974, Ann. Soc. Belge Med. Trop., v. 54 (4-5), 429-440
Naegleria, Acanthamoeba spp. causing disease in man and other vertebrates, selected bibliography and tabular survey of cases

Acanthamoeba [sp.], illus.
Jones, D. B.; Visvesvara, G. S.; and Robinson, N. M., 1975, Tr. Ophth. Soc. United Kingdom, v. 95 (2), 221-232
Acanthamoeba polyphaga keratitis and Acanthamoeba uveitis associated with fatal meningoencephalitis, 3 case reports, extensive clinical presentation, strain characteristics, pathology, diagnostic problems and possibly useful therapeutic agents: Texas

Acanthamoeba sp.
Kadlec, V., 1978, J. Protozool., v. 25 (2), 235-237
15 strains of free-living amebae found in variety of domestic animals, 3 isolates tested were not pathogenic for laboratory animals, some features of isolates differed from those previously known for members of these genera
bull (prepucial cavity)

Acanthamoeba sp., illus.
Lastovica, A. J., 1977, Proc. Electron Microsc. Soc. South. Africa, v. 7, 147-148
pathogenic and non-pathogenic Acanthamoeba cysts, transmission and scanning electron microscopy

Acanthamoeba astronyxis
Gullet, J.; et al., 1979, Am. J. Med., v. 67 (5), 891-896
Acanthamoeba astronyxis, Mexican woman, fatal case of disseminated granulomatous infection with skin lesion and focal encephalitis: San Francisco

Acanthamoeba astronyxis
Kasprzak, W.; Mazur, T.; and Rucka, A., 1974, Ann. Soc. Belge Med. Trop., v. 54 (4-5), 351-357
4 strains of free-living amoebae isolated from lakes in Poland, pathogenicity for mice, response to several drugs, identified as Acanthamoeba spp. on basis of morphology and protein disc electrophoretic patterns

Acanthamoeba castellani, illus.
Brown, T. J.; and Cursons, R. T. M., 1977, Scand. J. Infect. Dis., v. 9 (3), 237-240
Acanthamoeba castellani found in water samples taken from below ice levels of fresh water areas known to be used by swimmers in summer months, study shows that pathogenic-free living amoeba are not necessarily absent in low temperatures: Oslo, Norway

Acanthamoeba castellanii
Cursons, R. T. M.; et al., 1979, J. Parasitol., v. 65 (1), 189-191
Acanthamoeba, Naegleria, semidefined media for cultivation of pathogenic and nonpathogenic free-living amebae

Acanthamoeba castellanii, illus.
Cursons, R. T.; Brown, T. J.; and Keys, E. A., 1978, N. Zealand J. Med. Lab. Tech., v. 32 (1), 11-14
primary amoebic meningo-encephalitis, humans, differential diagnosis and identification of aetiologic agents

Acanthamoeba castellanii (Douglas, 1930) emend. Volkonsky, 1931
Derr-Harf, C.; et al., 1978, Ann. Parasitol., v. 53 (5), 467-477
isolation of 164 strains of free-living amoebae from public water supplies and swimming pools, some strains of Acanthamoeba polyphaga, A. castellanii, A. lenticulata, and A. comandoni were pathogenic for mice, none of the Naegleria isolated were pathogenic for mice, levels of bromine and chlorine used were insufficient to eliminate these amoebae, potential danger to users and suggested control measures: Strasbourg

Acanthamoeba castellani
Jones, B. R.; McGill, J. I.; and Steele, A. D. M., 1975, Tr. Ophth. Soc. United Kingdom, v. 95 (2), 210-213
Acanthamoeba castellani, farmer, recurrent suppurative kerato-uveitis with loss of eye, infection possibly result of handling barley from which Acanthamoeba was isolated: England

Acanthamoeba castellanii
Kasprzak, W.; Mazur, T.; and Rucka, A., 1974, Ann. Soc. Belge Med. Trop., v. 54 (4-5), 351-357
4 strains of free-living amoebae isolated from lakes in Poland, pathogenicity for mice, response to several drugs, identified as Acanthamoeba spp. on basis of morphology and protein disc electrophoretic patterns

Acanthamoeba castellanii, illus.
Markowitz, S. M.; et al., 1978, Am. J. Path. (436), v. 92 (3), 733-743
Acanthamoeba castellanii, mice (exper.), pretreated with methylprednisolone or tetracycline, increased host mortality due to depressed host immunity; potentially pathogenic role for naturally occurring Acanthamoeba sp. in immunosuppressed humans

Acanthamoeba castellanii
Nagington, J., 1975, Tr. Ophth. Soc. United Kingdom, v. 95 (2), 207-209
Acanthamoeba spp. isolated from human eye infections, clinical report, in vitro trials of compounds for possible amoebicidal and cysticidal activity

Acanthamoeba castellanii
Pernin, P.; and Riany, A., 1978, Ann. Parasitol., v. 53 (4), 333-344
survey of swimming pools for presence of free-living amebae, potential danger for swimmers: Lyon

Acanthamoeba castellanii
Seilhamer, J. J.; and Byers, T. J., 1978, J. Protozool., v. 25 (4), 486-489
Acanthamoeba castellanii, mutants resistant to erythromycin, chloramphenicol, and oligomycin

Acanthamoeba comandoni Pussard, 1964
Derr-Harf, C.; et al., 1978, Ann. Parasitol., v. 53 (5), 467-477
isolation of 164 strains of free-living amoebae from public water supplies and swimming pools, some strains of Acanthamoeba polyphaga, A. castellanii, A. lenticulata, and A. comandoni were pathogenic for mice, none of the Naegleria isolated were pathogenic for mice, levels of bromine and chlorine used were insufficient to eliminate these amoebae, potential danger to users and suggested control measures: Strasbourg

Acanthamoeba culbertsoni
Cursons, R. T. M.; et al., 1979, J. Parasitol., v. 65 (1), 189-191
Acanthamoeba, Naegleria, semidefined media for cultivation of pathogenic and nonpathogenic free-living amebae

Acanthamoeba culbertsoni
Cursons, R. T. M.; Brown, T. J.; and Keys, E. A., 1978, J. Parasitol., v. 64 (4), 744-745
Naegleria, Acanthamoeba, pathogenic vs. nonpathogenic strains, differences in level of production of phospholipase A may explain differences in invasiveness and virulence, likely that secretion of enzyme constitutes initial steps whereby host tissue is prepared for endocytosis by these amebae

Acanthamoeba culbertsoni, illus.
Cursons, R. T.; Brown, T. J.; and Keys, E. A., 1978, N. Zealand J. Med. Lab. Tech., v. 32 (1), 11-14
primary amoebic meningo-encephalitis, humans, differential diagnosis and identification of aetiologic agents

Acanthamoeba culbertsoni, illus.
Lastovica, A. J., 1977, Proc. Electron Microsc. Soc. South. Africa, v. 7, 147-148
pathogenic and non-pathogenic Acanthamoeba cysts, transmission and scanning electron microscopy

Acanthamoeba culbertsoni, illus.
Proca-Cioban, M., 1978, Bacteriol., Virusol., Parazitol., Epidemiol., Bucuresti, v. 23 (3), 129-140
Naegleria fowleri, Acanthamoeba culbertsoni, human primary amoebic meningoencephalitis, broad review

Acanthamoeba culbertsoni, illus.
Sarphie, T. G.; and Allen, D. J., 1978, Health Lab. Sc., v. 15 (4), 235-241
Acanthamoeba culbertsoni extracted from cerebrospinal fluid, surface topography, scanning electron microscopy, possible diagnostic application

Acanthamoeba culbertsoni
Willaert, E.; Stevens, A. R.; and Healy, G. R., 1978, J. Clin. Path., v. 31 (8), 717-720
Acanthamoeba culbertsoni, retrospective identification of organism causing fatal amoebic meningoencephalitis in immunosuppressed man, positive results obtained with anti-Acanthamoeba culbertsoni sera when brain sections were stained by indirect immunofluorescence antibody test

Acanthamoeba hatchetti
Simitzis, A. M.; Le Goff, F.; and L'Azou, M. T., 1979, Ann. Parasitol., v. 54 (2), 121-127
free-living amebae, 9 strains isolated from nasal mucosa of 1039 healthy humans sampled, pathogenicity for mice and in cell culture tested

Acanthamoeba lenticulata Molet et Ermolieff-Braun, 1976
  Derr-Harf, C.; et al., 1978, Ann. Parasitol., v. 53 (5), 467-477
    isolation of 164 strains of free-living amoebae from public water supplies and swimming pools, some strains of Acanthamoeba polyphaga, A. castellanii, A. lenticulata, and A. comandoni were pathogenic for mice, none of the Naegleria isolated were pathogenic for mice, levels of bromine and chlorine used were insufficient to eliminate these amoebae, potential danger to users and suggested control measures: Strasbourg

Acanthamoeba polyphaga
  Cursons, R. T.; Brown, T. J.; and Keys, E. A., 1978, N. Zealand J. Med. Lab. Tech., v. 32 (1), 11-14
    primary amoebic meningo-encephalitis, humans, differential diagnosis and identification of aetiologic agents

Acanthamoeba polyphaga
  De Jonckheere, J. F., 1979, Applied and Environment. Microbiol., v. 38 (4), 590-593
    Naegleria and Acanthamoeba, incidence in aquaria, may be source of human infection: Belgium

Acanthamoeba polyphaga (Puschkarew, 1913) emend. Page, 1967
  Derr-Harf, C.; et al., 1978, Ann. Parasitol., v. 53 (5), 467-477
    isolation of 164 strains of free-living amoebae from public water supplies and swimming pools, some strains of Acanthamoeba polyphaga, A. castellanii, A. lenticulata, and A. comandoni were pathogenic for mice, none of the Naegleria isolated were pathogenic for mice, levels of bromine and chlorine used were insufficient to eliminate these amoebae, potential danger to users and suggested control measures: Strasbourg

Acanthamoeba polyphaga, illus.
  Jones, D. B.; Visvesvara, G. S.; and Robinson, N. M., 1975, Tr. Ophth. Soc. United Kingdom, v. 95 (2), 221-232
    Acanthamoeba polyphaga keratitis and Acanthamoeba uveitis associated with fatal meningoencephalitis, 3 case reports, extensive clinical presentation, strain characteristics, pathology, diagnostic problems and possibly useful therapeutic agents: Texas

Acanthamoeba polyphaga
  Kadlec, V., 1978, J. Protozool., v. 25 (2), 235-237
    15 strains of free-living amebae found in variety of domestic animals, 3 isolates tested were not pathogenic for laboratory animals, some features of isolates differed from those previously known for members of these genera
    bull, cow (prepucial cavity, vagina)
    rabbit (liver)
    pigeon, domestic (intestine)
    turkey (intestine)

Acanthamoeba polyphaga
  Kasprzak, W.; Mazur, T.; and Rucka, A., 1974, Ann. Soc. Belge Med. Trop., v. 54 (4-5), 351-357
    4 strains of free-living amoebae isolated from lakes in Poland, pathogenicity for mice, response to several drugs, identified as Acanthamoeba spp. on basis of morphology and protein disc electrophoretic patterns

Acanthamoeba polyphaga
  Nagington, J., 1975, Tr. Ophth. Soc. United Kingdom, v. 95 (2), 207-209
    Acanthamoeba spp. isolated from human eye infections, clinical report, in vitro trials of compounds for possible amoebicidal and cysticidal activity

Acanthamoeba polyphaga
  Pernin, P.; and Riany, A., 1978, Ann. Parasitol., v. 53 (4), 333-344
    survey of swimming pools for presence of free-living amebae, potential danger for swimmers: Lyon

Acanthamoeba pustulosa
  Pernin, P.; and Riany, A., 1978, Ann. Parasitol., v. 53 (4), 333-344
    survey of swimming pools for presence of free-living amebae, potential danger for swimmers: Lyon

Acanthocephalus dujardini (Schneider, 1875) Balbiani, 1884
  Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
    as syn. of Chilogregarina dujardini (Schneider, 1875) comb. n.

Actinocephalidae Leger, 1892
  Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
    diagnosis, includes: Actinocephalinae

Actinocephalinae Leger, 1899
  Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
    Actinocephalidae
    diagnosis, includes: Chilogregarina gen. n.

Actinocephalus acanthaclisis n. sp., illus.
  Marques, A.; and Ormieres, R., 1978, Ztschr. Parasitenk., v. 56 (2), 159-165
    Acanthaclisis baetica
    A. occitanica
    (intestin of both): both from Cote mediterraneenne

Actinocephalus dujardini Schneider, 1875
  Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
    as syn. of Chilogregarina dujardini (Schneider, 1875) comb. n.

Actinocephalus stella (Leger, 1894) Ormieres, 1967
  Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
    as syn. of Chilogregarina stella (Leger, 1894) comb. n.

Actinocephalus striatus Leger & Duboscq, 1903
  Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
    as syn. of Chilogregarina striata (Leger & Duboscq, 1903) comb. n.

Actinocephalus tipulae
  Sherlock, P. L., 1979, Parasitology, v. 78 (2), 207-220
    Tipula paludosa (intestine)

Acuminata subgen. n.
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 23-24
    subgen. of Protoopalina
    type sp.: Protoopalina (Acuminata) regularis Metcalf, 1923

Acuminata
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
    subgen. of Protoopalina, key

Adelea Schneider, 1875
  Levine, N. D., 1977, Protozoology, v. 3, 173-182

Adelea sp. Leger, 1900
  Levine, N. D., 1977, Protozoology, v. 3, 173-182
    as syn. of Adelina legeri n. sp.

Adelea akidium Leger, 1898
  Levine, N. D., 1977, Protozoology, v. 3, 173-182
    as syn. of Adelina akidium (Leger, 1898) Hesse, 1911

Adelea dimidiata (Schneider, 1885) Labbe, 1896
  Levine, N. D., 1977, Protozoology, v. 3, 173-182
    synonymy

Adelea dimidiata var. coccidioides Leger and Duboscq, 1903
  Levine, N. D., 1977, Protozoology, v. 3, 173-182

Adelea (?) dorylaimi Micoletzky, 1921
  Levine, N. D., 1977, Protozoology, v. 3, 173-182
    nomen inquirendum

Adelea hartmanni Chagas, 1910
  Levine, N. D., 1977, Protozoology, v. 3, 173-182
    as syn. of Chagasella hartmanni (Chagas, 1910) Machado, 1911

Adelea hyalospora Narasimhamurti, 1960
  Levine, N. D., 1977, Protozoology, v. 3, 173-182

Adelea mesnili Perez, 1899
  Levine, N. D., 1977, Protozoology, v. 3, 173-182
    as syn. of Adelina mesnili (Perez, 1899) Hesse, 1911

Adelea ovata Schneider, 1875
  Levine, N. D., 1977, Protozoology, v. 3, 173-182

Adelea pachelebrae de Mello, 1921
  Levine, N. D., 1977, Protozoology, v. 3, 173-182

Adelea (?) pervulgata Micoletzky, 1921
  Levine, N. D., 1977, Protozoology, v. 3, 173-182
    nomen inquirendum

Adelea simplex (Schneider, 1885) Labbe, 1896
  Levine, N. D., 1977, Protozoology, v. 3, 173-182
    as syn. of Adelina simplex (Schneider, 1885) Hesse 1911

Adelea tipulae Leger, 1898
  Levine, N. D., 1977, Protozoology, v. 3, 173-182
    as syn. of Adelina tipulae (Leger, 1898) Hesse, 1911

Adelea transita Leger, 1904
  Levine, N. D., 1977, Protozoology, v. 3, 173-182
    as syn. of Adelina transita (Leger, 1904) Hesse, 1911

Adelea zonula Moroff, 1906
  Levine, N. D., 1977, Protozoology, v. 3, 173-182
    as syn. of Adelina zonula (Moroff, 1906) Hesse, 1911

Adeleidae Mesnil, 1903
  Levine, N. D., 1977, Protozoology, v. 3, 173-182
    review of genera and species
    includes: Adelea; Adelina; Chagasella; Ithania; Klossia; Orcheobius

Adelina Hesse, 1911
  Levine, N. D., 1977, Protozoology, v. 3, 173-182

Adelina sp. Barnard, Ernst and Dixon, 1974
  Levine, N. D., 1977, Protozoology, v. 3, 173-182

Adelina sp. Courtney, Forrester, Ernst and Nesbitt, 1975
  Levine, N. D., 1977, Protozoology, v. 3, 173-182

Adelina sp. Hall, Stewart, Arakawa and Strong, 1971
  Levine, N. D., 1977, Protozoology, v. 3, 173-182

Adelina sp. Loschiavo, 1969
  Levine, N. D., 1977, Protozoology, v. 3, 173-182

Adelina sp. Rioux, Leger, Manier and Croset, 1972
  Levine, N. D., 1977, Protozoology, v. 3, 173-182
    as syn. of Adelina riouxi n. sp.

Adelina sp. (Paichuk, 1953) nov. comb.
  Levine, N. D., 1977, Protozoology, v. 3, 173-182
    Syn.: Merocystis sp. Paichuk, 1953

Adelina akidium (Leger, 1898) Hesse, 1911
  Levine, N. D., 1977, Protozoology, v. 3, 173-182
    Syn.: Adelea akidium Leger, 1898

Adelina cryptocerci Yarwood, 1937
  Levine, N. D., 1977, Protozoology, v. 3, 173-182

Adelina deronis Hauschka and Pennypacker, 1942
  Levine, N. D., 1977, Protozoology, v. 3, 173-182

Adelina dimidiata (Schneider, 1885) Hesse, 1911
  Levine, N. D., 1977, Protozoology, v. 3, 173-182
    as syn. of Adelea dimidiata (Schneider, 1885) Labbe, 1896

Adelina legeri n. sp.
  Levine, N. D., 1977, Protozoology, v. 3, 173-182
    Syn.: Adelea sp. Leger, 1900
    Olocrates abbreviatus (hemocoel): Europe

Adelina melolonthae Tuzet, Vago, Ormieres and Robert, 1966
  Levine, N. D., 1977, Protozoology, v. 3, 173-182

Adelina mesnili (Perez, 1899) Hesse, 1911
  Levine, N. D., 1977, Protozoology, v. 3, 173-182
    Syn.: Adelea mesnili Perez, 1899

Adelina octospora Hesse, 1911
  Levine, N. D., 1977, Protozoology, v. 3, 173-182

Adelina riouxi n. sp.
Levine, N. D., 1977, Protozoology, v. 3, 173-182
Syn.: Adelina sp. Rioux, Leger, Manier and Croset, 1972
Sergentomyia m. minuta
Phlebotomus perniciosus
(hemocoel (fat body?) of all): all from Europe

Adelina schellacki Ray and Das Gupta, 1940
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Adelina sericesthis Weiser and Beard, 1959
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Adelina simplex (Schneider, 1885) Hesse 1911
Levine, N. D., 1977, Protozoology, v. 3, 173-182
synonymy

Adelina tenebrionis Sautet, 1930
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Adelina tipulae (Leger, 1898) Hesse, 1911
Levine, N. D., 1977, Protozoology, v. 3, 173-182
Syn.: Adelea tipulae Leger, 1898

Adelina transita (Leger, 1904) Hesse, 1911
Levine, N. D., 1977, Protozoology, v. 3, 173-182
Syn.: Adelea transita Leger, 1904

Adelina tribolii Bhatia, 1937
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Adelina tribolii
Listov, M. V., 1977, Entom. Obozr., v. 56 (4), 731-735
Nosema whitei, Adelina tribolii in Tribolium destructor (nat. and exper.) and T. confusum (exper.), % mortality, effect on metamorphosis, destruction of host fat body causes hormone imbalance

Adelina zonula (Moroff, 1906) Hesse, 1911
Levine, N. D., 1977, Protozoology, v. 3, 173-182
Syn.: Adelea zonula Moroff, 1906

Aduncuperistomatus subgen. n.
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139
subgen. of Nyctotheroides
tod of subgen.: N. (A.) tejerai (Pinto, 1926) Amaro & Sena, 1967

Aegyptianella pullorum
Bauer, F.; Raether, W.; and Seeger, K., 1978, Cahiers Bleus Vet. (27), 265-271
protozoal disease in exper. hosts, enhanced effect of berenil + reverin vs. berenil alone

Aegyptianella pullorum Carpano, 1928
Gothe, R.; and Hartmann, S., 1979, Ztschr. Parasitenk., v. 58 (2), 189-190
Aegyptianella pullorum, cryopreservation does not affect ability to propagate in Argas walkerae

Aegyptianella pullorum
Qureshi, M. I.; and Sheikh, A. H., 1978, Pakistan J. Sc., v. 30 (1-6), 165-167
chickens, Desi, white leghorns: Lahore district
ducks: Lahore Zoo

Aegyptianella pullorum
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Agamococcidiorida ord. n.
Levine, N. D., 1979, J. Protozool., v. 26 (2), 167-168
Coccidiasina
includes: Rhytidocystidae n. fam.

Agmasoma gen. n.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Thelohaniidae fam. n., key
tod: A. penaei (Sprague, 1950) comb. n.

Agmasoma penaei (Sprague, 1950) comb. n. (tod), illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
description, syn.: Thelohania penaei Sprague, 1950

Agmasoma penaei, illus.
Kelly, J. F., 1979, J. Invert. Path., v. 33 (3), 331-339
Thelohania duorara, Agmasoma penaei, and Pleistophora sp. in Penaeus duorarum, pathology, tissue specificity: southern Biscayne Bay

Akiba (or Leucocytozoon) [sp.]
Baker, J. R., 1974, J. Zool., London, v. 172 (2), 169-190
Corvus corone
C. frugilegus
C. monedula
Fringilla coelebs
Garrulus glandarius
Muscicapa striata
Parus caeruleus
Sturnus vulgaria
Turdus ericetorum
T. merula
Strix aluco
Gallinula chloropus
Columba palumbus
Lagopus scoticus
all from Britain

Akiba caulleryi (Leucocytozoon caulleryi)
Krishnan Nair, M.; Rajan, A.; and Sivadas, C. G., 1973, Kerala J. Vet. Sc., v. 4 (2), 181-189
Akiba caulleryi, chickens (blood), Bangkok haemorrhagic disease, pathology, clinical and experimental studies: Kerala

Amblyospora gen. n.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Thelohaniidae, key
tod: A. californica (Kellen and Lipa, 1960) comb. n.

Amblyospora sp., illus.
Andreadis, T. G.; and Hall, D. W., 1979, J. Protozool., v. 26 (3), 444-452
Amblyospora sp., development, ultrastructure, and mode of transmission in Culex salinarius

Amblyospora species, illus.
Hazard, E. I.; et al., 1979, J. Parasitol., v. 65 (1), 117-122
Amblyospora, meiotic configurations and synaptonemal complexes, Parathelohania, synaptonemal complexes; implications in life cycles
Culex salinarius

Amblyospora sp.
Kelly, J. F.; and Knell, J. D., 1979, J. Invert. Path., v. 33 (2), 252
Nosema algerae, Amblyospora sp., modification of Ludox gradient spore purification technique

Amblyospora amphipodae sp. n., illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Crangonyx richmondensis (hepatopancreas): east of Trenton, Florida, USA

Amblyospora benigna (Kellen and Wills, 1962) comb. n.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Syn.: Thelohania benigna Kellen and Wills, 1962

Amblyospora bicortex (Baudoin, 1969) comb. n.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Syn.: Thelohania bicortex Baudoin, 1969

Amblyospora bolinasae (Kellen and Wills, 1962) comb. n.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Syn.: Thelohania bolinasae Kellen and Wills, 1962

Amblyospora bracteata (Strickland, 1913) comb. n.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Syns.: Glugea bracteata Strickland, 1913; Thelohania bracteata: Debaisieux and Gastaldi (in part), 1919 [et auct.]

Amblyospora californica (Kellen and Lipa, 1960) comb. n., illus. (tod)
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
description
Syns.: Thelohania californica Kellen and Lipa, 1960; T. opacita: Weiser (in part), 1961; Nosema lunatum Kellen et al., 1967; Thelohania sp. Chapman, 1966; Tsai et al., 1969

Amblyospora callosa sp. n., illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Rhacophila fuscula (adipose tissue): stream near sewage plant east of Belchertown, Massachusetts

Amblyospora campbelli (Kellen and Wills, 1962) comb. n.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Syn.: Thelohania campbelli Kellen and Wills, 1962

Amblyospora canadensis (Wills and Beaudoin, 1965) comb. n., illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
brief description, syns.: Thelohania inimica canadensis Wills and Beaudoin, 1965; T. inimica: Bailey et al., 1967; T. nr. opacita Anderson, 1968; T. opacitor Fulton et al., 1974; Thelohania sp. Chapman et al., 1966

Amblyospora gigantea (Kellen and Wills, 1962) comb. n.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Syn.: Thelohania gigantea Kellen and Wills, 1962

Amblyospora inimica (Kellen and Wills, 1962) comb. n., illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
description
Syns.: Thelohania inimica Kellen and Wills, 1962; Thelohania sp. Chapman, 1966; Tsai et al., 1969

Amblyospora keenani sp. n., illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Aedeomyia squamipennis (adipose tissue): Chagras River, Canal Zone

Amblyospora khaliulini sp. n., illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Syns.: Thelohania opacita var. mariensis Khaliulin and Ivanov, 1971; T. opacita: Weiser (in part), 1947; Thelohania sp. Noeller, 1920 [et auct.]
Aedes communis (hemolymph, adipose tissue): Manitoba, Canada; Czechoslovakia; Germany; Alaska, U.S.A.; near South Deerfield, Massachusetts, U.S.A.; Mari, U.S.S.R.

Amblyospora lairdi (Weiser, 1965) comb. n.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Syn.: Thelohania lairdi Weiser, 1965

Amblyospora minuta (Kudo, 1924) comb. n., illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
description
Syns.: Thelohania minuta Kudo, 1924; T. rotunda Kudo, 1924; Thelohania sp. Chapman et al., 1967
Culex erraticus (adipose tissue): Florida

Amblyospora mojingensis sp. n., illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Anopheles eiseni (adipose tissue): Mojinga Swamp, Canal Zone

Amblyospora noxia (Kellen and Wills, 1962) comb. n.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Syn.: Thelohania noxia Kellen and Wills, 1962

Amblyospora opacita (Kudo, 1922) comb. n., illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
description, syns.: Thelohania opacita Kudo, 1922; Parathelohania opacita: Simmers, 1974; Thelohania opacitor Fulton et al., 1974; Thelohania sp. Chapman et al., 1967

Amblyospora trichostegiae (Baudoin, 1969) comb. n.
  Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
    Syns.: Thelohania trichostegiae Baudoin, 1969; T. bicortex Baudoin (in part), 1969

Amblyospora unica (Kellen and Wills, 1962) comb. n.
  Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
    Syn.: Thelohania unica Kellen and Wills, 1962

Ameba. See Amoeba.

Amebiasis. See Amoebiasis.

Ameson
  Vivares, C. P.; and Sprague, V., 1979, J. Invert. Path., v. 33 (1), 40-52
    transferred from Nosematidae to Unikaryonidae, distinguished from Perezia

Ameson nelsoni (Sprague, 1950) Sprague, 1977
  Vivares, C. P.; and Sprague, V., 1979, J. Invert. Path., v. 33 (1), 40-52
    as syn. of Perezia nelsoni (Sprague, 1950) comb. n.

Ameson pulvis (Perez, 1905) Sprague, 1977, illus.
  Vivares, C. P.; and Sprague, V., 1979, J. Invert. Path., v. 33 (1), 40-52
    Ameson pulvis, fine structure and its implications for classification and chromosome cycle
    Carcinus mediterraneus: Etang de Vic near
      Montpellier, France
    C. maenas: Roscoff, France
    (muscles of all)

Ameba histolytica
  Fernandez Hernandez, H.; Porro Abdo, Y.; and Ciolli Martinez, M., 1977, Rev. Cubana Med. Trop., v. 29 (3), 153-161
    Ameba histolytica, children with acute diarrhea, clinical trials testing efficacy of aminosidine sulfate: Cuba

Amoeba rotatoria Mayer, 1893
  Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
    as syn. of Trypanosoma rotatorium (Mayer, 1843)

Amoebae, free-living, possibly Hartmanella-Acanthamoeba group
  Campos, R.; et al., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (5), 349-351
    free-living amoebae in cerebrospinal fluid of 8-year-old boy with signs and symptoms of meningoencephalitis: zona rural do Municipio de Capao Bonito, Estado de Sao Paulo, Brasil

Amoebae, "Limax"
  Dive, D.; et al., 1978, Ann. Microbiol., v. 129 B (2), 225-244
    "Limax" amoebae in swimming pools, water purification systems, associated bacterial flora, temperature, pathogenicity to mice: North of France

Amoeba, free living (possibly Vahlkampfiidae), illus.
  Duma, R. J.; Helwig, W. B.; and Martinez, A. J., 1978, Ann. Int. Med., v. 88 (4), 468-473
    unidentified free living amoeba (appeared to be neither Naegleria or Acanthamoeba-Hartmannella, but possibly Vahlkampfiidae) causing fatal primary amoebic meningoencephalitis and brain abscess in diabetic woman, case report, discussion of identifying characteristics, classification and speciation, public health implications: rural Smithfield, Virginia

Amoeba, illus.
  Ferguson, H. W.; et al., 1978, Riv. Ital. Piscicolt. e Ittiopatol., v. 13 (3), 79-85
    amoeba isolated from trout, ultrastructure, scanning and transmission electron microscopy, appears to be ultrastructurally different from parasite associated with proliferative kidney disease: Italy

Amoebae, Limax-type
  Frank, W., 1974, Ann. Soc. Belge Med. Trop., v. 54 (4-5), 343-349
    Limax-amoebae, cold-blooded vertebrates as possible means of spreading parasite from one pond to another
    Boa constrictor
    Iguana iguana
    Coluber hippocrepis
    Morelia sp. (brain)
    Kinosternon leucostomum (")
    Ophiosaurus apodus (")

Amoebae, unidentified
  Kocan, A. A.; Hannon, L.; and Eve, J. H., 1979, Proc. Oklahoma Acad. Sc., v. 59, 20-22
    Colinus virginianus (intestine): Oklahoma

Amoebae, free living, possibly Acanthamoeba, illus.
  Mehta, A. P.; and Guirges, S. Y., 1979, J. Trop. Med. and Hyg., v. 82 (7), 134-136
    acute amoebic dysentery tentively diagnosed as caused by Acanthamoeba, 16-year-old-male, case report, successfully treated with metronidazole: India

Amoebiasis
  Adamali, N.; Wankya, B. M.; and Maneno, J., 1978, East African Med. J., v. 55 (9), 414-416
    amoebic liver abscesses, humans, safe and rapid diagnosis with infusion tomography, case reports: Kenya

Amoebiasis
  Ahumada Padilla, M.; and Montes de Oca M., F., 1975, Rev. Med., Mexico, v. 14 (3), 232-236
    human cutaneous and mucocutaneous amoebiasis, differential diagnosis, pathology, therapy, case reports: Mexico

Amoebiasis
  Aikat, B. K.; et al., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (2), 188-192
    amoebiasis, human hepatic infections, pathology and pathogenesis based on autopsies, mechanisms of evolution and extension of infections, vascular complications, immunological aberrations

Amoebiasis
  Amato Neto, V., 1973, Rev. Paul. Med., v. 81 (2), 93-104
    human protozoan infections, drugs in current use, dosage recommendations, review

Amoebiasis
Ambroise-Thomas, P.; and Desgeorges, P. T., 1978, Bull. World Health Organ., v. 56 (4), 609-613
parasitic diseases, diagnosis, enzyme-linked immunosorbent assay, modified micromethod

Amoebiasis
Anorve Carmona, I.; and Rodriguez Sanchez, A., 1974, Rev. Med. Hosp. Gen., Mexico, v. 37 (5), 331-335
human amoebiasis, clinical trials testing efficacy of emetine aminoxide, drug useful for both amoebiasis and trichocephaliasis

Amoebiasis
Armon, P. J., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (3), 284-288
maternal deaths in the Kilimanjaro region, survey, includes information on amoebic colitis and malaria: Tanzania

Amoebiasis
Arnold, K., 1978, Southeast Asian J. Trop. Med. and Pub. Health, v. 9 (2), 177-185
human parasitic diseases, trends in development of chemotherapy, review

Amoebiasis
Atakpa, T. O. U.; et al., 1978, Ann. Trop. Med. and Parasitol., v. 72 (2), 123-126
human amoebiasis, evaluation of the gel diffusion test for diagnostic purposes, definite seasonal variation in number of clinical cases and number of positive tests for amoebic precipitins (increase during wet season): Nigeria

Amoebiasis
Bieler, E. U.; et al., 1974, South African Med. J., v. 48 (8), 308-320
amoebiasis, human hepatic abscess, scintigraphic evaluation of disease processes and healing rate

Amoebiasis
Bieler, E. U.; Meyer, B. J.; and Jansen, C. R., 1972, South African Med. J., v. 46 (46), 1772-1781
liver diseases, humans, diagnosis using radio-active rose bengal, includes information on amoebic abscesses and echinococcosis

Amoebiasis
Biersack, H. J.; et al., 1977, Therapiewoche, v. 27 (20), 4033-4038
amoebic hepatic abscess, humans, diagnosis and treatment analysis using scintigraphy

Amoebiasis
Bobek, V., 1975, Rozhl. Chir., v. 54 (9), 659-661
amoebiasis, child, intestinal perforation, case report

Amoebiasis
Boero R., D.; and Schurmann, R., 1971, Rev. Chilena Pediat., v. 42 (2), 97-102
hepatic amoebic abscess in children, extensive analysis of pathologic findings of 20 autopsies: Chile

Amoebiasis
Boultbee, J. E.; and Lloyd, D. A., 1979, Brit. J. Radiol. (623), v. 52, 899-901
amoebic liver abscess rupturing into subphrenic space, child, diagnosis using radiography followed by ultrasound

Amoebiasis
Brown, R. C.; et al., 1978, Postgrad. Med. J., London (634), v. 54, 555-558
amoebic hepatic abscesses, human, case reports, potential causes of delay in diagnosis, value of ultrasonic scanning of liver in differential diagnosis

Amoebiasis
Buenemann, H.; Petersen, F.; and Mohr, W., 1976, ROEFO, v. 124 (2), 126-131
human hepatic amoebiasis, size, localization and course of hepatic abscesses evaluated by scintigraphy and compared with clinical symptoms, use in diagnosis

Amoebiasis
Capdevielle, P., 1979, Med. Trop., v. 39 (6), 643-649
digestive endoscopy in tropical environments, special aspects and problems, includes diagnosis of amoebiasis, oxyuriasis, and Schistosoma mansoni involving the gastric mucous membrane: Tananarive, Madagascar

Amoebiasis
Capron, A.; et al., 1974, Tijdschr. Gastroenterol., v. 17 (1), 17-23
hepato-biliary parasitic infections, humans, use of immunologic diagnostic techniques for post-therapeutic evaluations

Amoebiasis
Carrillo Mancero, L.; and Ontaneda Polit, M., 1974, Rev. Med., Quito, v. 1 (1), 16-24
human hepatic amoebic abscess, diagnosis by angiography

Amoebiasis
Cerecedo Cortina, V. B.; et al., 1975, Rev. Med. Hosp. Gen., Mexico, v. 38 (7), 447-457
human hepatic amoebic abscess, diagnosis, peritoneoscopy

Amoebiasis
Cerecedo Cortina, V. B.; et al., 1975, Rev. Med. Hosp. Gen., Mexico, v. 38 (8), 527-531
human hepatic amoebic abscess, ultrasonographic studies on 50 patients, usefulness of this diagnostic method

Amoebiasis
Cerecedo Cortina, V.; and Gaxiola Gaxiola, R., 1972, Rev. Fac. Med., Univ. Nac. Auton. Mexico, an. 15, v. 15 (1), 25-28
human hepatic amoebic abscess, side effects of currently used amoebicides (emetine, chloroquine, metronidazole, aminosidine)

Amoebiasis
Cervantes, L. F.; et al., 1975, Rev. Gastroenterol. Mexico, v. 40 (4), 185-193
human amoebiasis with hepatic abscess, clinical diagnosis confirmed by X-ray, laboratory studies and liver scan, therapy with tinidazole resulted in 93% cure rate: Mexico

Amoebiasis
Chang, W. Y.; and Chen, C. Y., 1970, Taiwan i Hsueh Hui Tsa Chih (J. Formosan Med. Ass.), v. 69 (12), 670-708
liver abscesses, human amoebic and pyogenic, extensive clinical review: Taiwan

Amoebiasis
Cornet, L.; et al., 1975, Rev. Med. Cote Ivoire (37), v. 11, 1. trimestre, 25-36
human gangrenous amoebic colitis, case reports, clinical findings, medical care, indications for surgical intervention

Amoebiasis
Cowie, R. L.; et al., 1972, South African Med. J., v. 46 (49), 1917-1920
amoebic liver abscess, human, prevalence, clinical aspects, diagnostic significance of erythrocyte sedimentation rate and aspiration biopsy: Cape Town

Amoebiasis
Cowie, R. L.; et al., 1979, South African Med. J., v. 55 (11), 402 [Letter]
amoebic liver abscess in patients presenting with jaundice or raised serum bilirubin, suggestions for management

Amoebiasis
Crichton, D.; and Knobel, J., 1973, South African Med. J., v. 47 (42), 2005-2010
prevention of avoidable maternal deaths, includes amoebiasis among causes, incidence survey in obstetric unit of King Edward VIII Hospital, Durban

Amebiasis
Cross, J. H., 1973, Taiwan i Hsueh Hui Tsa Chih (J. Formosan Med. Ass.), v. 72 (7), 407-412
parasitic diseases of human respiratory system, immunodiagnostic methods, review

Amoebiasis
Del Negro, G., 1979, AMB, Rev. Ass. Med. Brasil., v. 25 (3), 113-116
evaluation of the use of corticosteroids in infectious and parasitic diseases, generally contraindicated in parasitic diseases such as toxoplasmosis, malaria, amoebiasis, pneumocystosis

Amoebiasis
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Anaplasma marginale, splenectomized calves (exper.), 2 formulations of oxytetracycline (T-200 and T-50), doxycycline, drug efficacies, influence of treatment on parasitemia and packed cell volume

Anaplasma marginale
Kuttler, K. L.; Young, M. F.; and Simpson, J. E., 1978, Vet. Med. and Small Animal Clin., v. 73 (2), 187, 190-192
Anaplasma marginale, cows (exper.), relative efficacy of a long-acting oxytetracycline (terramycin) compared with a presently available oxytetracycline (liquamycin), both formulations highly effective and resulted in rapid recovery

Anaplasma marginale
Lopez V., G.; and Todorovic, R. A., 1978, Vet. Parasitol., v. 4 (1), 1-9
Babesia argentina, cattle (nat. and exper.), diagnosis by rapid latex agglutination test applicable in field, specificity relative to infections with Babesia bigemina and Anaplasma marginale, comparison of results with complement fixation and indirect fluorescent antibody tests

Anaplasma marginale, illus.
Magonigle, R. A.; and Eckblad, W. P., 1979, Cornell Vet., v. 69 (4), 402-410
Anaplasma marginale in non-splenectomized Cervus canadensis canadensis (exper.), hematologic, serologic, and clinical studies, diagnosis of carrier state by plasma and serum rapid card agglutination tests

Anaplasma marginale
Magonigle, R. A.; Simpson, J. E.; and Frank, F. W., 1978, Am. J. Vet. Research, v. 39 (9), 1407-1410
Anaplasma marginale, cattle (exper.), efficacy of new long-acting oxytetracycline formulation (Liquamycin/LA-200), 1 intramuscular injection, comparison with Liquamycin injectable L-50 administered on 2 consecutive days

Anaplasma marginale
Mallick, K. P.; Dwivedi, S. K.; and Malhotra, M. N., 1979, Indian Vet. J., v. 56 (8), 693-694
Anaplasma marginale, indigenous goats, 5 clinical cases, oxytetracycline hydrochloride: India

Anaplasma marginale, illus.
Mazzola, V.; Amerault, T. E.; and Roby, T. O., 1979, Am. J. Vet. Research, v. 40 (12), 1812-1815
Anaplasma marginale, ultrastructure within and outside Aedes albopictus cells in vitro, reproduction not seen

Anaplasma marginale
Michael, S. A.; and El Refaii, A. H., 1977, Bull. Animal Health and Prod. Africa, v. 25 (3), 229-231
Anaplasma marginale, buffaloes, use of both capillary tube agglutination test and blood-film examination recommended for survey and diagnostic purposes: Egypt

Anaplasma marginale
Minami, T., 1977, Japan Agric. Research Quart., v. 11 (4), 234-238
Japanese Babesia sp., cattle (exper.), capillary-tube agglutination test, discrimination from B. bigemina and B. argentina was almost possible, no cross reactions with other protozoa

Anaplasma marginale
Mishra, A. K.; and Sharma, N. N., 1979, Trop. Animal Health and Prod., v. 11 (4), 222-226
Anaplasma marginale, calves (exper.), comparative efficacy of several drugs

Anaplasma marginale
Molina, J. Q.; and Montenegro, M. M., 1976, Philippine J. Animal Indust., v. 31 (1-4), 87-92
crossbred cows, case report: Philippines

Anaplasma marginale, illus.
Molina, J. Q.; and Montenegro, M. M., 1977, Philippine J. Vet. Med., v. 16 (1-2), 181-186
Anaplasma marginale, case report, two crossbred cows (blood), clinical signs, hematological picture, pathology: Philippines

Anaplasma marginale
Obi, T. U., 1978, Trop. Animal Health and Prod., v. 10 (2), 87-90
Anaplasma marginale, incidence survey, Zebu trade cattle: Nigeria

Anaplasma marginale
Osorno, B. M.; et al., 1973, Tecn. Pecuaria Mexico (24), 57-63
Anaplasma marginale, cattle, University of Illinois attenuated vaccine highly effective and safe, challenged with virulent Mexican Anaplasma strain

Anaplasma marginale
Parker, R.; Parker, M. L.; and Wilson, A. J., 1978, Research Vet. Sc., v. 25 (3), 401-402
Anaplasma marginale complement fixation test antigens, liquid nitrogen storage by a multiple small aliquot technique

Anaplasma marginale
Patarroyo, J. H.; Villa, O.; and Diazgranados, H., 1978, Trop. Animal Health and Prod., v. 10 (3), 171-174
Anaplasma marginale, cattle, prevalence and distribution, card test: Colombia

Anaplasma marginale Theiler
Peterson, K. J.; Goulding, R. L.; and Turner, H. T., 1976, Proc. 80. Ann. Meet. U. S. Animal Health Ass., 66-77
Anaplasma marginale, inability to survive natural winter conditions on Dermacentor andersoni-infested pastures in absence of infected cattle, results suggest that anaplasmosis can be eliminated in selected herds: Oregon

Anaplasma marginale
Quinlan, J. F., 1978, Trop. Animal Health and Prod., v. 10 (3), 178
Babesia bovis, B. bigemina, Anaplasma marginale, presence in Friesian calves placed in Boophilus microplus infested environment and later splenectomised: Paraguay

Anaplasma marginale
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Anaplasma marginale
Renshaw, H. W.; et al., 1976, Proc. 80. Ann. Meet. U. S. Animal Health Ass., 79-88
Anaplasma marginale, cattle, some long-lasting immunity persists after elimination of carrier status with oxytetracycline hydrochloride

Anaplasma marginale
Rivas, A.; Rodriguez, O. N.; and Espaine, L., 1977, Rev. Cubana Cien. Vet., v. 8 (1), 13-20
Anaplasma marginale, bovine, immunofluorescence and complement fixation tests evaluated: Cuba

Anaplasma marginale
Roby, T. O.; Simpson, J. E.; and Amerault, T. E., 1978, Am. J. Vet. Research, v. 39 (7), 1115-1116
Anaplasma marginale, calves (exper.), elimination of carrier state using long-acting formulation of oxytetracycline

Anaplasma marginale, illus.
Rodriguez, O. N.; et al., 1975, Folia Vet., v. 19 (1-2), 221-231
ganado vacuno (eritrocitos): Cuba

Anaplasma marginale
Rodriquez, O. N.; et al., 1975, Folia Vet., v. 19 (1-2), 243-248
Anaplasma marginale, isolation of DNA and its basic components (adenin, cytosine, guanin, and thymin) by microspectro-photometric methods

Anaplasma marginale
Rodriguez, O. N.; et al., 1975, Folia Vet., v. 19 (1-2), 249-255
Anaplasma marginale, Paranaplasma caudata, antigenic differences determined by passive hemagglutination and hemagglutination inhibition test

Anaplasma marginale
Rodriguez, O. N.; et al., 1978, Rev. Cubana Cien. Vet., v. 9 (1), 87-94
Anaplasma marginale, Babesia argentina, B. bigemina, cattle of different breeds, serodiagnosis, complement fixation and capillary agglutination microtechniques: Cuba

Anaplasma marginale
Rogers, R. J.; Blight, G. W.; and Knott, S. G., 1978, Austral. Vet. J., v. 54 (3), 115-120
Anaplasma marginale in Bos taurus and B. indicus types, clinical outbreaks, serological survey, seasonal distribution, age, sex, and breed of host, high prevalence in Boophilus microplus infested areas: southern Queensland

Anaplasma marginale
Rose, J. E.; et al., 1978, Am. J. Vet. Research, v. 39 (5), 791-793
Anaplasma marginale, cattle (exper.), conglutinin, immunoconglutinin, and complement levels in peracute and acute stages of infection, study of disease process, possible improvement of card agglutination test

Anaplasma marginale, illus.
Sharma, S. K.; Banerjee, D. P.; and Gautam, O. P., 1978, Indian J. Animal Health, v. 17 (2), 105-110
Anaplasma marginale in Bubalus bubalis (exper.), clinical course, haematological changes, effect of immunosuppressants

Anaplasma marginale
Sinha, R. P.; and Dubey, R. K., 1978, Indian Vet. J., v. 55 (5), 372-376
Anaplasma marginale outbreak in non-preimmune Jersey cattle imported from United States and Denmark to farm in Bihar, clinical symptoms and pathological findings, epizootiological factors responsible for outbreak (high ambient temperature, stress of vaccination for rinderpest virus, presence of tick vectors), control achieved through chemotherapy of sick and healthy animals, removal of vectors, and housing in cool sheds; outbreaks in exotic herds could be avoided if cattle were imported in early winter: India

Anaplasma marginale
Stewart, C. G.; et al., 1979, J. South African Vet. Ass., v. 50 (2), 83-85
Anaplasma marginale, splenectomized calves (nat. and exper.), oxytetracycline, long- vs. short-acting formulation

Anaplasma marginale
Sweet, V. H.; and Stauber, E. H., 1978, J. Am. Vet. Med. Ass., v. 172 (11), 1310-1312
Anaplasma marginale, cattle, serologic survey, chlortetracycline, oxytetracycline, good results, concluded that test and treatment program would need to be continued for several years to achieve anaplasmosis-free status: northern Idaho; southeastern Washington

Anaplasma marginale
Swift, B. L.; and Paumer, R. J., 1978, Theriogenology, v. 10 (5), 395-403
Anaplasma marginale, heifers in third trimester of gestation (exper.), fetus and dam arterial blood gases and pH measured, death of fetus following progressive parasitic anemia in dam is attributed to fetal anoxia

Anaplasma marginale
Swift, B. L.; Settlemire, J., jr.; and Thomas, G. M., 1978, Theriogenology, v. 10 (6), 481-485
Anaplasma marginale, pregnant heifers (exper.), oxytetracycline hydrochloride, did not abort and transplacental transmission did not occur

Anaplasma marginale
Thompson, K. C.; et al., 1978, Trop. Animal Health and Prod., v. 10 (2), 75-81
Anaplasma marginale, Babesia argentina, B. bigemina, cattle under tropical conditions, immunization with virulent organisms followed by drug therapy (ganaseg; gloxazone; emicina) vs. chemoprophylaxis (imidocarb); tick and gastrointestinal parasite control without haemoparasitic control had advantage over no control system at all

Anaplasma marginale
Thompson, K. C.; et al., 1978, Trop. Animal Health and Prod., v. 10 (3), 141-144
Anaplasma marginale, Babesia bigemina, B. argentina (bovis), immunization and chemoprophylaxis of Bos taurus calves and subsequent challenge with Boophilus microplus, economic gain estimated: Colombia

A[naplasma] marginale
Todorovic, R. A.; Gonzalez, E. F.; and Garcia, O., 1979, Tropenmed. u. Parasitol., v. 30 (1), 43-52
anaplasmosis, babesiosis, cattle, immunization evaluated under field conditions, study indicates significant reduction in deaths and production losses and economic benefits for livestock producers when animals are vaccinated: Cauca River Valley, Colombia

Anaplasma marginale
Todorovic, R. A.; Gonzalez, E. F.; and Garcia, O., 1979, Tropenmed. u. Parasitol., v. 30 (2), 236-238
Anaplasma marginale, cattle (exper.), new long-acting injectable oxytetracycline (terramycin/LA) compared with commercial preparation (emicina)

Anaplasma marginale
Todorovic, R.; Gonzalez, E.; and Lopez, G., 1978, Tropenmed. u. Parasitol., v. 29 (2), 210-214
Anaplasma marginale, Babesia spp., calves, immunization, cryo-preserved vaccines, effects of dose, inoculation route, time, and temperature

Anaplasma marginale
Vizcaino, O.; et al., 1978, Am. J. Vet. Research, v. 39 (2), 229-233
Anaplasma marginale, response of calves inoculated with 3 different doses of attenuated A. marginale vaccine and subsequently challenged with a virulent strain; effects of field challenge exposure in calves inoculated with Anaplasma vaccine and premunized with both Babesia bigemina and B. argentina: Colombia

Anaplasma marginale
Wilson, A. J.; et al., 1978, Austral. Vet. J., v. 54 (8), 383-386
Anaplasma marginale, cattle, comparison of 4 serological tests for detection of humoral antibodies (capillary agglutination, complement fixation, plate agglutination, and indirect fluorescent antibody)

Anaplasma marginale
Wilson, A. J.; et al., 1979, Austral. Vet. J., v. 55 (2), 71-73
Anaplasma marginale, cattle and splenectomised calves (exper.), long-acting formulation of tetracycline compared with standard tetracycline and imidocarb dipropionate

Anaplasma marginale
Wilson, A. J.; and Trueman, K. F., 1978, Austral. Vet. J., v. 54 (3), 121-124
Anaplasma marginale, Bos indicus cross steers (exper.), effects of reduced energy intake on humoral antibody response, parasitaemia, body weight, packed cell volumes, and plasma protein values

Anaplasma marginale
Zaraza, H.; and Kuttler, K. L., 1976, Rev. Inst. Colomb. Agropec., v. 11 (4), 363-371
Anaplasma marginale, calves, comparative immunization systems using attenuated organism, a killed adjuvant, and virulent A. marginale

Anaplasma marginale
Zaraza O., H.; and Parra G., D., 1977, Rev. Inst. Colomb. Agropec, v. 14 (4), 457-471
Anaplasma marginale, calves, pathogenesis of a virulent vs. non-virulent Columbian strain, possible application for immunization

Anaplasma mesaeterum sp. n., illus.
Uilenberg, G.; van Vorstenbosch, C. J. A. H. V.; and Perie, N. M., 1979, Vet. Quart., v. 1 (1), 14-22
Anaplasma mesaeterum sp. n., sheep (nat. and exper.) and goats (exper.), pathogenicity, oxytetracycline treatment, cross-immunity tests with A. ovis: Ameland, the Netherlands

Anaplasma ovis
Charyev, O. Ch., 1977, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (6), 70-71
piroplasmids, sheep, pure and mixed infections: Turkmenia

Anaplasma ovis
Gupta, S. L.; and Gautam, O. P., 1978, Indian J. Animal Sc., v. 48 (12), 917
Anaplasma ovis, sheep (exper.), no apparent variation in blood sugar values observed during infection

Anaplasma ovis Lestoquard, 1924
Uilenberg, G.; van Vorstenbosch, C. J. A. H. V.; and Perie, N. M., 1979, Vet. Quart., v. 1 (1), 14-22
Anaplasma mesaeterum sp. n., sheep (nat. and exper.) and goats (exper.), pathogenicity, oxytetracycline treatment, cross-immunity tests with A. ovis: Ameland, the Netherlands

Anaplasmosis
Goodger, W. J.; Carpenter, T.; and Riemann, H., 1979, J. Am. Vet. Med. Ass., v. 174 (12), 1333-1336
anaplasmosis in beef cattle, estimation of economic loss in 1976: California

Anaplasmosis
Matthews, M., 1978, J. Zoo Animal Med., v. 9 (4), 148
anaplasmosis, Siberian ibex, case history: hunting ranch

Anaplasmosis
Miron, C. A., 1978, Agric. El Salvador, v. 17 (1), 12-24
anaplasmosis, bovine, prevalence, complement fixation test, no difference in regard to age, sex or breed of host: San Miguel, El Salvador

Anaplasmosis
Mordasov, P. M.; Golovnev, V. I.; and Zakharik, N. V., 1977, Vet. Nauka--Proizvod., Trudy, Minsk, v. 15, 93-95
anaplasmosis, bovine, treatment with oxytetracycline

Anaplasmosis
Norval, R. A. I., 1978, Rhodesian Vet. J., v. 9 (1-4), 9-16
ticks, cattle, significant increase in tick infestations and outbreaks of tick-borne diseases following collapse of dipping (1973-1978): African areas in Rhodesia

Anaplasmosis
Reshetniak, V. Z.; Bartenev, V. S.; and Firsov, N. F., 1977, Veterinariia, Moskva (10), 79
anaplasmosis, sheep outbreak, pathology, oxytetracycline effective treatment; Hyalomma scupense as probable vector, hexachlorane in creolin emulsion for control; no further anaplasmosis or H. scupense found: Rostovsk oblast

Anaplasmosis
Rodriguez, O. N.; et al., 1977, Rev. Cubana Cien. Vet., v. 8 (1), 71-77
anaplasmosis, bovine (exper.), clinical picture and course of infection, pure infection uncomplicated by other hemoparasites

Anaplasmosis
Rodriguez, O. N.; et al., 1977, Rev. Cubana Cien. Vet., v. 8 (2), 9-12
anaplasmosis, bovine, diagnosis by complement fixation in perspex plates

Anaplasmosis
Seifert, H., 1977, 3. Externe Veterinartagung, Ber. u. Arbeitsergebn. (40), 229-234
anaplasmosis, ruminants, epizootiology, immunity, prophylaxis, and pathology, review

Ancyrophora hispani n. sp., illus.
Massot, N.; and Ormieres, R., 1979, Ann. Parasitol., v. 54 (3), 267-275
Chrysocarabus hispanus (intestin): Martres (Aveyron)

Ancyrophora penetreti n. sp., illus.
Massot, N.; and Ormieres, R., 1979, Ann. Parasitol., v. 54 (3), 267-275
Penetretus rufipennis (intestin): Herault (St-Pons)

Andreula Kahl, 1934
Iankovskii, A. V., 1973, Parazitologiia, Leningrad, v. 7 (3), 214-219
Hymenostomatida, Tetrahymenina, Entodiscidae fam. n.

Anglasia Jean Delphy
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key

Angusta subgen. n.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34
subgen. of Opalina
type sp.: Opalina (Angusta) obtrigona Stein, 1864

Angusta
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
subgen. of Opalina, key

Angusta Amaro, 1966
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
subgen. of Opalina
key to species

Anoplophrya Stein
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key

Anoplophryidae
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key

Anoplophryimorphes
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key to families, subfamilies, and genera

Anthemosoma garnhami
Momen, H., 1979, Ann. Trop. Med. and Parasitol., v. 73 (2), 109-115
Plasmodium, Babesia, and Anthemosoma spp. in mouse erythrocytes, identification of enzymes of parasite origin using starch-gel electrophoresis

Anthemosoma garnhami
Momen, H., 1979, Ann. Trop. Med. and Parasitol., v. 73 (2), 117-121
Plasmodium, Babesia and Anthemosoma spp., comparative study of glucose catabolism by infected mouse erythrocytes, glucose utilization and lactate production of parasites

Anthemosoma garnhami
Momen, H.; Chance, M. L.; and Peters, W., 1979, Ann. Trop. Med. and Parasitol., v. 73 (3), 203-212
Babesia spp., human and animal strains and Anthemosoma garnhami, taxonomic differentiation using biochemical methods (variation in mobility of parasite enzymes on starch gels after electrophoresis, measurement of buoyant densities of DNA)

Anthonyella hoplites Rossolimo [nov. comb.] (tod)
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519

Anthorhynchus hanumanthi n. sp., illus.
Kalavati, C.; and Narasimhamurti, C. C., 1978, Proc. Indian Acad. Sc., Sect. B, Animal Sc., v. 87 (12), 397-404
Odontotermes sp. (midgut, faecal matter): Andhra University at Waltair

Antunesia subgen. n.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 25-27
subgen. of Zelleriella
type sp.: Zelleriella (Antunesia) antunesi Pessoa, 1934

Antunesia
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
subgen. of Zelleriella, key

Antunesia Amaro, 1966
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
subgen. of Zelleriella
key to species

Apicomplexa
Markus, M. B., 1978, South African J. Sc., v. 74 (3), 105-106
Sporozoa, subphylum Apicomplexa, terminology for invasive stages, only two basic terms (sporozoite and merozoite) are currently recognized

Apiosoma sp.
Armas, G., 1979, J. Fish Dis., v. 2 (6), 543-547
Mugil cephalus (body surface, gills): Rio Moche coastal lagoon, northern Peru

Apiosoma sp.
Iziumova, N. A.; Mashtakov, A. V.; and Kashkovskii, V. V., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36) 64-68
[Lucioperca lucioperca] (gills): Kamsk reservoir

Apiosoma sp.
Lester, R. J. G., 1974, Syesis, v. 7, 195-200
Gasterosteus aculeatus (gills, body surface): near Vancouver, British Columbia

Apiosoma amoebae
Chernysheva, N. B., 1975, Izvest. Gosudarstv. Nauchno-Issled. Inst. Ozer. i Rech. Ryb. Khoziaistva, Leningrad, v. 93, 117-120
protozoans of young predatory fish, extent of infection correlated with some environmental factors
[Esox lucius]: Vrevo Lake, Leningrad district

Apiosoma amurensis sp. n. Banina and Iukhimenko, illus.
Iunchis, O. N.; Banina, N. N.; and Iukhimenko, S. S., 1975, Parazitologiia, Leningrad, v. 9 (1), 93-95
[Phoxinus]: Kaban'ia river (Amura tributary)

Apiosoma baueri
Chernysheva, N. B., 1975, Izvest. Gosudarstv. Nauchno-Issled. Inst. Ozer. i Rech. Ryb. Khoziaistva, Leningrad, v. 93, 117-120
protozoans of young predatory fish, extent of infection correlated with some environmental factors
[Esox lucius]: Vrevo Lake, Leningrad district

Apiosoma constricta
Chernysheva, N. B., 1975, Izvest. Gosudarstv. Nauchno-Issled. Inst. Ozer. i Rech. Ryb. Khoziaistva, Leningrad, v. 93, 117-120
protozoans of young predatory fish, extent of infection correlated with some environmental factors
[Perca fluviatilis]: Vrevo Lake, Leningrad district

Apiosoma fungiformis sp. n. Banina and Iukhimenko, illus.
Iunchis, O. N.; Banina, N. N.; and Iukhimenko, S. S., 1975, Parazitologiia, Leningrad, v. 9 (1), 93-95
[Aspius aspius]: Far East

Apiosoma lopuchinae sp. n. Iunchis, illus.
Iunchis, O. N.; Banina, N. N.; and Iukhimenko, S. S., 1975, Parazitologiia, Leningrad, v. 9 (1), 93-95
[Rutilus rutilus]: lake Vrevo, Leningradsk oblast

Apiosoma megamicronucleata
Chernysheva, N. B., 1975, Izvest. Gosudarstv. Nauchno-Issled. Inst. Ozer. i Rech. Ryb. Khoziaistva, Leningrad, v. 93, 117-120
protozoans of young predatory fish, extent of infection correlated with some environmental factors
[Lota lota]
[Esox lucius]
all from Vrevo Lake, Leningrad district

Apiosoma minimicronucleata
Chernysheva, N. B., 1975, Izvest. Gosudarstv. Nauchno-Issled. Inst. Ozer. i Rech. Ryb. Khoziaistva, Leningrad, v. 93, 117-120
protozoans of young predatory fish, extent of infection correlated with some environmental factors
[Perca fluviatilis]: Vrevo Lake, Leningrad district

Apiosoma olae sp. n. Iunchis, illus.
Iunchis, O. N.; Banina, N. N.; and Iukhimenko, S. S., 1975, Parazitologiia, Leningrad, v. 9 (1), 93-95
[Rutilus rutilus]
[Alburnus alburnus]
[Leuciscus idus]
all from lake Vrevo, Leningradsk oblast

Apiosoma piscicola
Chernysheva, N. B., 1975, Izvest. Gosudarstv. Nauchno-Issled. Inst. Ozer. i Rech. Ryb. Khoziaistva, Leningrad, v. 93, 117-120
protozoans of young predatory fish, extent of infection correlated with some environmental factors
[Esox lucius]
[Perca fluviatilis]
all from Vrevo Lake, Leningrad district

Apiosoma schulmani
Chernysheva, N. B., 1975, Izvest. Gosudarstv. Nauchno-Issled. Inst. Ozer. i Rech. Ryb. Khoziaistva, Leningrad, v. 93, 117-120
protozoans of young predatory fish, extent of infection correlated with some environmental factors
[Lota lota]: Vrevo Lake, Leningrad district

Apodinium [sp.], illus.
Cachon, J.; and Cachon, M., 1979, Arch. Protistenk., v. 122 (3-4), 267-274
Apodinium [sp.], morphology, singular kinetochore structure

Apolocystis herculea (Bosanquet, 1894) Meier, 1956, illus.
Segun, A. O., 1978, J. Protozool., v. 25 (2), 157-162
description
Libyodrilus violaceus (perivisceral coelom): Nigeria

Apolocystis iridodrilii sp. n., illus.
Segun, A. O., 1978, J. Protozool., v. 25 (2), 157-162
Iridodrilus roseus (seminal vesicles): Oshogbo and Ile-Ife, Western Nigeria; Benin, Sapele, Warri, and Ughelli, Mid-western Nigeria
I. preussi (seminal vesicles): Vom and Jos, Benue Plateau, Nigeria

Apolocystis libyodrilii sp. n.
Segun, A. O., 1978, J. Protozool., v. 25 (2), 157-162
Libyodrilus violaceus (seminal vesicles): Ibadan, Western Nigeria

Arachnocystis gen. n.
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
Hirmocystidae
tod: A. arachnoidea (Devdhar & Gourishankar, 1971) comb. n.

Arachnocystis arachnoidea (Devdhar & Gourishankar, 1971) comb. n. (tod)
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
Syn.: Sycia arachnoidea Devdhar & Gourishankar, 1971
Opalnia sp. (intestine): India

Arachnocystis nitida (Geus, 1969) comb. n.
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
Syn.: Gregarina nitida Geus, 1969
Oribata clavipes (intestine): Germany

Arachnocystis oribatae (Geus, 1969) comb. n.
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
Syn.: Gregarina oribatae Geus, 1969
Oribata geniculata (intestine): Germany

Arachnocystis scutovertexi (Erhardova, 1955) comb. n.
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
Syn.: Gregarina scutovertexi Erhardova, 1955
Scutovertex minutus: Czechoslovakia

Ascocystis barretti
Miller, B. R.; and DeFoliart, G. R., 1979, Am. J. Trop. Med. and Hyg., v. 28 (6), 1064-1066
Ascocystis barretti does not seem to be mechanism for dispersal of La Crosse virus infection via Aedes triseriatus larvae nor does concomitant parasite infection increase virus infection in larvae

Astasia ovorum, illus.
Michajlow, W., 1977, Zhurnal Obsh. Biol., v. 38 (5), 657-675

Atoxoplasma sp.
Greiner, E. C.; and Mundy, P. J., 1979, J. Parasitol., v. 65 (1), 147-153
hematozoa from southern African vultures, prevalence, host age class, seasonal variation
Torgos tracheliotus
Gyps africanus
all from Rhodesia

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Auerbachiidae new family

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Eurysporea
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Trypanosoma, Plasmodium, Babesia, antigenic variation (nature, consequences for protective immunity, possible implications for other protozoan infections), colloquium presentation

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immune response to Babesia, review

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Sorex araneus
S. minutus
Talpa europaea
Pipistrellus pipistrellus
Apodemus sylvaticus
Arvicola amphibius
Clethrionomys glareolus
Micromys minutus
Microtus agrestis
all from Britain

Babesia sp. (Japanese Babesia sp.)
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Japanese Babesia sp., cattle (exper.), capillary-tube agglutination test, discrimination from B. bigemina and B. argentina was almost possible, no cross reactions with other protozoa

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Babesia bigemina and Japanese Babesia species can be serologically differentiated by the complement fixation and capillary-tube agglutination tests

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Johnston, L. A. Y.; Leatch, G.; and Jones, P. N., 1978, Austral. Vet. J., v. 54 (1), 14-18
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Babesia argentina
Lopez V., G.; and Todorovic, R. A., 1978, Vet. Parasitol., v. 4 (1), 1-9
Babesia argentina, cattle (nat. and exper.), diagnosis by rapid latex agglutination test applicable in field, specificity relative to infections with Babesia bigemina and Anaplasma marginale, comparison of results with complement fixation and indirect fluorescent antibody tests

Babesia argentina
Minami, T., 1977, Japan Agric. Research Quart., v. 11 (4), 234-238
Japanese Babesia sp., cattle (exper.), capillary-tube agglutination test, discrimination from B. bigemina and B. argentina was almost possible, no cross reactions with other protozoa

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Babesia argentina, cattle, experimental transmission through larvae of Boophilus microplus: Assam

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Rivas, A.; Rodriguez, O. N.; and Espaine, L., 1977, Rev. Cubana Cien. Vet., v. 8 (1), 1-11
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Anaplasma marginale, Babesia argentina, B. bigemina, cattle of different breeds, serodiagnosis, complement fixation and capillary agglutination microtechniques: Cuba

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Suh, M. D.; and Hoyte, H. M. D., 1977, Korean J. Vet. Research, v. 17 (2), 51-61
Babesia bigemina and B. argentina, morphological differentiation using smears and tissue sections of splenectomized calf organs (liver, lung, kidney, heart, lymph node, brain)

Babesia argentina
Thompson, K. C.; et al., 1978, Trop. Animal Health and Prod., v. 10 (2), 75-81
Anaplasma marginale, Babesia argentina, B. bigemina, cattle under tropical conditions, immunization with virulent organisms followed by drug therapy (ganaseg; gloxazone; emicina) vs. chemoprophylaxis (imidocarb); tick and gastrointestinal parasite control without haemoparasitic control had advantage over no control system at all

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Thompson, K. C.; et al., 1978, Trop. Animal Health and Prod., v. 10 (3), 141-144
Anaplasma marginale, Babesia bigemina, B. argentina (bovis), immunization and chemoprophylaxis of Bos taurus calves and subsequent challenge with Boophilus microplus, economic gain estimated: Colombia

Babesia argentina
Todorovic, R.; and Garcia, R., 1978, Tropenmed. u. Parasitol., v. 29 (1), 88-94
Babesia spp., cattle, diagnosis, indirect fluorescent antibody test, comparison between techniques of dried blood on filter paper and serum samples, sensitivity and specificity, laboratory and field conditions: Colombia

Babesia argentina
Todorovic, R.; Gonzalez, E.; and Lopez, G., 1978, Tropenmed. u. Parasitol., v. 29 (2), 210-214
Anaplasma marginale, Babesia spp., calves, immunization, cryo-preserved vaccines, effects of dose, inoculation route, time, and temperature

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Vizcaino, O.; et al., 1978, Am. J. Vet. Research, v. 39 (2), 229-233
Anaplasma marginale, response of calves inoculated with 3 different doses of attenuated A. marginale vaccine and subsequently challenged with a virulent strain; effects of field challenge exposure in calves inoculated with Anaplasma vaccine and premunized with both Babesia bigemina and B. argentina: Colombia

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Wright, I. G.; and Goodger, B. V., 1977, Ztschr. Parasitenk., v. 53 (1), 63-73
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Babesia bigemina
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Anaplasma marginale, Babesia bigemina, B. argentina, Boophilus microplus, cattle, prevalence and distribution: eastern plains of Colombia

Babesia bigemina
Corrier, D. E.; et al., 1979, Trop. Animal Health and Prod., v. 11 (4), 215-221
Babesia bigemina- and B. bovis-immunized Bos taurus calves transported to lowland tropics, exposed to heavy vs. light Boophilus microplus infestation, resulting B. bigemina, and B. bovis parasitemias, mortality, weight loss, and anemia: Caribbean Coast, Colombia

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Anaplasma marginale, Babesia bigemina, age at which calves in endemic area first become infected, packed cell volumes and clinical symptoms, mild clinical response and rapid recovery indicate that protection of native calves through immunization or other procedures prior to natural exposure would be of questionable value: north coast area of Colombia

Babesia bigemina
Dalgliesh, R. J.; and Stewart, N. P., 1978, Austral. Vet. J., v. 54 (9), 453-454 [Letter]
Babesia bigemina, B. bovis, development, in incubated Boophilus microplus eggs, and B. bigemina in unfed larval ticks held at 37°C, infectivity for calves, results indicate that high environmental temperature may be only stimulus required for development of infective Babesia within the tick

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Babesia bigemina, calves (exper.), indirect hemagglutination test, antibodies detected 14 days after infection, single and multiple exposures compared

Babesia bigemina
Dwivedi, S. K.; and Gautam, O. P., 1979, Indian J. Animal Sc., v. 49 (5), 395-396
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Babesia bigemina
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Babesia bigemina, acute cattle infection, plasma contains fibrin in monomer and high molecular weight forms, fibrin(ogen) degradation products not constantly detected, little or no evidence suggesting fibrinolysis or fibrin deposition; suggested that classic disseminated intravascular coagulation not present in B. bigemina infection

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Halik, J., 1977, Veterinarstvi, v. 27 (10), 469-471
Babesia, Theileria, Anaplasma, diagnostic problems, review: North Africa

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Babesia spp., analysis and sorting of red cells from infected mouse or calf blood by flow fluorimetry using 33258 Hoechst (DNA-binding fluorescent bisbenzimidazole dye)

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Johnston, L. A. Y.; Leatch, G.; and Jones, P. N., 1978, Austral. Vet. J., v. 54 (1), 14-18
Babesia argentina, B. bigemina, Droughtmaster and Hereford cattle, duration of latent infection and functional immunity following natural infection

Babesia bigemina
Kiurtov, N., 1977, Vet. Sbirka, v. 75 (5), 29-32
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Lopez V., G.; and Todorovic, R. A., 1978, Vet. Parasitol., v. 4 (1), 1-9
Babesia argentina, cattle (nat. and exper.), diagnosis by rapid latex agglutination test applicable in field, specificity relative to infections with Babesia bigemina and Anaplasma marginale, comparison of results with complement fixation and indirect fluorescent antibody tests

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Minami, T., 1977, Japan Agric. Research Quart., v. 11 (4), 234-238
Japanese Babesia sp., cattle (exper.), capillary-tube agglutination test, discrimination from B. bigemina and B. argentina was almost possible, no cross reactions with other protozoa

Babesia bigemina
Minami, T.; et al., 1979, Vet. Parasitol., v. 5 (1), 29-38
Babesia bigemina and Japanese Babesia species can be serologically differentiated by the complement fixation and capillary-tube agglutination tests

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Babesia major vermicules from Haemaphysalis punctata haemolymph and B. bigemina vermicules from Boophilus microplus haemolymph, length as differential diagnostic criterion

Babesia bigemina
Pal, M.; and Verma, J. D., 1975, U. P. Vet. J., v. 3 (4), 222-225
Babesia bigemina infection in Haryana cow following recovery from parturient paresis (milk fever), case report, complete recovery after treatment with babesan and nutrient supplements: India

Babesia bigemina
Pandey, N. N.; and Mishra, S. S., 1978, Indian Vet. J., v. 55 (2), 139-143
Babesia bigemina, cow calves (exper.), clinical symptoms as related to percentage of parasitaemia in various stages of disease as basis for timely diagnosis and treatment, highest parasitaemia at peak of fever

Babesia bigemina
Pandey, N. N.; and Mishra, S. S., 1978, Indian Vet. J., v. 55 (2), 144-148
Babesia bigemina, indigenous cow calves, berenil and acriflavin effective, sulfadimethoxine ineffective: India

Babesia bigemina
Pipano, E.; Jeruham, I.; and Frank, M., 1979, Trop. Animal Health and Prod., v. 11 (1), 13-16
Babesia bigemina, calves, pentamidine, sterilizing dose was at least 5 times as great as that needed for clinical recovery, promising agent for chemoimmunization

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Quinlan, J. F., 1978, Trop. Animal Health and Prod., v. 10 (3), 178
Babesia bovis, B. bigemina, Anaplasma marginale, presence in Friesian calves placed in Boophilus microplus infested environment and later splenectomised: Paraguay

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Rivas, A.; Rodriguez, O. N.; and Espaine, L., 1977, Rev. Cubana Cien. Vet., v. 8 (1), 1-11
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ganado vacuno (eritrocitos): Cuba

Babesia bigemina
Rodriguez, O. N.; et al., 1977, Rev. Cubana Cien. Vet., v. 8 (2), 1-7
Babesia argentina, B. bigemina, bovine, complement fixation test evaluated

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Rodriguez, O. N.; et al., 1978, Rev. Cubana Cien. Vet., v. 9 (1), 87-94
Anaplasma marginale, Babesia argentina, B. bigemina, cattle of different breeds, serodiagnosis, complement fixation and capillary agglutination microtechniques: Cuba

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Babesia bigemina, cattle (exper.), incubation period and clinical signs

Babesia bigemina
Roychoudhury, G. K.; and Gautam, O. P., 1979, Trop. Animal Health and Prod., v. 11 (2), 91-93
Babesia bigemina in Bubalus bubalis (exper.), infection with no clinical symptoms in splenectomized calves, no infection in non-splenectomized calves

Babesia bigemina, illus.
Suh, M. D.; and Hoyte, H. M. D., 1977, Korean J. Vet. Research, v. 17 (2), 51-61
Babesia bigemina and B. argentina, morphological differentiation using smears and tissue sections of splenectomized calf organs (liver, lung, kidney, heart, lymph node, brain)

Babesia bigemina
Thompson, K. C.; et al., 1978, Trop. Animal Health and Prod., v. 10 (2), 75-81
Anaplasma marginale, Babesia argentina, B. bigemina, cattle under tropical conditions, immunization with virulent organisms followed by drug therapy (ganaseg; gloxazone; emicina) vs. chemoprophylaxis (imidocarb); tick and gastrointestinal parasite control without haemoparasitic control had advantage over no control system at all

Babesia bigemina
Thompson, K. C.; et al., 1978, Trop. Animal Health and Prod., v. 10 (3), 141-144
Anaplasma marginale, Babesia bigemina, B. argentina (bovis), immunization and chemoprophylaxis of Bos taurus calves and subsequent challenge with Boophilus microplus, economic gain estimated: Colombia

Babesia bigemina
Thompson, K. C.; Todorovic, R. A.; and Hidalgo, R. J., 1978, Research Vet. Sc., v. 24 (2), 234-237
Babesia bigemina, immune response of calves to acute and chronic blood- and tick-borne infections with 4 stabilates, reduced immune response to homologous challenge but marked response to heterologous challenge indicated antigenic differences between isolates

Babesia bigemina
Todorovic, R.; and Garcia, R., 1978, Tropenmed. u. Parasitol., v. 29 (1), 88-94
Babesia spp., cattle, diagnosis, indirect fluorescent antibody test, comparison between techniques of dried blood on filter paper and serum samples, sensitivity and specificity, laboratory and field conditions: Colombia

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Todorovic, R. A.; Gonzalez, E. F.; and Garcia, O., 1979, Tropenmed. u. Parasitol., v. 30 (1), 43-52
anaplasmosis, babesiosis, cattle, immunization evaluated under field conditions, study indicates significant reduction in deaths and production losses and economic benefits for livestock producers when animals are vaccinated: Cauca River Valley, Colombia

Babesia bigemina
Todorovic, R.; Gonzalez, E.; and Lopez, G., 1978, Tropenmed. u. Parasitol., v. 29 (2), 210-214
Anaplasma marginale, Babesia spp., calves, immunization, cryo-preserved vaccines, effects of dose, inoculation route, time, and temperature

Babesia bigemina
Vizcaino, O.; et al., 1978, Am. J. Vet. Research, v. 39 (2), 229-233
Anaplasma marginale, response of calves inoculated with 3 different doses of attenuated A. marginale vaccine and subsequently challenged with a virulent strain; effects of field challenge exposure in calves inoculated with Anaplasma vaccine and premunized with both Babesia bigemina and B. argentina: Colombia

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Weber, G.; and Friedhoff, K. T., 1977, Ztschr. Parasitenk., v. 53 (1), 83-92
Babesia bigemina, ultrastructure of supposed sexual stages from gut of Boophilus microplus, comparison with developmental stage of Theileria annulata

Babesia bigemina, illus.
Weber, G.; and Friedhoff, K., 1979, Ztschr. Parasitenk., v. 58 (2), 191-194
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Wright, I. G.; and Goodger, B. V., 1977, Ztschr. Parasitenk., v. 53 (1), 63-73
Babesia bigemina, calves, plasma studies for changes in coagulation system and kallikrein levels; comparisons with B. argentina bovine infections discussed

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Ajayi, S. A., 1978, Vet. Rec., v. 103 (25), 564
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Bhat, U. K. M.; Mahoney, D. F.; and Wright, I. G., 1979, Experientia, v. 35 (6), 752-753
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Babesia bovis
Corrier, D. E.; et al., 1979, Trop. Animal Health and Prod., v. 11 (4), 215-221
Babesia bigemina- and B. bovis-immunized Bos taurus calves transported to lowland tropics, exposed to heavy vs. light Boophilus microplus infestation, resulting B. bigemina, and B. bovis parasitemias, mortality, weight loss, and anemia: Caribbean Coast, Colombia

Babesia bovis
Dalgliesh, R. J.; and Stewart, N. P., 1978, Austral. Vet. J., v. 54 (9), 453-454 [Letter]
Babesia bigemina, B. bovis, development, in incubated Boophilus microplus eggs, and B. bigemina in unfed larval ticks held at 37°C, infectivity for calves, results indicate that high environmental temperature may be only stimulus required for development of infective Babesia within the tick

Babesia bovis, illus.
Dalgliesh, R. J.; and Stewart, N. P., 1979, Internat. J. Parasitol., v. 9 (2), 115-120
Babesia bovis, morphology, development, and infectivity for cattle of parasites in unfed Boophilus microplus larvae after incubation at various temperatures

Babesia bovis
Dwivedi, S. K.; Mallick, K. P.; and Malhotra, M. N., 1979, Indian Vet. J., v. 56 (4), 333-335
Babesia bovis, Indian water buffaloes, 2 clinical case reports, treatment with berenil effective: Rithora, India

Babesia bovis, illus.
Erp, E. E.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (5), 1061-1064
Babesia bovis, growth in bovine erythrocyte cultures

Babesia bovis (= argentina)
Goodger, B. V., 1978, Ztschr. Parasitenk., v. 55 (1), 1-8
Babesia bovis-infected splenectomized and intact calves, changes in fibrinogen, plasminogen, and IgG_2 in saline eluates from sucrose-washed erythrocytes and in plasma, relationship to coagulation, fibrinolysis, and blood agglutination

Babesia bovis (argentina)
Goodger, B. V.; et al., 1978, Ztschr. Parasitenk., v. 58 (1), 3-13
Babesia bovis (argentina)-infected calves, cryofibrinogen complex in plasma, characterization, contribution to pathophysiology

Babesia bovis (argentina)
Goodger, B. V.; and Wright, I. G., 1977, Ztschr. Parasitenk., v. 54 (1), 9-27
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Babesia bovis, illus.
  Gravely, S. M.; et al., 1979, Internat. J. Parasitol., v. 9 (6), 591-598
    Babesia bovis, morphologic, biologic, and immunologic properties of corpuscular and soluble fractions of parasites purified from in vitro blood cultures

Babesia bovis
  Halik, J., 1977, Veterinarstvi, v. 27 (10), 469-471
    Babesia, Theileria, Anaplasma, diagnostic problems, review: North Africa

Babesia bovis
  Hashemi-Fesharki, R.; and Amjadi, A. R., 1977, Arch. Inst. Razi (29), 83-86
    Babesia bovis, outbreak in imported Holstein and Red-Danish cattle, imidocarb dihydrochloride more effective than quinorium sulphate: Rasht, northern Iran

Babesia bovis
  Howard, R. J.; and Rodwell, B. J., 1979, Exper. Parasitol., v. 48 (3), 421-431
    Babesia spp., analysis and sorting of red cells from infected mouse or calf blood by flow fluorimetry using 33258 Hoechst (DNA-binding fluorescent bisbenzimidazole dye)

Babesia bovis (syn. Babesia argentina)
  Mahoney, D. F.; et al., 1979, Internat. J. Parasitol., v. 9 (4), 297-306
    Babesia bovis, immune response in Bos taurus studied using passive transfer of serum from immune animals, results suggest effector mechanism is mediated by strain-specific antibody

Babesia bovis (syn. B. argentina)
  Mahoney, D. F.; and Mirre, G. B., 1979, Research Vet. Sc., v. 26 (2), 253-254
    Babesia bovis, transmission by Boophilus microplus, concluded that protozoan parasite did not persist in infective form in ticks beyond larval stage

Babesia bovis
  Mahoney, D. F.; Wright, I. G.; and Goodger, B. V., 1979, Austral. Vet. J., v. 55 (1), 10-12
    Babesia bovis, calves, single vaccination by attenuated and non-attenuated parasites was sufficient to prevent clinical babesiosis under conditions of reduced tick populations

Babesia bovis (B. argentina)
  Quinlan, J. F., 1978, Trop. Animal Health and Prod., v. 10 (3), 178
    Babesia bovis, B. bigemina, Anaplasma marginale, presence in Friesian calves placed in Boophilus microplus infested environment and later splenectomised: Paraguay

Babesia bovis
  Smith, R. D.; et al., 1978, Research Vet. Sc., v. 24 (3), 287-292
    Babesia bovis, calves, exposed by injection of infected blood or application of infected Boophilus microplus larvae, laboratory conditions, greater severity in tick-induced infections, severe reactions and high mortality occurring among older animals; tick transmission under laboratory conditions as useful research tool

Babesia bovis
  Smith, R. D.; et al., 1979, Am. J. Vet. Research, v. 40 (12), 1678-1682
    babesiosis, cattle, vaccination experiments to assess immunogenicity of and protection conferred by culture-derived Babesia bovis antigens against tick-borne infection

Babesia bovis (Babes, 1889), illus.
  Stewart, N. P., 1978, J. Protozool., v. 25 (4), 497-501
    Babesia bovis, vaccine (NT) strain, unmodified (T) strain, differential infectivity for Boophilus microplus, differences observed in parasite structure in gut contents following ingestion by tick, NT strain was incapable of penetrating epithelial cells of tick gut

B[abesia] bovis
  Todorovic, R. A.; Gonzalez, E. F.; and Garcia, O., 1979, Tropenmed. u. Parasitol., v. 30 (1), 43-52
    anaplasmosis, babesiosis, cattle, immunization evaluated under field conditions, study indicates significant reduction in deaths and production losses and economic benefits for livestock producers when animals are vaccinated: Cauca River Valley, Colombia

Babesia bovis
  Trueman, K. F.; and Blight, G. W., 1978, Austral. Vet. J., v. 54 (6), 301-305
    Babesia bovis, susceptible cattle of different ages, innate immunity, aged cattle highly susceptible compared to other groups aged up to 2 years, animals of all ages had solid resistance to subsequent heterologous challenge

Babesia bovis
  Wright, I. G., 1978, Vet. Parasitol., v. 4 (4), 393-398
    Babesia bovis-infected Bos taurus, biogenic amine levels in plasma and whole blood, probable role in cardiovascular disturbances

Babesia bovis
  Wright, I. G., 1979, Gen. Pharmacol., v. 10 (4), 319-325
    Babesia, Plasmodium, Trypanosoma, kallikrein-kinin system, mechanisms of activation (parasite enzymes, immune complexes), role in hypotensive shock syndrome of infected animals, review

Babesia bovis
  Wright, I. G.; and Goodger, B. V., 1979, Ztschr. Parasitenk., v. 59 (2), 115-119
    Babesia bovis, splenectomised calves (exper.), urine analysis, kidney histopathology

Babesia caballi
  Amerault, T. E.; Frerichs, W. M.; and Stiller, D., 1979, Am. J. Vet. Research, v. 40 (4), 529-531
    Babesia caballi, horses, card test using newly developed antigen evaluated and compared with complement-fixation test

Babesia caballi
  Bertoldini, G.; Codazza, D.; and Redaelli, G., 1978, Clin. Vet., Milano, v. 101 (8), 490-492
    Babesia equi, B. caballi, horses, diagnosis, complement fixation test

Babesia caballi
Chevrier, L., 1978, Prat. Vet. Equine, v. 10 (4), 171-173
inapparent equine piroplasmosis, clinical and serological methods of diagnosis reviewed briefly, comparison of incidence in 1976 and 1978 at Alfort

Babesia caballi
Sharma, R. D.; and Gautam, O. P., 1977, Haryana Vet., v. 16 (1), 19-26
Babesia caballi, B. equi, horses, case reports, haematological changes, berenil, some adverse reactions: India

Babesia canis
Bauer, F.; Raether, W.; and Seeger, K., 1978, Cahiers Bleus Vet. (27), 265-271
protozoal disease in exper. hosts, enhanced effect of berenil + reverin vs. berenil alone

Babesia canis, illus.
Liebisch, A.; and Gillani, S., 1979, Deutsche Tierarztl. Wchnschr., v. 86 (4), 149-152
Babesia canis, dogs, experimental transmission by native German Rhipicephalus sanguineus, enzootic foci could develop in Germany

Babesia canis
Okoh, A. E. J., 1978, Bull. Animal Health and Prod., Africa, v. 26 (2), 118-119
Babesia canis, dog (hippocampus and cerebrum filling lumen of small capillaries and small arterioles), case report: Nigeria

Babesia canis, illus.
Osorno E., B. M.; and Ristic, M., 1974, Tecn. Pecuaria Mexico (26), 36-40
dogs (nat. and exper.) (blood): central regions, Veracruz, Mexico

Babesia canis
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Babesia canis
Sasum, H., 1979, Kleintier-Praxis, v. 24 (4), 176
Babesia canis, dog, diagnosis

Babesia canis
Weiland, G.; and zu Dohna, H. G., 1978, Berl. u. Munchen. Tierarztl. Wchnschr., v. 91 (2), 33-35
Babesia spp., mice, dogs, rabbits, enzyme-linked immunosorbent assay suitable for detection of antibodies in field surveys, cross-reactions occurred although distinct titer differences could be observed

Babesia capreoli
Christensson, D.; and Jaerplid, B., 1979, Svensk Vet.-Tidn., v. 31 (8), 231-232
Capreolus capreolus: Sweden

B[abesia] colchica
Gadzhiev, K. Sh., 1978, Veterinariia, Moskva (11), 67
B[abesia] colchica, cattle, course of infection simultaneous with P[asteurella] multocida infection: Azizbekov Shamkhorsk region, Azerbaidzhansk SSR

Babesia divergens
Adam, K. M. G.; et al., 1978, Brit. Vet. J., v. 134 (5), 428-433
babesiasis, cattle, 2 outbreaks compared (clean cattle introduced to tick-infested endemic area; tick-infested cattle introduced to clean area), percentage seropositive for babesial antibody, degree of manifest disease, treatment with amicarbalide, results indicate that vaccination against Babesia divergens is feasible and desirable: Scotland

Babesia divergens
Adam, K. M. G.; and Blewett, D. A., 1978, Ann. Trop. Med. and Parasitol., v. 72 (5), 417-428
Babesia divergens, cattle, serological survey using indirect fluorescent antibody test, occurrence of antibody in the population and distribution of infected herds: Scotland

Babesia divergens
Bidwell, D. E.; et al., 1978, Vet. Rec., v. 103 (20), 446-449
Babesia divergens, B. major, cattle (exper.), serological diagnosis, comparison between microplate enzyme linked immunosorbent assay, indirect fluorescent antibody, and complement fixation tests, ELISA may be preferable

Babesia divergens
Blewett, D. A.; and Adam, K. M. G., 1978, Ann. Trop. Med. and Parasitol., v. 72 (5), 405-415
Babesia divergens, cattle, serological survey using indirect fluorescent antibody test, critical assessment of sampling and testing methods: Scotland

Babesia divergens
Blewett, D. A.; and Adam, K. M. G., 1978, Ann. Trop. Med. and Parasitol., v. 72 (6), 513-522
Babesia divergens, cattle, data for host age and antibody incidence and titre analyzed using 3 epidemiological models to estimate rates of gain and loss of antibody: Scotland

Babesia divergens
Chermette, R., 1979, Point Vet. (45), v. 9, 31-40
Babesia divergens, bovine, as cause of icterus, review: life cycle, epidemiology, clinical aspects, pathogenesis, immunity, diagnosis, treatment, prophylaxis

Babesia divergens
Entrican, J. H.; et al., 1979, Brit. Med. J. (6188), v. 2, 474
Babesia divergens in man who had been splenectomized and treated for Hodgkin's disease by irradiation, case report: Inverness, Scotland

Babesia divergens
Irvin, A. D.; and Young, E. R., 1979, Internat. J. Parasitol., v. 9 (2), 109-114
Babesia spp. of cattle and mice, uptake and metabolism of tritiated nucleic acid precursors

Babesia divergens
Irvin, A. D.; Young, E. R.; and Osborn, G. D., 1978, Research Vet. Sc., v. 25 (2), 245-246
Babesia divergens, B. major, attempt to infect mice (nu/nu, nu/+, nu/nu splenectomized, and Lasat), neither parasite became established, B. divergens persisted up to 10 days, B. major lasted only 1 day, B. divergens persisted longer in splenectomized mice but absence of thymus made no apparent difference

Babesia divergens
Irvin, A. D.; Young, E. R.; and Purnell, R. E., 1978, Internat. J. Parasitol., v. 8 (1), 19-24
Babesia spp. of cattle and mice, in vitro uptake of tritiated nucleic acid precursors by intra-erythrocytic stages of parasites

Babesia divergens
Lewis, D.; Purnell, R. E.; and Brocklesby, D. W., 1979, Research Vet. Sc., v. 26 (2), 220-222
Babesia divergens, splenectomised calves, immunization using irradiated piroplasms

Babesia divergens, illus.
Lewis, D.; and Williams, H., 1979, Nature, London (5700), v. 278, 170-171
Babesia divergens, successful experimental infection of Meriones unguiculatus by intraperitoneal inoculation of infected bovine blood, parasite is then transmissible to further gerbils and can be used to infect splenectomised calves once again after 2 gerbil passages

Babesia divergens
Purnell, R. E.; et al., 1978, J. Comp. Path., v. 88 (3), 419-423
Babesia divergens, reactions of splenectomized calves to inoculation of infected blood taken from a calf during its reaction and carrier phases, parasite virulence, possible role in immunization

Babesia divergens
Trees, A. J., 1978, Research Vet. Sc., v. 24 (1), 126-128
Babesia divergens, cattle (exper.), indirect fluorescent antibody levels not markedly different in splenectomized calves vs. nonsplenectomized adult cattle

Babesia equi
Barnett, D.; Proctor, S. J.; and Chieves, L., 1977, Proc. 14. Ann. Electron Micr. Colloq., 166-171
Babesia equi, ultrastructure, alterations of parasitized equine erythrocytes

Babesia equi
Bertoldini, G.; Codazza, D.; and Redaelli, G., 1978, Clin. Vet., Milano, v. 101 (8), 490-492
Babesia equi, B. caballi, horses, diagnosis, complement fixation test

Babesia equi
Chevrier, L., 1978, Prat. Vet. Equine, v. 10 (4), 171-173
inapparent equine piroplasmosis, clinical and serological methods of diagnosis reviewed briefly, comparison of incidence in 1976 and 1978 at Alfort

Babesia equi, illus.
Correa, W. M.; Correa, C. N. M.; and Fanton, E. B., 1978, Arq. Escola Vet. Univ. Fed. Minas Gerais, v. 30 (3), 303-305
fetal Babesia equi as cause of abortion: Brazil, imported from Florida, USA

Babesia equi
Halik, J., 1977, Veterinarstvi, v. 27 (10), 469-471
Babesia, Theileria, Anaplasma, diagnostic problems, review: North Africa

Babesia equi
Kalra, I. S.; Verma, B. B.; and Sinha, B. P., 1976, Indian Vet. J., v. 53 (12), 967-968
Babesia equi, equine, 2 case reports, clinical symptoms of colic and chronic enteritis, successfully treated with babesan

Babesia equi
Malhotra, D. V.; Banerjee, D. P.; and Gautam, O. P., 1978, Equine Vet. J., v. 10 (1), 24-26
Babesia equi, horses, prevalence of latent cases as measured by capillary agglutination test: north west India

Babesia equi
Malhotra, D. V.; Gautam, O. P.; and Banerjee, D. P., 1979, Indian J. Animal Sc., v. 49 (1), 75-77
Babesia equi, donkeys (exper), diminazene aceturate as effective as diminazene aceturate + rolitetracycline, quinuronium sulphate ineffective

Babesia equi
Malhotra, D. V.; Gautam, O. P.; and Banerjee, D. P., 1979, Trop. Animal Health and Prod., v. 11 (2), 95-97
Babesia equi, failure to transmit in laboratory animals

Babesia equi
Sharma, R. D.; and Gautam, O. P., 1977, Haryana Vet., v. 16 (1), 19-26
Babesia caballi, B. equi, horses, case reports, haematological changes, berenil, some adverse reactions: India

Babesia equi
Singh, B.; et al., 1978, Indian Vet. J., v. 55 (9), 677-679
Babesia equi, donkeys, horses, diagnosis, capillary tube-agglutination test, no cross-reactions with related haemoprotozoan antisera

Babesia ernestoi sp. n., illus.
Freire, N. M. da S., 1979, Zentralbl. Vet.-Med., Reihe B, v. 26 (8), 614-622
Didelphis marsupialis (nat. and exper.) (blood, adrenals, aorta, spleen, cerebrum, cerebellum, liver, bone marrow, lungs, kidney): Brazil
D. albiventris (nat. and exper.) (blood): Brazil

Babesia felis (Davis, 1929), illus.
El-Shabrawy, M. N., 1972, Vet. Med. J., Giza, v. 20 (20), 205-209
Babesia felis in Felis catus (blood), incidence, description of developmental and reproductive forms: Egypt

Babesia galagolata
Weiland, G.; and zu Dohna, H. G., 1978, Berl. u. Munchen. Tierarztl. Wchnschr., v. 91 (2), 33-35
Babesia spp., mice, dogs, rabbits, enzyme-linked immunosorbent assay suitable for detection of antibodies in field surveys, cross-reactions occurred although distinct titer differences could be observed

Babesia gibsoni, illus.
Anderson, J.; et al., 1979, Science (4400), v. 204, 1431-1432
dogs (nat. and exper.): Ridgefield, Connecticut (had also been in New York and Massachusetts)

Babesia gibsoni
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Babesia gibsoni, illus.
Sonoda, M.; et al., 1978, Japan. J. Vet. Sc., v. 40 (3), 335-341
Haemobartonella canis, presence in blood of intact and splenectomized dogs experimentally infected with Babesia gibsoni, scanning and transmission electron microscopy: Japan

Babesia hylomysci sp. nov., illus.
Bafort, J. M.; Timperman, G.; and Molyneux, D. H., 1970, Ann. Soc. Belges Med. Trop. Parasitol., v. 50 (3), 301-318
Hylomyscus stella: Ndjili (Kinshasa), Dem. Rep. of the Congo
albino mouse (exper.)
" rat (")
Mesocricetus auratus (")
Sigmodon hispidus hispidus (")
Mastomys natalensis (")
Clethrionomys glareolus (")
Meriones unguiculatus (")
M. shawi (")
Lagurus lagurus (")
Cricetulus griseus (")

Apodemus sylvaticus (")
Peromyscus maniculatus (")
Acomys camirinus (")
Eutamias asiaticus (")
Macaca mulatta (")

Babesia hylomysci
Hussein, H. S., 1979, Exper. Parasitol., v. 47 (1), 1-12
Babesia microti, B. hylomysci, mice, role of spleen during infection, erythrophagocytosis, determination of phagocytic activity of reticuloendothelial system

Babesia hylomysci
Momen, H.; Chance, M. L.; and Peters, W., 1979, Ann. Trop. Med. and Parasitol., v. 73 (3), 203-212
Babesia spp., human and animal strains and Anthemosoma garnhami, taxonomic differentiation using biochemical methods (variation in mobility of parasite enzymes on starch gels after electrophoresis, measurement of buoyant densities of DNA)

Babesia major
Bidwell, D. E.; et al., 1978, Vet. Rec., v. 103 (20), 446-449
Babesia divergens, B. major, cattle (exper.), serological diagnosis, comparison between microplate enzyme linked immunosorbent assay, indirect fluorescent antibody, and complement fixation tests, ELISA may be preferable

Babesia major
Irvin, A. D.; and Young, E. R., 1978, Research Vet. Sc., v. 25 (2), 211-214
Babesia spp., drug inhibition of hypoxanthine uptake in vitro could be used as primary screen for babesicidal drugs but drugs showing in vitro activity are not necessarily active in vivo

Babesia major, illus.
Irvin, A. D.; and Young, E. R., 1979, Internat. J. Parasitol., v. 9 (2), 109-114
Babesia spp. of cattle and mice, uptake and metabolism of tritiated nucleic acid precursors

Babesia major
Irvin, A. D.; Young, E. R.; and Osborn, G. D., 1978, Research Vet. Sc., v. 25 (2), 245-246
Babesia divergens, B. major, attempt to infect mice (nu/nu, nu/+, nu/nu splenectomized, and Lasat), neither parasite became established, B. divergens persisted up to 10 days, B. major lasted only 1 day, B. divergens persisted longer in splenectomized mice but absence of thymus made no apparent difference

Babesia major
Irvin, A. D.; Young, E. R.; and Purnell, R. E., 1978, Internat. J. Parasitol., v. 8 (1), 19-24
Babesia spp. of cattle and mice, in vitro uptake of tritiated nucleic acid precursors by intra-erythrocytic stages of parasites

Babesia major, illus.
Morzaria, S. P.; Bland, P.; and Brocklesby, D. W., 1978, Ztschr. Parasitenk., v. 55 (2), 119-125
Babesia major vermicules from Haemaphysalis punctata haemolymph, ultrastructure, negative staining

Babesia major
Morzaria, S. P.; and Brocklesby, D. W., 1977, Ztschr. Parasitenk., v. 52 (3), 241-243
Babesia major vermicules from Haemaphysalis punctata haemolymph and B. bigemina vermicules from Boophilus microplus haemolymph, length as differential diagnostic criterion

Babesia major
Payne, R. C., 1978, Research Vet. Sc., v. 24 (3), 375
Babesia major, cryopreservation of infected bovine blood, preserved in liquid nitrogen for use in indirect fluorescent antibody test

Babesia major
Purnell, R. E.; et al., 1979, Brit. Vet. J., v. 135 (1), 44-49
Babesia major, intact calves, reactions to inoculation with varying numbers of infected erythrocytes, immunity to homologous challenge

Babesia major
Purnell, R. E.; Brocklesby, D. W.; and Stark, A. J., 1978, Research Vet. Sc., v. 25 (3), 388-390
Babesia major, splenectomised calves, protection by inoculation of irradiated piroplasms

Babesia major
  Purnell, R. E.; Lewis, D.; and Brocklesby, D. W., 1979, Internat. J. Parasitol., v. 9 (1), 69-71
    Babesia major, protection of intact calves against homologous challenge by injection of irradiated piroplasms

Babesia microti
  Anderson, J. F.; Magnarelli, L. A.; and Kurz, J., 1979, J. Parasitol., v. 65 (4), 599-604
    Babesia microti antibody found in Peromyscus leucopus from Connecticut, failure to detect organisms following splenectomy or subinoculation of whole blood into susceptible hamsters or in direct blood smears; successful exper. infection of Connecticut P. leucopus with parasite strain from New York

Babesia microti
  Bautista, C. R.; and Kreier, J. P., 1979, Infect. and Immun., v. 25 (1), 470-472
    Babesia microti, short-term in vitro culture in hamster erythrocytes, inhibitory effect of immune serum on growth in vitro

Babesia microti
  Benach, J. L.; et al., 1979, Am. J. Trop. Med. and Hyg., v. 28 (4), 643-648
    Babesia microti, 1 human-derived and 2 rodent-derived isolates from Long Island, immunological relationships
    albino hamsters (exper.)
    Sprague-Dawley rats (exper.)
    Swiss mice (exper.)

Babesia microti
  Benach, J. L.; White, D. J.; and McGovern, J. P., 1978, Am. J. Trop. Med. and Hyg., v. 27 (6), 1073-1078
    Babesia microti, rodent- and human-derived Long Island isolates in Mesocricetus auratus, course of infection, effect of splenectomy and inoculum size, pathogenesis

Babesia microti, illus.
  Chisholm, E. S.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (1, pt. 1), 14-19
    Babesia microti, human, diagnosis, indirect immunofluorescent antibody test is sensitive, specific, and reproducible; cross-reactivity did occur with other Babesia spp. and with Plasmodium spp.

Babesia microti
  Clark, I. A., 1978, Lancet, London (8080), v. 2, 75-77
    acute malaria and babesiosis, hypothesis that endotoxin (lipopolysaccharide) causes both the disease and the parasite death, experiments in mice

Babesia microti
  Clark, I. A., 1979, Infect. and Immun., v. 24 (2), 319-325
    Babesia spp., Plasmodium vinckei petteri, mice, protection conferred by pretreatment with extract of Coxiella burnetii, possible involvement of interferon or tumor necrosis factor

Babesia microti
  Clark, I. A., 1979, Parasite Immunol., v. 1 (3), 179-196
    Babesia microti, mice, protection against subsequent infection by injection of cord factor, COAM, zymosan, glucan, Salmonella, or Listeria

Babesia microti
  Cox, F. E. G., 1977, Protozoology, v. 3, 129-134
    Trypanosoma musculi, T. b. brucei, Babesia microti, mice, interactions between parasites
    Apodemus sylvaticus
    Microtus agrestis
    Clethrionomys glareolus

Babesia microti
  Cox, F. E. G., 1978, Parasitology, v. 76 (1), 55-60
    Plasmodium vinckei, Babesia microti, heterologous immunity, simultaneous elimination of the two species from blood of doubly infected mice

Babesia microtia
  Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
    Sorex araneus (Blut, Herz)
    Apodemus flavicollis (Blut, Herz, Lunge)
    A. sylvaticus (Blut, Herz, Lunge)
    Clethrionomys glareolus (Herz, Leber)
    Microtus arvalis (Lunge)
    all from Neusiedlerseegebiet, nordlichen Burgenland

Babesia microti, illus.
  Herod, E.; Clark, I. A.; and Allison, A. C., 1978, Clin. and Exper. Immunol., v. 31 (3), 518-523
    mice infected with attenuated vaccine strain of Brucella abortus are strongly protected against infection with Babesia microti, this non-specific immunity seems to be best explained by stimulation of macrophages so as to release mediator which limits intracellular replication

Babesia microti
  Hsu, N. H.; and Cross, J. H., 1977, Taiwan i Hsueh Hui Tsa Chih (J. Formosan Med. Ass.), v. 76 (12), 950-954
    Babesia microti, serologic survey of human rural populations for presence of antibodies using indirect fluorescent antibody test; known prevalence in rodents, findings of survey suggest low human prevalence: Taiwan

Babesia microti
  Hussein, H. S., 1979, Exper. Parasitol., v. 47 (1), 1-12
    Babesia microti, B. hylomysci, mice, role of spleen during infection, erythrophagocytosis, determination of phagocytic activity of reticuloendothelial system

Babesia microti
  Irvin, A. D.; and Young, E. R., 1979, Internat. J. Parasitol., v. 9 (2), 109-114
    Babesia spp. of cattle and mice, uptake and metabolism of tritiated nucleic acid precursors

Babesia microti, illus.
  Irvin, A. D.; Young, E. R.; and Purnell, R. E., 1978, Internat. J. Parasitol., v. 8 (1), 19-24
    Babesia spp. of cattle and mice, in vitro uptake of tritiated nucleic acid precursors by intra-erythrocytic stages of parasites

Babesia microti (Franca, 1912), illus.
Krampitz, H. E.; and Baeumler, W., 1978, Ztschr. Parasitenk., v. 58 (1), 15-33
Babesia microti in Microtus agrestis, prevalence by host sex and weight and by months, splenomegaly, only Ixodes ricinus found on mammals and vegetation of surveyed area; experimental host range
Microtus agrestis (nat. and exper.): westlich von Munchen
M. arvalis (exper.)
Clethrionomys glareolus (exper.)
Mesocricetus auratus (exper.)
Cricetulus griseus (exper.)
Meriones unguiculatus (exper.)

Babesia microti
Momen, H., 1979, Ann. Trop. Med. and Parasitol., v. 73 (2), 117-121
Plasmodium, Babesia and Anthemosoma spp., comparative study of glucose catabolism by infected mouse erythrocytes, glucose utilization and lactate production of parasites

Babesia microti
Momen, H.; Chance, M. L.; and Peters, W., 1979, Ann. Trop. Med. and Parasitol., v. 73 (3), 203-212
Babesia spp., human and animal strains and Anthemosoma garnhami, taxonomic differentiation using biochemical methods (variation in mobility of parasite enzymes on starch gels after electrophoresis, measurement of buoyant densities of DNA)

Babesia microti
Oliveira, M. R.; and Kreier, J. P., 1979, J. Parasitol., v. 65 (5), 816-817
Babesia microti, transmission to hamsters and gerbils by Ixodes pacificus and I. scapularis but not by Dermacentor andersoni or D. variabilis

Babesia microti
Piesman, J.; et al., 1979, J. Med. Entom., v. 15 (5-6), 537-540
Babesia microti not found in Odocoileus virginianus naturally or experimentally inoculated, nor in vector Ixodes dammini: Nantucket and Naushon Islands, Massachusetts

Babesia microti
Piesman, J.; and Spielman, A., 1979, Ann. Entom. Soc. Am., v. 72 (6), 829-832
immature Ixodes dammini, host association and seasonal abundance, role as vectors of Babesia microti: southeastern Massachusetts

Babesia microti
Rosenberg, Y. J.; and Evans, C. B., 1979, Nature, London (5729), v. 281, 302-304
Babesia microti, mice suppressed for IgM production, resistance to infection as reflected by virtual absence of parasites in peripheral circulation

Babesia microti
Ruebush, M. J.; and Hanson, W. L., 1979, J. Parasitol., v. 65 (3), 430-433
Babesia microti, susceptibility of 5 strains of mice to parasites of human origin

Babesia microti
Ruebush, T. K. II; et al., 1979, Am. J. Trop. Med. and Hyg., v. 28 (2), 184-189
Babesia microti, 65-year-old man, treated with diminazene aceturate after failure to respond to chloroquine therapy, development of acute idiopathic polyneuritis: Nantucket Island

Babesia microti
Ruebush, T. K. II; et al., 1979, J. Parasitol., v. 65 (1), 144-146
Babesia microti, course of infection in nonsplenectomized Macaca mulatta (exper.)

Babesia microti
Spielman, A.; Piesman, J.; and Etkind, P., 1977, J. N. York Entom. Soc., v. 85 (4), 214-216
Babesia microti, human, survey of reservoir hosts and tick vector Ixodes sp. nr. scapularis: Nantucket and nearby islands

Babesia motasi
Prabhakar, K. S.; and Hiregoudar, L. S., 1977, Indian J. Animal Sc., v. 47 (12), 848-850
sheep (blood): Karnataka

Babesia muratovi
Momen, H.; Chance, M. L.; and Peters, W., 1979, Ann. Trop. Med. and Parasitol., v. 73 (3), 203-212
Babesia spp., human and animal strains and Anthemosoma garnhami, taxonomic differentiation using biochemical methods (variation in mobility of parasite enzymes on starch gels after electrophoresis, measurement of buoyant densities of DNA)

Babesia ovis
Charyev, O. Ch., 1977, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (6), 70-71
piroplasmids, sheep, pure and mixed infections: Turkmenia

Babesia ovis
Halik, J., 1977, Veterinarstvi, v. 27 (10), 469-471
Babesia, Theileria, Anaplasma, diagnostic problems, review: North Africa

Babesia ovis
Khalacheva, M.; and Kararizova, L., 1977, Vet. Med. Nauki, v. 14 (3), 32-38
Babesia ovis, effect of ionizing radiation on virulence and immunogenic properties

Babesia ovis
Khalacheva, M.; and Pavlov, N., 1977, Vet. Med. Nauki, v. 14 (2), 50-56
Babesia ovis, splenectomized sheep (exper.), histopathology

Babesia ovis
Kiurtov, N., 1977, Vet. Med. Nauki, v. 14 (4), 25-30
Babesia ovis, lambs, vaccination with live vaccine

Babesia procyonis, illus.
Schaffer, G. D.; et al., 1978, J. Am. Vet. Med. Ass., v. 173 (9), 1148-1151
hematotropic parasites of Procyon lotor, carrier potential as related to translocation and release for hunting purposes, practice is considered biologically hazardous: southeastern United States
Procyon lotor: Hawkins County, Tennessee; Ohio County, West Virginia; Glades County, Florida; Hillsborough County, Florida; Liberty County, Georgia; Brown County, Texas; James City County, Virginia

Babesia rodhaini
Abdalla, H. S.; Hussein, H. S.; and Kreier, J. P., 1978, J. Parasitol., v. 64 (3), 554-556
Babesia rodhaini, centrifugation times and speeds to obtain maximum yield of free parasites following passage of infected blood through sonic oscillation field, infectivity of free parasites in comparison to that of infected red blood cells

Babesia rodhaini
Abdalla, H. S.; Hussein, H. S.; and Kreier, J. P., 1978, Tropenmed. u. Parasitol., v. 29 (3), 295-306
Babesia rodhaini, intact and splenectomized mice, immune serum (IS) delayed onset of parasitemia but neither prevented development of infection nor protected mice from death even with further supplementation of IS, effect of IS greater when parasitized cells incubated in IS were administered with additional IS

Babesia rodhaini
Bauer, F.; Raether, W.; and Seeger, K., 1978, Cahiers Bleus Vet. (27), 265-271
protozoal disease in exper. hosts, enhanced effect of berenil + reverin vs. berenil alone

Babesia rodhaini
Clark, I. A., 1978, Lancet, London (8080), v. 2, 75-77
acute malaria and babesiosis, hypothesis that endotoxin (lipopolysaccharide) causes both the disease and the parasite death, experiments in mice

Babesia rodhaini
Clark, I. A., 1979, Infect. and Immun., v. 24 (2), 319-325
Babesia spp., Plasmodium vinckei petteri, mice, protection conferred by pretreatment with extract of Coxiella burnetii, possible involvement of interferon or tumor necrosis factor

Babesia rodhaini
Gillet, J.; and Herman, F., 1973, Ann. Soc. Belge Med. Trop., v. 53 (1), 1-6
Babesia rodhaini, inoculation into mouse embryos on 15th day of gestation, delayed development

Babesia rodhaini
Howard, R. J.; and Rodwell, B. J., 1979, Exper. Parasitol., v. 48 (3), 421-431
Babesia spp., analysis and sorting of red cells from infected mouse or calf blood by flow fluorimetry using 33258 Hoechst (DNA-binding fluorescent bisbenzimidazole dye)

Babesia rodhaini
Irvin, A. D.; and Young, E. R., 1978, Research Vet. Sc., v. 25 (2), 211-214
Babesia spp., drug inhibition of hypoxanthine uptake in vitro could be used as primary screen for babesicidal drugs but drugs showing in vitro activity are not necessarily active in vivo

Babesia rodhaini
Irvin, A. D.; and Young, E. R., 1979, Internat. J. Parasitol., v. 9 (2), 109-114
Babesia spp. of cattle and mice, uptake and metabolism of tritiated nucleic acid precursors

Babesia rodhaini
Irvin, A. D.; Young, E. R.; and Purnell, R. E., 1978, Internat. J. Parasitol., v. 8 (1), 19-24
Babesia spp. of cattle and mice, in vitro uptake of tritiated nucleic acid precursors by intra-erythrocytic stages of parasites

Babesia rodhaini
Mitchell, G. F.; Handman, E.; and Howard, R. J., 1978, Austral. J. Exper. Biol. and Med. Sc., v. 56 (5), 553-559
Plasmodium berghei, Babesia rodhaini, mice, attempts to raise host-protective sera using variety of immunization manipulations (BCG injection, P. yoelii infection, others)

Babesia rodhaini
Momen, H., 1979, Ann. Trop. Med. and Parassitol., v. 73 (2), 109-115
Plasmodium, Babesia, and Anthemosoma spp. in mouse erythrocytes, identification of enzymes of parasite origin using starch-gel electrophoresis

Babesia rodhaini
Momen, H., 1979, Ann. Trop. Med. and Parasitol., v. 73 (2), 117-121
Plasmodium, Babesia and Anthemosoma spp., comparative study of glucose catabolism by infected mouse erythrocytes, glucose utilization and lactate production of parasites

Babesia rodhaini
Momen, H.; Chance, M. L.; and Peters, W., 1979, Ann. Trop. Med. and Parasitol., v. 73 (3), 203-212
Babesia spp., human and animal strains and Anthemosoma garnhami, taxonomic differentiation using biochemical methods (variation in mobility of parasite enzymes on starch gels after electrophoresis, measurement of buoyant densities of DNA)

Babesia rodhaini
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Babesia rodhaini
Thoongsuwan, S.; Cox, H. W.; and Patrick, R. A., [1979], J. Parasitol., v. 64 (6), 1978, 1050-1056
Trypanosoma lewisi, Babesia rodhaini, Plasmodium chabaudi, rats, acquired nonspecific resistance associated with recovery from various infectious anemias, association with immunoconglutinin activity, infections also stimulated production of cold-active hemagglutinin

Babesia rodhaini
Thoongsuwan, S.; Cox, H. W.; and Patrick, R. A., [1979], J. Parasitol., v. 64 (6), 1978, 1060-1066
Plasmodium chabaudi, Babesia rodhaini, rats, serologic specificity of immunoconglutinin associated with infectious anemia and its role in nonspecific acquired resistance

Babesia rodhaini
Thoongsuwan, S.; Cox, H. W.; and Rickman, W. J., 1979, J. Parasitol., v. 65 (3), 426-429
Plasmodium chabaudi, Babesia rodhaini, Trypanosoma lewisi, rats, presence of antibody to fibrinogen/fibrin products, association with cold-activated hemagglutinin and immunoconglutinin, temporal relationships with anemia and parasitemia

Babesia rodhaini
  Weiland, G.; and zu Dohna, H. G., 1978, Berl. u. Munchen. Tierarztl. Wchnschr., v. 91 (2), 33-35
    Babesia spp., mice, dogs, rabbits, enzyme-linked immunosorbent assay suitable for detection of antibodies in field surveys, cross-reactions occurred although distinct titer differences could be observed

Babesia vesperuginis
  Abranches, P.; and Magalhaes, C., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (6), 361-365
    Miniopterus schreibersii: Olhos de Agua, near Santarem, Portugal
    Pipistrellus pipistrellus: Tapada de Mafra, Portugal

Babesia vogeli
  Halik, J., 1977, Veterinarstvi, v. 27 (10), 469-471
    Babesia, Theileria, Anaplasma, diagnostic problems, review: North Africa

Babesiasis, probably Babesia divergens
  Bindseil, E.; and Ishøy, P., 1978, Dansk Vet.-Tidsskr., v. 61 (20), 980-982
    babesiasis, probably Babesia divergens, cattle, case histories, acaprin treatment, Ixodes ricinus identified on one cow: Laesø

Babesiasis
  Dubovyi, S. Z.; et al., 1977, Veterinariia, Moskva (3), 71-72
    babesiasis, piroplasmosis, cattle, dimidine as effective chemoprophylaxis under pasture conditions with presence of vector, Boophilus calcaratus, comparison with azidine

Babesiasis
  Euzeby, J. A., 1979, Rev. Med. Vet., Toulouse, v. 130 (5), 685-694, 697-698, 701-702, 705-712
    babesiosis, cattle, up-to-date review of present knowledge: epidemiology, host susceptibility, distribution, clinical manifestations, diagnosis, control, treatment

Babesiasis
  Hourrigan, J. L., 1977, J. N. York Entom. Soc., v. 85 (4), 217-220
    bovine babesiosis, epizootiology, history and current status of Boophilus spp. eradication in Texas

Babesiasis
  Norval, R. A. I., 1978, Rhodesian Vet. J., v. 9 (1-4), 9-16
    ticks, cattle, significant increase in tick infestations and outbreaks of tick-borne diseases following collapse of dipping (1973-1978): African areas in Rhodesia

Babesiasis
  O'Neill, A. R., 1979, Irish Vet. J., v. 33 (1), 1, 4-7
    babesiosis, cattle, blood transfusion therapy for severe anemia, criteria for use, technique

Babesiasis
  Smith, R. D.; et al., 1979, Am. J. Vet. Research, v. 40 (12), 1678-1682
    babesiosis, cattle, vaccination experiments to assess immunogenicity of and protection conferred by culture-derived Babesia bovis antigens against tick-borne infection

Babesiosis. See Babesiasis.

Babesiosoma
  Krylov, M. V.; and Krylova, N. P., 1972, Parazitologiia, Leningrad, v. 6 (6), 493-505
    Piroplasmida, analysis of host specificity

Babesiosoma Jakowska and Nigrelli
  Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102
    as syn. of Haemohormidium Henry

Babesiosoma tetragonis Becker & Katz
  Ollenschlaeger, B., 1975, Tieraerztl. Prax., v. 3 (1), 99-107
    blood and other parasites of commercial fish, pathology, transmission, therapy, clinical review

Bacillidium bacilliforme (Leger et Hesse, 1922)
  Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135
    as syn. of Toxoglugea bacilliformis (Leger et Hesse, 1922) comb. nov.

Bacillidium tetrasporum (Leger et Hesse, 1922)
  Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135
    as syn. of Toxoglugea tetraspora (Leger et Hesse, 1922) comb. nov.

Baculea daphniae Loubes et Akbarieh, sous presse [nom. nud.?]
  Loubes, C.; and Akbarieh, M., 1977, Ztschr. Parasitenk., v. 54 (2), 125-137

Baculea daphniae Loubes & Akbarieh, illus.
  Loubes, C., 1979, J. Protozool., v. 26 (2), 200-208
    Microsporida, synaptonemal complexes demonstrated in 6 genera but not in Nosema, implications for life cycles
    Daphnia pulex: environs de Montpellier

Balantidiasis
  Nikulin, T. G.; Shpak, G. E.; and Savchenko, V. F., 1977, Veterinariia, Moskva (10), 80-81
    balantidiasis, oesophagostomiasis, mixed infection in swine (exper.), changes in carboanhydrase and alkaline phosphatase activities

[Balantidium] balantidiiami
  Lipnitskii, S. S.; and Iakubovskii, M. V., 1975, Vet. Nauka--Proizvod., Trudy, Minsk, v. 13, 143-147
    helminths, protozoa, cattle, influence of micro-elements in host diet on infectivity

Balantidium
  Yu, M.; and Kin, T. H., 1970, Singapore Med. J., v. 11 (1), 55-56
    Endamoeba histolytica, Balantidium, Trichomonas, culture, new simple method

Balantidium spp.
  Holt, P. E.; Cooper, J. E.; and Needham, J. R., 1979, J. Small Animal Practice, v. 20 (5), 269-286
    Testudo: Britain

Balantidium-like sp.
  Keymer, I. F., 1978, Vet. Rec., v. 103 (25), 548-552
    parasitic and other diseases of tortoises, necropsy survey
    Testudo tornieri (rectum)

Balantidium spp.
Meyer, R. C., 1978, Advances Vet. Sc. and Comp. Med., v. 22, 133-158
swine dysentery, review, possible role of Balantidium spp.

Balantidium coli
Bischofova, N.; and Zajicek, D., 1976, Veterinarstvi, v. 26 (11), 506-507
Balantidium coli, pigs, large-scale fattening farms, Lincospectin treatment

Balantidium coli
Botero R., D., 1978, Ann. Rev. Pharmacol. and Toxicol., v. 18, 1-15
chemotherapy of common intestinal protozoan and helminth infections in humans, review of antiparasitic drugs in current use

Balantidium coli
Buck, A. A.; Anderson, R. I.; and MacRae, A. A., 1978, Tropenmed. u. Parasitol., v. 29 (3), 253-268
polyparasitism, humans, epidemiology, assessment of combined effects of multiple infections on an individual's state of health, comparative analysis of field data obtained from several tropical villages

Balantidium coli
de Carneri, I., 1972, Rev. Inst. Med. Trop. S. Paulo, v. 14 (5), 321-325
Balantidium coli in culture, nitrimidazine and metronidazole tested

Balantidium coli
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Sus scrofa (Jejunum, Ileum, Colon): Neusiedlerseegebiet, nordlichen Burgenland

Balantidium coli, illus.
Goldsmid, J. M.; and Rogers, S., 1978, J. South African Vet. Ass., v. 49 (2), 109-111
Papio ursinus (feces): Rhodesia, imported from Northern Transvaal

Balantidium coli
Gutierrez Manay, J.; Garcia Calderon, J.; and Hidalgo Pallete, A., 1973, Rev. Med. Caja Nac. Seguro Soc., Lima, v. 22 (3), 402-404
Enterobius vermicularis, Balantidium coli, human, cause of appendicitis, search of surgical case records: Peru

Balantidium coli, illus.
Kim, J. C. S.; Abee, C. R.; and Wolf, R. H., 1978, Lab. Animals, v. 12 (4), 231-233
balantidiosis, colony-born splenectomized Pan troglodytes (mucosal and submucosal layers of colon and caecum; faeces) concomitantly infected with Strongyloides stercoralis and Shigella flexneri, fatal infection, case report

Balantidium coli
Lumbreras, H.; and Uyema, N., 1972, Rev. Peruana Med. Trop., v. 1 (2), 95-98
Balantidium coli, in vitro, comparative action of dehydroemetine and emetine hydrochloride

Balantidium coli
Reddy, R. G., 1979, Livestock Advis., v. 4 (3), 37-40
Balantidium coli and mixed infection with amphistomes, cattle and buffaloes, incidence and treatment

Balantidium coli, illus.
Rees, R. G. P.; and Shelley, A. J., 1977, Acta Amazonica, v. 7 (1), 47-49
Balantidium coli, Yanomama Indian, serious dysentery, case report, Ascaris lumbricoides and Trichuris trichiura also present, chlortetracycline, improved condition: Toototobi, norte do Estado do Amazonas, Brasil

Balantidium coli
Rijksen, H. D., 1978, Mededel. Landbouwhogeschool, Wageningen, v. 78 (2), 420 pp.
Pongo pygmaeus abelii (caecum, colon): Ketambe area, Sumatra

Balantidium coli
Rostkowska, J., 1970, Acta Parasitol. Polon., v. 18 (27-41), 377-392
Balantidium coli, effect of various bacteria on propagation in vitro, on erythrophagocytic capability of balantidia, and on susceptibility of balantidia to atebrin, entobex, mexaform, and protargol; Trichomonas hominis, Chilomastix mesnili, and Dientamoeba fragilis found to be without effect; effect of balantidia on bacteria

Balantidium coli
Supperer, R., 1973, Tieraerztl. Prax., v. 1 (1), 33-42
parasitic diseases of swine, intensive rearing, treatment and control measures, review

Balantidium ctenopharyngodonis
Astakhova, T. V.; and Stepanova, G. A., 1972, Parazitologiia, Leningrad, v. 6 (4), 364-368
Ctenopharyngodon idella (intestine): pond and spawning-nursery fisheries, Volga delta

Balantidium ctenopharyngodonis Chen, 1955, illus.
Molnar, K.; and Reinhardt, M., 1978, J. Fish Dis., v. 1 (2), 151-156
Balantidium ctenopharyngodonis in Ctenopharyngodon idella, gross and microscopic intestinal lesions

Balantidium duodeni (Stein, 1862)
Frandsen, F., 1974, Acta Parasitol. Polon., v. 22 (1-11), 49-66
Rana esculenta (intestine): Denmark

Balantidium duodeni Stein, 1862, illus.
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
synonymy, description
Rana esculenta
R. ridibunda
R. temporaria
(strevo of all): all from CSSR

Balantidium elongatum (Stein, 1862)
Frandsen, F., 1974, Acta Parasitol. Polon., v. 22 (1-11), 49-66
Triturus cristatus
T. vulgaris
(intestine of all): all from Denmark

Balantidium elongatum Stein, 1862, illus.
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
description
Bombina variegata
Pelobates fuscus
Rana esculenta
R. ridibunda
R. temporaria
Triturus vulgaris
all from CSSR

Balantidium entozoon (Ehrenberg, 1838)
Frandsen, F., 1974, Acta Parasitol. Polon., v. 22 (1-11), 49-66
Rana esculenta
Bombina bombina
(rectum of all): all from Denmark

Balantidium entozoon (Ehrenberg, 1838), illus.
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
description
Bombina bombina
B. variegata
Bufo viridis
B. calamita

Balantidium hyalinum Dobell, 1910
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
as syn. of B. duodeni Stein, 1862

Balantidium rotundum Bezzenberger, 1903
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
as syn. of B. duodeni Stein, 1862

Balbiania mucosa Blanchard, 1885
Levine, N. D., 1979, Ann. Trop. Med. and Parasitol., v. 73 (1), 91-92
Balbiania mucosa, descriptive history, erroneously indicated as Sarcocystis; "Until it is studied further, its correct name should be considered Eimeria (?) mucosa (Blanchard, 1885) nov. comb."

Besnoitia
Beier, T. V.; Shibalova, T. A.; and Kostenko, L. A., 1978, [Cytology of coccidia], 185 pp., illus.

Besnoitia
Heydorn, A. O., 1979, Berl. u. Munchen. Tierarztl. Wchnschr., v. 92 (11), 214-220
coccidia of cats, life cycle, epidemiology, review

Besnoitia
Rommel, M., 1978, Ztschr. Parasitenk., v. 57 (3), 269-283
cyst-forming coccidia, life cycle, taxonomy, comparative review

[Besnoitia] beznoitoze
Uvaliev, I. U.; and Baigaziev, K. K., 1979, Vestnik Sel'skokhoz. Nauki Kazakhstana (1), 75-78
besnoitiosis, bovine, disinfection of animals or hides by sodium hydroxide solution spray; disinfection of premises by sodium hydroxide or chlorine solution sprays

Besnoitia [sp.], illus.
Cheema, A. H.; and Toofanian, F., 1979, Cornell Vet., v. 69 (3), 159-168
Besnoitia [sp.], gross and histopathological changes in wild and domestic goats
Capra aegagrus (skin, blood vessels, epididymis, testes)
C. hircus (subcutaneous tissues)
all from Fars Province, Iran

Besnoitia cysts
Keep, M. E.; and Basson, P. A., 1973, J. South African Vet. Ass., v. 44 (3), 287
Phacochoerus aethiopicus (lung, testes, epididymis, lymph node): Hluhluwe Game Reserve, Zululand

Besnoitia sp., illus.
Wobeser, G., 1976, J. Wildlife Dis., v. 12 (4), 566-571
Rangifer tarandus caribou: northern Saskatchewan

Besnoitia besnoiti
Kaggwa, E.; Weiland, G.; and Rommel, M., 1979, Bull. Animal Health and Prod. Africa, v. 27 (2), 127-137
Besnoitia besnoiti, B. jellisoni, rabbits, mice, diagnosis, indirect immunofluorescent antibody technique and enzyme-linked immunosorbent assay compared

Besnoitia besnoiti (Marotel, 1912), illus.
Lee, H. S.; Lee, H. B.; and Moon, M. H., 1979, Korean J. Animal Sc., v. 21 (4), 281-288
Besnoitia besnoiti, goats (exper.), antimony potassium tartrate, pacprim

Besnoitia besnoiti, illus.
Nobel, T. A.; et al., 1977, Bull. Acad. Vet. France, n.s., v. 50 (4), 569-574
Besnoitia besnoiti, cows, pathology, vaginal mucosa and endometrium

Besnoitia besnoiti, illus.
Oduye, O. O., 1975, Bull. Animal Health and Prod. Africa, v. 23 (1), 45-50
Demodex folliculorum bovis, cattle, clinical observations and gross pathology, histopathology, concomitant infections with Dermatophilus congolensis and Besnoitia besnoiti: Nigeria

Besnoitia besnoiti, illus.
Poluboiarova, G. V.; and Iaps, V. V., 1978, Vestnik Sel'skokhoz. Nauki Kazakhstana (10), 89-91
Besnoitia besnoiti, maintained in tissue culture vs. fresh from cutaneous cysts, equally virulent for susliks, similar pathological changes

Besnoitia jellisoni
Chinchilla, M.; and Frenkel, J. K., 1978, Infect. and Immun., v. 19 (3), 999-1012
Toxoplasma gondii, Besnoitia jellisoni, antigen-treated lymphocytes from immune hamsters can directly confer immunity not only to macrophages but also to fibroblasts and kidney cells, infection-specific mediators can be derived from immune lymphocytes in presence of antigen and also confer immunity to macrophages and somatic cells

Besnoitia jellisoni
Dubey, J. P., 1978, J. Protozool., v. 25 (3, pt. 2), 382-384
Hammondia hammondi, Besnoitia jellisoni, Toxoplasma gondii, BCG, comparison of cross-protection in hamsters

Besnoitia jellisoni
Frenkel, J. K., 1978, Fed. Proc., v. 37 (7), 2063-2064
immune responses to nonviral intracellular infections of hamsters, symposium presentation

Besnoitia jellisoni
Frenkel, J. K.; and Caldwell, S. A., 1975, J. Infect. Dis., v. 131 (3), 201-209
Toxoplasma gondii, Besnoitia jellisoni, Listeria, and virus infections in mice and hamsters, challenge with homologous and heterologous species, components of specific immunity and nonspecific resistance

Besnoitia jellisoni
Kaggwa, E.; Weiland, G.; and Rommel, M., 1979, Bull. Animal Health and Prod. Africa, v. 27 (2), 127-137
Besnoitia besnoiti, B. jellisoni, rabbits, mice, diagnosis, indirect immunofluorescent antibody technique and enzyme-linked immunosorbent assay compared

Besnoitia wallacei
Fayer, R.; and Frenkel, J. K., 1979, J. Parasitol., v. 65 (5), 756-762
6 spp. of feline coccidia, oocysts fed to calves, calf tissue then fed to cats: neither Besnoitia, Hammondia, nor Sarcocystis were pathogenic for calves nor did they establish patent infections which could be transmitted back to cats; Cystoisospora spp. were not pathogenic for calves but could be transmitted back to cats; Toxoplasma strains were slightly to moderately pathogenic for calves and could be transmitted back to cats

Bezzenbergia Earl, 1973
Earl, P. R., 1979, Tr. Am. Micr. Soc., v. 98 (4), 549-557
Protoopalininae

Biggaria Kahl, 1934
Iankovskii, A. V., 1973, Parazitologiia, Leningrad, v. 7 (3), 214-219
Hymenostomatida, Tetrahymenina, Entodiscidae fam. n.

Blastocrithidia [sp.]
da Rocha e Silva, E. O.; et al., 1977, Rev. Saude Pub., S. Paulo, v. 11 (1), 87-96
Triatoma sordida: Estado de Sao Paulo, Brasil

Blastocrithidia sp., illus.
da Rocha e Silva, E. O.; et al., 1977, Rev. Saude Pub., S. Paulo, v. 11 (1), 87-96
Panstrongylus megistus (tubo intestinal): Bairro Santo Antonio, municipio de Sao Joao da Boa Vista

Blastocrithidia [sp.], similar to B. triatomae
da Rocha e Silva, E. O.; et al., 1977, Rev. Saude Pub., S. Paulo, v. 11 (1), 87-96
Triatoma infestans
T. sordida
Panstrongylus megistus
P. lignarius
all from insect colony, Moji Guacu

Blastocrithidia culicis
Camargo, E. P.; Itow, S.; and Alfieri, S. C., [1979], J. Parasitol., v. 64 (6), 1978, 1120-1121
Trypanosomatidae, 18 spp. of 6 genera, proteolytic activities in cell extracts

Blastocrithidia culicis
Chang, K. P.; et al., 1978, J. Protozool., v. 25 (1), 145-149
methylglyoxal bis(guanylhydrazone) (MGBG), little in vitro effect on Blastocrithidia culicis, Crithidia oncopelti, and Leishmania spp. but complete inhibition of growth of Trypanosoma brucei, reduced parasitemia of T. brucei and T. congolense in rats but infections relapsed, tracer studies with T. brucei showed that MGBG interfered with nucleoside incorporation

Blastocrithidia culicis
Fish, W. R.; Holz, G. G., jr.; and Beach, D. H., 1978, J. Parasitol., v. 64 (3), 546-547
26 trypanosomatid species, cultivation in new chemically-defined medium RE III

Blastocrithidia culicis
Galinari, S.; and Camargo, E. P., 1979, J. Parasitol., v. 65 (1), 88
Blastocrithidia culicis, symbiont-harboring wild strain has complete set of urea cycle enzymes and acetylornithinase, corresponding aposymbiotic strain lacks some of these enzymes

Blastocrithidia culicis
Goncalves de Lima, V. M. Q.; Roitman, I.; and Kilgour, V., 1979, J. Protozool., v. 26 (4), 648-652
trypanosomatids, 7 species distinguished by electrophoretic mobilities of some isoenzymes

Blastocrithidia culicis
Mendes, N. F.; et al., 1979, Transplant. Proc., v. 11 (2), 1304-1305
cross-reactions between Trypanosomatidae cell extracts and HLA antigens

Blastocrithidia culicis
Yoshida, N.; and Camargo, E. P., 1978, J. Bacteriol., v. 136 (3), 1184-1186
trypanosomatids, excretion of urea or ammonia or both, varies according to genus, may be of taxonomic use

Blastocrithidia triatomae, illus.
da Rocha e Silva, E. O.; et al., 1977, Rev. Saude Pub., S. Paulo, v. 11 (1), 87-96
[host uncertain]: insect colony, Moji Guacu

Blastocrithidia triatomae galvaoi n. subsp., illus.
Carvalho, A. L. de M., 1973, Rev. Patol. Trop., v. 2 (2), 223-274
Zelus leucogrammus (tubo digestivo, orgaos reprodutores femininos, fezes): Goias, Minas Gerais and Santa Catarina, Brasil

Blastocrithidia vagoi, n. sp., illus.
Tuzet, O.; and Laporte, M., 1965, Arch. Zool. Exper. et Gen., v. 105 (2), 77-81
Eurydema ventralis (intestin)

Blastocystis sp., illus.
Burden, D. J.; Anger, H. S.; and Hammet, N. C., 1979, Vet. Microbiol., v. 3 (3), 227-234
Blastocystis sp., pigs (feces), pathogenicity discussed: Berkshire and East Anglia, England

Blastocystis hominis, illus.
Silard, R.; et al., 1977, Arch. Roumaines Path. Exper. et Microbiol., v. 36 (1), 55-60
E[ntamoeba] histolytica, intrahepatic inoculation in Cricetus auratus of human strain associated with Blastocystis hominis, liver abscess, diagnosis, results demonstrate the tissue adaptability of B. hominis and its potential as a conditioned pathogen

Blastocystis h[ominis]
Soto Travieso, R.; et al., 1977, Rev. Cubana Med. Trop., v. 29 (3), 115-120
human intestinal parasites, diagnosis, modified Baermann's technique compared with standard diagnostic methods

Bodo decipiens Diesing (1850)
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
as syn. of Hexamita intestinalis Dujardin, 1841

Bodo intestinalis Ehrenberg (1838)
  Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
    as syn. of Hexamita intestinalis Dujardin, 1841

Buchneriella pheretima, nova sp., illus.
  de Puytorac, P.; and Rakotoarivelo, R., 1965, Arch. Zool. Exper. et Gen., v. 105 (2), 229-240
    Pheretima sp. (gesier): Singapour

Burenella gen. n.
  Jouvenaz, D. P.; and Hazard, E. I., 1978, J. Protozool., v. 25 (1), 24-29
    Burenellidae fam. n.
    mt: B. dimorpha sp. n.

Burenella dimorpha sp. n. (mt), illus.
  Jouvenaz, D. P.; and Hazard, E. I., 1978, J. Protozool., v. 25 (1), 24-29
    pathologic manifestations
    Solenopsis geminata (hypodermis, fat cells) (nat. and exper.): State Road 26 east of Interstate Highway 75, Gainesville, Alachua Co., Florida
    S. invicta (exper.)
    S. richteri (exper.)
    S. xyloni (exper.)

Burenellidae fam. n.
  Jouvenaz, D. P.; and Hazard, E. I., 1978, J. Protozool., v. 25 (1), 24-29
    Microsporida
    includes: Burenella gen. n.

Caudicula subgen. n.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 23-24
subgen. of Protoopalina
type sp.: Protoopalina (Caudicula) caudata (Zeller, 1877) Metcalf, 1923

Caudicula
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
subgen. of Protoopalina, key

Caudiculata subgen. n.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 23-24
subgen. of Protoopalina
type sp.: Protoopalina (Caudiculata) diplocarya Metcalf, 1923

Caudiculata
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
subgen. of Protoopalina, key

Cepedea Metcalf, 1920
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31
Opalinidae
includes: Paucinucleata subgen. n.: Cepedea Metcalf, 1920; Spinifera subgen. n.; Pulchra subgen. n.; Phrynomantidis subgen. n.; Longa subgen. n.; Plata subgen. n.; Globoides subgen. n.

Cepedea Metcalf, 1920
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
Opalininae, key
key to subgenera

Cepedea Metcalf, 1920
Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (4), 205-209
Cepedea, subgenera Cepedea, Longa, and Plata, synopsis of Brazilian species

Cepedea Metcalf, 1920
Earl, P. R., 1979, Tr. Am. Micr. Soc., v. 98 (4), 549-557
Protoopalininae

Cepedea Metcalf, 1920
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31
subgen. of Cepedea
type sp.: Cepedea (Cepedea) dimidiata (Stein, 1860) Metcalf, 1923

Cepedea
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
subgen. of Cepedea, key

Cepedea Metcalf, 1920
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
subgen. of Cepedea
key to species

Cepedea sp.
Amaro, A.; Sena, S.; and dos Santos, E., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 123
Atelopus moreirae: Veu da Noiva, Maromba, em Itatiaia, no Estado do Rio de Janeiro, Brasil

Cepedea sp.
Amaro, A.; Sena, S.; and dos Santos, E., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 123
Hyla cuspidata: Recreio dos Bandeirantes, Lagoinha das Taxas, na cidade do Rio de Janeiro, Estado da Guanabara, Brasil

Cepedea (Longa) ciliata Metcalf, 1940
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Cepedea (Longa) daloalensis Tuzet & Zuber-Vogeli, 1954
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31

Cepedea (Cepedea) dimidiata (Stein, 1860) Metcalf, 1923 (tod of subgen.), illus.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31

Cepedea dimidiata (Stein, 1860), illus.
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
description
Bombina variegata
Bufo bufo
Pelobates fuscus
Rana esculenta
(kloaka of all): all from CSSR

Cepedea (Cepedea) dimidiata cottoniana Mello, 1944
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31

Cepedea (Cepedea) dimidiata naglavensis Uttangi, 1951
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31

Cepedea (Cepedea) dimidiata saigonensis Boisson, 1957
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31

Cepedea (Longa) dracuncula Boisson, 1957
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31

Cepedea (Plata) foliacea Boisson, 1959
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31

Cepedea (Globoides) globosa Metcalf, 1923 (tod of subgen.), illus.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31

Cepedea (Longa) hylae Khan, 1962
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31

Cepedea (Paucinucleata) lanceolata (Bezzenberger, 1904) Metcalf, 1923 (tod of subgen.), illus.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31

Cepedea (Longa) longa (Bezzenberger, 1904) Metcalf, 1923 (tod of subgen.), illus.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31

Cepedea longa (Bezzenberger)
Mukherjee, R., 1976, J. Zool. Soc. India, v. 26 (1-2), 1974, 142-143
brief description
Bufo melanostictus (rectal contents): Dum Dum near Calcutta, India

Cepedea (Cepedea) marginata Amaro, 1964
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31

Cepedea (Cepedea) marginata Amaro, 1964
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Cepedea (Cepedea) mogyana (Carini, 1937) Metcalf, 1940
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Cepedea (Cepedea) mogyana (Carini, 1937) Metcalf, 1940, illus.
Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (4), 205-209
synonymy

Cepedea (Longa) multiformis Metcalf, 1923
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Cepedea (Cepedea) parva Lu, 1945
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31

Cepedea (Longa) philauti Uttangi, 1951
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31

Cepedea (Phrynomantidis) phrynomantidis Metcalf, 1923 (tod of subgen.), illus.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31

Cepedea (Plata) plata Metcalf, 1940 (tod of subgen.), illus.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31

Cepedea (Pulchra) pulchra Metcalf, 1923 (tod of subgen.), illus.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31

Cepedea (Cepedea) rubra (Carini, 1937) Metcalf, 1940
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Cepedea (Cepedea) rubra (Carini, 1937) Metcalf, 1940, illus.
Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (4), 205-209
synonymy

Cepedea (Cepedea) rugosa (Carini, 1937) Metcalf, 1940
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Cepedea (Cepedea) rugosa (Carini, 1937) Metcalf, 1940, illus.
Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (4), 205-209
synonymy

Cepedea (Longa) seychellensis angusta Mello, 1932
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31

Cepedea (Spinifera) spinifera Metcalf, 1923 (tod of subgen.), illus.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31

Cepedella Poyarkof
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key

Cephaloidophora
Theodorides, J., 1979, Compt. Rend. Acad. Sc., Paris, v. 289, s. D, Sc. Nat. (3), 363-365
intestinal eugregarines of Brachyura, distribution among hosts confirms validity of new classification system for hosts

Ceratomyxa sp.
Lester, R. J. G., 1974, Syesis, v. 7, 195-200
Gasterosteus aculeatus (intestine): near Vancouver, British Columbia

Ceratomyxa spp.
Paperna, I.; and Baudin Laurencin, F., 1979, Aquaculture, v. 16 (2), 173-175
Dicentrarchus labrax (gall bladder): marine cultures in France

Ceratomyxa sp., illus.
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
Scorpaena porcus (gall bladder): Black Sea

Ceratomyxa anoplopoma sp. n., illus.
Moser, M., 1976, Canad. J. Zool., v. 54 (8), 1405-1406
Anoplopoma fimbria (gallbladder): off southern California

Ceratomyxa arcuata Thelohan, 1895, illus.
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
Uranoscopus scaber (gall bladder): Black Sea

Ceratomyxa asymmetrica sp. n., illus.
Moser, M.; and Noble, E. R., 1976, Canad. J. Zool., v. 54 (9), 1535-1537
Coryphaenoides cinereus (gall bladder): waters off Alaska (Shumagin Island)
Ventrifossa macropogon (gall bladder): waters off Venezuela

Ceratomyxa australis sp. n., illus.
Gaevskaia, A. V.; and Kovaleva, A. A., 1979, Biol. Moria, Vladivostok (3), 80-83
Trachurus trachurus capensis (gall bladder): Namibia region, southeastern Atlantic

Ceratomyxa coryphaenoida sp. n., illus.
Moser, M.; and Noble, E. R., 1976, Canad. J. Zool., v. 54 (9), 1535-1537
Coryphaenoides cinereus (gall bladder): waters off Canada (Triangle Island)

Ceratomyxa elegans Jameson, 1929
  Moser, M.; and Noble, E. R., 1976, Canad. J. Zool., v. 54 (9), 1535-1537
    Coryphaenoides fernandezianus
    Ventrifossa potronus
    (gall bladder of all): all from waters off northern Chile

Ceratomyxa globulifera Thelohan, 1895
  Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
    Ophidion rochei (gall bladder): Black Sea

Ceratomyxa herouardi Georgevitch, 1916, illus.
  Siau, Y., 1979, Compt. Rend. Acad. Sc., Paris, v. 288, s. D, Sc. Nat. (4), 403-404
    Myxobolus exiguus, Ceratomyxa herouardi, synaptonemal complexes, electron microscopic observations, implications for life cycle and classification of myxosporidians

Ceratomyxa hokarari Meglitsch, 1960
  Moser, M.; and Noble, E. R., 1976, Canad. J. Zool., v. 54 (9), 1535-1537
    Coryphaenoides acrolepis (gall bladder, bile ducts): waters off northern Mexico; southern, central, and northern California; Oregon; British Columbia
    C. fernandezianus (gall bladder): waters off northern Chile

Ceratomyxa hopkinsi Jameson, 1929
  Moser, M.; and Noble, E. R., 1976, Canad. J. Zool., v. 54 (9), 1535-1537
    Nezumia stelgidolepis (gall bladder, bile ducts): waters off southern California

Ceratomyxa laxa Meglitsch, 1960
  Moser, M.; and Noble, E. R., 1976, Canad. J. Zool., v. 54 (9), 1535-1537
    Trachyrincus sp. (gall bladder): waters off northern Chile

Ceratomyxa merlangi sp. nov., illus.
  Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
    Odontogadus merlangus euxinus (gall bladder): Black Sea (region near Sevastopol)

Ceratomyxa obesa Jameson, 1929
  Moser, M.; and Noble, E. R., 1976, Canad. J. Zool., v. 54 (9), 1535-1537
    Coryphaenoides marginatus (gall bladder): waters off Japan (Goto Island)
    Malacocephalus occidentalis (gall bladder): waters off Angola

Ceratomyxa parva (Thelohan, 1895)
  Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
    synonymy

Ceratomyxa shasta
  Markiw, M. E.; and Wolf, K., 1978, J. Fish. Research Bd. Canada, v. 35 (6), 828-832
    Myxosoma cerebralis, rabbits immunized with antigens extracted from mature spores or prespore stages, antisera and globulins used in fluorescent antibody techniques, direct fluorescent antibody test showed higher specificity than indirect FAT in cross reactions with other species of myxosporidians

Ceratomyxa shasta (Noble, 1950), illus.
  Yamamoto, T.; and Sanders, J. E., 1979, J. Fish Dis., v. 2 (5), 411-428
    Ceratomyxa shasta, stages of development leading to sporogenesis, light and electron microscopy
    Salmo clarki (exper.)

Ceratomyxa subtilis Meglitsch, 1960
  Moser, M.; and Noble, E. R., 1976, Canad. J. Zool., v. 54 (9), 1535-1537
    Coelorinchus australis (gall bladder): waters off New Zealand (Wellington)
    C. innotabilis (gall bladder, bile duct): waters off New Zealand (East Cape)
    C. occa (gall bladder): waters off Lesser Antilles (Dominica Island)

Cercomonas sp., illus.
  Szidat, L., 1973, Ang. Parasitol., v. 14 (1), 1-10
    Paragyrodactylus superbus n. g., n. sp. (Darm): Aquarium des Naturhistorischen Museums, 'Bernardino Rivadavia', Buenos Aires, Argentinien

Cercomonas ranarum Perty (1852)
  Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
    as syn. of Hexamita intestinalis Dujardin, 1841

Chagas disease
  Albuquerque, R. D. R.; et al., 1972, Rev. Inst. Med. Trop. S. Paulo, v. 14 (1), 1-5
    Chagas disease, human, serial hemoculture in Warren's medium, valuable diagnostic technique

Chagas disease
  Almeida, H. de O.; et al., 1974, Rev. Soc. Brasil. Med. Trop., v. 8 (1), 37-39
    human Chagas disease, complement fixation test performed on pericardial fluid of patients with chronic infection, fewer positive results obtained in patients with severe congestive failure than in those with no or minimal failure

Chagas disease
  Almeida, H. de O.; and Chapadeiro, E., 1978, Rev. Inst. Med. Trop. S. Paulo, v. 20 (5), 293-299
    Chagas disease, humans, endocardial changes in apical region

Chagas disease
  Almeida, H. O.; Pereira, F. E. L.; and Tafuri, W. L., 1975, Rev. Inst. Med. Trop. S. Paulo, v. 17 (1), 5-9
    human Chagas disease, high number of mast cells in myocardium, possibly responsible for sclerosing characteristic of myocarditis

Chagas disease
  Amato Neto, V., 1973, Rev. Paul. Med., v. 81 (2), 93-104
    human protozoan infections, drugs in current use, dosage recommendations, review

Chagas disease
  Andrade, J. C. S.; et al., 1976, Rev. Brasil. Med., v. 33, special no., 129-135
    Chagas disease, human, tachycardia and aneurysms, case reports, medical and surgical management

Chagas disease
Apt, W.; Arribada, A.; and Maubens, S., 1971, Recent Advances Stud. Cardiac Struct. and Metab., v. 2, 95-105
human cardiomyopathies resulting from Chagas disease or Toxoplasma gondii, etiology, clinical aspects, prognosis, clinicopathological correlations with autopsies, review: Chile

Chagas disease
Assini, M. T. T.; Otero, G. O.; and Bertona, C., 1970, Rev. Brasil. Pesqui. Med. e Biol., v. 3 (3-4), 45-49
Chagas disease, woman, pregnancy diagnosed shortly after parasitemia discovered, spontaneous abortion after 5 months of pregnancy, evidence of transplacental transmission to fetus, case report

Chagas disease
Barreto, J. C. de C., 1979, Brit. J. Anaesth., v. 51 (12), 1189 [Letter]
patients with Chagas cardiomyopathy and Stokes-Adams syndrome with total atrioventricular block, modifications of anaesthesia technique for surgical implantation of epicardial pacemakers

Chagas disease
Baruffa, G., 1975, Rev. Soc. Brasil. Med. Trop., v. 9 (2), 95-104
human Chagas disease, survey of randomly selected rural people for evidence of infection using complement fixation and electrocardiograms, no correlations could be established between the two tests: county of Encruzilhada do Sul, State of Rio Grande do Sul, Brazil

Chagas disease
Baruffa, G.; and Alcantara Filho, A., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (2), 117-123
Chagas disease, humans, prevalence survey using complement fixation test: southern zone, State of Rio Grande do Sul, Brazil

Chagas disease
Bastaroli, J. C.; et al., 1975, Rev. Argent. Cardiol., v. 43 (6), 421-437
Chagasic sinus node disease, human, clinical manifestations, case report: Argentina

Chagas disease
Becker, P. F. L.; et al., 1979, Rev. Paul. Med., v. 93 (5-6), 91-96
Chagas disease, comparative study of weight and metric features of diaphragm between chagasic (hypertrophic) and nonchagasic cardiopathies, autopsy study of 50 males: Brasil

Chagas disease
Bittencourt, A. L.; et al., 1972, Rev. Inst. Med. Trop. S. Paulo, v. 14 (2), 131-134
Chagas disease, incidence of congenital transmission in premature infants: Salvador, Bahia

Chagas disease
Bittencourt, A. L.; et al., 1974, Rev. Inst. Med. Trop. S. Paulo, v. 16 (4), 197-199
human Chagas disease, 400 term pregnancies, incidence in mothers and infants, no congenital transmission

Chagas disease
Bittencourt, A. L.; and Barbosa, H. S., 1972, Rev. Inst. Med. Trop. S. Paulo, v. 14 (4), 257-259
Chagas disease, human, survey of aborted fetal remains for incidence of congenital transmission: Brazil

Chagas disease
Bittencourt, A. L.; and Barbosa, H. S., 1972, Rev. Inst. Med. Trop. S. Paulo, v. 14 (4), 260-263
Chagas disease, congenital, diagnosis through study of fetal remains

Chagas disease
Blandon, R.; Guevara, J. F.; and Johnson, C. M., 1976, Rev. Med. Panama, v. 1 (3), 153-162
Chagas disease in children, diagnosis, pathology, therapeutic trials with various drugs, metronidazole was well tolerated and therefore most promising therapy: Panama

Chagas disease
Boainain, E.; Rassi, A.; and Jatene, A. D., 1975, Rev. Goiana Med., v. 21 (3-4), 129-130
Chagas disease, prevalence survey of patients at the Instituto de Cardiologia in Sao Paulo state, Brasil

Chagas disease
Boehm, G. M., 1972, Rev. Inst. Med. Trop. S. Paulo, v. 14 (4), 277-289
Chagas disease, rats, pulmonary vascular permeability changes suggested to be due to aging, not disease

Chagas disease
Bosque, C.; and Rabinovich, J. E., 1979, Canad. Entom., v. 111 (2), 171-180
Telenomus fariai parasitizing eggs of Triatoma phyllosoma pallidipennis, vector of Chagas' disease, oviposition behavior and host discrimination

Chagas disease
Brandalise, N. A.; et al., 1974, Rev. Paul. Med., v. 83 (4), 169-172
Chagas disease, man, development of esophageal carcinoma shortly after surgical repair of Chagasic megaesophagus, case report: Brazil

Chagas disease
de Britto Costa, R.; and Gallina, R. A., 1971, Rev. Inst. Med. Trop. S. Paulo, v. 13 (2), 92-98
Chagas disease, human, hypothalamus, pathologic changes

Chagas disease
de Britto-Costa, R.; Martins, E. (Neto Junior); and Mabtum, J., 1973, Rev. Inst. Med. Trop. S. Paulo, v. 15 (4), 227-234
Chagas disease, human, biochemical alterations in mucous secretions of digestive tract and bronchus attributed to disturbances of autonomic regulation resulting from infection

Chagas disease
Campana, J. O., 1977, Rev. Paul. Med., v. 90 (3-4), 37-44
Chagas disease, decreased lipoprotein lipase activity in persons with chronic infection

Chagas disease
Canese, A., 1974, Rev. Paraguaya Microbiol., v. 9 (1), 45-51
Chagas disease, statistics of epidemiologic and vector survey: Paraguay

Chagas disease
Canese, A., 1978, Rev. Paraguaya Microbiol., v. 13 (1), 7-19
Chagas disease, current epidemiologic statistics, vectors, suggestions for further investigations: Paraguay

Chagas disease
Canese, A., 1978, Rev. Paraguaya Microbiol., v. 13 (1), 21-24
Chagas disease, bibliography: Paraguay

Chagas disease
Canese, A.; et al., 1975, Rev. Paraguaya Microbiol., v. 10 (1), 47-54
Chagas disease, statistics of epidemiologic survey taken during 1974 in various areas of Paraguay

Chagas disease
Canese, A.; and Canese, J., 1976, Rev. Paraguaya Microbiol., v. 11 (1), 35
Chagas disease, vector survey: Paraguay

Chagas disease
Canese, J.; and Brice, E., 1977, Rev. Paraguaya Microbiol., v. 12 (1), 9-10
Chagas disease, Indians of the Paraguayan Chaco, antibody prevalence survey: Paraguay

Chagas disease
Canese, J.; and Brice, E., 1978, Rev. Paraguaya Microbiol., v. 13 (1), 3-6
Chagas disease, incidence survey in Indian populations of the Chaco area, search for antibodies using indirect immunofluorescence and direct agglutination tests: Paraguay

Chagas disease
Castagnino, H. E.; Cicco, J. A.; and Thompson, A., 1976, Semana Med. (4970), an. 83, v. 149 (21), 701-706, 725
Chagas disease, humans, chronic myocardiopathy, clinical aspects, differential diagnosis, pathology, review

Chagas disease
Chiari, E.; and Dias, J. C. P., 1975, Rev. Soc. Brasil. Med. Trop., v. 9 (3), 133-136
human Chagas disease, diagnosis using Lit medium and Mourao and Mello technique for blood cultures

Chagas disease
Cotarelli, D. A.; et al., 1972, Rev. Patol. Trop., v. 1 (4), 421-429
Chagas disease, human, prevalence survey: Londrina

Chagas disease
Dain, A. L., 1973, Rev. Fac. Cien. Med. Univ. Nac. Cordoba, v. 31 (3), 227-235
Chagas disease, statistics of survey of potential blood donors found to have positive sera, recommendations for possible control measures: Cordoba, Argentina

Chagas disease
Dantas, A. N. M.; et al., 1975, Rev. Brasil. Pesqui. Med. e Biol., v. 8 (5-6), 427-431
human Chagas disease with resulting megaesophagus, surgical procedure for successful repair of diseased area

Chagas disease
Del Nero, E., jr.; et al., 1974, Rev. Hosp. Clin., S. Paulo, v. 29, suppl., 383-389
human Chagas cardiac disease, phonomechanocardiographic parameter of left ventricular systole in pre-clinical infection

Chagas disease
Del Nero, E., jr.; et al., 1975, Rev. Hosp. Clin., S. Paulo, v. 30 (4), suppl., 281-285
human Chagas cardiac disease, relationship between the preejection and ejection phases of the left ventricle in chronic infections

Chagas disease
Esteso, S. C.; and Ferrari, I., 1973, Rev. Fac. Cien. Med. Univ. Nac. Cordoba, v. 31 (2), 117-123
human Chagas disease, recommendations for therapy of chronic cardiopathies with imolamine chlorhydrate

Chagas disease
Etcheverry, D., 1976, Semana Med. (4962), an. 83, v. 149 (13), 444-447
Chagas disease, humans, statistics of incidence survey: Province of Buenos Aires

Chagas disease
Feliciangeli, M. D., 1973, Rev. Inst. Med. Trop. S. Paulo, v. 15 (4), 235-238
Chagas disease vector Rhodnius prolixus eggs found naturally parasitized by endophagous Microhymenoptera, possible importance as biological control: Venezuela

Chagas disease
Feliciangeli, D.; and Rabinovich, J. E., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (1), 21-34
effects of population density on Ooencyrtus trinidadensis, parasite of Rhodnius prolixus eggs: Venezuela

Chagas disease
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key to triatomids that are vectors of human Chagas disease in Brazil

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Chagas disease, human, demonstration of degenerative lesions in cardiac autonomic fibers and interstitial cells supports theory that cardiopathy is of neurogenic origin

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human Chagas disease, first autochthonous cases reported from the State of Piaui, Brazil

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Fiorillo, A. M.; and Muccillo, G., 1975, Rev. Brasil. Pesqui. Med. e Biol., v. 8 (1), 1-8
Chagas disease in rats, excretion of urinary catecholamines under basal conditions, after insulin hypoglycemia and under reserpine stimulation, comparison with normal controls

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Giannini, S. D.; et al., 1975, Rev. Hosp. Clin., S. Paulo, v. 30 (4), suppl., 257-263
Chagas disease, aspects of lipid metabolism, comparison study of persons with chronic infections, chronic infections with cardiopathy, and normal controls, results imply that persons with chronic Chagas and cardiopathy may have lowered triglyceride synthesis

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Giannini, S. D.; et al., 1976, Rev. Hosp. Clin., S. Paulo, v. 31 (2), 109-115
Chagas disease, human chronic infections, activity of lipoprotein lipase, compared with normal controls

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altered inotropic and chronotropic effects of noradrenaline on isolated rat atria exposed to chagasic sera, possible significance in alterations observed in chronic Chagas heart disease

Chagas disease
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Chagas disease, patients, acute dilatation of chagasic megacolon, clinical aspects, medical and surgical management, differential diagnosis: Brasil

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de Gutierrez, M. V.; et al., 1977, Rev. Neurol. Argentina, v. 3 (3), 452-457
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Chagas disease
Hedge, E. C.; Moody, A. H.; and Ridley, D. S., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (4), 445 [Letter]
Leishmania donovani, human, screening of sera for kala-azar using Crithidia sp. as antigen in indirect fluorescent antibody test, some positive reactions with L. tropica, cross-reactions at low titer with Chagas' disease and African trypanosomiasis

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nitroblue tetrazolium test of limited diagnostic value in areas with high incidence of human intestinal parasites associated with Chagas disease or bilharziosis

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Kuenzle, J. E.; and Ziliotto, A., jr., 1975, Rev. Goiana Med., v. 21 (1-2), 29-36
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Lemos, B. C., 1975, Rev. Brasil. Med., v. 32 (7), 503-505
Chagas disease, human, clinical evaluation of equine specific beta gammaglobulin as therapy

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gastric emptying time, patients with Chagas disease or gastric ulcer

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Chagas disease, intracardiac autonomic nervous system pathology compared with pathology of rheumatic heart disease and hypertensive heart disease

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Chagas disease, humans, chronic infections, heart weights at death compared with heart weights in deaths from other causes, legal medicine aspects

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Panstrongylus megistus leucofasciatus and Rhodnius nasutus, first encounter in interior of houses in State of Pernambuco, Brazil

Chagas disease
Ludwig, A. E., 1972, Rev. Ass. Med. Rio Grande do Sul, v. 16 (2), 193-208
new type of file card designed for the selection of blood donors and for computerized data processing in order to identify potential donors who are carriers of Chagas disease and malaria

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latent Chagas disease, humans, evaluation of physical capacity and myocardial function

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Chagas disease, human, diagnosis in rural areas using immunofluorescence, recommendations for storage of serum absorbed on filter paper samples

Chagas disease
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Telenomus fariai, parasite of Chagas' disease vectors, population dynamics, laboratory conditions

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Chagas disease, human chronic cardiomyopathy amiodarone hydrochloride as therapy for extra ventricular systole

Chagas disease
Nunesmaia, H. G.; and Azevedo, E. S., 1973, Rev. Inst. Med. Trop. S. Paulo, v. 15 (1), 10-13
Chagas disease, autopsy survey of cardiac associated infections, possible association between Chagas infection and the Negro race: Bahia, Brasil

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de Oliveira, A. R., 1972, Rev. Patol. Trop., v. 1 (3), 429-492
Chagas disease, humans, physiopathology of megaesophagus resulting from parasitic infection, extensive report

Chagas disease
Pagano, M. A.; O'Neill, E.; and Aristimuno, G., 1977, Rev. Neurol. Argentina, v. 3 (3), 467-470
human Chagas disease, nifurtimox therapy, development in some persons of an axonal neuropathy involving mainly the lower limbs

Chagas disease
Pagano, M. A.; O'Neill, E.; and Ratusnu, A., 1977, Rev. Neurol. Argentina, v. 3 (3), 464-466
Chagas disease, electromyographic study of persons with chronic infections, finding that there is some involvement of motor neurons but that these lesions have been adequately compensated by healthy neurons

Chagas disease
Pagniez, G.; et al., 1976, Rev. Asoc. Bioquim. Argent. (224), v. 41, 38-42
Chagas disease, efficacy of the direct agglutination test for diagnosis (with and without the addition of 2-mercaptoethanol), complement fixation test used as reference; concluded that test useful mainly in areas such as blood banks

Chagas disease
Pareja, J. C.; et al., 1978, AMB, Rev. Ass. Med. Brasil., v. 24 (2), 66-68
Chagas disease, humans with advanced megaesophagus, subtotal esophagectomy, new surgical technique: Brazil

Chagas disease
de Paula-Costa, M. D.; and de Rezende, J. M., 1978, AMB, Rev. Ass. Med. Brasil., v. 24 (8), 269-272
Chagas megaesophagus, humans, abnormally increased pressure of lower esophageal sphincter

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Chagas disease, males, necropsy study, increased incidence of tracheal deviations: Brazil

Chagas disease
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human Chagas cardiac disease, phonocardiographic systolic parameters and complete heart block

Chagas disease
Perlowagora-Szumlewicz, A., 1973, Rev. Soc. Brasil. Med. Trop., v. 7 (4), 243-260
Triatominae vectors of Chagas disease, spontaneous morphogenetic juvenilization observed in laboratory populations

Chagas disease
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Chagas disease, hybridization experiments with triatomine vectors, sterile males produced, genetic mechanisms, possible control method

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Triatominae (Chagas disease vectors), sex identification of immature forms, possible application in sterile male release

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Pfeferman, A.; et al., 1974, AMB, Rev. Ass. Med. Brasil., v. 20 (5), 191-193
Chagas disease, humans, tachycardia-bradycardia syndrome, 2 case reports, differential diagnosis: Brazil

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Chagas' disease, slow-release juvenile hormone formulations for control of triatomine vectors

Chagas disease
Pinotti, H. W.; et al., 1974, AMB, Rev. Ass. Med. Brasil., v. 20 (9), 331-334
Chagas disease, humans with megaesophagus, new surgical treatment

Chagas disease
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Chagas disease, humans, nifurtimox, R07-1051, comparative study

Chagas disease
Pugliese, C.; Lessa, I.; and Santos Filho, A., 1976, Rev. Inst. Med. Trop. S. Paulo, v. 18 (3), 191-201
Chagas disease, survival time of humans with decompensated chronic myocardiopathy, congestive heart failure may be important limiting factor in survival time

Chagas disease
de Rezende, J. M.; and Moreira, H., 1975, Rev. Gastroenterol. Mexico, v. 40 (1), 12-17
Chagas disease, humans, megacolon resulting from parasitic infection, diagnosis, clinical management, surgical treatment with the Duhamel-Haddad operation being the preferred surgical method: Goiania, State of Goias

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Rivera Gallegos, B. T.; and Arrarte Congrains, R., 1973, Rev. Med. Caja Nac. Seguro Soc., Lima, v. 22 (2), 281-285
human Chagasic cardiomyopathy with A-V heart block, implantation of permanent pacemaker, case report: Arequipa

Chagas disease
Ruggieri, H.; and Piserchia, C. N., 1977, Rev. Neurol. Argentina, v. 3 (3), 474-476
Chagas disease, humans with psychiatric disturbances, symptoms intensified by nifurtimox therapy, disappearance or diminution of symptoms when drug was discontinued

Chagas disease
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Chagas disease, xenodiagnosis and complement fixation test compared: district of Bambui, Minas Gerais, Brazil

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Salgado Filho, W.; et al., 1976, AMB, Rev. Ass. Med. Brasil., v. 22 (7), 237-240
Chagas disease, outpatients, review of cases from 1960-1970, decreased incidence of cardiopathy: Hospital das Clinicas de Ribeirao Preto

Chagas disease
Sanz Malaga, G.; Morales Gonzales, W.; and Boza Revilla, A., 1975, Rev. Peruana Cardiol., v. 18 (1), 5-21
Chagas disease, chronic cardiomyopathy, pathology

Chagas disease
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Triatoma infestans, Rhodnius prolixus (vectors of Chagas' disease), assembly pheromone of nymphs

Chagas disease
Siqueira, L. A.; and Ayala, M. A. R., 1972, Rev. Soc. Brasil. Med. Trop., v. 6 (5), 251-255
Chagas disease, humans, investigation of pulmonary hemosiderosis in subjects with Chagasic and non-Chagasic myocarditis, concluded that entity results from passive congestion and does not represent a feature of Chagas disease

Chagas disease
Tafuri, W. L.; Lopes, E. R.; and Nunan, B., 1973, Rev. Inst. Med. Trop. S. Paulo, v. 15 (5), 322-330
Chagas disease, congenital case report, infant with cardiopathy and esophageal disperistalsis from birth: Belo Horizonte, Brazil

Chagas disease
Teruel, J. R.; and Nogueira, J. L., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (4), 239-244
Chagas disease, highly endemic area for chronic infection, comparison of infected and non-infected women showed no differences in fetal losses: nordeste do Estado de Sao Paûlo

Chagas disease
Vieira, C. B.; and Meneghelli, U. G., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (3), 175-178
Chagas disease, human, chronic infections, frequent evidence of hypoglycemia

Chagas disease
Viviani, C. A.; et al., 1977, Rev. Neurol. Argentina, v. 3 (3), 471-473
Chagas disease, humans with neurologic disturbances, symptoms aggravated by nifurtimox therapy

Chagasella Machado, 1911
Levine, N. D., 1977, Protozoology, v. 3, 173-182
Syn.: Chagasia Leger, 1911

(?)Chagasella sp. Kirby, 1932
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Chagasella sp. Gibbs, 1944
Levine, N. D., 1977, Protozoology, v. 3, 173-182
as syn. of Chagasella gibbsi n. sp.

Chagasella alydi Machado, 1913
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Chagasella ganapatii Narasimhamurti and Kalavati, 1968
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Chagasella gibbsi n. sp.
Levine, N. D., 1977, Protozoology, v. 3, 173-182
Syn.: Chagasella sp. Gibbs, 1944
Cenaeus carnifex (salivary glands): Africa

Chagasella hartmanni (Chagas, 1910) Machado, 1911
Levine, N. D., 1977, Protozoology, v. 3, 173-182
synonymy

Chagasia Leger, 1911
Levine, N. D., 1977, Protozoology, v. 3, 173-182
as syn. of Chagasella Machado, 1911

Chagasia hartmanni (Chagas, 1910) Leger, 1911
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as syn. of Chagasella hartmanni (Chagas, 1910) Machado, 1911

Chapmanium gen. n.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Thelohaniidae fam. n., key
tod: C. cirritus sp. n.

Chapmanium cirritus sp. n. (tod), illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Syn.: Thelohania sp. Chapman et al., 1969
Corethrella brakeleyi (adipose tissue): near Lake Charles, Louisiana, U.S.A.

Chapmanium macrocystis (Gurley, 1893) comb. n.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Syns.: Thelohania macrocystis Gurley, 1893; Sarcosporidi Garbini, 1891; Sarcosporidae Henneguy and Thelohan, 1892

Chapmanium nepae (Lipa, 1966) comb. n.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Syn.: Thelohania nepae Lipa, 1966

Chilodonella cyprini
Astakhova, T. V.; and Stepanova, G. A., 1972, Parazitologiia, Leningrad, v. 6 (4), 364-368
Ctenopharyngodon idella (skin): pond and spawning-nursery fisheries, Volga delta

Chilodonella cyprini (Moroff, 1902), illus.
Heckmann, R.; and Liebelt, J., 1970, J. Wildlife Dis., v. 6 (3), 174
Catostomus catostomus (gill): Yellowstone River, Livingston, Montana

Chilodonella cyprini (Morof, 1902)
Vlasenko, M. I.; and Meshcheriakova, A. A., 1977, Veterinariia, Moskva (4), 75-78
Chilodonella cyprini, Ichthyophthirius multifiliis, [Costia], fish, formalin solution for control, toxicity tested and safe levels established

Chilodonella cyprini (Moroff, 1902)
Wierzbicki, K., 1970, Acta Parasitol. Polon., v. 18 (1-12), 45-55
Perca fluviatilis (gills): Lake Dargin, Mazurian Lakeland, Poland

Chilodonella cyprini Moroff
Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
parasites of fishes: China

Chilodonella hexasticha (Kiernik, 1909), illus.
Hoffman, G. L.; et al., 1979, J. Fish Dis., v. 2 (2), 153-157
Chilodonella hexasticha on Ictalurus punctatus and Carassius auratus (gills of both), occurrence at new water temperature optimum, histopathology: North America

Chilogregarina gen. n.
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
Actinocephalidae; Actinocephalinae
tod: C. striata (Leger & Duboscq, 1903) comb. n.

Chilogregarina brasiliensis (Pinto, 1918) comb. n.
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
Syn.: Gregarina brasiliensis Pinto, 1918
Scolopendra sp. (intestine): Brazil

Chilogregarina dujardini (Schneider, 1875) comb. n.
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
Syns.: Actinocephalus dujardini Schneider, 1875; Acanthocephalus dujardini (Schneider, 1875) Balbiani, 1884
Lithobius forficatus (intestine): Europe; U.S.S.R.; North America

Chilogregarina stella (Leger, 1894) comb. n.
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
Syns.: Rhopalonia stella Leger, 1894; Actinocephalus stella (Leger, 1894) Ormieres, 1967
Geophilus gabrielis (digestive tract)
G. longicornis (digestive tract): France; Germany

Chilogregarina striata (Leger & Duboscq, 1903) comb. n. (tod)
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
Syn.: Actinocephalus striatus Leger & Duboscq, 1903
Scolopendra cingulata (intestine): France

Chilomastix
Zierdt, W. S., 1978, Am. J. Clin. Path., v. 70 (1), 89-93
human intestinal parasites, description of simple method for recovery of eggs, larva and cysts from feces by concentration device, comparison with other methods of parasite recovery

Chilomastix sp.
Kocan, A. A.; Hannon, L.; and Eve, J. H., 1979, Proc. Oklahoma Acad. Sc., v. 59, 20-22
Colinus virginianus (intestine): Oklahoma

Chilomastix sp., illus.
Koesters, J.; Cubillos, A.; and Zuelch, S., 1978, Bol. Chileno Parasitol., v. 33 (3-4), 69-72
Chilomastix sp., cultivation in Dobell Laidlaw medium; in vitro and in vivo (chickens) testing of metronidazole
gansos
pollitos (exper.)

Chilomastix gallinarum
Koesters, J.; et al., 1979, Berl. u. Munchen. Tierarztl. Wchnschr., v. 92 (13), 266-268
Chilomastix gallinarum in chickens (exper.) and in vitro, metronidazole

Chilomastix mesnili
Akhtaruzzaman, K. M.; et al., 1978, Tropenmed. u. Parasitol., v. 29 (4), 427-431
comparison of different methods for detection of intestinal protozoa and helminths in human stool

C[hilomastix] mesnili
Apt, W.; et al., 1977, Rev. Med. Chile, v. 105 (6), 388-389
human intestinal protozoa, modified Teleman and PAFS methods compared for diagnostic purposes

Chilomastix mesnili
Bartlett, M. S.; et al., 1978, J. Clin. Microbiol., v. 7 (6), 524-528
modified zinc sulfate flotation technique evaluated in comparison with formalin-ether concentration method for recovery of protozoan cysts and helminth eggs and larvae from feces preserved in formalin less than and longer than 1 month, results suggest that (except for schistosomes) F-$ZnSO_4$ compares favorably to FE method for detecting infections of clinical significance

Chilomastix mesnili
Blecka, L. J., 1978, J. Parasitol., v. 64 (2), 362-363
patients infected with intestinal parasites, attempt to quantitate immunoglobulin levels in fecal extracts with radial immunodiffusion, mean IgA levels higher than in controls, other immunoglobulin classes rarely detectable

Chilomastix mesnili
Goldsmid, J. M.; and Rogers, S., 1978, J. South African Vet. Ass., v. 49 (2), 109-111
Papio ursinus (feces): Rhodesia, imported from Northern Transvaal

Chilomastix mesnili, illus.
Harvey, J., 1973, Saguenay Med., v. 20 (3), 146-156
parasitic flagellates, life cycle, infection in man, clinical signs, therapy, brief review

Chilomastix mesnili
Rostkowska, J., 1970, Acta Parasitol. Polon., v. 18 (27-41), 377-392
Balantidium coli, effect of various bacteria on propagation in vitro, on erythrophagocytic capability of balantidia, and on susceptibility of balantidia to atebrin, entobex, mexaform, and protargol; Trichomonas hominis, Chilomastix mesnili, and Dientamoeba fragilis found to be without effect; effect of balantidia on bacteria

Chilomastix m[esnili]
Soto Travieso, R.; et al., 1977, Rev. Cubana Med. Trop., v. 29 (3), 115-120
human intestinal parasites, diagnosis, modified Baermann's technique compared with standard diagnostic methods

Chloromyxum sp.
Arthur, J. R.; Margolis, L.; and Arai, H. P., 1976, J. Fish. Research Bd. Canada, v. 33 (11), 2489-2499
Thymallus arcticus (gall bladder): Aishihik Lake, Yukon Territory
Coregonus clupeaformis (gall bladder): Aishihik Lake and Stevens Lake, Yukon Territory

Chloromyxum catostomi Kudo, 1919, illus.
Mitchell, L. G., 1978, J. Protozool., v. 25 (1), 100-105
diagnosis
Campostoma anomalum
Notropis dorsalis
Pimephales notatus
P. promelas
Semotilus atromaculatus
(gall bladder of all): all from Iowa

Chloromyxum coregoni
Makhovenko, E. T., 1972, Parazitologiia, Leningrad, v. 6 (4), 369-375
Salvelinus alpinus inhabiting different ecological niches, possible use of differences in parasite fauna between groups as biological tags: Lake Azabach'e, Kamchatka

Chloromyxum cristatum Leger
Iziumova, N. A.; Mashtakov, A. V.; and Timoshechkina, L. G., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36), 71-76
[Abramis brama]: Upper Kama

Chloromyxum dubium Auerbach, 1908
Arthur, J. R.; Margolis, L.; and Arai, H. P., 1976, J. Fish. Research Bd. Canada, v. 33 (11), 2489-2499
Lota lota (gall bladder): Aishihik Lake, Yukon Territory

Chloromyxum dubium
Chernysheva, N. B., 1975, Izvest. Gosudarstv. Nauchno-Issled. Inst. Ozer. i Rech. Ryb. Khoziaistva, Leningrad, v. 93, 117-120
protozoans of young predatory fish, extent of infection correlated with some environmental factors
[Lota lota]: Vrevo Lake, Leningrad district

Chloromyxum esocinum Dogiel, 1934
Arthur, J. R.; Margolis, L.; and Arai, H. P., 1976, J. Fish. Research Bd. Canada, v. 33 (11), 2489-2499
Esox lucius (gall bladder): Stevens Lake, Yukon Territory

Chloromyxum legeri Touraine
Iziumova, N. A.; Mashtakov, A. V.; and Timoshechkina, L. G., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36), 71-76
[Abramis brama]
[Rutilus rutilus]
all from Upper Kama

Chloromyxum mucronatum
Chernysheva, N. B., 1975, Izvest. Gosudarstv. Nauchno-Issled. Inst. Ozer. i Rech. Ryb. Khoziaistva, Leningrad, v. 93, 117-120
protozoans of young predatory fish, extent of infection correlated with some environmental factors
[Lota lota]: Vrevo Lake, Leningrad district

Chloromyxum quadratum Thelohan, 1895
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
as syn. of Kudoa quadratum (Thelohan, 1895)

Chloromyxum trijugum Kudo, 1919, illus.
Mitchell, L. G., 1978, J. Protozool., v. 25 (1), 100-105
Lepomis cyanellus
L. macrochirus
Pomoxis annularis
P. nigromaculatus
(gall bladder of all): all from Iowa

Chloromyxum trijugum Kudo, 1919, illus.
Mitchell, L. G., 1978, Northwest Sc., v. 52 (3, pt. 2), 236-242
Chloromyxum trijugum, description, host specificity
Lepomis gibbosus (gall bladders): western Montana

Chloromyxum truttae Leger, 1906, illus.
Mitenev, V. K., 1971, Parazitologiia, Leningrad, v. 5 (6), 556-558
[Salmo salar]
[Salmo trutta]
(gall bladder of all): all from mouth of Ponoy River

Chloromyxum wardi Kudo, 1919, illus.
Mitchell, L. G., 1978, Northwest Sc., v. 52 (3, pt. 2), 236-242
Chloromyxum wardi, description, host specificity
Salmo clarki
S. gairdneri
Salvelinus fontinalis
Salvelinus malma
(gall bladders of all): all from western Montana

Ciliate
Gupta, P. P.; and Kalra, I. S., 1978, Indian Vet. J., v. 55 (11), 916
ciliate protozoa, bullock (lung), death appeared to be due to pulmonary echinococcosis

Ciliate, unidentified, illus.
Ishii, A., 1973, Snake, v. 5 (1-2), 133-140
Trimeresurus flavoviridis (intestine): Amami-oshima, southern Japan

Ciliates, illus.
Newman, M. W., 1971, Proc. 1970 National Shellfish. Ass., v. 61, 59-63
Crassostrea virginica (gills): New Haven Harbor, Connecticut

Ciliates, resembling Ancistrocomidae [sp.], illus.
  Newman, M. W., 1971, Proc. 1970 National Shellfish. Ass., v. 61, 59-63
    Crassostrea virginica (digestive diverticula, gills): New Haven Harbor, Connecticut

Ciliates
  Wright, C. A.; Rollinson, D.; and Goll, P. H., 1979, Parasitology, v. 79 (1), 95-105
    Bulinus senegalensis and associated parasites, isoelectric focusing studies on enzymes, differences in prevalence and variety of infections in 7 host populations: south bank of Gambia River

Ciliophora Doflein, phylum
  Corliss, J. O., 1977, Tr. Am. Micr. Soc., v. 96 (1), 104-140
    Ciliophora, Corlissian scheme of higher classification, annotated assignment of families and genera to orders and classes

Ciliophora
  Lynn, D. H., 1979, J. Protozool., v. 26 (3), 359-366
    Ciliophora, changes in classification of several major taxa on basis of numerical phenetic analysis

Ciliophora [sp.]
  Keymer, I. F., 1978, Vet. Rec., v. 103 (25), 548-552
    parasitic and other diseases of tortoises, necropsy survey
    Testudo graeca
    Kinixys belliana
    T. nigriti
    T. hermanni
    Geochelone pardalis
    G. gigantea
    T. elegans
    T. horsfieldi
    (intestinal tract of all)

Ciliophora [sp.]
  Keymer, I. F., 1978, Vet. Rec., v. 103 (26-27), 577-582
    parasitic and other diseases of terrapins and turtles, necropsy survey
    Clemmys caspica leprosa
    Staurotypus triporcatus
    (large intestine of all)

Cirrigregarina gen. n.
  Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
    Gregarinidae
    tod: C. spissa (Henry, 1938) comb. n.

Cirrigregarina kamenote (Hoshide, 1951) comb. n.
  Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
    Syn.: Gregarina kamenote Hoshide, 1951
    Mitella mitella (intestine): Japan; Asia

Cirrigregarina spissa (Henry, 1938) comb. n. (tod)
  Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
    Syn.: Gregarina spissa Henry, 1938
    Balanus crenatus
    B. cariosus
    B. glandula
    (intestine of all): all from Washington State, North America

[Coccidia] koktsidii
  Antonenko, A. E., 1975, Vet. Nauka--Proizvod., Trudy, Minsk, v. 13, 121-126
    nematode eggs, coccidian oocysts, non-litter pig manure, substances tested as disinfectants

[Coccidia] koktsidii
  Antonenko, A. E., 1975, Vet. Nauka--Proizvod., Trudy, Minsk, v. 13, 126-129
    nematode eggs, coccidial oocysts, non-litter pig manure, disinfecting by electrohydraulic effect (high voltage electric impulses)

Coccidia
  Antonenko, A. E., 1976, Vet. Nauka--Proizvod., Trudy, Minsk, v. 14, 179-182
    helminth eggs and coccidia oocysts, effect of ultraviolet rays

Coccidi[a] coccidian oocysts, illus.
  Ashford, R. W., 1979, Ann. Trop. Med. and Parasitol., v. 73 (5), 497-500
    previously undescribed coccidian oocysts, occurrence in 3 humans from very diverse backgrounds: Papua New Guinea

Coccidia
  Beier, T. V.; Shibalova, T. A.; and Kostenko, L. A., 1978, [Cytology of coccidia], 185 pp., illus.

Coccidia
  Blanco, C.; and Aznar, E., 1977, Rev. Cubana Cien. Vet., v. 8 (1), 37-46
    parasites and other agents, diarrhea, calves, clinical aspects, etiology: Cuba

Coccidia
  Hudson, R.; Kitts, W. D.; and Bandy, P. J., 1971, J. Wildlife Dis., v. 7 (3), 171-174
    Ovis canadensis, immunoglobulin response, effects of individual variation, season, and parasite activity

Coccidia
  Joyner, L. P.; et al., 1978, Parasitology, v. 77 (1), 27-31
    coccidia, particularly of the genus Eimeria, proposed terminology which provides for discussion of variation at infrasubspecific levels, guidelines for designation of strains and lines

[Coccidia] koktsidiiami
  Lipnitskii, S. S.; and Iakubovskii, M. V., 1975, Vet. Nauka--Proizvod., Trudy, Minsk, v. 13, 143-147
    helminths, protozoa, cattle, influence of micro-elements in host diet on infectivity

[Coccidia] koktsidiami
  Matsuev, V. M., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 119-124
    nematodes, coccidia, chickens, comparative infection levels and egg productivity and quality under various maintenance conditions (with and without enclosures, deep litter, uncovered or asphalt surfaces)

[Coccidia] koktsidii
  Matsuev, V. M., 1973, Vet. Nauka--Proizvod., Trudy, Minsk, v. 11, 159-162
    nematodes, coccidia, age and seasonal dynamics of infection rates of chickens

Coccidia
  Orekhova, M. M., 1970, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 8, 120-124
    ascarid eggs, coccidian oocysts in poultry feces in litter being composted in heaps for biothermic treatment, not surviving after three months

Coccidia, illus.
  Sangster, L. T.; Seibold, H. R.; and Mitchell, F. E., 1976, Proc. 19. Ann. Meet., Am. Ass. Vet. Lab. Diagn., 51-55
    Coccidia, suckling pigs 5-14 days old (intestines), diarrhea, histopathology: southern Georgia; northern Florida

Coccidia oocysts
Smith, J. P., 1979, Feline Pract., v. 9 (2), 14, 16, 18
intestinal parasites, cats, efficacy of parenteral aqueous levamisole

Coccidi[a] coccidial oocysts
Wescott, R. B.; Shelton, T. A.; and Gates, N. L., 1979, West. Veterinarian, v. 17 (2), 22-23
gastrointestinal nematodes and cestodes, sheep, 3 field trials with oxfendazole, effective; no apparent effect on coccidial oocysts

Coccidi[a] coccidial oocysts
Yazwinski, T. A.; and Brown, A. H., 1979, Vet. Med. and Small Animal Clin., v. 74 (8), 1156-1158, 1160
intestinal parasites, bulls, levels of infection, variation of growth performance factors in relation to levels of eggs per gram of feces, regression analysis: Arkansas

[Coccidia] koktsidii
Zharikov, I. S.; and Antonenko, A. E., 1978, Veterinariia, Moskva (12), 63-64
helminth eggs, coccidia oocysts, disinfection of manure with penetrating electron radiation, dosages necessary

Coccidia [sp.], illus.
Bemrick, W. J.; and O'Leary, T. P., 1979, Vet. Med. and Small Animal Clin., v. 74 (3), 389-390
Coccidia [sp.] in 1-week-old Ursus horribilis (small intestine), severe diarrhea of unknown etiologic origin prior to death: Como Park Zoo, St. Paul, Minn.

Coccidia [sp.]
Carpenter, J. W.; et al., 1979, J. Am. Vet. Med. Ass., v. 175 (9), 948-951
Grus canadensis (intestines): Patuxent Wildlife Research Center

Coccidia [sp.]
Carpenter, J. W.; and Hillman, C. N., 1979, Ann. Proc., Am. Ass. Zoo Vet., 36-47
Mustela nigripes: South Dakota, transported to Patuxent Wildlife Research Center, Laurel, Maryland

Coccidia [sp.], resembling immature Eimeria cani
Eaton, R. D. P.; and Secord, D. C., 1979, Canad. J. Comp. Med., v. 43 (2), 229-230
Arctic fox (ileum): Banks Island, Northwest Territories

Coccidia [sp.]
Frank, C., 1976, Acta Vet. Brno, v. 45 (4), 263-270
Podiceps cristatus
P. ruficollis
(small intestine of all): all from southeastern "Seewinkel", Burgenland

Coccidia [sp.] (presumably Eimeria stieda)
Jacobson, H. A.; Kirkpatrick, R. L.; and McGinnes, B. S., 1978, Wildlife Monogr. (60), 53 pp.
disease and physiologic characteristics of cottontail rabbits in 2 study areas in relation to population density, includes data on seasonal and sex differences
Sylvilagus floridanus: Virginia

Coccidia spp.
Juniper, I., 1978, Canad. Field-Naturalist, v. 92 (2), 186-189
Ursus americanus: north of Montreal, Province of Quebec

Coccidia [sp.]
Thompson, E. J.; and Wright, I. G. A., 1978, N. Zealand Vet. J., v. 26 (6), 167 [Letter]
Apteryx australia mantelli (kidneys, feces): bred in captivity, New Zealand

Coccidiosis
Avakian, A. A.; et al., 1978, Veterinariia, Moskva (11), 76-77
coccidiosis, chickens (broilers), prophylactic control by various preparations, pharmcoccide recommended, treatment economics: Krymsk oblast

Coccidiosis
Bedrnik, P.; et al., 1979, Veterinarstvi, v. 29 (8), 353-355
coccidiosis, broiler chicken fattening, lasalocid compared with amprol plus and monensin: Czechoslovakia

Coccidiosis
Borgsteede, F. H. M.; and Koenig, C. D. W., 1979, Tijdschr. Diergeneesk., v. 104 (21), 825-828
Nematodirus battus definitely established in Dutch sheep after detection in sheep imported from Britain; clinical symptoms difficult to differentiate from coccidiosis: Netherlands

Coccidiosis
Brasitus, T. A., 1979, Am. J. Med., v. 67 (6), 1058-1065
parasitic infections, association with malabsorption in man

Coccidiosis
Brunnthaler, F., 1977, Prakt. Tierarzt, v. 58 (11), 849-851
coccidiosis, dogs, treatment with various drugs, best results with amprolium

Coccidiosis
Fox, J. E., 1978, Mod. Vet. Pract., v. 59 (8), 599-603
bovine coccidiosis, review, emphasis on prevention and control; field tests, decoquinate against Eimeria bovis and E. zuernii prevented clinical signs of disease with no observable signs of toxicity

Coccidiosis
Gobzem, V. R.; and Zibitsker, D. E., 1973, Vet. Nauka--Proizvod., Trudy, Minsk, v. 11, 155-156
coccidiosis, rabbits, polysaccharide T as nonspecific prophylaxis

Coccidiosis
Kasai, K.; Miyamoto, M.; and Sakaguchi, T., 1979, Jikken Dobutsu (Exper. Animals), v. 28 (4), 491-495
method of establishing colony of Japanese white rabbits free from coccidia, host breeding performance

Coccidiosis
Kolabskii, N. A.; and Pashkin, P. I., 1974, [Coccidiosis of farm animals], 160 pp.
coccidiosis, farm animals, handbook

Coccidiosis
Kremer, M.; and Molet, B., 1975, Ann. Soc. Belge Med. Trop., v. 55 (5), 427-430
intestinal parasites, human, diagnosis, Kato technique

Coccidiosis
Kuchin, A. S., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 94-96
parasites of lambs, distribution in the course of the year

Coccidiosis
Long, P. L.; and Millard, B. J., 1978, Avian Path., v. 7 (3), 373-381
coccidiosis, broiler chickens, effect on oocyst output of various treatment regimens

Coccidiosis
Manafova, Sh. G., 1978, Izvest. Akad. Nauk Azerbaidzhan. SSR, s. Biol. Nauk. (4), 75-79
coccidiosis, cattle, zebu, buffalo, age and seasonal dynamics: Azerbaidzhan

Coccidiosis
Manuel, M. F.; and Buduan, R. J., 1972, Philippine J. Vet. Med., v. 11 (2), 73-91
battery-raised broilers, effects of amprol plus or cycostat with or without payzone on weight gain and feed efficiency, no coccidiosis found

Coccidiosis
Martinez, J. A., 1978, Rev. Med. Vet., Buenos Aires, v. 59 (5), 294-295
coccidiosis, avian, control, brief review

Coccidiosis
Mohan, K.; et al., 1978, Indian Vet. J., v. 55 (12), 976-981
coccidiosis and other diseases, White Leghorn chicks, four strains, incidence and chick mortality patterns: India

Coccidiosis
Orekhova, M. M.; Arestov, I. G.; and Orekhov, P. V., 1975, Vet. Nauka--Proizvod., Trudy, Minsk, v. 13, 149-151
coccidiosis, chickens, levels of sulfhydryl groups in blood as measure of functional status of host

Coccidiosis
Paliwal, O. P.; Krishna, L.; and Kulshrestha, S. B., 1978, Indian Vet. Med. J., v. 2 (4), 191-196
lambs and kids, incidence of mortality correlated with age, sex, season, and pathological conditions, including coccidiosis, amphistomiasis, and cysticercosis: organised farm, [India]

Coccidiosis, illus.
Parihar, N. S.; Charan, K.; and Chakravarty, I. B., 1978, Indian Vet. J., v. 55 (11), 920
coccidiosis, histopathology
Gazella subgutturosa (intestine): Delhi Zoological Park, brought from Iran

Coccidiosis
Sangster, L. T.; et al., 1978, Vet. Med. and Small Animal Clin., v. 73 (10), 1317-1319
coccidiosis, baby pigs, cause of scours, clinical and pathologic features, treatment with amprolium: southern Georgia

Coccidiosis
Sherkov, Sh.; et al., 1978, Vet. Sbirka, v. 76 (6), 39-41
coccidiosis, calves, disinfectants against oocysts, sulfaquinoxaline as chemoprophylactic, elancoban-100 as coccidiostat

Coccidiosis
Strakova, J.; Sevcik, B.; and Dvorak, M., 1978, Biol. a Chem. Zivoc. Vyroby, Vet., v. 14 (v. 20) (2), 171-180
coccidiosis, chicks, sulphacombine, acute and subacute toxicity studies

Coccidiosis
Supperer, R., 1973, Tieraerztl. Prax., v. 1 (1), 33-42
parasitic diseases of swine, intensive rearing, treatment and control measures, review

Coccidiosis
Thiery, G., 1978, Ann. Immunol., v. 129C (4), 503-522
rabbit's appendix, immunological model applied to study of epithelial immunity, including that against coccidiosis

Coccidiosis
Thiery, G., 1978, Bull. Acad. Vet. France, n.s., v. 51 (2), 211-216
coccidiosis, rabbit, mechanism of epithelial immunity in caecal appendix

Coccidiosis
Vivo Rodriguez, R., 1977, Noticias Neosan (186), v. 35, 35-38, 41-43
coccidiosis, lambs, excretion and development of oocysts, humidity of bedding, and growth of hosts

Coccidiosis
Voeten, A. C.; et al., 1978, Tijdschr. Diergeneesk., v. 103 (23), 1284-1289
coccidiosis, broilers, anticoccidials, floor pen trials

Coccidiosis
Zaionts, V. I.; et al., 1976, Khimiko-Farm. Zhurnal, v. 10 (11), 20-29
molecular mechanisms of action of some anticoccidial preparations, review

Coccidium bigeminum (Stiles, 1891) Luehe, 1906 pro parte
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
as syn. of Sarcocystis muris (Blanchard, 1885) Labbe, 1899

Coccidium bigeminum Stiles, 1891
Tadros, W.; and Laarman, J. J., 1978, Proc. K. Nederl. Akad. Wetensch., s. C, Biol. and Med. Sc., v. 81 (4), 466-468
as syn. of Sarcocystis miescheriana (Kuehn, 1865) Lankester, 1882

Coccidium bigeminum bigeminum
Tadros, W.; and Laarman, J. J., 1978, Proc. K. Nederl. Akad. Wetensch., s. C, Biol. and Med. Sc., v. 81 (4), 466-468
C. bigeminum var. canis selected as nominate subspecies of C. bigeminum and thus designated C. bigeminum bigeminum
as syn. of Sarcocystis miescheriana (Kuehn, 1865) Lankester, 1882

Cocccidium bigeminum var. canis Railliet and Lucet, 1891
Tadros, W.; and Laarman, J. J., 1978, Proc. K. Nederl. Akad. Wetensch., s. C, Biol. and Med. Sc., v. 81 (4), 466-468
selected as nominate subspecies of C. bigeminum and thus designated C. bigeminum bigeminum
as syn. of Sarcocystis miescheriana (Kuehn, 1865) Lankester, 1882

Coccidium bigeminum var. cati Railliet and Lucet, 1891 pro parte
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
as syn. of Sarcocystis muris (Blanchard, 1885) Labbe, 1899

Coccidium bigeminum var. putorii Railliet and Lucet, 1891
Tadros, W.; and Laarman, J. J., 1978, Proc. K. Nederl. Akad. Wetensch., s. C, Biol. and Med. Sc., v. 81 (4), 466-468
as syn. of Sarcocystis putorii (Railliet and Lucet, 1891) comb. nov.

Coccidium variabile Thelohan, 1893
Davies, A. J., 1978, J. Protozool., v. 25 (1), 15-21
as syn. of Eimeria variabilis (Thelohan, 1893) Reichenow, 1921

Cognettia gen. n.
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
Metameridae fam. n.
tod: C. legeri (Cognetti de Martiis, 1911) comb. n.

Cognettia legeri (Cognetti de Martiis, 1911) comb. n. (tod)
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
Syn.: Taeniocystis legeri Cognetti de Martiis, 1911
Kynotus pitarelli (coelom): Madagascar, Africa

Colligocineta affinis n. sp., illus.
Kozloff, E. N., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 622-627
Sabella pavonina (prostomial cirri): Salstone, near Salcombe, England

Colligocineta finleyi n. sp., illus.
Kozloff, E. N., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 622-627
Sabella pavonina (prostomial cirri): Salstone, near Salcombe, England

Colligocineta scololepidis n. sp., illus.
Kozloff, E. N., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 622-627
Scololepis fuliginosa (anterior tentacles, dorsal branchiae, dorsal lamellae): Point Perharidic (Perharidy), near Roscoff

Colponema spp.
Paperna, I.; and Baudin Laurencin, F., 1979, Aquaculture, v. 16 (2), 173-175
Dicentrarchus labrax
Sparus aurata
(gills of all): all from marine cultures in France

Cometoides pechumani sp. n., illus.
Anderson, J. F.; and Magnarelli, L. A., 1978, J. Invert. Path., v. 31 (3), 324-328
incidence of infection throughout year, distribution within alimentary canal
Chrysops fuliginosus
C. atlanticus
all from mouth of Housatonic River, Milford, Connecticut

Conispora gen. nov.
Sankurathri, C. S., 1977, Canad. J. Zool., v. 55 (7), 1147-1150
Wardiidae
tod: C. renalis sp. nov.

Conispora abyssa (Yoshino and Moser 1974) n. comb.
Sankurathri, C. S., 1977, Canad. J. Zool., v. 55 (7), 1147-1150
Syn.: Myxoproteus abyssus Yoshino and Moser 1974

Conispora renalis sp. nov. (tod), illus.
Sankurathri, C. S., 1977, Canad. J. Zool., v. 55 (7), 1147-1150
Merluccius productus (kidney tubules): Stuart Channel, British Columbia, Canada

Conorhynchus nov. gen.
Greeff, R., 1879, Nova Acta Acad. Nat. Curios., v. 41, pt. 2 (1), 1-172
gregarine
mt: C. gibbosus nov. gen. et nov. spec.

Conorhynchus gibbosus nov. gen. et nov. spec. (mt), illus.
Greeff, R., 1879, Nova Acta Acad. Nat. Curios., v. 41, pt. 2 (1), 1-172
Syn.: Gregarina echiuri
Echiurus pallasii (Darmkanal)

Contospora gen. nov.
Devdhar, M. J.; and Amoji, S. D., 1978, Arch. Protistenk., v. 120 (1-2), 182-189
Actinocephalidae, Acanthosporinae
mt: C. opalnia sp. nov.

Contospora opalnia gen. nov., sp. nov. (mt), illus.
Devdhar, M. J.; and Amoji, S. D., 1978, Arch. Protistenk., v. 120 (1-2), 182-189
Opalnia sp. (mid-gut, caeca): Shomeshwar and Kalaghatagi (Dharwar District), Karnataka State, India

Cornucopioides subgen. n.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 25-27
subgen. of Zelleriella
type sp.: Zelleriella (Cornucopioides) cornucopia Carini, 1933

Cornucopioides
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
subgen. of Zelleriella, key

Cornucopioides Amaro, 1966
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
subgen. of Zelleriella
key to species

[Costia] kostioze
Vlasenko, M. I.; and Meshcheriakova, A. A., 1977, Veterinariia, Moskva (4), 75-78
Chilodonella cyprini, Ichthyophthirius multifiliis, [Costia], fish, formalin solution for control, toxicity tested and safe levels established

Costia necatrix
Astakhova, T. V.; and Stepanova, G. A., 1972, Parazitologiia, Leningrad, v. 6 (4), 364-368
Ctenopharyngodon idella (skin): pond and spawning-nursery fisheries, Volga delta

Costia necatrix
Iashchuk, V. D., 1977, Veterinariia, Moskva (9), 71-73
Costia necatrix, rainbow trout, severe outbreak, economic benefits of treatment with malachite green: Vinnitsk fish farm

Costia necatrix (Henneguy)
Lester, R. J. G., 1974, Syesis, v. 7, 195-200
Gasterosteus aculeatus (gills): near Vancouver, British Columbia

Crithidia
Evans, D. A., 1978, Methods Cultiv. Parasites in Vitro, 55-88
Kinetoplastida, cultivation, review

Crithidia
Gutteridge, W. E.; and Coombs, G. H., 1977, Biochemistry of parasitic protozoa, 172 pp., illus.
biochemistry of parasitic protozoa, textbook: methodology; catabolism and generation of energy; nucleic acid metabolism; protein metabolism; lipid metabolism; biochemical mechanism of drug action; isolation of parasitic protozoa from infected animals; culture of parasitic protozoa

Crithidia sp.
Bos, H. J.; et al., 1978, Acta Leidensia, v. 46, 17-30
cryopreservation of parasitic protozoa

Crithidia sp.
Carvalho, A. L. de M., 1973, Rev. Patol. Trop., v. 2 (2), 223-274
Zelus leucogrammus: Brasil

Crithidia-like
Fish, W. R.; Holz, G. G., jr.; and Beach, D. H., 1978, J. Parasitol., v. 64 (3), 546-547
26 trypanosomatid species, cultivation in new chemically-defined medium RE III

Crithidia sp.
Hedge, E. C.; Moody, A. H.; and Ridley, D. S., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (4), 445 [Letter]
Leishmania donovani, human, screening of sera for kala-azar using Crithidia sp. as antigen in indirect fluorescent antibody test, some positive reactions with L. tropica, cross-reactions at low titer with Chagas' disease and African trypanosomiasis

Crithidia sp.
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Crithidia acanthocephali
Camargo, E. P.; Itow, S.; and Alfieri, S. C., [1979], J. Parasitol., v. 64 (6), 1978, 1120-1121
Trypanosomatidae, 18 spp. of 6 genera, proteolytic activities in cell extracts

Crithidia acanthocephali
Figueiredo, E. N.; et al., 1978, J. Protozool., v. 25 (4), 546-549
Crithidia spp., enzymes of ornithine-arginine metabolism, existence of common enzymatic pattern for species of this genus

Crithidia acanthocephali, illus.
Fouts, D. L.; and Wolstenholme, D. R., 1979, Nucleic Acids Research, v. 6 (12), 3785-3804
Crithidia acanthocephali, evidence for partial RNA transcript of small circular component of kinetoplast DNA

Crithidia acanthocephali
Fouts, D. L.; Wolstenholme, D. R.; and Boyer, H. W., 1978, J. Cell. Biol., v. 79 (2, pt. 1), 329-341
Crithidia acanthocephali, circular kinetoplast DNA molecules, heterogeneity in sensitivity to cleavage by 2 bacterial restriction endonucleases

Crithidia acanthocephali
Galinari, S.; and Camargo, E. P., 1978, Exper. Parasitol., v. 46 (2), 277-282
trypanosomatid protozoa, 16 spp., survey for acetylornithinase and ornithine acetyltransferase, metabolic and nutritional implications

Crithidia acanthocephali
Mendes, N. F.; et al., 1979, Transplant. Proc., v. 11 (2), 1304-1305
cross-reactions between Trypanosomatidae cell extracts and HLA antigens

Crithidia acanthocephali
Yoshida, N.; and Camargo, E. P., 1978, J. Bacteriol., v. 136 (3), 1184-1186
trypanosomatids, excretion of urea or ammonia or both, varies according to genus, may be of taxonomic use

Crithidia arili
Figueiredo, E. N.; et al., 1978, J. Protozool., v. 25 (4), 546-549
Crithidia spp., enzymes of ornithine-arginine metabolism, existence of common enzymatic pattern for species of this genus

Crithidium cunninghami Carter, 1909
Gardener, P. J., 1977, Trop. Dis. Bull., v. 74 (12), 1069-1088
as syn. of Leishmania tropica (Wright, 1903) Luehe, 1906

Crithidia deanai [lapsus p. 246 for C. deanei n. sp.]
Carvalho, A. L. de M., 1973, Rev. Patol. Trop., v. 2 (2), 223-274

Crithidia deanei n. sp., illus.
Carvalho, A. L. de M., 1973, Rev. Patol. Trop., v. 2 (2), 223-274
[lapsus p. 246 as C. deanai]
Zelus leucogrammus (tubo digestivo)
Musca domestica
all from Brasil

Crithidia deanei
Camargo, E. P.; Itow, S.; and Alfieri, S. C., [1979], J. Parasitol., v. 64 (6), 1978, 1120-1121
Trypanosomatidae, 18 spp. of 6 genera, proteolytic activities in cell extracts

Crithidia deanei, aposymbiotic
Camargo, E. P.; Itow, S.; and Alfieri, S. C., [1979], J. Parasitol., v. 64 (6), 1978, 1120-1121
Trypanosomatidae, 18 spp. of 6 genera, proteolytic activities in cell extracts

Crithidia deanei
Ceron, C. R.; et al., 1979, J. Protozool., v. 26 (3), 479-483
Trypanosoma cruzi, Crithidia deanei, aposymbiotic C. deanei, purine metabolism

Crithidia deanei
Figueiredo, E. N.; et al., 1978, J. Protozool., v. 25 (4), 546-549
Crithidia spp., enzymes of ornithine-arginine metabolism, existence of common enzymatic pattern for species of this genus

Crithidia deanei
Fish, W. R.; Holz, G. G., jr.; and Beach, D. H., 1978, J. Parasitol., v. 64 (3), 546-547
26 trypanosomatid species, cultivation in new chemically-defined medium RE III

Crithidia deanei
Galinari, S.; and Camargo, E. P., 1978, Exper. Parasitol., v. 46 (2), 277-282
trypanosomatid protozoa, 16 spp., survey for acetylornithinase and ornithine acetyltransferase, metabolic and nutritional implications

Crithidia deanei
Goncalves de Lima, V. M. Q.; Roitman, I.; and Kilgour, V., 1979, J. Protozool., v. 26 (4), 648-652
trypanosomatids, 7 species distinguished by electrophoretic mobilities of some isoenzymes

Crithidia deanei
Mendes, N. F.; et al., 1979, Transplant. Proc., v. 11 (2), 1304-1305
cross-reactions between Trypanosomatidae cell extracts and HLA antigens

Crithidia deanei
Previato, J. O.; Mendonca-Previato, L.; and Gorin, P. A. J., 1979, Carbohydrate Research, v. 70 (1), 172-174
Crithidia deanei, novel β-D-(1→2)-linked D-mannopyranan

Crithidia deanei, illus.
Souto-Padron, T.; and de Souza, W., 1979, J. Protozool., v. 26 (4), 551-557
trypanosomatids, various developmental stages, basic proteins, localization at fine-structural level with ethanolic phosphotungstic acid technique

Crithidia deanei
Yoshida, N.; and Camargo, E. P., 1978, J. Bacteriol., v. 136 (3), 1184-1186
trypanosomatids, excretion of urea or ammonia or both, varies according to genus, may be of taxonomic use

Crithidia fasciculata
Bowman, I. B. R., 1974, Ciba Found. Symp., n.s. (20), 255-284
trypanosomes, intermediary metabolism, review

Crithidia fasciculata Leger, illus.
Brooks, A. S., 1978, Tr. Am. Micr. Soc., v. 97 (3), 287-296
Crithidia fasciculata, Trypanosoma equiperdum, T. lewisi, ultrastructure of flagellar attachment site, thin section, freeze-etch, and cytochemical techniques

Crithidia fasciculata
Camargo, E. P.; Itow, S.; and Alfieri, S. C., [1979], J. Parasitol., v. 64 (6), 1978, 1120-1121
Trypanosomatidae, 18 spp. of 6 genera, proteolytic activities in cell extracts

Crithidia fasciculata
Cosgrove, W. B.; Skeen, M. J.; and Hajduk, S. L., 1979, J. Protozool., v. 26 (4), 643-648
Crithidia fasciculata in vitro, effect of hydroxyurea on cell numbers, structure, macromolecular synthesis, and degree of synchrony

Crithidia fasciculata
Dewey, V. C.; Kidder, G. W.; and Nolan, L. L., 1978, Biochem. Pharmacol., v. 27 (10), 1479-1485
Crithidia fasciculata, mechanism of inhibition of growth by adenosine and adenosine analogs

Crithidia fasciculata
Englund, P. T., 1978, Cell, v. 14 (1), 157-168
Crithidia fasciculata, replication of kinetoplast DNA networks

Crithidia fasciculata
Englund, P. T., 1979, J. Biol. Chem., v. 254 (11), 4895-4900
Crithidia fasciculata, free minicircles of kinetoplast DNA

Crithidia fasciculata
Figueiredo, E. N.; et al., 1978, J. Protozool., v. 25 (4), 546-549
Crithidia spp., enzymes of ornithine-arginine metabolism, existence of common enzymatic pattern for species of this genus

Crithidia fasciculata
Fish, W. R.; Holz, G. G., jr.; and Beach, D. H., 1978, J. Parasitol., v. 64 (3), 546-547
26 trypanosomatid species, cultivation in new chemically-defined medium RE III

Crithidia fasciculata
Fukushima, T.; et al., 1978, Analyt. Biochem., v. 89 (1), 71-79
differential microdetermination for various forms of biopterin, methods include Crithidia bioassay

Crithidia fasciculata
Galinari, S.; and Camargo, E. P., 1978, Exper. Parasitol., v. 46 (2), 277-282
trypanosomatid protozoa, 16 spp., survey for acetylornithinase and ornithine acetyltransferase, metabolic and nutritional implications

Crithidia fasciculata
Garvin, R. T.; Hill, R. C.; and Weber, M. M., 1978, Arch. Biochem. and Biophys., v. 191 (2), 774-781
Crithidia fasciculata, atypical RNA components of cytoplasmic ribosomes

Crithidia fasciculata
Goncalves de Lima, V. M. Q.; Roitman, I.; and Kilgour, V., 1979, J. Protozool., v. 26 (4), 648-652
trypanosomatids, 7 species distinguished by electrophoretic mobilities of some isoenzymes

Crithidia fasciculata
Gorin, P. A. J.; et al., 1979, J. Protozool., v. 26 (3), 473-478
Crithidia fasciculata, D-mannan and D-arabino-D-galactan, structure, changes in proportion with age of culture

Crithidia fasciculata
Gottlieb, M., 1978, Biochim. et Biophys. Acta, v. 541 (4), 444-458
Crithidia fasciculata, carbohydrates, further characterization of previously identified mannan, isolation and partial characterization of arabinose- and galactose-containing polysaccharide, immunological evidence suggests cell surface nature of arabinogalactan

Crithidia fasciculata
Gray, M. W., 1979, Canad. J. Biochem., v. 57 (6), 914-926
Crithidia fasciculata, ribosomal RNA, physical characteristics and methylated sequences

Crithidia fasciculata
Gutteridge, W. E.; Dave, D.; and Richards, W. H. G., 1979, Biochim. et Biophys. Acta, v. 582 (3), 390-401
Kinetoplastida spp., Plasmodium spp., conversion of dihydroorotate to orotate, mechanism of reaction different in these 2 groups of protozoa, possible target of chemotherapeutic attack

Crithidia fasciculata, illus.
Hajduk, S. L., 1979, J. Cell Sc., v. 35, 185-202
Crithidia fasciculata, Trypanosoma equiperdum, observations on dyskinetoplasty, possible mechanisms of acriflavine action

Crithidia fasciculata
Higa, A. I.; de Cazzulo, B. M. F.; and Cazzulo, J. J., 1979, J. Gen. Microbiol., v. 113 (2), 429-432
Crithidia fasciculata, NAD-specific glutamate dehydrogenase, partial purification and some properties

Crithidia fasciculata
Kidder, G. W.; Dewey, V. C.; and Nolan, L. L., 1978, J. Cell. Physiol., v. 96 (2), 165-170
Crithidia fasciculata, transport and accumulation of purine bases

Crithidia fasciculata
Kidder, G. W.; and Nolan, L. L., 1979, Proc. National Acad. Sc., v. 76 (8), 3670-3672
Crithidia fasciculata, Leishmania spp., adenine aminohydrolase, occurrence and possible significance

Crithidia fasciculata
Kidder, G. W.; Nolan, L. L.; and Dewey, V. C., 1979, J. Parasitol., v. 65 (4), 520-525
Crithidia fasciculata, purine phosphoribosyltransferases, identification, some properties

Crithidia fasciculata
Kutzman, R. S.; and Roberts, J. F., 1978, Comp. Biochem. and Physiol., v. 61C (1), 141-145
Crithidia fasciculata, adaptation to growth in presence of carbonyl cyanide m-chlorophenylhydrazone is apparently a physiological and not a genetic phenomenon, retention of this adaptive ability reported only in free-living protozoa is of interest from evolutionary standpoint and when considering drug resistance to parasitic protozoa

Crithidia fasciculata
Kutzman, R. S.; and Roberts, J. F., 1979, Comp. Biochem. and Physiol., v. 62B (4), 449-453
Crithidia fasciculata, metabolic changes accompanying physiological adaptation to growth in presence of carbonyl cyanide m-chlorophenylhydrazone

Crithidia fasciculata
Lopes, J. N.; et al., 1978, Ann. Trop. Med. and Parasitol., v. 72 (6), 523-531
Trypanosoma cruzi, 1,4-naphthoquinone and 1,2-naphthoquinone derivatives, in vitro and in vivo (mice) evaluation of effects on growth, viability, and infectivity; in vitro studies also on Crithidia fasciculata

Crithidia fasciculata
Mendes, N. F.; et al., 1979, Transplant. Proc., v. 11 (2), 1304-1305
cross-reactions between Trypanosomatidae cell extracts and HLA antigens

Crithidia fasciculata
Midgley, M., 1978, Biochem. J., v. 174 (1), 191-202
Crithidia fasciculata, α-aminoisobutyrate transport: effect of incubation medium composition, kinetic studies, effects of inhibitors, studies on respiration, metabolic effects of inhibitors

Crithidia fasciculata
Morales, N. M.; and Roberts, J. F., 1978, Comp. Biochem. and Physiol., v. 59B (1), 1-4
Crithidia fasciculata, Leishmania tropica, Trypanosoma brucei, comparison of ribosomal RNAs, possible evolutionary significance

Crithidia fasciculata
Morales, N. M.; and Roberts, J. F., 1978, J. Protozool., v. 25 (1), 140-144
Crithidia fasciculata, ribonucleic acids, characterization

Crithidia fasciculata
Nolan, L. L.; and Kidder, G. W., 1979, Biochem. and Biophys. Research Commun., v. 91 (1), 253-262
Crithidia fasciculata, action of caffeine on purine metabolizing enzymes

Crithidia fasciculata
Rembold, H.; and Langenbach, T., 1978, J. Protozool., v. 25 (3, pt. 2), 404-408
Crithidia fasciculata, effect of colchicine on cell membrane and on biopterin transport

Crithidia fasciculata
Shapiro, A.; et al., 1978, J. Protozool., v. 25 (4), 530-535
Crithidia fasciculata, growth in vitro, improvements in defined medium, survey of efficiency of chelators, chelator-metal ion relations, Fe, Cu, Mo requirements for hemin sparing, potential uses of high-yield media

Crithidia fasciculata
Simpson, L., 1979, Proc. National Acad. Sc., v. 76 (4), 1585-1588
hemoflagellate protozoa, method for isolation of maxicircle component of kinetoplast DNA

Crithidia fasciculata
de Souza, M. do C. M., 1974, Rev. Patol. Trop., v. 3 (3), 291-332
Leptomonas pessoai, antigenic relationships with other trypanosomatids, cross-protection of mice against Trypanosoma cruzi

Crithidia fasciculata
de Souza, M. do C. M.; and Barbosa, W., 1972, Rev. Patol. Trop., v. 1 (4), 415-419
antigens of Crithidia fasciculata, Trypanosoma cruzi and Leishmania brasiliensis showed cross-reacting precipitating bands with the antigen of Leptomonas pessoai as demonstrated by the agar gel diffusion technique

Crithidia fasciculata
Trager, W., 1974, Ciba Found. Symp., n.s. (20), 225-254
trypanosomes, leishmanias, nutrition and biosynthetic capabilities, problems of in vitro cultivation and differentiation, review

Crithidia fasciculata
Vakirtzi-Lemonias, C.; and Gregoriadis, G., 1978, Biochem. Soc. Tr., v. 6 (6), 1241-1244
Crithidia fasciculata, uptake of liposome-entrapped agents

Crithidia fasciculata
Yoshida, N.; and Camargo, E. P., 1978, J. Bacteriol., v. 136 (3), 1184-1186
trypanosomatids, excretion of urea or ammonia or both, varies according to genus, may be of taxonomic use

Crithidia fasciculata
Zaitseva, G. N.; Mett, I. L.; and Mett, A. L., 1979, Tsitologiia, v. 21 (3), 310-317
Crithidia oncopelti, C. fasciculata, ribosomal RNA synthesis in kinetoplasts

Crithidia fasciculata var. culicis
Figueiredo, E. N.; et al., 1978, J. Protozool., v. 25 (4), 546-549
Crithidia spp., enzymes of ornithine-arginine metabolism, existence of common enzymatic pattern for species of this genus

Crithidia fasciculata var. noelleri
Figueiredo, E. N.; et al., 1978, J. Protozool., v. 25 (4), 546-549
Crithidia spp., enzymes of ornithine-arginine metabolism, existence of common enzymatic pattern for species of this genus

Crithidia fasciculata var. noguchii
Figueiredo, E. N.; et al., 1978, J. Protozool., v. 25 (4), 546-549
Crithidia spp., enzymes of ornithine-arginine metabolism, existence of common enzymatic pattern for species of this genus

Crithidia harmosa
Daggett, P. M.; Decker, J. E.; and Janovy, J., 1978, Comp. Biochem. and Physiol., v. 59A (4), 363-366
6 insect trypanosomatids, attempted adaptation to mice, alteration of component elements of excreted factors (EF) produced by the 3 adapted species, EF components of Leishmania donovani isolates differ with varying infectivity for hamsters

Crithidia harmosa
Figueiredo, E. N.; et al., 1978, J. Protozool., v. 25 (4), 546-549
Crithidia spp., enzymes of ornithine-arginine metabolism, existence of common enzymatic pattern for species of this genus

Crithidia harmosa
Fish, W. R.; Holz, G. G., jr.; and Beach, D. H., 1978, J. Parasitol., v. 64 (3), 546-547
26 trypanosomatid species, cultivation in new chemically-defined medium RE III

Crithidia hutneri
Figueiredo, E. N.; et al., 1978, J. Protozool., v. 25 (4), 546-549
Crithidia spp., enzymes of ornithine-arginine metabolism, existence of common enzymatic pattern for species of this genus

Crithidia luciliae
Arai, H.; et al., 1978, Rinsho Meneki (Clin. Immunol.), v. 10 (3), 404-413
Crithidia luciliae kinetoplast used as substrate (Crithidia method) for immunofluorescence test for antibody to native DNA (ds-DNA), method compared with radioimmunoassay by testing sera from persons with systemic sclerosis, rheumatoid arthritis, and other conditions

Crithidia luciliae
Ballou, S. P.; and Kushner, I., 1979, Arthritis and Rheum., v. 22 (4), 321-327
Crithidia luciliae method for detecting anti-native DNA, diagnosis and clinical management of systemic lupus erythematosus

Crithidia luciliae (Strickland, 1911), illus.
Carvalho, A. L. de M., 1973, Rev. Patol. Trop., v. 2 (2), 223-274
Zelus leucogrammus (tubo digestivo)
Musca domestica
all from Brasil

Crithidia luciliae
Daggett, P. M.; Decker, J. E.; and Janovy, J., 1978, Comp. Biochem. and Physiol., v. 59A (4), 363-366
6 insect trypanosomatids, attempted adaptation to mice, alteration of component elements of excreted factors (EF) produced by the 3 adapted species, EF components of Leishmania donovani isolates differ with varying infectivity for hamsters

Crithidia luciliae
Figueiredo, E. N.; et al., 1978, J. Protozool., v. 25 (4), 546-549
Crithidia spp., enzymes of ornithine-arginine metabolism, existence of common enzymatic pattern for species of this genus

Crithidia luciliae
Hoeijmakers, J. H. J.; and Borst, P., 1978, Biochim. et Biophys. Acta, v. 521 (1), 407-411
Crithidia luciliae, RNA contains transcripts of maxi-circle and not mini-circle component of kinetoplast DNA

Crithidia luciliae
Huber, O.; Greenberg, M. L.; and Huber, J., 1979, J. Lab. and Clin. Med., v. 93 (1), 32-39
Crithidia luciliae, use in detection of anti-dsDNA antibodies in patients with systemic lupus erythematosus

Crithidium luciliae
Jarzabek-Chorzelska, M.; et al., 1978, Arch. Immunol. et Therap. Exper., v. 26 (1-6), 791-794
Crithidium luciliae, use as antigen substrate in immunofluorescence test for detecting anti-native DNA antibodies

Crithidia luciliae
Jonsson, J.; and Norberg, R., 1976, Scand. J. Rheumatol., v. 5 (4), 221-226
Crithidia luciliae, effect of enzyme treatment on reactivity of kinetoplast with anti-DNA sera, concluded that results of Crithidia test for demonstration of anti-DNA antibodies are not completely unambiguous

Crithidia luciliae, illus.
Lastovica, A. J., 1978, Proc. Electron Microsc. Soc. South. Africa, v. 8, 71-72
Crithidia luciliae, ultrastructure

Crithidia luciliae
Monier, J. C.; et al., 1978, Ann. Immunol., v. 129C (4), 463-474
Trypanosoma gambiense, Crithidia luciliae, use as antigen for detection of antibodies to double-stranded DNA in immunofluorescence tests

Crithidia luciliae, illus.
Scheinberg, M. A.; Santos, L.; and Diniz, R., 1979, AMB, Rev. Ass. Med. Brasil., v. 25 (6), 199-200
Crithidia luciliae, detection of antibodies against native DNA, comparison of radioimmunoassay and direct immunofluorescence, rat livers

Crithidia luciliae, illus.
Sontheimer, R. D.; and Gilliam, J. N., 1978, J. Lab. and Clin. Med., v. 91 (4), 550-558
immunofluorescence assay for double-stranded DNA antibodies using Crithidia luciliae kinetoplast as double-stranded DNA substrate

Crithidia luciliae, illus.
Sontheimer, R. D.; and Gilliam, J. N., 1979, J. Invest. Dermat., v. 72 (1), 29-32
Crithidia luciliae, use in immunofluorescence anti-DNA assay in patients with systemic lupus erythematosus

Crithidia luciliae
Stefanovic, Z.; Arneric, S.; and Mijailovic, B., 1979, Period. Biol., v. 81 (2), 499-500
Crithidia luciliae used as antigen substrate in immunofluorescence test for detection of antibodies against dsDNA

Crithidia luciliae
Whiteside, T. L.; and Dixon, J. A., 1979, Am. J. Clin. Path., v. 72 (5), 829-835
Crithidia luciliae test for antibodies to native DNA in patients with systemic lupus erythematosus

Crithidia mellificae
Fish, W. R.; Holz, G. G., jr.; and Beach, D. H., 1978, J. Parasitol., v. 64 (3), 546-547
26 trypanosomatid species, cultivation in new chemically-defined medium RE III

Crithidia oncopelti
Camargo, E. P.; Itow, S.; and Alfieri, S. C., [1979], J. Parasitol., v. 64 (6), 1978, 1120-1121
Trypanosomatidae, 18 spp. of 6 genera, proteolytic activities in cell extracts

Crithidia oncopelti
Chang, K. P.; et al., 1978, J. Protozool., v. 25 (1), 145-149
methylglyoxal bis(guanylhydrazone) (MGBG), little in vitro effect on Blastocrithidia culicis, Crithidia oncopelti, and Leishmania spp. but complete inhibition of growth of Trypanosoma brucei, reduced parasitemia of T. brucei and T. congolense in rats but infections relapsed, tracer studies with T. brucei showed that MGBG interfered with nucleoside incorporation

Crithidia oncopelti
Figueiredo, E. N.; et al., 1978, J. Protozool., v. 25 (4), 546-549
Crithidia spp., enzymes of ornithine-arginine metabolism, existence of common enzymatic pattern for species of this genus

Crithidia oncopelti
Fish, W. R.; Holz, G. G., jr.; and Beach, D. H., 1978, J. Parasitol., v. 64 (3), 546-547
26 trypanosomatid species, cultivation in new chemically-defined medium RE III

Crithidia oncopelti
Galinari, S.; and Camargo, E. P., 1978, Exper. Parasitol., v. 46 (2), 277-282
trypanosomatid protozoa, 16 spp., survey for acetylornithinase and ornithine acetyltransferase, metabolic and nutritional implications

Crithidia oncopelti
Holwill, M. E. J.; and Wais, J., 1979, J. Exper. Biol., London, v. 82, 177-195
Crithidia oncopelti, motile response of flagellum to changes in temperature, pressure, and viscosity of environment, results provide information about mechanochemical cycle which bends flagellum

Crithidia oncopelti, illus.
Johnston, D. N.; Silvester, N. R.; and Holwill, M. E. J., 1979, J. Exper. Biol., London, v. 80, 299-315
Crithidia oncopelti, analysis of shape and propagation of waves on flagellum

Crithidia oncopelti
Keller, R. M.; Picot, D.; and Wuethrich, K., 1979, Biochim. et Biophys. Acta, v. 580 (2), 259-265
Crithidia oncopelti, individual assignments of heme resonances in 360 MHz ^{1}H NMR spectra of cytochrome c-557

Crithidia oncopelti
McCormack, J. J.; et al., 1979, Biochem. Pharmacol., v. 28 (21), 3227-3229
inhibition of dihydrofolate reductases by derivatives of 2,4-diaminopyrroloquinazoline, Crithidia oncopelti used as one source of reductases

Crithidia oncopelti, illus.
Shirshov, A. T.; et al., 1975, Izvest. Akad. Nauk SSSR, s. Biol. (5), 759-765
Crithidia oncopelti, acriflavine, effect on structure of kinetoplast, kinetoplast DNA, protein synthesis in kinetoplast and cytoplasmic ribosomes; suggests that information required for synthesis of kinetoplast ribosomes is contained in kinetoplasts

Crithidia oncopelti, illus.
Sukhareva-Nemakova, N. N.; Titova, T. S.; and Avakian, A. A., 1975, Izvest. Akad. Nauk SSSR, s. Biol. (2), 275-280
Crithidia oncopelti, comparative study of ultrastructure, cultures differing in sensitivity to olivomycin; lipid drops in cytoplasm of resistant protozoa; nature of action of olivomycin on sensitive parasites

Crithidia oncopelti
Zaitseva, G. N.; Aseev, V. V.; and Kunavina, I. Iu., 1978, Biokhimiia, v. 43 (2), 283-289
Crithidia oncopelti nucleoli, DNA-dependent RNA polymerase activity

Crithidia oncopelti
Zaitseva, G. N.; Mett, I. L.; and Mett, A. L., 1979, Tsitologiia, v. 21 (3), 310-317
Crithidia oncopelti, C. fasciculata, ribosomal RNA synthesis in kinetoplasts

Cryptobia sp., illus.
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
Odontogadus merlangus euxinus (blood): Black Sea

Cryptobia agitata Chen
Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
parasites of fishes: China

Cryptobia branchialis Nie
Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
parasites of fishes: China

Cryptobia bullocki
Daily, D. D., 1978, J. Parasitol., v. 64 (2), 361-362
Hippoglossoides platessoides: Whiting Bay, Maine

Cryptobia bullocki Strout, 1965
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102
brief description
Liopsetta putnami
Pseudopleuronectes americanus
(blood of all): all from St. Andrews, New Brunswick

Cryptobia cataractae
Ollenschlaeger, B., 1975, Tieraerztl. Prax., v. 3 (1), 99-107
blood and other parasites of commercial fish, pathology, transmission, therapy, clinical review

Cryptobia catostomi n. sp., illus.
Bower, S. M.; and Woo, P. T. K., 1977, Canad. J. Zool., v. 55 (7), 1082-1092
Cryptobia catostomi n. sp., morphological variation, host specificity
Catostomus commersoni (blood) (nat. and exper.): southern Lake Huron within a 15-mile radius of Sarnia, Ontario, Canada
C. catostomus: Nottawasaga River near Alliston, Ontario

Cryptobia catostomi, illus.
Bower, S. M.; and Woo, P. T. K., 1977, Canad. J. Zool., v. 55 (7), 1093-1099
Cryptobia catostomi in Catostomus commersoni (blood), division and morphogenesis, explanation for variation in parasite size

Cryptobia catostomi
Muzzall, P. M., 1979, Tr. Am. Micr. Soc., v. 98 (3), 472-473
Catostomus commersoni (blood): Oyster River, Strafford County, New Hampshire

Cryptobia lomakini sp. n., illus.
Khaibulaev, K. Kh., 1971, Parazitologiia, Leningrad, v. 5 (6), 551-555
Gobius melanostomus affinis
G. kessleri gorlap
G. fluviatilis pallasi
Benthophillus macrocephalus
(blood of all): all from Caspian Sea

Cryptobia salmositica
Lowe-Jinde, L., 1979, J. Fish Biol., v. 14 (3), 297-302
Cryptobia salmositica in Salmo gairdneri (exper.), plasma glucose and proteins and haematocrit levels during course of infection, changes produced in host metabolism will undoubtedly affect host growth and population size

Cryptobia salmositica
Ollenschlaeger, B., 1975, Tieraerztl. Prax., v. 3 (1), 99-107
blood and other parasites of commercial fish, pathology, transmission, therapy, clinical review

Cryptobia salmositica, illus.
Woo, P. T. K., 1978, Canad. J. Zool., v. 56 (7), 1514-1518
Cryptobia salmositica in Salmo gairdneri (exper.), division process
Piscicola salmositica: Vancouver Island
Oncorhynchus kisutch (exper.)

Cryptocaryon gen. n.
Brown, E. M., 1951, Agenda and Abstr. Scient. Meet. Zool. Soc. London (11), 1-2
mt: C. irritans gen. et sp. n.

Cryptocaryon irritans gen. et sp. n. (mt)
Brown, E. M., 1951, Agenda and Abstr. Scient. Meet. Zool. Soc. London (11), 1-2
Canthigaster rostratus
Scarus cretensis
Diplodus vulgaris

Cryptocaryon irritans Brown, 1951
Cheung, P. J.; Nigrelli, R. F.; and Ruggieri, G. D., 1979, J. Fish Dis., v. 2 (2), 93-97
Cryptocaryon irritans, effect of temperature and salinity on reproductive cycle

Cryptochilum Maupas, 1883
Iankovskii, A. V., 1973, Parazitologiia, Leningrad, v. 7 (3), 214-219
Hymenostomatida, Tetrahymenina, Entodiscidae fam. n.

Cryptochilum cuenoti Florentin, 1898
Iankovskii, A. V., 1973, Parazitologiia, Leningrad, v. 7 (3), 214-219
as syn. of Symbionecta cuenoti (Florentin, 1898) comb. n.

Cryptosporidia
Brasitus, T. A., 1979, Am. J. Med., v. 67 (6), 1058-1065
parasitic infections, association with malabsorption in man

Cryptosporidium, illus.
Moon, H. W.; et al., 1978, J. Am. Vet. Med. Ass., v. 173 (5, pt. 2), 577-583
cryptosporidia and Eimeria found among other agents in newborn calves that had naturally occurring diarrheal disease: Iowa

Cryptosporidium
Pohlenz, J.; Moon, H. W.; and Bemrick, B., 1977, Proc. 14. Ann. Electron Micr. Colloq., 14-15
Cryptosporidium, bovine, ultrastructure of life cycle stages

Cryptosporidium sp., illus.
Berg, I. E.; Peterson, A. C.; and Freeman, T. P., 1978, J. Am. Vet. Med. Ass., v. 173 (12), 1586-1587
lambs (small intestine)

Cryptosporidium [sp.], illus.
Doster, A. R.; Mahaffey, E. A.; and McClearen, J. R., 1979, Avian Dis., v. 23 (3), 654-661
morphologic features, host tissue changes
Amazona autumnalis (cloacal coprodeum)

Cryptosporidium sp., illus.
Hoerr, F. J.; Ranck, F. M., jr.; and Hastings, T. F., 1978, J. Am. Vet. Med. Ass., v. 173 (12), 1591-1593
turkeys (respiratory tract): Indiana

Cryptosporidium sp.
McKenzie, R. A.; et al., 1978, Austral. Vet. J., v. 54 (7), 365-366 [Letter]
Pseudechis porphyriacus (stomach mucosa): zoological park near Brisbane, Australia

Cryptosporidium [sp.], illus.
Pearson, G. R.; and Logan, E. F., 1978, Vet. Rec., v. 103 (10), 212-213
Cryptosporidium [sp.], calf (brush border of villous epithelial cells in distal small intestine), light, transmission and scanning electron microscopy: Great Britain

Cryptosporidium sp., illus.
Pohlenz, J.; et al., 1978, Vet. Path., v. 15 (3), 417-427
Cryptosporidium sp., calves (free in lumen and attached to epithelium of ileum), life cycle, morphology, pathology, diarrhea, transmission and scanning electron microscopy

Cryptosporidium [sp.], illus.
Snyder, S. P.; England, J. J.; and McChesney, A. E., 1978, Vet. Path., v. 15 (1), 12-17
Cryptosporidium [sp.] in Arabian foals with inherited combined immunodeficiency, mixed infection with adenovirus, difficult to separate effects of both disease agents
Arabian foals (intestine, stomach, pancreatic ducts, gall bladder): Colorado State University

Cryptosporidium [sp.], illus.
Weisburger, W. R.; et al., 1979, Am. J. Clin. Path., v. 72 (3), 473-478
Cryptosporidium [sp.], human, case report, immunosuppressed renal-transplant recipient with IgA deficiency: Johns Hopkins Hospital

Cryptosporidium cuniculus [n. sp.?], illus.
Inman, L. R.; and Takeuchi, A., 1979, Vet. Path., v. 16 (1), 89-95
Cryptosporidium cuniculus [n. sp.?] in asymptomatic Oryctolagus cuniculus (ileum), electron microscopy (transmission and scanning), altered intestinal microvilli in attachment of parasite

Cryptosporidium cuniculus, illus.
Rehg, J. E.; Lawton, G. W.; and Pakes, S. P., 1979, Lab. Animal Sc., v. 29 (5), 656-660
Oryctolagus cuniculus (jejunum, ileum): rabbitry, California

Cryptosporina gen. n.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Thelohaniidae fam. n., key
tod: C. brachyfila sp. n.

Cryptosporina brachyfila sp. n., (tod), illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Piona sp. (adipose tissue): West swamp, Payne's Prairie, State Road 121, near Gainesville, Florida, U.S.A.

Curviperistomatus subgen. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120
subgen. of Nyctotherus
tod of subgen.: N. (C.) ochoterenai Schouten, 1937

Cygnicollum Bogolepova, 1953
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702
as syn. of Lecudina Mingazzini, 1891

Cygnicollum attenuatum Bogolepova, 1953
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702
as syn. of Lecudina attenuata (Bogolepova, 1953) Theodorides, 1969

Cystocephalus devdharii n. sp., illus.
Patil, C. C.; and Amoji, S. D., 1979, Arch. Protistenk., v. 122 (3-4), 333-339
Rhytinota impolita (intestine): near Gulbarga, Karnataka, India

Cystoisospora
Rommel, M., 1978, Ztschr. Parasitenk., v. 57 (3), 269-283
cyst-forming coccidia, life cycle, taxonomy, comparative review

Cystoisospora canis
Boch, J.; Boehm, A.; and Weiland, G., 1979, Berl. u. Munchen. Tierarztl. Wchnschr., v. 92 (12), 240-243
coccidia, dogs (feces), coprological and serological survey: South Germany

Cystoisospora felis
Fayer, R.; and Frenkel, J. K., 1979, J. Parasitol., v. 65 (5), 756-762
6 spp. of feline coccidia, oocysts fed to calves, calf tissue then fed to cats: neither Besnoitia, Hammondia, nor Sarcocystis were pathogenic for calves nor did they establish patent infections which could be transmitted back to cats; Cystoisospora spp. were not pathogenic for calves but could be transmitted back to cats; Toxoplasma strains were slightly to moderately pathogenic for calves and could be transmitted back to cats

Cystoisospora ohioensis
Boch, J.; Boehm, A.; and Weiland, G., 1979, Berl. u. Munchen. Tierarztl. Wchnschr., v. 92 (12), 240-243
coccidia, dogs (feces), coprological and serological survey: South Germany

Cystoisospora rivolta
Fayer, R.; and Frenkel, J. K., 1979, J. Parasitol., v. 65 (5), 756-762
6 spp. of feline coccidia, oocysts fed to calves, calf tissue then fed to cats: neither Besnoitia, Hammondia, nor Sarcocystis were pathogenic for calves nor did they establish patent infections which could be transmitted back to cats; Cystoisospora spp. were not pathogenic for calves but could be transmitted back to cats; Toxoplasma strains were slightly to moderately pathogenic for calves and could be transmitted back to cats

Cystoisosporinae new subfamily
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
Coccidia

Cytamoeba bacterifera, illus.
da Costa, S. C. G.; da Silva, A. M.; and Pereira, N. M., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 157-159
Cytamoeba bacterifera, taxonomy, geographic distribution, cytology, life cycle, review Leptodactylus ocellatus: Estado do Rio de Janeiro; Estado da Guanabara, Brasil

Cytamoeba bacterifera Labbe, 1894
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
Rana esculenta (krev): CSSR

Cytauxzoon sp., illus.
Ferris, D. H., 1979, Comp. Immunol., Microbiol. and Infect. Dis., v. 1 (4), 269-276
Cytauxzoon sp., cats (blood, lungs, spleen, liver, lymph nodes) (nat. and exper.), disease syndrome, differential diagnosis, prognosis, epizootiology, review: Missouri; Arkansas; Texas; Georgia

Cytauxzoon sp., illus.
Kier, A. B.; et al., 1977, Proc. 20. Ann. Meet., Am. Ass. Vet. Lab. Diagn. (Minneapolis, Minnesota, Oct. 16-18), 123-130
Cytauxzoon sp., domestic cats, diagnosis, gross and microscopic lesions

Cytauxzoon-like parasite
Shindel, N.; Dardiri, A. H.; and Ferris, D. H., 1978, Canad. J. Comp. Med., v. 42 (4), 460-465
Cytauxzoon-like parasite, cats (exper.), detection in frozen spleen sections by indirect fluorescent antibody test

Cytauxzoon [sp.]
Wagner, J. E.; Kier, A. B.; and Morehouse, L. G., 1976, Missouri Vet., v. 26 (2), 12-13
Cytauxzoon [sp.], domestic cats (blood), case reports: southwestern Missouri

Cytauxzoon felis, illus.
Kier, A. B.; et al., 1977, Proc. 20. Ann. Meet., Am. Ass. Vet. Lab. Diagn. (Minneapolis, Minnesota, Oct. 16-18), 123-130

Dactylosoma
Krylov, M. V.; and Krylova, N. P., 1972, Parazitologiia, Leningrad, v. 6 (6), 493-505
Piroplasmida, analysis of host specificity

Dactylosoma ranarum (Lankester, 1882) Labbe 1894, illus.
da Costa, S. C. G.; et al., 1970, Atas Soc. Biol. Rio de Janeiro, v. 12, Suppl., 7-13
Dactylosoma ranarum, life cycle forms related to Toxoplasma
Leptodactylus ocellatus: Rio Bonito, Alcantara (Estado do Rio de Janeiro), Brazil

Dactylosoma ranarum (Krusse, 1890) Labbe, 1894, illus.
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
synonymy, description
Rana esculenta (krev): CSSR

Davisia donecae sp. n., illus.
Gaevskaia, A. V.; and Kovaleva, A. A., 1979, Biol. Moria, Vladivostok (3), 80-83
Trachurus trachurus capensis (urinary bladder): Namibia region, southeastern Atlantic

Davisia ophidioni sp. nov., illus.
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
Ophidion rochei (urinary bladder): Black Sea (region near Sevastopol)

Degiustia gen. n.
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
Gregarinidae
tod: D. hyalella (Batten & DeGiusti, 1949) comb. nov.

Degiustia hyalella (Batten & DeGiusti, 1949) comb. nov. (tod)
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
Syn.: Gregarina hyalella Batten & DeGiusti, 1949
Hyalella azteca (intestine): Michigan, North America

Dehornia Porchet-Hennere, 1972
Levine, N. D., 1979, J. Protozool., v. 26 (2), 167-168
as syn. of Rhytidocystis Henneguy, 1907

Dehornia sthenelais Porchet-Hennere, 1972
Levine, N. D., 1979, J. Protozool., v. 26 (2), 167-168
as syn. of Rhytidocystis sthenelais (Porchet-Hennere, 1972) comb. n.

Dermocystidium, illus.
Koerting, W., 1977, Fisch u. Umwelt (4), 37-48
fish parasites, histopathological changes

Dermocystidium marinum Mackin, Owen, and Collier, 1950
Levine, N. D., 1978, J. Parasitol., v. 64 (3), 549
as syn. of Perkinsus marinus (Mackin, Owen, and Collier, 1950) comb. n.

Dermocystidium percae
Iskov, M. P., 1979, Gidrobiol. Zhurnal, v. 15 (4), 68-72
parasites causing intensive epizootic disease of food fishes: Kremenchug reservoir

Dermocystidium percae Reichenbach-Klinke
Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
parasites of fishes: China

Dermocystidium ranae Guyenot et Naville, 1922, illus.
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
description
Rana temporaria (kuze): CSSR

Dermocystidium salmonis
Makhovenko, E. T., 1972, Parazitologiia, Leningrad, v. 6 (4), 369-375
Salvelinus alpinus inhabiting different ecological niches, possible use of differences in parasite fauna between groups as biological tags: Lake Azabach'e, Kamchatka

Dermosporidium granulosum Broz et Privora, 1952, illus.
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
description
Rana temporaria (kuze): CSSR

Dermosporidium multigranulare Broz, Kulda, 1954, illus.
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
description
Rana esculenta (kuze): CSSR

Deuteromera Bhatia & Setna, 1938
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
Metameridae fam. n.
diagnosis

Dicauda gen. nov.
Hoffman, G. L.; and Walker, R., 1978, J. Fish Dis., v. 1 (4), 343-349
Myxobolidae
tod: D. atherinoidi sp. nov.

Dicauda atherinoidi sp. nov. (tod), illus.
Hoffman, G. L.; and Walker, R., 1978, J. Fish Dis., v. 1 (4), 343-349
Dicauda atherinoidi sp. nov., pathology
Notropis atherinoides (subdermally, fins, lining of mouth): Lake Erie (Sterling State Park), Michigan; lower Mohawk and adjacent Hudson River, New York

Dicercomonas intestinalis Grassi (1882)
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
as syn. of Hexamita intestinalis Dujardin, 1841

Didymophyes gigantea, illus.
Hildebrand, H. F., 1978, Ztschr. Parasitenk., v. 55 (1), 9-27
Didymophyes gigantea, electron microscopy of developmental stages of trophozoite, fine structure of deutomerite, nuclear division, Golgi apparatus
Oryctes nasicornis (Darm): Prafrance (Cevennen) and 'La Forestiere Lesquinoise', Lesquin (Departement Nord), France

Dientamoeba fragilis
Akhtaruzzaman, K. M.; et al., 1978, Tropenmed. u. Parasitol., v. 29 (4), 427-431
comparison of different methods for detection of intestinal protozoa and helminths in human stool

Dientamoeba fragilis
Blecka, L. J., 1978, J. Parasitol., v. 64 (2), 362-363
patients infected with intestinal parasites, attempt to quantitate immunoglobulin levels in fecal extracts with radial immunodiffusion, mean IgA levels higher than in controls, other immunoglobulin classes rarely detectable

Dientamoeba fragilis
de Carneri, I.; and Grassi, L., 1974, Parassitologia, v. 16 (1), 79-81
Dientamoeba fragilis, Entamoeba histolytica, E. coli, prevalence in 500 individuals, taxonomic implications of differential drug sensitivity

Dientamoeba fragilis
Felman, Y. M.; and Ricciardi, N. B., 1979, Bull. N. York Acad. Med., v. 55 (6), 533-539
enteric human pathogens, sexual transmission, higher prevalence among homosexual males, public health aspects

Dientamoeba fragilis
Rostkowska, J., 1970, Acta Parasitol. Polon., v. 18 (27-41), 377-392
Balantidium coli, effect of various bacteria on propagation in vitro, on erythrophagocytic capability of balantidia, and on susceptibility of balantidia to atebrin, entobex, mexaform, and protargol; Trichomonas hominis, Chilomastix mesnili, and Dientamoeba fragilis found to be without effect; effect of balantidia on bacteria

Dientamoeba fragilis
Sargeaunt, P. G.; and Williams, J. E., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (2), 225-227
pathogenic and non-pathogenic intestinal amoeba of man, all species can easily be distinguished by characteristic electrophoretic isoenzyme patterns

Dientamoeba fragilis, illus.
Spencer, M. J.; Garcia, L. S.; and Chapin, M. R., 1979, Am. J. Dis. Child., v. 133 (4), 390-393
Dientamoeba fragilis, children with acute and chronic gastrointestinal symptoms, clinical findings, symptomatic recovery after treatment with diiodohydroxyquin or metronidazole indicates pathogenic role

Dientamoeba fragilis
William, D. C.; et al., 1978, Sex. Transmit. Dis., v. 5 (4), 155-157
enteric protozoa, Enterobius vermicularis, high rate of infection in homosexual men who practice anilingus: New York City

Dinema Perty, Huber, Pestalozzi
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
Peranemidae, key
key to species

Dinema sp.
Michajlow, W., 1978, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 26 (4), 269-272
Microcyclops varicans: water body of Botanical Garden, Brisbane, Australia

Dinema aegypticum Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Dinema agile Michajlow, 1965
Michajlow, W., 1970, Acta Parasitol. Polon., v. 18 (42-50), 473-482
Acanthocyclops vernalis

Dinema agile Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Dinema amoeboidale Michajlow, 1970
Michajlow, W., 1970, Acta Parasitol. Polon., v. 18 (42-50), 473-482
Acanthocyclops vernalis

Dinema amoeboidale Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Dinema antarcticum sp. n., illus.
Michajlow, W., 1978, Bull. Acad. Polon. Sc., Cl. V, s. Sc. Biol., v. 26 (1), 51-54
Pseudoboeckella silvestri (eggs): reservoir on King George Island, Southern Shetland Islands, Antarctica

Dinema australiense Mich.
Michajlow, W., 1978, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 26 (4), 269-272
Mesocyclops leuckarti: water body of Botanical Garden, Australia

Dinema bangalorense sp. n., illus.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (7), 475-478
Mesocyclops leuckarti
M. rylovi
Eucyclops serrulatus
all from Botanical Gardens Lal-Bagh, Bangalore, India (Karnataka)

Dinema cyclopis Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Dinema dispicatum Michajlow, 1970
Michajlow, W., 1970, Acta Parasitol. Polon., v. 18 (42-50), 473-482
Eucyclops serrulatus

Dinema dispicatum Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Dinema eudiaptomi Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Dinema hanoiense Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Dinema harpacticolens Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Dinema helveticum Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Dinema italicum Mich.
  Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
    key

Dinema mazuriense Mich.
  Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
    key

Dinema mazuriense, illus.
  Michajlow, W., 1977, Zhurnal Obsh. Biol., v. 38 (5), 657-675
    Acanthocyclops vernalis

Dinema metabolicum Michajlow, 1970
  Michajlow, W., 1970, Acta Parasitol. Polon., v. 18 (42-50), 473-482
    Acanthocyclops vernalis
    Mesocyclops leuckarti
    Eucyclops serrulatus

Dinema metabolicum Mich.
  Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
    key

Dinema minimum Mich.
  Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
    key

Dinema mongolicum Mich.
  Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
    key

Dinema naupliorum Mich.
  Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
    key

Dinema ocelli Mich.
  Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
    key

Dinema ocelli, illus.
  Michajlow, W., 1977, Zhurnal Obsh. Biol., v. 38 (5), 657-675

Dinema parvum Mich.
  Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
    key

Dinema penetrans Mich.
  Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
    key

Dinema pseudoboeckellae sp. n., illus.
  Michajlow, W., 1978, Bull. Acad. Polon. Sc., Cl. V, s. Sc. Biol., v. 26 (1), 51-54
    Pseudoboeckella silvestri (eggs): reservoir on King George Island, Southern Shetland Islands, Antarctica

Dinema rostratum Mich.
  Michajlow, W., 1978, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 26 (4), 269-272
    Mesocyclops leuckarti: water body of Botanical Garden, Australia

Dinema rotans Mich.
  Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
    key

Dinema rotundum Mich.
  Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
    key

Dinema rotundum Mich.
  Michajlow, W., 1978, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 26 (4), 269-272
    Mesocyclops leuckarti: water body of Botanical Garden, Melbourne, Australia

Dinema semiparasiticum Michajlow, 1970
  Michajlow, W., 1970, Acta Parasitol. Polon., v. 18 (42-50), 473-482
    Acanthocyclops vernalis

Dinema semiparasiticum Mich.
  Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
    key

Dinema semiparasiticum Mich, illus.
  Michajlow, W., 1977, Zhurnal Obsh. Biol., v. 38 (5), 657-675
    Acanthocyclops vernalis

Dinema symmetricum Mich.
  Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
    key

Dinema trakayense Michajlow, 1970
  Michajlow, W., 1970, Acta Parasitol. Polon., v. 18 (42-50), 473-482
    Macrocyclops albidus

Dinema trakayense Mich.
  Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
    key

Dinema undulaflagellatum Monch.
  Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
    key

Dinema unispicatum Michajlow, 1970
  Michajlow, W., 1970, Acta Parasitol. Polon., v. 18 (42-50), 473-482
    Eucyclops serrulatus

Dinema unispicatum Mich.
  Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
    key

Dinema vaksalense Mich.
  Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
    key

Dinema velox Mich.
  Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
    key

Dinema vietnamiense Mich.
  Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
    key

Dinema violentum Michajlow, 1970
  Michajlow, W., 1970, Acta Parasitol. Polon., v. 18 (42-50), 473-482
    Acanthocyclops vernalis

Dinema violentum Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Dinemina Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
Peranemidae, key
key to species

Dinemina ocelli Michajlow, 1970
Michajlow, W., 1970, Acta Parasitol. Polon., v. 18 (42-50), 473-482
Mesocyclops leuckarti
Acanthocyclops vernalis

Dinemina ocelli Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Dinemula Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
Peranemidae, key
key to species, includes: Dinemula celer Mich.

Dinemula celer, illus.
Michajlow, W., 1977, Zhurnal Obsh. Biol., v. 38 (5), 657-675

Dipartiella sp.?, illus.
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
Crenilabrus tinca (gills): Black Sea

Diplauxis [sp.]
Theodorides, J., 1978, Ann. Parasitol., v. 53 (4), 437-438
Laeonereis glauca (coelomique): Villefranche-sur-Mer (Alpes-Maritimes)

Diplauxis schreveli
Theodorides, J., 1978, Ann. Parasitol., v. 53 (4), 437-438
syzygy from Platynereis dumerilii at Villefranche-sur-Mer identified by Theodorides, 1969 a, as Lecudina platynereidis actually belongs to D. schreveli

Diplocystis tipulae sp. nov., illus.
Sherlock, P. L., 1979, Parasitology, v. 78 (2), 207-220
Diplocystis tipulae sp. nov. from Tipula spp., morphology, life cycle, relationship with intestinal gregarines, synchrony between host and parasite developmental rates, pathology, host reactions
Tipula paludosa (nat. and exper.): Northumberland and Cumbria
T. oleracea (exper.)
(larval midgut surface and throughout haemocoele of pupae and adults of all)

Doliocystis Leger, 1893
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702
as syn. of Lecudina Mingazzini, 1891

Doliocystis criodrilii Sciacchitano, 1931
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702
as syn. of Lecudina criodrilii (Sciacchitano, 1931) n. comb.

Drepanidium ranarum Krusse, 1890
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
as syn. of Dactylosoma ranarum (Krusse, 1890) Labbe, 1894

Duttonella, subgenus
Godfrey, D. G., 1977, Protozoology, v. 3, 33-49
identification of morphologically similar trypanosomes of mammals

East Coast fever
Shannon, D., 1977, Trop. Animal Health and Prod., v. 9 (1), 29-35
East Coast fever, cattle, 65 field cases, clinical symptoms; recovery of 20 animals appeared to be unrelated to severity of symptoms or to drug treatment: Uganda

Eimeria
Barker, I. K.; Munday, B. L.; and Presidente, P. J. A., 1979, J. Parasitol., v. 65 (3), 451-456
Syn.: Ileocystis

Eimeria
Beier, T. V.; Shibalova, T. A.; and Kostenko, L. A., 1978, [Cytology of coccidia], 185 pp., illus.

Eimeria
Godfrey, D. G., 1978, Nature, London (5664), v. 273, 600-604
identification of economically important parasites (use of anatomical, biochemical, and behavioral tests), brief review

Eimeria
Gutteridge, W. E.; and Coombs, G. H., 1977, Biochemistry of parasitic protozoa, 172 pp., illus.
biochemistry of parasitic protozoa, textbook: methodology; catabolism and generation of energy; nucleic acid metabolism; protein metabolism; lipid metabolism; biochemical mechanism of drug action; isolation of parasitic protozoa from infected animals; culture of parasitic protozoa

Eimeria
Joyner, L. P.; et al., 1978, Parasitology, v. 77 (1), 27-31
coccidia, particularly of the genus Eimeria, proposed terminology which provides for discussion of variation at infrasubspecific levels, guidelines for designation of strains and lines

Eimeri[a], illus.
Koerting, W., 1977, Fisch u. Umwelt (4), 37-48
fish parasites, histopathological changes

Eimeria
Lumsden, W. H. R., 1978, Handb. Exper. Immunol. (Weir), 3. ed., 42.1-42.47
demonstration of antibodies to Protozoa, extensive review

Eimeria
McCallister, G. L., 1979, J. Parasitol., v. 65 (1), 24
Eimeria, use of commercial soft drinks as source of carbon dioxide for excystation of oocysts

Eimeria
Moon, H. W.; et al., 1978, J. Am. Vet. Med. Ass., v. 173 (5, pt. 2), 577-583
cryptosporidia and Eimeria found among other agents in newborn calves that had naturally occurring diarrheal disease: Iowa

Eimeria
Ogilvie, B. M.; and Rose, M. E., 1978, Colloque Immun. Parasit. Dis. (Thiverval-Grignon, Sept. 5-9, 1977), 237-248
intestinal nematodes and coccidia compared in terms of host response and immunity, colloquium presentation

Eimeria
Rose, M. E., 1972, Immun. Animal Parasites, 365-388
immune response to Eimeria, review

Eimeria
Samoil, H. P.; and Samuel, W. M., 1977, Canad. J. Zool., v. 55 (10), 1671-1683
key to species parasitizing members of genus Lepus
includes: Eimeria septentrionalis; E. holmesi n. sp.; E. punjabensis; E. hungarica; E. ruficaudati; E. keithi n. sp.; E. rowani n. sp.; E. athabascensis n. sp.; E. robertsonii; E. stefanskii; E. americana; E. europea; E. stiedai; E. belorussica; E. townsendi; E. semisculpta; E. sculpta

Eimeria [sp.], presumably E. neosciuri
Britt, D.; and Molyneux, D. H., 1979, J. Parasitol., v. 65 (3), 408
Sciurus carolinensis: Delamere Forest, Cheshire, England

Eimeria sp.
Byman, D.; et al., 1977, Canad. J. Zool., v. 55 (2), 376-380
Canis lupus (feces): northeastern Minnesota

Eimeria spp. oocysts
Carpenter, J. W.; et al., 1979, J. Am. Vet. Med. Ass., v. 175 (9), 948-951
Grus canadensis (feces): Patuxent Wildlife Research Center

Eimeria sp.
Davronov, O., 1973, Parazitologiia, Leningrad, v. 7 (2), 190-191
Eimeria sp., virulence for Mus musculus, possible use in biological control of house mice

Eimeria sp.
Dissanaike, A. S.; and Poopalachelvam, M., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (3), 420
coccidian oocysts from stools of crows examined, "After sporulation, however, it was clear that they were oocysts of Isospora and not Eimeria ..." as reported by Bisseru and Lim in 1971: Klang, Malaysia

Eimeria sp. oocysts
Dubey, J. P.; Fayer, R.; and Seesee, F. M., 1978, J. Am. Vet. Med. Ass., v. 173 (9), 1167-1170
coyotes (feces): Montana

Eimeria spp.
Horton, G. M. J.; and Stockdale, P. H. G., 1979, Am. J. Vet. Research, v. 40 (7), 966-970
Eimeria spp., lambs, amprolium and monensin, oocyst discharge, feed utilization, and ruminal metabolism

Eimeria spp.
Jacobson, H. A.; Kirkpatrick, R. L.; and McGinnes, B. S., 1978, Wildlife Monogr. (60), 53 pp.
disease and physiologic characteristics of cottontail rabbits in 2 study areas in relation to population density, includes data on seasonal and sex differences
Sylvilagus floridanus: Virginia

Eimeria sp.
Kocan, A. A.; Hannon, L.; and Eve, J. H., 1979, Proc. Oklahoma Acad. Sc., v. 59, 20-22
Colinus virginianus (intestine): Oklahoma

E[imeria spp.]
Kolev, G., 1978, Vet. Sbirka, v. 76 (6), 41-43
E[imeria spp.], calves, heifers, and cows, age and seasonal dynamics: Loveshk okrug

Eimeria sp.
Krishnamurthy, R.; and Kshirsagar, H. S., 1976, Marathwada Univ. J. Sc. (Nat. Sc.), v. 15 (8), 153-156
Eimeria spp., goats (feces), prevalence and seasonal incidence: slaughter house at Parbhani, Mahrasthra state

Eimeria sp., illus.
Manuel, M. F.; and Guerzon, V. A., 1978, Philippine J. Vet. Med., v. 17 (1-2), 121-128
Histomonas meleagridis, absence in ceca of Coturnix coturnix japonicum; incidental finding of developmental stages of Eimeria sp.: Philippines

Eimeria spp., illus.
Selim, M. K.; et al., 1970, Vet. Med. J., Giza, v. 17 (18), 173-193
cattle (intestine)
sheep
all imported to United Arab Republic

Eimeria sp.
Tarazona, J. M.; and Fernandez, J. M., 1974, An. Inst. Nac. Invest. Agrar., s. Hig. y San. Animal (1), 55-62
Eimeria spp., sheep, incidence, distribution, review of clinical aspects, pathology, epizootiology: Spain

Eimeria sp. n. Brada, 1966
Todd, K. S., jr.; Gallina, A. M.; and Schmidt, J. M., 1977, Proc. Helminth. Soc. Washington, v. 44 (2), 188-190
as syn. of Eimeria dunsingi Farr, 1960

Eimeria sp., unidentified
Woolf, A.; and Harder, J. D., 1979, Wildlife Monogr. (67), 53 pp.
Odocoileus virginianus: Rachelwood Wildlife Research Preserve, New Florence, Pennsylvania

Eimeria sp., illus.
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
Symphodus scina (urinary bladder): Black Sea

Eimeria abidzhanovi sp. n., illus.
Davronov, O., 1973, Parazitologiia, Leningrad, v. 7 (1), 79-86
Rhombomys opimus: Karshinsk and Ul'ianovsk regions, Uzbek SSR

E[imeria] acervulina
Balauca, N., 1978, Arch. Vet., Inst. Cercet. Vet. si Bioprep. Pasteur, v. 13, 127-141
experimental reproduction of necrotic enteritis in chicks with mixed infections of Clostridium perfringens and coccidia

E[imeria] acervulina
Bedrnik, P., 1977, Veterinarstvi, v. 27 (10), 458-459
coccidiosis, chickens, current prevalence, amprol plus treatment

Eimeria acervulina
Bedrnik, P.; et al., 1979, Arch. Geflugelk., v. 43 (1), 7-10
Eimeria spp., chickens (exper.), comparative efficiency of anticoccidials in combination with growth promotant nitrovin

Eimeria acervulina
Chapman, H. D., 1978, Ztschr. Parasitenk., v. 56 (2), 115-121
Eimeria spp., excystation, roles of various enzymes

Eimeria acervulina, illus.
Chappel, L. R., 1979, J. Parasitol., v. 65 (1), 137-143
Eimeria spp., chicks, site of action of salinomycin

Eimeria acervulina
Chappel, L. R.; and Babcock, W. E., 1979, Poultry Science, v. 58 (2), 304-307
Eimeria spp., broilers (exper.), salinomycin, monensin, lasalocid, drug toleration and anticoccidial efficacy compared in 5 field trials, commercial facilities

Eimeria acervulina
Cruthers, L. R.; et al., 1978, Poultry Science, v. 57 (5), 1227-1233
Eimeria spp., broiler chicks (exper.), lonomycin in feed, high degree of anticoccidial activity, comparison with lasalocid and monensin

Eimeria acervulina
Edgar, S. A.; and Flanagan, C., 1979, Poultry Science, v. 58 (6), 1469-1475
Eimeria spp. (recent field isolates resistant to various drugs), chickens, halofuginone, action cidal rather than static

Eimeria acervulina
Edgar, S. A.; and Flanagan, C., 1979, Poultry Science, v. 58 (6), 1476-1482
Eimeria spp. (recent field isolates resistant to various drugs), halofuginone with roxarsone and/or bacitracin MD

Eimeria acervulina
Fernando, M. A.; and McCraw, B. M., 1977, Ztschr. Parasitenk., v. 52 (3), 213-218
Eimeria acervulina-infected chickens, reduced time of generation cycle of duodenal crypt cells as measured by $[^3H]$thymidine, increased population of dividing cells within each duodenal crypt; changes seem to result from induced changes in functional activity

Eimeria acervulina
Greuel, E.; and Kuehnhold, W., 1977, Prakt. Tierarzt, v. 58 (5), 338-341
Eimeria spp., chickens (exper.), synergistic effect of metichlorpindol and methylbenzoquate, rotation program with other coccidiostats discussed

Eimeria acervulina
  Ishii, T.; and Onaga, H., 1978, Nippon Zyuisi-Kai Zassi (J. Japan Vet. Med. Ass.), v. 31 (9), 534-538
    Eimeria spp., chicks (exper.) and cell cultures, activity of chlorobromofebrifugine

Eimeria acervulina
  Kechik, I. T.; and Sykes, A. H., 1979, Brit. J. Nutrition, v. 42 (1), 97-103
    Eimeria acervulina, chicks (exper.), stress of intestinal infection results in depletion of ascorbic acid in blood plasma and tissues, addition of dietary ascorbic acid prevents depletion

Eimeria acervulina
  Kellogg, F. E.; Doster, G. L.; and Johnson, J. K., 1971, J. Wildlife Dis., v. 7 (3), 186-187
    Indian red junglefowl, pen-raised (small intestine)

Eimeria acervulina
  Kilgore, R. L.; et al., 1978, Poultry Science, v. 57 (4), 907-911
    Eimeria spp., broiler chickens (exper.), arprinocid, effects on weight, feed efficiency, and histological lesions

Eimeria acervulina
  Kilgore, R. L.; et al., 1979, Poultry Science, v. 58 (1), 67-71
    Eimeria spp., chickens, floor-pen trials evaluating 4 methods of induced exposure to coccidiosis suitable for use in drug research operations, laboratory sporulated oocysts spread over litter most satisfactory method

Eimeria acervulina
  Kucera, J.; and Lom, J., 1979, Veterinarstvi, v. 29 (8), 351-353
    Eimeria spp., Heterakis gallinae, incidence at large scale poultry farms using cage breeding methods: Cechach

Eimeria acervulina
  Kutzer, E.; et al., 1979, Wien. Tierarztl. Monatsschr., v. 66 (6-7), 197-202
    Eimeria spp., broiler chickens, arprinocid, drug efficacy compared with monensin

Eimeria acervulina
  Latif, B. M. A.; Ali, S. R.; and Versenyi, L., 1974, Acta Vet. Beograd, v. 24 (3), 135-138
    Eimeria spp., chickens (feces), rate of infection: Baghdad area, Iraq

Eimeria acervulina
  Lee, E. H.; and Onderka, D. K., 1978, Canad. Vet. J., v. 19 (9), 237-243
    Eimeria spp., chickens, incidence of clinical coccidiosis by month of year and age of host in the Ontario Veterinary Services Branch records 1973-1977, possible use of data to synchronize drug rotation with change in incidence

E[imeria] acervulina
  Long, P. L.; and Millard, B. J., 1978, Avian Path., v. 7 (3), 373-381
    coccidiosis, broiler chickens, effect on oocyst output of various treatment regimens

Eimeria acervulina
  Long, P. L.; and Millard, B. J., 1979, Avian Path., v. 8 (3), 213-228
    Eimeria spp., young chickens kept in litter pens, immunization, response to challenge with homologous and heterologous strains, effect of host age and of immunizing dose, timing of onset of immunity, longevity of immunity

Eimeria acervulina
  Long, P. L.; Millard, B. J.; and Smith, K. M., 1979, Avian Path., v. 8 (4), 453-467
    Eimeria spp., chickens, effect of 4 anticoccidial drugs on development of immunity, field and laboratory conditions

Eimeria acervulina
  Long, P. L.; and Rowell, J. G., 1975, Brit. Poultry Sc., v. 16 (6), 583-592
    Eimeria spp., method of sampling surface litter of commercial broiler houses for laboratory estimation of numbers of coccidial oocysts

Eimeria acervulina
  McDougald, L. R.; Karlsson, T.; and Reid, W. M., 1979, Avian Dis., v. 23 (4), 999-1005
    coccidiosis, chickens (exper.), natural outbreak of infectious bursal disease (IBD) during comparison of anticoccidials for their effect on development of immunity, interaction between diseases, immunity to coccidiosis not blocked by IBD

Eimeria acervulina
  Major, J. R., jr.; and Ruff, M. D., 1978, Exper. Parasitol., v. 45 (2), 234-240
    Eimeria spp., effects of coccidiosis on digestive capacity (amylase) of broiler chickens, changes in pancreatic, luminal, and surface-bound amylolytic activity, reduction in amylolytic activity as pH went below 5.0

Eimeria acervulina
  Major, J. R., jr.; and Ruff, M. D., 1978, J. Parasitol., v. 64 (4), 706-711
    Eimeria spp.-infected broilers, reduced disaccharidase activity in region of intestine with maximum infection, this reduction is related to both time and severity of infection and can contribute to overall reduction in nutrient absorption

Eimeria acervulina, illus.
  Michael, E., 1978, Ztschr. Parasitenk., v. 57 (3), 221-228
    Eimeria acervulina, formation and final structure of oocyst wall, transmission and scanning electron microscopy

Eimeria acervulina
  Migaki, T. T.; Chappel, L. R.; and Babcock, W. E., 1979, Poultry Science, v. 58 (5), 1192-1196
    Eimeria spp., chicks (exper.), salinomycin, monensin, lasalocid, efficacy in battery trials

Eimeria acervulina
  Mørch, J., 1978, Nord. Vet. Med., v. 30 (6), 253-259
    Eimeria spp., chickens (exper.), efficacy of coccidiostats in feed, better production efficiency of medicated groups

Eimeria acervulina
Morozova, T. I.; and Kravtsov, V. A., 1978, Tsitologiia, v. 20 (7), 817-819
Eimeria tenella, E. acervulina, sporozoites, ratio of rhoptry-microneme elements to anterior pole cytoplasm, such ratios may be species specific

Eimeria acervulina
Morrison, W. D.; Ferguson, A. E.; and Leeson, S., 1979, Poultry Science, v. 58 (5), 1160-1166
Eimeria spp., chicks (exper.), salinomycin and stenorol compared with other anticoccidials, efficacy and effect on chick performance

Eimeria acervulina
Noblet, G. P.; and Turk, D. E., 1979, Poultry Science, v. 58 (2), 392-403
Eimeria spp., chicks (exper.), no interference with overall intestinal absorption of ^{14}C-glucose, results suggest probable compensatory absorption by sections of intestine unaffected by a specific coccidial infection

Eimeria acervulina
Oikawa, H.; and Kawaguchi, H., 1975, Kiseichugaku Zasshi (Japan. J. Parasitol.), v. 24 (3), 99-106
Eimeria acervulina, E. tenella, chickens, effect of single vs. repeated vs. successive infections of mixed species on manifestation of symptoms, food intake and body weight gain, and oocyst production

Eimeria acervulina
Olson, G.; et al., 1978, Poultry Science, v. 57 (5), 1245-1250
Eimeria spp. field isolates, chickens (exper.), arprinocid in comparison trials with marketed drugs, effective against all isolates tested including those refractory to many of the other products

Eimeria acervulina
Onaga, H.; et al., 1978, Nippon Zyuisi-Kai Zassi (J. Japan Vet. Med. Ass.), v. 31 (10), 592-596
Eimeria spp., chicks and cell cultures, monensin

Eimeria acervulina
Panitz, E., 1979, Parasitology, v. 78 (1), 33-40
Eimeria spp., chicks, anticoccidial efficacy and cross-resistance patterns of N,N'-bis (3,4 ditrifluoromethylphenyl) methylmalonamide compound have no practical application because of weight gain depression

Eimeria acervulina
Parre, J.; and Olkonen, E., 1977, Eesti Pollumaj. Akad. Teadusl. Toode Kogum. (104), 100-108
Eimeria tenella, E. acervulina, E. brunetti, polyvaccine tested, chicks maintained in battery cages or deep litter; simultaneous zoalene treatment efficacious in deep litter maintenance

E[imeria] acervulina
Pujic, P.; Krecov, M.; and Simic, V., 1977, Vet. Glasnik, v. 31 (4), 277-280
E[imeria] spp., chickens, stenorol prevents coccidiosis under laboratory and field conditions, during fattening period

Eimeria acervulina
Quarles, C. L.; and Fagerberg, D. J., 1979, Poultry Science, v. 58 (2), 465-468
Eimeria acervulina, chicks (exper.), ammonia stress, mortality and growth performance

Eimeria acervulina
Rose, M. E.; and Hesketh, P., 1979, Infect. and Immun., v. 26 (2), 630-637
Eimeria spp. infections in normal animals vs. in animals with functional deficiencies in either T-lymphocytes or B-lymphocytes

Eimeria acervulina
Rose, M. E.; Hesketh, P.; and Ogilvie, B. M., 1979, Immunology, v. 36 (1), 71-79
Eimeria maxima, chickens, E. nieschulzi, rats, primary and secondary infections, E. maxima-immunized chickens challenged with E. acervulina: peripheral blood leucocyte response, correlation with resistance to reinfection

Eimeria acervulina
Ruff, M. D., 1978, Proc. 1978 Maryland Nutrition Conf. Feed Mfr. (University of Maryland, Mar. 16-17), 32-37
Eimeria spp., chickens, anticoccidials, safe withdrawal times, effect on nutrient malabsorption

Eimeria acervulina
Ruff, M. D.; et al., 1978, Avian Dis., v. 22 (1), 32-41
Eimeria spp., broilers (exper.), arprinocid, drug efficacy in both battery and floor-pen trials

Eimeria acervulina
Ruff, M. D.; et al., 1979, Poultry Science, v. 58 (2), 298-303
Eimeria spp., battery raised broilers (exper.), narasin compared with monensin

Eimeria acervulina
Ruff M. D.; Anderson, W. I.; and Reid, W. M., 1978, J. Parasitol., v. 64 (2), 306-311
Eimeria spp. in broilers, arprinocid decreased number of oocysts produced, fewer of the oocysts sporulated, and those oocysts which did sporulate were less infective than those from unmedicated birds

Eimeria acervulina
Ruff, M. D.; and Wyatt, R. D., 1978, Avian Dis., v. 22 (3), 471-480
Eimeria acervulina, 5 strains, broiler chicks, dietary aflatoxin increased severity of coccidiosis (body weight, plasma pigment, blood parameters)

Eimeria acervulina
Ruff, M. D.; Wyatt, R. D.; and Witlock, D. R., 1978, J. Parasitol., v. 64 (1), 23-26
Eimeria spp. in broilers, severe infection increased prothrombin times compared with uninfected birds

Eimeria acervulina
Ryley, J. F.; and Hardman, L., 1978, J. Parasitol., v. 64 (5), 878-881
Eimeria acervulina, E. mivati, speciation studies (cross-immunity and drug resistance studies), some immunological relationship was demonstrated but the failure of the 2 organisms to interbreed in the drug resistance studies lends support to status of E. mivati as distinct species

Eimeria acervulina
Ryley, J. F.; and Hardman, L., 1978, Parasitology, v. 76 (1), 11-20
Eimeria spp., chicks (exper.), effects of dietary vitamin K on severity of disease with particular attention to effects of vitamin K on response to anticoccidial drugs, concluded that use of vitamin K deficient diet for experimental work is quite justified

Eimeria acervulina
Ryley, N. G.; and Ryley, J. F., 1978, Parasitology, v. 77 (1), 33-39
Eimeria, 10 spp., survival and viability of oocysts following prolonged exposure to saturated sodium chloride solution, additional observations on virulence of Eimeria tenella for chickens following such exposure, no evidence obtained to contraindicate use of salt-flotation methods for separation of oocysts from feces

Eimeria acervulina
Schindler, P.; et al., 1979, Poultry Science, v. 58 (1), 23-27
Eimeria spp., broiler chicken pen trials, arprinocid in feed highly effective prophylaxis, comparison with halofuginone, monensin, nicarbazin, and pancoxin: England; France; Germany

Eimeria acervulina
Sevcik, B.; et al., 1979, Biol. a Chem. Zivoc. Vyroby, Vet., v. 15 (v. 21), (4), 375-383
coccidiosis, broilers, field trials of anticoccidials: CSSR

Eimeria acervulina
Shirley, M. W., 1979, Avian Path., v. 8 (4), 469-475
Eimeria mivati, chickens (exper.), 3 strains (2 chicken-maintained, 1 embryo-adapted), pathogenicity compared with E. acervulina, cross-protection between virulent and attenuated strains

Eimeria acervulina
Shirley, M. W., 1979, Parasitology, v. 78 (2), 221-237
Eimeria mivati, enzyme electrophoresis, cross-immunity, and other studies suggest that confusion over taxonomic status arose because some cultures were contaminated with E. acervulina; possibility exists that E. mivati and E. mitis are same species; E. diminuta or E. acervulina var. diminuta "should . . . be referred to as E. mivati (diminuta)."

Eimeria acervulina
Takhar, B. S.; and Farrell, D. J., 1979, Brit. Poultry Sc., v. 20 (2), 197-211
Eimeria acervulina- or Eimeria tenella-infected chickens, energy and nitrogen metabolism

Eimeria acervulina
Takhar, B. S.; and Farrell, D. J., 1979, Brit. Poultry Sc., v. 20 (2), 213-224
Eimeria acervulina, chickens, single infection provides protection against adverse effects on energy and nitrogen metabolism of further similar infection

Eimeria acervulina
Tamas, T.; et al., 1978, Poultry Science, v. 57 (2), 381-385
Eimeria acervulina, E. maxima, E. necatrix, E. tenella, arprinocid and dichloro analog L-628,914 in feed, decrease in oocysts' capacity to sporulate

Eimeria acervulina
Wang, C. C.; et al., 1979, Biochem. Pharmacol., v. 28 (15), 2249-2260
arprinocid inhibits hypoxanthine-guanine transport, may be mode of anticoccidial action

Eimeria acervulina var. diminuta
Joyner, L. P.; et al., 1978, Parasitology, v. 77 (1), 27-31
"The term 'variety' is considered unnecessary: it is proposed that the above parasites should be given the status of strains. Parasites previously described as E. maxima var. indentata for example, would be better referred to as E. maxima (indentata)."

Eimeria acervulina var. diminuta
Shirley, M. W., 1979, Parasitology, v. 78 (2), 221-237
Eimeria mivati, enzyme electrophoresis, cross-immunity, and other studies suggest that confusion over taxonomic status arose because some cultures were contaminated with E. acervulina; possibility exists that E. mivati and E. mitis are same species; E. diminuta or E. acervulina var. diminuta "should . . . be referred to as E. mivati (diminuta)."

Eimeria acervulina var. mivati
Schindler, P.; et al., 1979, Poultry Science, v. 58 (1), 23-27
Eimeria spp., broiler chicken pen trials, arprinocid in feed highly effective prophylaxis, comparison with halofuginone, monensin, nicarbazin, and pancoxin: England; France; Germany

Eimeria acervulina var. mivati
Shirley, M. W., 1979, Parasitology, v. 78 (2), 221-237
Eimeria mivati, enzyme electrophoresis, cross-immunity, and other studies suggest that confusion over taxonomic status arose because some cultures were contaminated with E. acervulina; possibility exists that E. mivati and E. mitis are same species; E. diminuta or E. acervulina var. diminuta "should . . . be referred to as E. mivati (diminuta)."

Eimeria adenoides, illus.
Bemrick, W. J.; and Hammer, R. F., 1979, Avian Dis., v. 23 (4), 812-820
Eimeria adenoides, turkeys (exper.), damage to cecal mucosa, scanning electron microscopy

Eimeria adenoeides
Edgar, S. A.; and Flanagan, C., 1979, Poultry Science, v. 58 (6), 1483-1489
Eimeria spp., turkeys, halofuginone effective

Eimeria adenoeides
Hamet-Foure, N.; Macar, C.; and Robin, B., 1979, Avian Path., v. 8 (1), 107-113
Eimeria meleagrimitis, E. adenoeides, turkeys, activity of clopidol with methylbenzoquate and amprolium with ethopabate: France

Eimeria adenoeides
McDougald, L. R., 1979, Poultry Science, v. 58 (1), 76-80
coccidiosis, histomoniasis, turkeys, tests for efficacy and compatibility indicate that amprolium and carbarsone can be used in combination

Eimeria adenoeides
McDougald, L. R.; and Johnson, J. K., 1979, Poultry Science, v. 58 (1), 72-75
Eimeria spp., turkeys, efficacy of arprinocid compared with amprolium, floor pen studies

Eimeria adenoeides
McDougald, L. R.; and McQuistion, T. E., 1978, Avian Dis., v. 22 (4), 765-770
Eimeria spp., turkeys, coccidiosis management, innate (age) resistance and acquired immunity vs. anticoccidial medication

Eimeria adenoeides, illus.
Madden, P. A.; and Ruff, M. D., 1979, J. Parasitol., v. 65 (2), 234-242
Eimeria spp. in turkeys, effects on structural integrity of intestinal and cecal mucosa, scanning electron microscopy, comparison of damage with parasite distribution as seen by light microscopy

Eimeria adenoeides
Mitrovic, M.; Schildknecht, E. G.; and Marusich, W. L., 1979, Poultry Science, v. 58 (5), 1154-1159
Eimeria spp., turkey poults (exper.), single and mixed infections, lasalocid sodium, controlled battery trials

Eimeria adenoides
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Eimeria adenoeides
Reid, W. M.; Anderson, W. I.; and McDougald, L. R., 1978, Avian Path., v. 7 (4), 569-576
Eimeria spp., turkey poults, anticoccidial protection and development of immunity while using monensin

Eimeria adenoides
Weber, L. J., 1978, Canad. Vet. J., v. 19 (1), 21 [Letter]
Eimeria meleagrimitis, E. adenoides, turkeys, outbreak causing great economic loss despite anticoccidial drug in medicated feed; need for recognizing development of resistant strains of coccidia: Beamsville, Ontario

Eimeria adenoeides
Yvore, P.; et al., 1978, Ann. Recherches Vet., v. 9 (3), 531-539
Eimeria adenoeides, turkeys (exper.), single and multiple infections, pathology, suggested role of bacteria in pathogenic potential

Eimeria ahsata
Ajayi, J. A.; and Todd, A. C., 1977, Bull. Animal Health and Prod. Africa, v. 25 (3), 257-261
prevalence, prepatent period
sheep (feces): Wisconsin

Eimeria ah-sa-ta
Ermatova, D. U., 1978, Veterinariia, Moskva (3), 69-70
Eimeria spp., sheep and goats, occurrence, seasonality, host age: Kashkadar'insk oblast

Eimeria ahsata
Fitzgerald, P. R.; and Mansfield, M. E., 1978, Am. J. Vet. Research, v. 39 (1), 7-10
Eimeria spp., lambs (nat. and exper.), effect of monensin on oocyst discharge, feed consumption, fecal output, and weight gain (by sex of host): Illinois

Eimeria ahsata (Honess, 1942), illus.
Gobzem, V. R., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 124-130
[Ovis aries] (feces): Belorussia

Eimeria ahsata, illus.
Golemanski, V. G., 1977, Priroda, Sofiia, v. 26 (1), 71-74
Ovis musimon: Bulgaria

Eimeria ahsata
Horton, G. M. J.; and Stockdale, P. H. G., 1979, Am. J. Vet. Research, v. 40 (7), 966-970
Eimeria spp., lambs, amprolium and monensin, oocyst discharge, feed utilization, and ruminal metabolism

Eimeria ahsata, illus.
McDougald, L. R., 1979, J. Protozool., v. 26 (1), 109-113

Eimeria ahsata
McDougald, L. R.; and Dunn, W. J., 1978, Am. J. Vet. Research, v. 39 (9), 1459-1462
Eimeria spp., lambs (exper.), efficacy of monensin in feed

Eimeria ahsata
Samizadeh-Yazd, A.; et al., 1979, Am. J. Vet. Research, v. 40 (8), 1107-1109
Eimeria spp., lambs, efficacy of monensin and aureomycin separately and combined

Eimeria ah-sa-ta
Santiago, M. A. M.; da Costa, U. C.; and da Silva, O. L., 1977, Rev. Centro Cien. Rurais, v. 7 (3), 297-301
Eimeria spp., sheep, natural infection, amprolium, furoxone, not sufficient control for sheep grazing on pasture

Eimeria ahsata
Tarazona, J. M.; and Fernandez, J. M., 1974, An. Inst. Nac. Invest. Agrar., s. Hig. y San. Animal (1), 55-62
Eimeria spp., sheep, incidence, distribution, review of clinical aspects, pathology, epizootiology: Spain

Eimeria alabamensis
Graefner, G.; Graubmann, H. D.; and Kron, A., 1978, Monatsh. Vet.-Med., v. 33 (23), 910-912
Eimeria spp., cattle on rearing and fattening farms, intensity and extensity of infection in relation to certain environmental conditions: DDR

Eimeria anatis
Golovneva, L. F., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 116-119
helminths and protozoa of domestic waterfowl, prevalence under intensive rearing conditions

Eimeria anseris
  Golovneva, L. F., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 116-119
    helminths and protozoa of domestic waterfowl, prevalence under intensive rearing conditions

Eimeria apodemi
  Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
    Apodemus flavicollis (vorderes Jejunum, Ileum, Rectum): Neusiedlerseegebiet, nordlichen Burgenland

Eimeria arctica Yakimoff, Matschoulsky, Spartansky, 1939, illus.
  Nukerbaeva, K. K., 1978, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (3), 16-21
    Cervus axis (feces): Alma-Ata zoopark

Eimeria arkhari Yakimoff, Matschoulsky, 1934, illus.
  Nukerbaeva, K. K., 1978, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (3), 16-21
    Ovis ammon (feces): Alma-Ata zoopark

Eimeria arloingi
  Ajayi, J. A.; and Todd, A. C., 1977, Bull. Animal Health and Prod. Africa, v. 25 (3), 257-261
    prevalence, prepatent period
    sheep (feces): Wisconsin

Eimeria arloingi
  Ermatova, D. U., 1978, Veterinariia, Moskva (3), 69-70
    Eimeria spp., sheep and goats, occurrence, seasonality, host age: Kashkadar'insk oblast

Eimeria arloingi (Marotel, 1910), illus.
  Gobzem, V. R., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 124-130
    [Ovis aries] (feces): Belorussia

Eimeria arloingi
  Krishnamurthy, R.; and Kshirsagar, H. S., 1976, Marathwada Univ. J. Sc. (Nat. Sc.), v. 15 (8), 153-156
    Eimeria spp., goats (feces), prevalence and seasonal incidence: slaughter house at Parbhani, Mahrasthra state

Eimeria arloingi, Martin, 1909, illus.
  Nukerbaeva, K. K., 1978, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (3), 16-21
    Capra ibex sibirica (feces): Alma-Ata zoopark

Eimeria arloingi
  Santiago, M. A. M.; da Costa, U. C.; and da Silva, O. L., 1977, Rev. Centro Cien. Rurais, v. 7 (3), 297-301
    Eimeria spp., sheep, natural infection, amprolium, furoxone, not sufficient control for sheep grazing on pasture

Eimeria arloingi
  Seidel, B., 1979, Zool. Garten, n. F., v. 49 (2), 131-160
    Ovibos moschatus, management in zoological park, includes information on parasites and their treatment: Tierpark Berlin

Eimeria arloingi
  Tarazona, J. M.; and Fernandez, J. M., 1974, An. Inst. Nac. Invest. Agrar., s. Hig. y San. Animal (1), 55-62
    Eimeria spp., sheep, incidence, distribution, review of clinical aspects, pathology, epizootiology: Spain

Eimeria arundeli sp. n., illus.
  Barker, I. K.; Munday, B. L.; and Presidente, P. J. A., 1979, J. Parasitol., v. 65 (3), 451-456
    Vombatus ursinus (feces, tissues): Northeastern Tasmania, Australia; Victoria, Australia

Eimeria athabascensis sp. n., illus.
  Samoil, H. P.; and Samuel, W. M., 1977, Canad. J. Zool., v. 55 (10), 1671-1683
    key
    Lepus americanus (feces): near Rochester, north-central Alberta

E[imeria] auburnensis
  Bobkova, A. F., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 90-94
    coccidia and helminths of calves, comparison of stall-enclosure and pasture maintenance, economics

Eimeria auburnensis
  Cotteleer, C.; and Fameree, L., 1978, Schweiz. Arch. Tierh., v. 120 (3), 149-156
    Eimeria spp., cattle, rate of infestation: Belgium

Eimeria auburnensis
  Evplov, N. N.; and Nazarov, V. G., 1977, Veterinariia, Moskva (6), 65-66
    Eimeria spp., calves, chemococcide effective; compared with biomycin and norsulfazol: Belgorodsk oblast

Eimeria auburnensis
  Fitzgerald, P. R.; and Mansfield, M. E., 1979, J. Parasitol., v. 65 (5), 824-825
    Eimeria spp., calves (exper.), lasalocid

Eimeria auburnensis
  Graefner, G.; Graubmann, H. D.; and Kron, A., 1978, Monatsh. Vet.-Med., v. 33 (23), 910-912
    Eimeria spp., cattle on rearing and fattening farms, intensity and extensity of infection in relation to certain environmental conditions: DDR

Eimeria auburnensis
  Hiepe, T.; Romeyke, D.; and Jungmann, R., 1978, Monatsh. Vet.-Med., v. 33 (23), 904-910
    Eimeria spp., calves reared under conditions of industrialized cattle farming, course of infection, clinical symptoms, monthly distribution, control measures: DDR

Eimeria auburnensis
  Ward, J. K.; Ferguson, D. L.; and Parkhurst, A. M., 1979, J. Animal Sc., v. 49 (2), 306-309
    gastrointestinal parasites, beef cows (feces), level of infection, effect of animal age and season of year: Mead, Nebraska

Eimeria audubonii
  Wiggins, J. P.; and Rothenbacher, H., 1979, J. Parasitol., v. 65 (3), 393-394
    Sylvilagus floridanus (feces): central Pennsylvania

Eimeria azul sp. n., illus.
  Wiggins, J. P.; and Rothenbacher, H., 1979, J. Parasitol., v. 65 (3), 393-394
    Sylvilagus floridanus (feces): central Pennsylvania

Eimeria badchisica
  Davronov, O., 1973, Parazitologiia, Leningrad, v. 7 (1), 79-86
    Rhombomys opimus: Uzbek SSR

Eimeria bandipurensis, illus.
  Chowattukunnel, J. T., 1979, J. Protozool., v. 26 (1), 36-38
    redescription
    Funambulus tristriatus (feces): Kottayam district, Kerala State, India

Eimeria bareillyi Gill, Chhabra & Lal, 1963 illus.
  Krishnamurthy, R.; and Shastry, U. V., 1976, Riv. Parassitol., Roma, v. 37 (1), 71-77
    Eimeria bareillyi, morphology of oocyst
    Bubalus bubalis: Parbhani, Maharashtra State, India

Eimeria? beauchampi, illus.
  Benito, J., 1978, Bol. R. Soc. Espan. Hist. Nat., Secc. Biol., v. 75, 1977, 267-273
    Eimeria beauchampi, cytochemistry, pathology
    Glossobalanus minutus (intestinal epithelium): Luanco

Eimeria bigemina Labbe, 1896
  Levine, N. D., 1977, Protozoology, v. 3, 173-182
    as syn. of Klossia bigemina (Labbe, 1896) Labbe, 1899

Eimeria bovis
  Benz, G. W.; and Ernst, J. V., 1979, Am. J. Vet. Research, v. 40 (8), 1180-1186
    Eimeria bovis, calves (exper.), efficacy of salinomycin

E[imeria] bovis
  Bobkova, A. F., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 90-94
    coccidia and helminths of calves, comparison of stall-enclosure and pasture maintenance, economics

Eimeria bovis
  Cotteleer, C.; and Fameree, L., 1978, Schweiz. Arch. Tierh., v. 120 (3), 149-156
    Eimeria spp., cattle, rate of infestation: Belgium

Eimeria bovis, "resembled"
  Desser, S. S., 1978, J. Parasitol., v. 64 (5), 933-935
    evidence of extraintestinal development
    Rupicapra rupicapra (liver): Westland game farm

Eimeria bovis, illus.
  Dubremetz, J. F.; and Elsner, Y. Y., 1979, J. Protozool., v. 26 (3), 367-376
    Eimeria bovis, schizogony and merogenesis in cell culture, ultrastructural study

Eimeria bovis
  Evplov, N. N.; and Nazarov, V. G., 1977, Veterinariia, Moskva (6), 65-66
    Eimeria spp., calves, chemococcide effective; compared with biomycin and norsulfazol: Belgorodsk oblast

Eimeria bovis, illus.
  Farquhar, A. S.; Anthony, W. B.; and Ernst, J. V., 1979, J. Animal Sc., v. 49 (5), 1331-1336
    Eimeria bovis oocysts in manure-blended diet, adequate ensiling prevents sporulation

Eimeria bovis
  Fitzgerald, P. R.; and Mansfield, M. E., 1979, J. Parasitol., v. 65 (5), 824-825
    Eimeria spp., calves (exper.), lasalocid

Eimeria bovis, illus.
  Fox, J. E., 1978, Mod. Vet. Pract., v. 59 (8), 599-603
    bovine coccidiosis, review, emphasis on prevention and control; field tests, decoquinate against Eimeria bovis and E. zuernii prevented clinical signs of disease with no observable signs of toxicity

E[imeria] bovis
  Gobzem, V. R., 1970, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 8, 118-120
    E[imeria] bovis, E. cylindrica, E. ellipsoidalis, calves, safe prophylaxis with biovetin

Eimeria bovis
  Gobzem, V. R.; and Nazarov, V. G., 1978, Veterinariia, Moskva (3), 67-69
    Eimeria spp., calves, diagnostic difficulties, clinical symptoms, chemoprophylactic substances tested at various dosages and in various combinations

Eimeria bovis
  Graefner, G.; Graubmann, H. D.; and Kron, A., 1978, Monatsh. Vet.-Med., v. 33 (23), 910-912
    Eimeria spp., cattle on rearing and fattening farms, intensity and extensity of infection in relation to certain environmental conditions: DDR

Eimeria bovis
  Hiepe, T.; Romeyke, D.; and Jungmann, R., 1978, Monatsh. Vet.-Med., v. 33 (23), 904-910
    Eimeria spp., calves reared under conditions of industrialized cattle farming, course of infection, clinical symptoms, monthly distribution, control measures: DDR

Eimeria bovis
  Klesius, P. H.; et al., 1975, Transplant. Proc., v. 7 (3), 449-452
    Eimeria bovis, calves, delayed hypersensitivity (DH) response, passive transfer to other calves via lymphocytes and via cell-free transfer factor (TF), acquired immunity found in some calves receiving TF; DH skin reactivity for coccidian oocyst antigen and diphtheria toxoid was also passively transferred to rabbits, dogs, and rhesus monkeys with calf TF

Eimeria bovis
  Klesius, P. H.; et al., 1978, Clin. Immunol. and Immunopathol., v. 10 (2), 214-221
    Eimeria ferrisi, cell-mediated immunity stimulated in mice by prophylactic treatment with bovine transfer factor (TFd) prepared from lymph node lymphocytes of cattle immune to E. bovis, lymphocyte stimulation and protection against clinical infection, susceptible mice given lymphocytes from donor mice treated with bovine TFd were also partly protected against clinical infection

Eimeria bovis
  McDougald, L. R., 1978, Am. J. Vet. Research, v. 39 (11), 1748-1749
    Eimeria bovis, Holstein-Friesian calves (exper.), monensin added to complete feed ration as preventive medication, control excellent

Eimeria bovis (Zublin, 1908) Fiebiger, 1912, illus.
  Musaev, M. A.; and Manafova, Sh. G., 1971, Parazitologiia, Leningrad, v. 5 (4), 310-315
    Eimeria smithi, valid species, not a synonym of Eimeria bovis

Eimeria bovis
  Ryley, N. G.; and Ryley, J. F., 1978, Parasitology, v. 77 (1), 33-39
    Eimeria, 10 spp., survival and viability of oocysts following prolonged exposure to saturated sodium chloride solution, additional observations on virulence of Eimeria tenella for chickens following such exposure, no evidence obtained to contraindicate use of salt-flotation methods for separation of oocysts from feces

Eimeria bovis, illus.
  Supperer, R., 1973, Tieraerztl. Prax., v. 1 (4), 403-412
    parasites of calves, diagnosis, pathology, therapy, review

Eimeria bovis
  Ward, J. K.; Ferguson, D. L.; and Parkhurst, A. M., 1979, J. Animal Sc., v. 49 (2), 306-309
    gastrointestinal parasites, beef cows (feces), level of infection, effect of animal age and season of year: Mead, Nebraska

Eimeria bovis
  Zajicek, D.; et al., 1977, Veterinarstvi, v. 27 (11), 507-508
    Eimeria spp., calves, prevalence in large-scale breeding

Eimeria brasiliensis
  Cotteleer, C.; and Fameree, L., 1978, Schweiz. Arch. Tierh., v. 120 (3), 149-156
    Eimeria spp., cattle, rate of infestation: Belgium

Eimeria brunetti, illus.
  Black, J.; and Smith, B. F., 1977, Microbios Letters (23-24), v. 6, 111-116
    Eimeria brunetti, microgametogony and macrogametogony, ultrastructure

Eimeria brunetti
  Chapman, H. D., 1978, Avian Path., v. 7 (2), 269-277
    Eimeria maxima, E. brunetti, E. tenella, chickens, effect of monensin on development of immunity acquired by repeated low-level infections

Eimeria brunetti
  Chapman, H. D., 1978, Ztschr. Parasitenk., v. 56 (2), 115-121
    Eimeria spp., excystation, roles of various enzymes

Eimeria brunetti
  Chappel, L. R.; and Babcock, W. E., 1979, Poultry Science, v. 58 (2), 304-307
    Eimeria spp., broilers (exper.), salinomycin, monensin, lasalocid, drug toleration and anticoccidial efficacy compared in 5 field trials, commercial facilities

Eimeria brunetti
  Cruthers, L. R.; et al., 1978, Poultry Science, v. 57 (5), 1227-1233
    Eimeria spp., broiler chicks (exper.), lonomycin in feed, high degree of anticoccidial activity, comparison with lasalocid and monensin

Eimeria brunetti
  Edgar, S. A.; and Flanagan, C., 1979, Poultry Science, v. 58 (6), 1469-1475
    Eimeria spp. (recent field isolates resistant to various drugs), chickens, halofuginone, action cidal rather than static

Eimeria brunetti, illus.
  Ferguson, D. J. P.; et al., 1978, Acta Path. et Microbiol. Scand., v. 86B (1), 1-11
    Eimeria brunetti, sporulation of oocysts, development of zygote and formation of sporoblasts, light and electron microscopy

Eimeria brunetti, illus.
  Ferguson, D. J. P.;et al., 1978, Acta Path. et Microbiol. Scand., v. 86B (1), 13-24
    Eimeria brunetti, sporulation of oocysts, development into sporocyst, formation of sporozoite, light and electron microscopy

Eimeria brunetti
  Greuel, E.; and Kuehnhold, W., 1977, Prakt. Tierarzt, v. 58 (5), 338-341
    Eimeria spp., chickens (exper.), synergistic effect of metichlorpindol and methylbenzoquate, rotation program with other coccidiostats discussed

Eimeria brunetti
  Ishii, T.; and Onaga, H., 1978, Nippon Zyuisi-Kai Zassi (J. Japan Vet. Med. Ass.), v. 31 (9), 534-538
    Eimeria spp., chicks (exper.) and cell cultures, activity of chlorobromofebrifugine

Eimeria brunetti
  Kilgore, R. L.; et al., 1978, Poultry Science, v. 57 (4), 907-911
    Eimeria spp., broiler chickens (exper.), arprinocid, effects on weight, feed efficiency, and histological lesions

Eimeria brunetti
  Kilgore, R. L.; et al., 1979, Poultry Science, v. 58 (1), 67-71
    Eimeria spp., chickens, floor-pen trials evaluating 4 methods of induced exposure to coccidiosis suitable for use in drug research operations, laboratory sporulated oocysts spread over litter most satisfactory method

Eimeria brunetti
  Kucera, J.; and Lom, J., 1979, Veterinarstvi, v. 29 (8), 351-353
    Eimeria spp., Heterakis gallinae, incidence at large scale poultry farms using cage breeding methods: Cechach

Eimeria brunetti
  Kutzer, E.; et al., 1979, Wien. Tierarztl. Monatsschr., v. 66 (6-7), 197-202
    Eimeria spp., broiler chickens, arprinocid, drug efficacy compared with monensin

Eimeria brunetti
Latif, B. M. A.; Ali, S. R.; and Versenyi, L., 1974, Acta Vet. Beograd, v. 24 (3), 135-138
Eimeria spp., chickens (feces), rate of infection: Baghdad area, Iraq

Eimeria brunetti
Lee, E. H.; and Onderka, D. K., 1978, Canad. Vet. J., v. 19 (9), 237-243
Eimeria spp., chickens, incidence of clinical coccidiosis by month of year and age of host in the Ontario Veterinary Services Branch records 1973-1977, possible use of data to synchronize drug rotation with change in incidence

Eimeria brunetti
Long, P. L.; and Millard, B. J., 1979, Avian Path., v. 8 (3), 213-228
Eimeria spp., young chickens kept in litter pens, immunization, response to challenge with homologous and heterologous strains, effect of host age and of immunizing dose, timing of onset of immunity, longevity of immunity

Eimeria brunetti
Long, P. L.; Millard, B. J.; and Smith, K. M., 1979, Avian Path., v. 8 (4), 453-467
Eimeria spp., chickens, effect of 4 anticoccidial drugs on development of immunity, field and laboratory conditions

Eimeria brunetti
Long, P. L.; and Rowell, J. G., 1975, Brit. Poultry Sc., v. 16 (6), 583-592
Eimeria spp., method of sampling surface litter of commercial broiler houses for laboratory estimation of numbers of coccidial oocysts

Eimeria brunetti
McDougald, L. R.; Karlsson, T.; and Reid, W. M., 1979, Avian Dis., v. 23 (4), 999-1005
coccidiosis, chickens (exper.), natural outbreak of infectious bursal disease (IBD) during comparison of anticoccidials for their effect on development of immunity, interaction between diseases, immunity to coccidiosis not blocked by IBD

Eimeria brunetti
Major, J. R., jr.; and Ruff, M. D., 1978, Exper. Parasitol., v. 45 (2), 234-240
Eimeria spp., effects of coccidiosis on digestive capacity (amylase) of broiler chickens, changes in pancreatic, luminal, and surface-bound amylolytic activity, reduction in amylolytic activity as pH went below 5.0

Eimeria brunetti
Major, J. R., jr.; and Ruff, M. D., 1978, J. Parasitol., v. 64 (4), 706-711
Eimeria spp.-infected broilers, reduced disaccharidase activity in region of intestine with maximum infection, this reduction is related to both time and severity of infection and can contribute to overall reduction in nutrient absorption

Eimeria brunetti
Maxey, B. W.; and Page, R. K., 1977, Poultry Science, v. 56 (6), 1909-1913
Eimeria brunetti, E. maxima, chickens, lincomycin feed medication for control of necrotic enteritis

Eimeria brunetti
Migaki, T. T.; Chappel, L. R.; and Babcock, W. E., 1979, Poultry Science, v. 58 (5), 1192-1196
Eimeria spp., chicks (exper.), salinomycin, monensin, lasalocid, efficacy in battery trials

Eimeria brunetti
Mladenovic, Z.; Movsesijan, M.; and Borojevic, D., 1978, Vet. Glasnik, v. 32 (10), 829-834
Eimeria spp., chickens (exper.), mixed infections, cycostat, nitryl, and stenorol

Eimeria brunetti
Mørch, J., 1978, Nord. Vet. Med., v. 30 (6), 253-259
Eimeria spp., chickens (exper.), efficacy of coccidiostats in feed, better production efficiency of medicated groups

Eimeria brunetti
Noblet, G. P.; and Turk, D. E., 1979, Poultry Science, v. 58 (2), 392-403
Eimeria spp., chicks (exper.), no interference with overall intestinal absorption of ^{14}C-glucose, results suggest probable compensatory absorption by sections of intestine unaffected by a specific coccidial infection

Eimeria brunetti
Olson, G.; et al., 1978, Poultry Science, v. 57 (5), 1245-1250
Eimeria spp. field isolates, chickens (exper.), arprinocid in comparison trials with marketed drugs, effective against all isolates tested including those refractory to many of the other products

Eimeria brunetti
Onaga, H.; et al., 1978, Nippon Zyuisi-Kai Zassi (J. Japan Vet. Med. Ass.), v. 31 (10), 592-596
Eimeria spp., chicks and cell cultures, monensin

Eimeria brunetti
Parre, J.; and Olkonen, E., 1977, Eesti Pollumaj. Akad. Teadusl. Toode Kogum. (104), 100-108
Eimeria tenella, E. acervulina, E. brunetti, polyvaccine tested, chicks maintained in battery cages or deep litter; simultaneous zoalene treatment efficacious in deep litter maintenance

Eimeria brunetti
Ruff, M. D., 1978, Proc. 1978 Maryland Nutrition Conf. Feed Mfr. (University of Maryland, Mar. 16-17), 32-37
Eimeria spp., chickens, anticoccidials, safe withdrawal times, effect on nutrient malabsorption

Eimeria brunetti
Ruff, M. D.; et al., 1978, Avian Dis., v. 22 (1), 32-41
Eimeria spp., broilers (exper.), arprinocid, drug efficacy in both battery and floor-pen trials

Eimeria brunetti
Ruff, M. D.; et al., 1979, Poultry Science, v. 58 (2), 298-303
Eimeria spp., battery raised broilers (exper.), narasin compared with monensin

Eimeria brunetti
Ruff, M. D.; Anderson, W. I.; and Reid, W. M., 1978, J. Parasitol., v. 64 (2), 306-311
Eimeria spp. in broilers, arprinocid decreased number of oocysts produced, fewer of the oocysts sporulated, and those oocysts which did sporulate were less infective than those from unmedicated birds

Eimeria brunetti
Ryley, J. F.; and Hardman, L., 1978, Parasitology, v. 76 (1), 11-20
Eimeria spp., chicks (exper.), effects of dietary vitamin K on severity of disease with particular attention to effects of vitamin K on response to anticoccidial drugs, concluded that use of vitamin K deficient diet for experimental work is quite justified

Eimeria brunetti
Ryley, N. G.; and Ryley, J. F., 1978, Parasitology, v. 77 (1), 33-39
Eimeria, 10 spp., survival and viability of oocysts following prolonged exposure to saturated sodium chloride solution, additional observations on virulence of Eimeria tenella for chickens following such exposure, no evidence obtained to contraindicate use of salt-flotation methods for separation of oocysts from feces

Eimeria brunetti
Schindler, P.; et al., 1979, Poultry Science, v. 58 (1), 23-27
Eimeria spp., broiler chicken pen trials, arprinocid in feed highly effective prophylaxis, comparison with halofuginone, monensin, nicarbazin, and pancoxin: England; France; Germany

Eimeria brunetti
Thanikachalam, M.; Sundararaj, A.; and Manohar, B. M., 1979, Poultry Adviser, v. 12 (7), 63-65
Eimeria brunetti, fowls, pathology of ileum and rectum: near Madras

Eimeria bukidnonensis
Cotteleer, C.; and Fameree, L., 1978, Schweiz. Arch. Tierh., v. 120 (3), 149-156
Eimeria spp., cattle, rate of infestation: Belgium

Eimeria bukidnonensis
Evplov, N. N.; and Nazarov, V. G., 1977, Veterinariia, Moskva (6), 65-66
Eimeria spp., calves, chemococcide effective; compared with biomycin and norsulfazol: Belgorodsk oblast

Eimeria bukidnonensis
Graefner, G.; Graubmann, H. D.; and Kron, A., 1978, Monatsh. Vet.-Med., v. 33 (23), 910-912
Eimeria spp., cattle on rearing and fattening farms, intensity and extensity of infection in relation to certain environmental conditions: DDR

Eimeria bukidnonensis
Hiepe, T.; Romeyke, D.; and Jungmann, R., 1978, Monatsh. Vet.-Med., v. 33 (23), 904-910
Eimeria spp., calves reared under conditions of industrialized cattle farming, course of infection, clinical symptoms, monthly distribution, control measures: DDR

Eimeria bukidnonensis
Ward, J. K.; Ferguson, D. L.; and Parkhurst, A. M., 1979, J. Animal Sc., v. 49 (2), 306-309
gastrointestinal parasites, beef cows (feces), level of infection, effect of animal age and season of year: Mead, Nebraska

Eimeria bukidnonensis
Zajicek, D.; et al., 1977, Veterinarstvi, v. 27 (11), 507-508
Eimeria spp., calves, prevalence in large-scale breeding

Eimeria cameli, illus.
Ipczynski, V., 1978, Berl. u. Munchen. Tierarztl. Wchnschr., v. 91 (1), 11-12
Eimeria cameli, sporulation time, measurements
Kamel (Kot): Zoologischen Garten Berlin

Eimeria canadensis
Graefner, G.; Graubmann, H. D.; and Kron, A., 1978, Monatsh. Vet.-Med., v. 33 (23), 910-912
Eimeria spp., cattle on rearing and fattening farms, intensity and extensity of infection in relation to certain environmental conditions: DDR

Eimeria canadensis
Hiepe, T.; Romeyke, D.; and Jungmann, R., 1978, Monatsh. Vet.-Med., v. 33 (23), 904-910
Eimeria spp., calves reared under conditions of industrialized cattle farming, course of infection, clinical symptoms, monthly distribution, control measures: DDR

Eimeria canadensis Bruce, 1921, illus.
Musaev, M. A.; and Manafova, Sh. G., 1971, Parazitologiia, Leningrad, v. 5 (4), 310-315
Eimeria zurnabadensis, valid species, not a synonym of Eimeria canadensis

Eimeria canadensis
Ward, J. K.; Ferguson, D. L.; and Parkhurst, A. M., 1979, J. Animal Sc., v. 49 (2), 306-309
gastrointestinal parasites, beef cows (feces), level of infection, effect of animal age and season of year: Mead, Nebraska

Eimeria canis, illus.
Nukerbaeva, K. K., 1978, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (3), 16-21
Canis lupus (feces): Alma-Ata zoopark

Eimeria carassiusaurati n. sp., illus.
Romero Rodriguez, J., 1978, Rev. Iber. Parasitol., v. 38 (3-4), 775-781
Eimeria carassiusaurati n. sp., pathology
Carassius auratus (intestino): Granada, Espana

Eimeria carpelli
Tesarcik, J., 1971, Prace VURH Vodnany (9), 99-132
Neoechinorhynchus rutili, Eimeria spp., carp, tetrafinol, nitrofurazon, prophylaxis and therapy under aquarium conditions

Eimeria christenseni Levine, Ivens & Fritz, illus.
McDougald, L. R., 1979, J. Protozool., v. 26 (1), 109-113
Eimeria christenseni oocysts from goat, failure to infect sheep; E. ninakohlyakimovae-like oocysts from sheep, failure to infect goats, E. ninakohlyakimovae-like coccidia of the 2 hosts belong to separate species, E. ovinoidalis sp. n. proposed for species from sheep, E. ninakohlyakimovae retained for species from goats

Eimeria citelli
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Citellus citellus (Jejunum, Rectum): Neusiedlerseegebiet, nordlichen Burgenland

Eimeria clethrionomyis sp. n., illus.
Straneva, J. E.; and Kelley, G. L., 1979, J. Protozool., v. 26 (4), 530-532
Clethrionomys gapperi (feces): Brush Valley, Indiana County, Pennsylvania

Eimeria coecicola Cheissin, 1947, illus.
Catchpole, J.; and Norton, C. C., 1979, Parasitology, v. 79 (2), 249-257
Eimeria spp. in rabbits from 3 commercial rabbitries, species identification, level of infection, % occurrence and % predominance of different spp.: South East England

Eimeria colchici
Bejsovec, J., 1978, Ang. Parasitol., v. 19 (2), 76-85
Eimeria phasiani and E. colchici in Phasianus colchicus, dynamics of incidence dependent upon host biotope, host movements, season, temperature, and humidity: Mittelbohmen

Eimeria colchici
Williams, R. B., 1978, Vet. Parasitol., v. 4 (2), 193-197
Eimeria colchici causing coccidiosis in intensively raised pheasants, controlled by sulphaquinoxaline and diaveridine (Saquadil) in drinking water
Pavo cristatus: Essex, Great Britain

Eimeria columbarum
Cotteleer, C.; and Fameree, L., 1978, Schweiz. Arch. Tierh., v. 120 (4), 181-187
domestic and semi-feral pigeons, survey of intestinal parasites, presence of anti-Toxoplasma antibodies in serum, possible role in epidemiology of toxoplasmosis: Belgium

Eimeria conevi
Davronov, O., 1973, Parazitologiia, Leningrad, v. 7 (1), 79-86
Rhombomys opimus: Uzbek SSR

Eimeria crandallis
Ajayi, J. A.; and Todd, A. C., 1977, Bull. Animal Health and Prod. Africa, v. 25 (3), 257-261
prevalence, prepatent period
sheep (feces): Wisconsin

Eimeria crandallis
Ermatova, D. U., 1978, Veterinariia, Moskva (3), 69-70
Eimeria spp., sheep and goats, occurrence, seasonality, host age: Kashkadar'insk oblast

Eimeria crandallis
Foreyt, W. J.; Gates, N. L.; and Wescott, R. B., 1979, Am. J. Vet. Research, v. 40 (1), 97-100
Eimeria spp., confinement-reared lambs (exper.) from weaning to market weight, monensin and lasalocid highly effective in eliminating oocysts, weight gains and feed conversion rates measured

Eimeria crandallis (Honess, 1942), illus.
Gobzem, V. R., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 124-130
Syn.: Eimeria punctata (Landers, 1955)
[Ovis aries] (feces): Belorussia

Eimeria crandallis, illus.
Golemanski, V. G., 1977, Priroda, Sofiia, v. 26 (1), 71-74
Ovis musimon: Bulgaria

Eimeria crandallis
Mason, P., 1977, N. Zealand Vet. J., v. 25 (1-2), 30-33
Eimeria spp., lambs, development of naturally acquired infection: Otago, New Zealand

Eimeria crandallis
Samizadeh-Yazd, A.; et al., 1979, Am. J. Vet. Research, v. 40 (8), 1107-1109
Eimeria spp., lambs, efficacy of monensin and aureomycin separately and combined

Eimeria crandallis Honess 1942
Seidel, B., 1979, Zool. Garten, n. F., v. 49 (2), 131-160
Ovibos moschatus, management in zoological park, includes information on parasites and their treatment: Tierpark Berlin

Eimeria crandallis
Tarazona, J. M.; and Fernandez, J. M., 1974, An. Inst. Nac. Invest. Agrar., s. Hig. y San. Animal (1), 55-62
Eimeria spp., sheep, incidence, distribution, review of clinical aspects, pathology, epizootiology: Spain

Eimeria criceti
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Cricetus cricetus (Rectum): Neusiedlerseegebiet, nordlichen Burgenland

E[imeria] cylindrica
Bobkova, A. F., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 90-94
coccidia and helminths of calves, comparison of stall-enclosure and pasture maintenance, economics

Eimeria cylindrica
Cotteleer, C.; and Fameree, L., 1978, Schweiz. Arch. Tierh., v. 120 (3), 149-156
Eimeria spp., cattle, rate of infestation: Belgium

Eimeria cylindrica
Evplov, N. N.; and Nazarov, V. G., 1977, Veterinariia, Moskva (6), 65-66
Eimeria spp., calves, chemococcide effective; compared with biomycin and norsulfazol: Belgorodsk oblast

Eimeria cylindrica
Fitzgerald, P. R.; and Mansfield, M. E., 1979, J. Parasitol., v. 65 (5), 824-825
Eimeria spp., calves (exper.), lasalocid

E[imeria] cylindrica
Gobzem, V. R., 1970, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 8, 118-120
E[imeria] bovis, E. cylindrica, E. ellipsoidalis, calves, safe prophylaxis with biovetin

Eimeria cylindrica
Graefner, G.; Graubmann, H. D.; and Kron, A., 1978, Monatsh. Vet.-Med., v. 33 (23), 910-912
Eimeria spp., cattle on rearing and fattening farms, intensity and extensity of infection in relation to certain environmental conditions: DDR

Eimeria cylindrica
Hiepe, T.; Romeyke, D.; and Jungmann, R., 1978, Monatsh. Vet.-Med., v. 33 (23), 904-910
Eimeria spp., calves reared under conditions of industrialized cattle farming, course of infection, clinical symptoms, monthly distribution, control measures: DDR

Eimeria cylindrica
Ward, J. K.; Ferguson, D. L.; and Parkhurst, A. M., 1979, J. Animal Sc., v. 49 (2), 306-309
gastrointestinal parasites, beef cows (feces), level of infection, effect of animal age and season of year: Mead, Nebraska

Eimeria cylindrica
Zajicek, D.; et al., 1977, Veterinarstvi, v. 27 (11), 507-508
Eimeria spp., calves, prevalence in large-scale breeding

Eimeria debliecki group
Desser, S. S., 1978, J. Parasitol., v. 64 (5), 933-935
evidence of extraintestinal development
pig (liver)

Eimeria debliecki
Napravnik, J.; and Fajkos, A., 1978, Sborn. Vysoke Skoly Zemedelske Praze, Fak. Agron., Rada B, Zivoc. Vyroba (1), 3-19
Eimeria spp., nematodes, pigs, soft runs as source of contamination, weather and climatic conditions

Eimeria debliecki
Upadhyay, A. N.; and Ahluwalia, S. S., 1976, Indian Vet. J., v. 53 (12), 972-973
mixed coccidial infection, pig, first infection did not confer any immunity, pig became resistant to challenge after second infection

Eimeria debliecki, illus.
Upadhyay, A. N.; and Ahluwalia, S. S., 1978, Indian Vet. J., v. 55 (6), 458-461
coccidia, pigs (exper.), mixed infection with 4 spp., macro- and microgametocytic stages, mostly not identified to species

Eimeria debliecki
Upadhyay, A. N.; and Ahluwalia, S. S., 1978, Indian Vet. J., v. 55 (10), 829-830
4 spp. of coccidia, pigs (exper.), only slightly pathogenic, no gross observable lesions

Eimeria deserticola sp. n., illus.
Davronov, O., 1973, Parazitologiia, Leningrad, v. 7 (1), 79-86
Citellus fulvus: Karshinsk and Ul'ianovsk regions, Uzbek SSR

Eimeria diminuta
Shirley, M. W., 1979, Parasitology, v. 78 (2), 221-237
Eimeria mivati, enzyme electrophoresis, cross-immunity, and other studies suggest that confusion over taxonomic status arose because some cultures were contaminated with E. acervulina; possibility exists that E. mivati and E. mitis are same species; E. diminuta or E. acervulina var. diminuta "should . . . be referred to as E. mivati (diminuta)."

Eimeria dingleyi sp. n., illus.
Davies, A. J., 1978, J. Protozool., v. 25 (1), 15-21
cytochemistry
Blennius pholis (epithelium of intestine): Aberystwyth and Porth Cwyfan, Wales, U. K.

Eimeria dispersa Tyzzer 1929
Doran, D. J., 1978, J. Parasitol., v. 64 (5), 882-885
Eimeria dispersa, turkey strain, life cycle compared in chickens, Alectoris graeca, Phasianus colchicus, and Colinus virginianus (all exper.)

Eimeria dispersa Tyzzer, 1929, illus.
Doran, D. J., 1978, J. Protozoool., v. 25 (3, pt. 2), 293-297
Eimeria dispersa, life cycle in turkeys

Eimeria dispersa
Edgar, S. A.; and Flanagan, C., 1979, Poultry Science, v. 58 (6), 1483-1489
Eimeria spp., turkeys, halofuginone effective

Eimeria dispersa Tyzzer, 1929, illus.
Fisher, J. W.; Kelley, G. L.; and Youssef, N. N., 1979, Ztschr. Parasitenk., v. 59 (3), 203-210
Eimeria dispersa, development in bovine kidney cell cultures

Eimeria dispersa Tyzzer 1929, illus.
Long, P. L.; and Millard, B. J., 1979, Parasitology, v. 78 (1), 41-51
Eimeria dispersa, isolation from turkeys in Britain, life cycle and reproduction, cross-protection against American strain, electrophoretic analysis of enzymes, host specificity studies, in vitro growth studies, gross pathology, pathogenicity, immunogenicity

Eimeria dispersa, illus.
Long, P. L.; Millard, B. J.; and Lawn, A. M., 1979, Ztschr. Parasitenk., v. 60 (2), 193-195
Eimeria dispersa, turkey (exper.), presence of spine-like structures attached to outside of parasitophorous vacuole of second generation schizonts within host intestinal epithelial cells

Eimeria dispersa, illus.
Madden, P. A.; and Ruff, M. D., 1979, J. Parasitol., v. 65 (2), 234-242
Eimeria spp. in turkeys, effects on structural integrity of intestinal and cecal mucosa, scanning electron microscopy, comparison of damage with parasite distribution as seen by light microscopy

Eimeria dispersa
Mitrovic, M.; Schildknecht, E. G.; and Marusich, W. L., 1979, Poultry Science, v. 58 (5), 1154-1159
Eimeria spp., turkey poults (exper.), single and mixed infections, lasalocid sodium, controlled battery trials

Eimeria dispersa
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Eimeria dunsingi Farr, 1960, illus.
Todd, K. S., jr.; Gallina, A. M.; and Schmidt, J. M., 1977, Proc. Helminth. Soc. Washington, v. 44 (2), 188-190
description
Syn.: Eimeria sp. n. Brada, 1966
Melopsittacus undulatus (feces, small intestine)

Eimeria duodenalis
Williams, R. B., 1978, Vet. Parasitol., v. 4 (2), 193-197
Pavo cristatus: Essex, Great Britain

E[imeria] ellipsoidalis
Bobkova, A. F., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 90-94
coccidia and helminths of calves, comparison of stall-enclosure and pasture maintenance, economics

Eimeria ellipsoidalis
Cotteleer, C.; and Fameree, L., 1978, Schweiz. Arch. Tierh., v. 120 (3), 149-156
Eimeria spp., cattle, rate of infestation: Belgium

Eimeria ellipsoidalis
Evplov, N. N.; and Nazarov, V. G., 1977, Veterinariia, Moskva (6), 65-66
Eimeria spp., calves, chemococcide effective; compared with biomycin and norsulfazol: Belgorodsk oblast

Eimeria ellipsoidalis
Fitzgerald, P. R.; and Mansfield, M. E., 1979, J. Parasitol., v. 65 (5), 824-825
Eimeria spp., calves (exper.), lasalocid

E[imeria] ellipsoidalis
Gobzem, V. R., 1970, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 8, 118-120
E[imeria] bovis, E. cylindrica, E. ellipsoidalis, calves, safe prophylaxis with biovetin

Eimeria ellipsoidalis
Gobzem, V. R.; and Nazarov, V. G., 1978, Veterinariia, Moskva (3), 67-69
Eimeria spp., calves, diagnostic difficulties, clinical symptoms, chemoprophylactic substances tested at various dosages and in various combinations

Eimeria ellipsoidalis
Graefner, G.; Graubmann, H. D.; and Kron, A., 1978, Monatsh. Vet.-Med., v. 33 (23), 910-912
Eimeria spp., cattle on rearing and fattening farms, intensity and extensity of infection in relation to certain environmental conditions: DDR

Eimeria ellipsoidalis
Hiepe, T.; Romeyke, D.; and Jungmann, R., 1978, Monatsh. Vet.-Med., v. 33 (23), 904-910
Eimeria spp., calves reared under conditions of industrialized cattle farming, course of infection, clinical symptoms, monthly distribution, control measures: DDR

Eimeria ellipsoidalis
Ward, J. K.; Ferguson, D. L.; and Parkhurst, A. M., 1979, J. Animal Sc., v. 49 (2), 306-309
gastrointestinal parasites, beef cows (feces), level of infection, effect of animal age and season of year: Mead, Nebraska

Eimeria environ
Wiggins, J. P.; and Rothenbacher, H., 1979, J. Parasitol., v. 65 (3), 393-394
Sylvilagus floridanus (feces): central Pennsylvania

Eimeria erythrourica Musajev et Alijeva, 1961, illus.
Ismailov, S. G., 1974, Parazitologiia, Leningrad, v. 8 (3), 261-265
Eimeria erythrourica in Meriones erythrourus (exper.), endogenous stages of life cycle

Eimeria europaea, illus.
Golemanski, V. G., 1976, Priroda, Sofiia, v. 25 (2), 64-67
Lepus europaeus: Bulgaria

Eimeria europaea
Gottschalk, C., 1973, Ang. Parasitol., v. 14 (1), 44-54
endo-parasites of Lepus europaeus, seasonal dynamics, distribution according to locality, sex and age of host, economic importance of parasitism for regional hunting: Ostthuringen, DDR

Eimeria europaea Pellerdy 1956, illus.
Romero-Rodriguez, J., 1976, Rev. Iber. Parasitol., v. 36 (1-2), 131-134
description
Lepus granatensis (fecal material from parts of intestine having macroscopic lesions): Granada, Spain

Eimeria exigua
Pav, J., 1978, Veterinarstvi, v. 28 (2), 84-86
coccidia, cestodes, prevalence in hares: Czechoslovakia

Eimeria falciformis
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Rattus norvegicus: Neusiedlerseegebiet, nordlichen Burgenland

Eimeria falciformis var. pragensis, illus.
Mesfin, G. M.; and Bellamy, J. E. C., 1978, J. Parasitol., v. 64 (4), 696-705
Eimeria falciformis var. pragensis in Mus musculus (exper.), prepatent and patent periods, description of oocysts, location in host, merogony, first- to fourth-generation schizonts, microgametogony, macrogametogony; development in Rattus norvegicus progressed only as far as mature 1st-generation schizonts; comparison with other murine Eimeria

Eimeria falciformis var pragensis, illus.
Mesfin, G. M.; and Bellamy, J. E. C., 1979, J. Parasitol., v. 65 (3), 469-471
Eimeria falciformis var pragensis, migration of sporozoites from absorptive to crypt epithelium of mouse colon

Eimeria falciformis var. pragensis
Mesfin, G. M.; and Bellamy, J. E. C., 1979, Infect. and Immun., v. 23 (1), 108-114
Eimeria falciformis var. pragensis, mice, (i) effects of immune response on life cycle, (ii) relative immunizing ability of different doses of oocysts, (iii) duration of acquired resistance; possibility that cell-mediated immune mechanism is responsible for arrest in schizogony

Eimeria falciformis var. pragensis, illus.
Mesfin, G. M.; and Bellamy, J. E. C., 1979, Infect. and Immun., v. 23 (2), 460-464
Eimeria falciformis var. pragensis, mice, thymic dependence of immunity

Eimeria falciformis var. pragensis, illus.
Mesfin, G. M.; Bellamy, J. E. C.; and Stockdale, P. H. G., 1978, Canad. J. Comp. Med., v. 42 (4), 496-510
Eimeria falciformis var. pragensis, mice (exper.), clinical signs and pathological changes, effects of varying infective dose

Eimeria faurei
Ajayi, J. A.; and Todd, A. C., 1977, Bull. Animal Health and Prod. Africa, v. 25 (3), 257-261
prevalence, prepatent period
sheep (feces): Wisconsin

Eimeria faurei
Ermatova, D. U., 1978, Veterinariia, Moskva (3), 69-70
Eimeria spp., sheep and goats, occurrence, seasonality, host age: Kashkadar'insk oblast

Eimeria faurei
Fitzgerald, P. R.; and Mansfield, M. E., 1978, Am. J. Vet. Research, v. 39 (1), 7-10
Eimeria spp., lambs (nat. and exper.), effect of monensin on oocyst discharge, feed consumption, fecal output, and weight gain (by sex of host): Illinois

Eimeria faurei (Moussu et Marotell, 1901), illus.
Gobzem, V. R., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 124-130
synonymy
[Ovis aries] (feces): Belorussia

Eimeria faurei, illus.
Golemanski, V. G., 1977, Priroda, Sofiia, v. 26 (1), 71-74
Ovis musimon: Bulgaria

Eimeria faurei
Krishnamurthy, R.; and Kshirsagar, H. S., 1976, Marathwada Univ. J. Sc. (Nat. Sc.), v. 15 (8), 153-156
Eimeria spp., goats (feces), prevalence and seasonal incidence: slaughter house at Parbhani, Mahrasthra state

Eimeria faurei Moussi, Marotel, 1905, illus.
Nukerbaeva, K. K., 1978, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (3), 16-21
Ammotragus lervia
Capra ibex sibirica
Ovis canadensis canadensis
(feces of all): all from Alma-Ata zoopark

Eimeria faurei
Samizadeh-Yazd, A.; et al., 1979, Am. J. Vet. Research, v. 40 (8), 1107-1109
Eimeria spp., lambs, efficacy of monensin and aureomycin separately and combined

Eimeria faurei
Santiago, M. A. M.; da Costa, U. C.; and da Silva, O. L., 1977, Rev. Centro Cien. Rurais, v. 7 (3), 297-301
Eimeria spp., sheep, natural infection, amprolium, furoxone, not sufficient control for sheep grazing on pasture

Eimeria faurei
Seidel, B., 1979, Zool. Garten, n. F., v. 49 (2), 131-160
Ovibos moschatus, management in zoological park, includes information on parasites and their treatment: Tierpark Berlin

Eimeria faurei
Tarazona, J. M.; and Fernandez, J. M., 1974, An. Inst. Nac. Invest. Agrar., s. Hig. y San. Animal (1), 55-62
Eimeria spp., sheep, incidence, distribution, review of clinical aspects, pathology, epizootiology: Spain

Eimeria felina
Cotteleer, C.; and Fameree, L., 1976, Rev. Med. Liege, v. 31 (23), 729-734
Eimeriidae, incidence in cats: Belgique

Eimeria ferrisi Levine and Ivens, 1956, illus.
Blagburn, B. L.; Chobotar, B.; and Smith, R. T., 1979, Ztschr. Parasitenk., v. 59 (1), 1-14
Eimeria ferrisi in Mus musculus, clinical and histologic study of actively induced resistance

Eimeria ferrisi
Klesius, P. H.; et al., 1978, Clin. Immunol. and Immunopathol., v. 10 (2), 214-221
Eimeria ferrisi, cell-mediated immunity stimulated in mice by prophylactic treatment with bovine transfer factor (TFd) prepared from lymph node lymphocytes of cattle immune to E. bovis, lymphocyte stimulation and protection against clinical infection, susceptible mice given lymphocytes from donor mice treated with bovine TFd were also partly protected against clinical infection

Eimeria ferrisi
Klesius, P. H.; et al., 1979, Clin. Immunol. and Immunopathol., v. 12 (2), 143-149
Eimeria ferrisi, C57BL/6 mice, effects of immunization and treatment with transfer factor, results suggest this host strain has genetically determined defect in cell-mediated immune response to this infection

Eimeria ferrisi
Klesius, P. H.; and Hinds, S. E., 1979, Infect. and Immun., v. 26 (3), 1111-1115
Eimeria ferrisi, comparison of susceptibility in various inbred and F_1 hybrid mouse strains and in nu/nu and nu/+ BALB/c mice, effect of treatment with rabbit anti-mouse thymocyte serum

Eimeria filamentifera sp. n., illus.
Wacha, R. S.; and Christiansen, J. L., 1979, J. Protozool., v. 26 (3), 353-354
Chelydra serpentina (intestinal contents): Shimek State Forest, Lee Co., Iowa

Eimeria flavescens Marotel & Guilhon, 1941, illus.
  Catchpole, J.; and Norton, C. C., 1979, Parasitology, v. 79 (2), 249-257
    Eimeria spp. in rabbits from 3 commercial rabbitries, species identification, level of infection, % occurrence and % predominance of different spp.: South East England

Eimeria flavescens Marotel & Guilhon, 1941, illus.
  Norton, C. C.; Catchpole, J.; and Joyner, L. P., 1979, Parasitology, v. 79 (2), 231-248
    Eimeria irresidua, E. flavescens, redescription, sporulation time, schizogony and gametogony, pathogenicity and oocyst production, immunogenicity, geographic distribution, prevalence; "the name E. hakei should be suppressed in favour of E. flavescens"
    Oryctolagus cuniculus (caecum, colon, ileum) (exper.)

Eimeria gallatii sp. n., illus.
  Straneva, J. E.; and Kelley, G. L., 1979, J. Protozool., v. 26 (4), 530-532
    Clethrionomys gapperi (feces): Brush Valley, Indiana County, Pennsylvania

Eimeria gallopavonis
  Edgar, S. A.; and Flanagan, C., 1979, Poultry Science, v. 58 (6), 1483-1489
    Eimeria spp., turkeys, halofuginone effective

Eimeria gallopavonis
  McDougald, L. R., 1979, Poultry Science, v. 58 (1), 76-80
    coccidiosis, histomoniasis, turkeys, tests for efficacy and compatibility indicate that amprolium and carbarsone can be used in combination

Eimeria gallopavonis
  McDougald, L. R.; and Johnson, J. K., 1979, Poultry Science, v. 58 (1), 72-75
    Eimeria spp., turkeys, efficacy of arprinocid compared with amprolium, floor pen studies

Eimeria gallopavonis
  McDougald, L. R.; and McQuistion, T. E., 1978, Avian Dis., v. 22 (4), 765-770
    Eimeria spp., turkeys, coccidiosis management, innate (age) resistance and acquired immunity vs. anticoccidial medication

Eimeria gallopavonis
  Mitrovic, M.; Schildknecht, E. G.; and Marusich, W. L., 1979, Poultry Science, v. 58 (5), 1154-1159
    Eimeria spp., turkey poults (exper.), single and mixed infections, lasalocid sodium, controlled battery trials

Eimeria gallopavonis
  Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
    parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Eimeria gallopavonis
  Reid, W. M.; Anderson, W. I.; and McDougald, L. R., 1978, Avian Path., v. 7 (4), 569-576
    Eimeria spp., turkey poults, anticoccidial protection and development of immunity while using monensin

Eimeria gasterostei (Thelohan)
  Lester, R. J. G., 1974, Syesis, v. 7, 195-200
    Gasterosteus aculeatus (liver): near Vancouver, British Columbia

Eimeria gaviae n. sp., illus.
  Montgomery, R. D.; Novilla, M. N.; and Shillinger, R. B., 1978, Avian Dis., v. 22 (4), 809-814
    Eimeria gaviae n. sp. in Gavia immer (kidney), pathology

Eimeria granulosa
  Ajayi, J. A.; and Todd, A. C., 1977, Bull. Animal Health and Prod. Africa, v. 25 (3), 257-261
    prevalence
    sheep (feces): Wisconsin

Eimeria granulosa (Christensen, 1938)
  Gobzem, V. R., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 124-130
    [Ovis aries] (feces): Belorussia

Eimeria granulosa
  Krishnamurthy, R.; and Kshirsagar, H. S., 1976, Marathwada Univ. J. Sc. (Nat. Sc.), v. 15 (8), 153-156
    Eimeria spp., goats (feces), prevalence and seasonal incidence: slaughter house at Parbhani, Mahrasthra state

Eimeria granulosa
  Samizadeh-Yazd, A.; et al., 1979, Am. J. Vet. Research, v. 40 (8), 1107-1109
    Eimeria spp., lambs, efficacy of monensin and aureomycin separately and combined

Eimeria granulosa
  Tarazona, J. M.; and Fernandez, J. M., 1974, An. Inst. Nac. Invest. Agrar., s. Hig. y San. Animal (1), 55-62
    Eimeria spp., sheep, incidence, distribution, review of clinical aspects, pathology, epizootiology: Spain

Eimeria grenieri, illus.
  Long, P. L.; and Millard, B. J., 1978, Parasitology, v. 76 (1), 1-9
    Eimeria grenieri in Numida meleagris (intestine, caeca) (nat. and exper.), life cycle, reproduction rate, pathogenicity (severe depression of body weight gain), immunity to reinfection, treatment with sulphaquinoxaline in drinking water and robenidine in food: Britain

Eimeria grenieri
  Long, P. L.; and Millard, B. J., 1979, Parasitology, v. 78 (2), 239-247
    Eimeria maxima, rejection by 'foreign' host (Numida meleagris); E. tenella, E. grenieri, survival of sporozoites in peritoneal macrophages from 'foreign' vs. normal hosts in vitro

Eimeria gundii n. sp., illus.
  Mishra, G. S.; and Gonzalez, J. P., 1978, Ann. Parasitol., v. 53 (2), 241-243
    Ctenodactylus gundi (intestinal mucosa): southern region of Tunisia

Eimeria hagani
  Kucera, J.; and Lom, J., 1979, Veterinarstvi, v. 29 (8), 351-353
    Eimeria spp., Heterakis gallinae, incidence at large scale poultry farms using cage breeding methods: Cechach

Eimeria hagani
  Latif, B. M. A.; Ali, S. R.; and Versenyi, L., 1974, Acta Vet. Beograd, v. 24 (3), 135-138
    Eimeria spp., chickens (feces), rate of infection: Baghdad area, Iraq

Eimeria hakei
  Norton, C. C.; Catchpole, J.; and Joyner, L. P., 1979, Parasitology, v. 79 (2), 231-248
    "the name E. hakei should be suppressed in favour of E. flavescens"

Eimeria holmesi sp. n., illus.
  Samoil, H. P.; and Samuel, W. M., 1977, Canad. J. Zool., v. 55 (10), 1671-1683
    key
    Lepus americanus (feces): near Rochester, north-central Alberta

Eimeria honessi
  Wiggins, J. P.; and Rothenbacher, H., 1979, J. Parasitol., v. 65 (3), 393-394
    Sylvilagus floridanus (feces): central Pennsylvania

Eimeria hungarica, illus.
  Golemanski, V. G., 1976, Priroda, Sofiia, v. 25 (2), 64-67
    Lepus europaeus: Bulgaria

Eimeria hungarica
  Gottschalk, C., 1973, Ang. Parasitol., v. 14 (1), 44-54
    endo-parasites of Lepus europaeus, seasonal dynamics, distribution according to locality, sex and age of host, economic importance of parasitism for regional hunting: Ostthuringen, DDR

Eimeria hypophthalmichthys Dogiel et Achmerov
  Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
    parasites of fishes: China

Eimeria illinoisensis
  Hiepe, T.; Romeyke, D.; and Jungmann, R., 1978, Monatsh. Vet.-Med., v. 33 (23), 904-910
    Eimeria spp., calves reared under conditions of industrialized cattle farming, course of infection, clinical symptoms, monthly distribution, control measures: DDR

Eimeria indentata
  Lee, E. H.; and Fernando, M. A., 1977, Ztschr. Parasitenk., v. 53 (1), 129-131
    Eimeria indentata, E. maxima, characterization of DNA in oocysts by weight and buoyant density, "E. indentata from the Malayan jungle fowl is not a species distinct from E. maxima.", suggests use of technique for making species distinctions between morphologically similar, closely related species of Coccidia

Eimeria intestinalis Cheissin, 1948
  Catchpole, J.; and Norton, C. C., 1979, Parasitology, v. 79 (2), 249-257
    Eimeria spp. in rabbits from 3 commercial rabbitries, species identification, level of infection, % occurrence and % predominance of different spp.: South East England

Eimeria intestinalis
  Coudert, P., 1977, Compt. Rend. Acad. Sc., Paris, v. 285, s. D, Sc. Nat. (8), 885-887
    Eimeria pellerdyi n. sp., E. intestinalis, pure isolates distinguished in coccidia-free rabbits on basis of duration of life cycle, site of development, and strict specificity of acquired immunity; pathogenicity
    Oryctolagus cuniculus (ileon) (exper.)

E[imeria] intestinalis
  Coudert, P.; Vaissaire, J.; and Licois, D., 1978, Rec. Med. Vet., v. 154 (5), 437-440
    E[imeria] pellerdyi, E[imeria] intestinalis, young rabbits, hematological changes

Eimeria intestinalis
  Gallazzi, D., 1977, Folia Vet. Latina, v. 7 (4), 371-380
    Eimeria spp., rabbits, cyclical variations in excretion of fecal oocysts, seasonal effects, effect of pregnancy, parturition, and lactation, removal of mother from litter, infection of litters, relationship between maternal coccidial levels and those in the young, performance

Eimeria intestinalis (Cheissin, 1948)
  Licois, D.; Coudert, P.; and Mongin, P., 1978, Ann. Recherches Vet., v. 9 (1), 1-10
    Eimeria intestinalis, E. pellerdyi, rabbits (exper.), changes in water metabolism in diarrhoeic hosts

Eimeria intestinalis
  Owen, D., 1978, Lab. Animals, v. 12 (2), 49-50
    Eimeria stiedae, E. intestinalis, standard rabbit diet heavily contaminated with oocysts, sterilization by autoclaving and irradiation was satisfactory for elimination of oocysts from diet but pelleting even at 68° was unsatisfactory

Eimeria intestinalis
  Peeters, J. E.; Halen, P.; and Meulemans, G., 1979, Brit. Vet. J., v. 135 (4), 349-354
    Eimeria spp., rabbits (nat. and exper.), efficacy of robenidine

Eimeria intestinalis
  Ryley, N. G.; and Ryley, J. F., 1978, Parasitology, v. 77 (1), 33-39
    Eimeria, 10 spp., survival and viability of oocysts following prolonged exposure to saturated sodium chloride solution, additional observations on virulence of Eimeria tenella for chickens following such exposure, no evidence obtained to contraindicate use of salt-flotation methods for separation of oocysts from feces

Eimeria intricata
  Ajayi, J. A.; and Todd, A. C., 1977, Bull. Animal Health and Prod. Africa, v. 25 (3), 257-261
    prevalence, prepatent period
    sheep (feces): Wisconsin

Eimeria intricata Spiegl, 1926, illus.
  Bhatia, B. B.; et al., 1978, Indian J. Animal Sc., v. 48 (9), 688-691
    Eimeria intricata, sheep (exper.), sporogony

Eimeria intricata
  Ermatova, D. U., 1978, Veterinariia, Moskva (3), 69-70
    Eimeria spp., sheep and goats, occurrence, seasonality, host age: Kashkadar'insk oblast

Eimeria intricata
  Fitzgerald, P. R.; and Mansfield, M. E., 1978, Am. J. Vet. Research, v. 39 (1), 7-10
    Eimeria spp., lambs (nat. and exper.), effect of monensin on oocyst discharge, feed consumption, fecal output, and weight gain (by sex of host): Illinois

Eimeria intricata (Spiegel, 1925), illus.
Gobzem, V. R., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 124-130
[Ovis aries] (feces): Belorussia

Eimeria intricata, illus.
Golemanski, V. G., 1977, Priroda, Sofiia, v. 26 (1), 71-74
Ovis musimon: Bulgaria

Eimeria intricata
Krishnamurthy, R.; and Kshirsagar, H. S., 1976, Marathwada Univ. J. Sc. (Nat. Sc.), v. 15 (8), 153-156
Eimeria spp., goats (feces), prevalence and seasonal incidence: slaughter house at Parbhani, Mahrasthra state

Eimeria intricata
McDougald, L. R.; and Dunn, W. J., 1978, Am. J. Vet. Research, v. 39 (9), 1459-1462
Eimeria spp., lambs (exper.), efficacy of monensin in feed

Eimeria intricata
Samizadeh-Yazd, A.; et al., 1979, Am. J. Vet. Research, v. 40 (8), 1107-1109
Eimeria spp., lambs, efficacy of monensin and aureomycin separately and combined

Eimeria intricata
Santiago, M. A. M.; da Costa, U. C.; and da Silva, O. L., 1977, Rev. Centro Cien. Rurais, v. 7 (3), 297-301
Eimeria spp., sheep, natural infection, amprolium, furoxone, not sufficient control for sheep grazing on pasture

Eimeria intricata
Seidel, B., 1979, Zool. Garten, n. F., v. 49 (2), 131-160
Ovibos moschatus, management in zoological park, includes information on parasites and their treatment: Tierpark Berlin

Eimeria intricata
Tarazona, J. M.; and Fernandez, J. M., 1974, An. Inst. Nac. Invest. Agrar., s. Hig. y San. Animal (1), 55-62
Eimeria spp., sheep, incidence, distribution, review of clinical aspects, pathology, epizootiology: Spain

Eimeria irresidua Kessel & Jankiewicz, 1931, illus.
Catchpole, J.; and Norton, C. C., 1979, Parasitology, v. 79 (2), 249-257
Eimeria spp. in rabbits from 3 commercial rabbitries, species identification, level of infection, % occurrence and % predominance of different spp.: South East England

Eimeria irresidua
Danek, J.; et al., 1978, Biol. a Chem. Zivoc. Vyroby, Vet., v. 14 (v. 20) (2), 151-169
Eimeria spp., rabbits (nat. and exper.), sulphacombine, controlled test, subacute toxicity, no negative effect on followed indicators

Eimeria irresidua
Gallazzi, D., 1977, Folia Vet. Latina, v. 7 (4), 371-380
Eimeria spp., rabbits, cyclical variations in excretion of fecal oocysts, seasonal effects, effect of pregnancy, parturition, and lactation, removal of mother from litter, infection of litters, relationship between maternal coccidial levels and those in the young, performance

Eimeria irresidua, illus.
Norton, C. C.; Catchpole, J.; and Joyner, L. P., 1979, Parasitology, v. 79 (2), 231-248
Eimeria irresidua, E. flavescens, redescription, sporulation time, schizogony and gametogony, pathogenicity and oocyst production, immunogenicity, geographic distribution, prevalence
Oryctolagus cuniculus (jejunum and ileum) (exper.)

Eimeria irresidua
Pav, J., 1978, Veterinarstvi, v. 28 (2), 84-86
coccidia, cestodes, prevalence in hares: Czechoslovakia

Eimeria jersenica sp. n., illus.
Davronov, O., 1973, Parazitologiia, Leningrad, v. 7 (1), 79-86
Meriones erythrourus: Chustsk, Zadar'insk, Tiurakurgansk, and Karshinsk regions, Uzbek SSR

Eimeria karschinica sp. n., illus.
Davronov, O., 1973, Parazitologiia, Leningrad, v. 7 (1), 79-86
Meriones meridianus: Guzarsk and Ul'ianovsk regions, Uzbek SSR

Eimeria keithi sp. n., illus.
Samoil, H. P.; and Samuel, W. M., 1977, Canad. J. Zool., v. 55 (10), 1671-1683
key
Lepus americanus (feces): near Rochester, north-central Alberta

Eimeria kofoidi
Zaprianov, M. Ia., 1978, Vet. Med. Nauki, v. 15 (4), 103-107
Eimeria kofoidi in Alectoris graeca cypriatis (nat. and exper.), amprolium, DOT-soluble, and Esb$_3$ tested

Eimeria kostencovi sp. n., illus.
Davronov, O., 1973, Parazitologiia, Leningrad, v. 7 (1), 79-86
Meriones meridianus: Karshinsk region, Uzbek SSR

Eimeria kotlani Graefner et Graubmann, 1964, illus.
Shibalova, T. A.; and Antukhaev, I. K., 1979, Tsitologiia, v. 21 (3), 300-303
Eimeria kotlani-infected goose intestinal epithelial cells, ultrastructure

Eimeria labbeana
Cotteleer, C.; and Fameree, L., 1978, Schweiz. Arch. Tierh., v. 120 (4), 181-187
domestic and semi-feral pigeons, survey of intestinal parasites, presence of anti-Toxoplasma antibodies in serum, possible role in epidemiology of toxoplasmosis: Belgium

Eimeria larimerensis
Broda, R. J.; and Schmidt, G. D., 1978, J. Helminth., v. 52 (4), 323-326
Spermophilus spilosoma: Colorado

Eimeria leachi
Timchenko, A. D., 1972, Parazitologiia, Leningrad, v. 6 (6), 509-512
Eimeria spp., survival of non-sporulated and sporulated oocysts on soil surface and at various depths under conditions prevailing in southern Ukraine during winter to spring period, implications for prophylaxis and control of coccidiosis

Eimeria leporis, illus.
Golemanski, V. G., 1976, Priroda, Sofiia, v. 25 (2), 64-67
Lepus europaeus: Bulgaria

Eimeria leporis
Gottschalk, C., 1973, Ang. Parasitol., v. 14 (1), 44-54
endo-parasites of Lepus europaeus, seasonal dynamics, distribution according to locality, sex and age of host, economic importance of parasitism for regional hunting: Ostthuringen, DDR

Eimeria leporis Nieschulz 1923, illus.
Hernandez Rodriguez, S.; et al., 1978, Rev. Iber. Parasitol., v. 38 (3-4), 515-526
Eimeria leporis, life cycle in Lepus capensis (exper.), electron microscopy

Eimeria leporis
Pav, J., 1978, Veterinarstvi, v. 28 (2), 84-86
coccidia, cestodes, prevalence in hares: Czechoslovakia

Eimeria leporis Nieschulz, 1923, illus.
Romero-Rodriguez, J., 1976, Rev. Iber. Parasitol., v. 36 (1-2), 131-134
description
Lepus granatensis (fecal material from parts of intestine having macroscopic lesions): Granada, Spain

Eimeria leporis Nieschulz 1923, illus.
Samoil, H. P.; and Samuel, W. M., 1977, Canad. J. Zool., v. 55 (10), 1671-1683
redescription, key
Lepus americanus (feces): near Rochester, north-central Alberta

Eimeria leuckarti, illus.
Bemrick, W. J.; O'Leary, T. P.; and Barnes, D. M., 1979, Vet. Med. and Small Animal Clin., v. 74 (1), 77-80
Eimeria leuckarti, horses, 5 necropsy reports, possible pathogenic role: Minnesota

Eimeria leuckarti
Chineme, C. N.; Tulpule, S. S.; and Jamdar, M. N., 1979, Vet. Rec., v. 105 (6), 126
Eimeria leuckarti, donkeys (jejunum, ileum), enteritis: Nigeria

Eimeria leuckarti, illus.
Tomanovic, B., 1977, Vet. Glasnik, v. 31 (4), 281-285
morphology, biology: Yugoslavia

Eimeria leucuri sp. n., illus.
Stabler, R. M.; et al., 1979, J. Parasitol., v. 65 (2), 272-274
Lagopus leucurus (feces): Alpine meadows of Colorado Rockies

Eimeria mccordocki
Woolf, A.; and Harder, J. D., 1979, Wildlife Monogr. (67), 53 pp.
Odocoileus virginianus: Rachelwood Wildlife Research Preserve, New Florence, Pennsylvania

Eimeria madisonensis
Woolf, A.; and Harder, J. D., 1979, Wildlife Monogr. (67), 53 pp.
Odocoileus virginianus: Rachelwood Wildlife Research Preserve, New Florence, Pennsylvania

Eimeria magna Perard, 1925, illus.
Catchpole, J.; and Norton, C. C., 1979, Parasitology, v. 79 (2), 249-257
Eimeria spp. in rabbits from 3 commercial rabbitries, species identification, level of infection, % occurrence and % predominance of different spp.: South East England

Eimeria magna
Danek, J.; et al., 1978, Biol. a Chem. Zivoc. Vyroby, Vet., v. 14 (v. 20) (2), 151-169
Eimeria spp., rabbits (nat. and exper.), sulphacombine, controlled test, subacute toxicity, no negative effect on followed indicators

Eimeria magna
Gallazzi, D., 1977, Folia Vet. Latina, v. 7 (4), 371-380
Eimeria spp., rabbits, cyclical variations in excretion of fecal oocysts, seasonal effects, effect of pregnancy, parturition, and lactation, removal of mother from litter, infection of litters, relationship between maternal coccidial levels and those in the young, performance

Eimeria magna
Pav, J., 1978, Veterinarstvi, v. 28 (2), 84-86
coccidia, cestodes, prevalence in hares: Czechoslovakia

Eimeria magna
Peeters, J. E.; Halen, P.; and Meulemans, G., 1979, Brit. Vet. J., v. 135 (4), 349-354
Eimeria spp., rabbits (nat. and exper.), efficacy of robenidine

Eimeria magna
Ryley, N. G.; and Ryley, J. F., 1978, Parasitology, v. 77 (1), 33-39
Eimeria, 10 spp., survival and viability of oocysts following prolonged exposure to saturated sodium chloride solution, additional observations on virulence of Eimeria tenella for chickens following such exposure, no evidence obtained to contraindicate use of salt-flotation methods for separation of oocysts from feces

Eimeria magna, illus.
Speer, C. A., 1979, J. Parasitol., v. 65 (4), 591-598
Eimeria magna, development of gamonts and oocysts in cell cultures inoculated with merozoites, extent of sporulation and infectiousness of oocysts developing in such cultures

Eimeria maior
Wiggins, J. P.; and Rothenbacher, H., 1979, J. Parasitol., v. 65 (3), 393-394
Sylvilagus floridanus (feces): central Pennsylvania

Eimeria malabaricas sp. n., illus.
Chowattukunnel, J. T., 1979, J. Protozool., v. 26 (1), 36-38
Funambulus tristriatus (feces): Kottayam district, Kerala State, India

Eimeria marconii sp. n., illus.
Straneva, J. E.; and Kelley, G. L., 1979, J. Protozool., v. 26 (4), 530-532
Clethrionomys gapperi (feces): Brush Valley, Indiana County, Pennsylvania

E[imeria] maxima
Bedrnik, P., 1977, Veterinarstvi, v. 27 (10), 458-459
coccidiosis, chickens, current prevalence, amprol plus treatment

Eimeria maxima
Bedrnik, P.; et al., 1979, Arch. Geflugelk., v. 43 (1), 7-10
Eimeria spp., chickens (exper.), comparative efficiency of anticoccidials in combination with growth promotant nitrovin

Eimeria maxima
Chapman, H. D., 1978, Avian Path., v. 7 (2), 269-277
Eimeria maxima, E. brunetti, E. tenella, chickens, effect of monensin on development of immunity acquired by repeated low-level infections

Eimeria maxima
Chapman, H. D., 1978, Ztschr. Parasitenk., v. 56 (2), 115-121
Eimeria spp., excystation, roles of various enzymes

Eimeria maxima
Chapman, H. D., 1979, Avian Path., v. 8 (2), 181-186
Eimeria maxima in chickens (exper.), sensitivity to monensin of field isolates from monensin-exposed vs. unexposed sites, low level resistance may have developed as a result of previous drug exposure

Eimeria maxima
Chappel, L. R., 1979, J. Parasitol., v. 65 (1), 137-143
Eimeria spp., chicks, site of action of salinomycin

Eimeria maxima
Chappel, L. R.; and Babcock, W. E., 1979, Poultry Science, v. 58 (2), 304-307
Eimeria spp., broilers (exper.), salinomycin, monensin, lasalocid, drug toleration and anticoccidial efficacy compared in 5 field trials, commercial facilities

Eimeria maxima
Cruthers, L. R.; et al., 1978, Poultry Science, v. 57 (5), 1227-1233
Eimeria spp., broiler chicks (exper.), lonomycin in feed, high degree of anticoccidial activity, comparison with lasalocid and monensin

Eimeria maxima
Edgar, S. A.; and Flanagan, C., 1979, Poultry Science, v. 58 (6), 1469-1475
Eimeria spp. (recent field isolates resistant to various drugs), chickens, halofuginone, action cidal rather than static

Eimeria maxima
Edgar, S. A.; and Flanagan, C., 1979, Poultry Science, v. 58 (6), 1476-1482
Eimeria spp. (recent field isolates resistant to various drugs), halofuginone with roxarsone and/or bacitracin MD

Eimeria maxima
Greuel, E.; and Kuehnhold, W., 1977, Prakt. Tierarzt, v. 58 (5), 338-341
Eimeria spp., chickens (exper.), synergistic effect of metichlorpindol and methylbenzoquate, rotation program with other coccidiostats discussed

Eimeria maxima
Ishii, T.; and Onaga, H., 1978, Nippon Zyuisi-Kai Zassi (J. Japan Vet. Med. Ass.), v. 31 (9), 534-538
Eimeria spp., chicks (exper.) and cell cultures, activity of chlorobromofebrifugine

Eimeria maxima
Joyner, L. P.; and Norton, C. C., 1978, Parasitology, v. 76 (3), 369-377
Eimeria maxima, activity of methyl benzoquate and clopidol, synergy shown to be supra-additive, collateral sensitivity could not be demonstrated in resistant lines, effect of Lerbek against standard and drug-resistant lines, resistance transfer experiments with clopidol- and methyl benzoquate-resistant lines, preparation of bi-resistant lines, attempts to develop Lerbek-resistant strain

Eimeria maxima
Kellogg, F. E.; Doster, G. L.; and Johnson, J. K., 1971, J. Wildlife Dis., v. 7 (3), 186-187
Indian red junglefowl, pen-raised (small intestine)

Eimeria maxima
Kilgore, R. L.; et al., 1978, Poultry Science, v. 57 (4), 907-911
Eimeria spp., broiler chickens (exper.), arprinocid, effects on weight, feed efficiency, and histological lesions

Eimeria maxima
Kilgore, R. L.; et al., 1979, Poultry Science, v. 58 (1), 67-71
Eimeria spp., chickens, floor-pen trials evaluating 4 methods of induced exposure to coccidiosis suitable for use in drug research operations, laboratory sporulated oocysts spread over litter most satisfactory method

Eimeria maxima
Kucera, J.; and Lom, J., 1979, Veterinarstvi, v. 29 (8), 351-353
Eimeria spp., Heterakis gallinae, incidence at large scale poultry farms using cage breeding methods: Cechach

Eimeria maxima
Kutzer, E.; et al., 1979, Wien. Tierarztl. Monatsschr., v. 66 (6-7), 197-202
Eimeria spp., broiler chickens, arprinocid, drug efficacy compared with monensin

Eimeria maxima
Latif, B. M. A.; Ali, S. R.; and Versenyi, L., 1974, Acta Vet. Beograd, v. 24 (3), 135-138
Eimeria spp., chickens (feces), rate of infection: Baghdad area, Iraq

Eimeria maxima
Lee, E. H.; and Fernando, M. A., 1977, Ztschr. Parasitenk., v. 53 (1), 129-131
Eimeria indentata, E. maxima, characterization of DNA in oocysts by weight and buoyant density, "E. indentata from the Malayan jungle fowl is not a species distinct from E. maxima.", suggests use of technique for making species distinctions between morphologically similar, closely related species of Coccidia

Eimeria maxima
Lee, E. H.; and Fernando, M. A., 1978, J. Parasitol., v. 64 (3), 483-485
Eimeria maxima, chickens, single sporocyst infections give rise to infective oocysts and confer partial protective immunity, results suggest that sporozoites of this species are probably sexually undifferentiated

Eimeria maxima
Lee, E. H.; and Onderka, D. K., 1978, Canad. Vet. J., v. 19 (9), 237-243
Eimeria spp., chickens, incidence of clinical coccidiosis by month of year and age of host in the Ontario Veterinary Services Branch records 1973-1977, possible use of data to synchronize drug rotation with change in incidence

E[imeria] maxima
Long, P. L.; and Millard, B. J., 1978, Avian Path., v. 7 (3), 373-381
coccidiosis, broiler chickens, effect on oocyst output of various treatment regimens

Eimeria maxima
Long, P. L.; and Millard, B. J., 1979, Parasitology, v. 78 (2), 239-247
Eimeria maxima, rejection by 'foreign' host (Numida meleagris); E. tenella, E. grenieri, survival of sporozoites in peritoneal macrophages from 'foreign' vs. normal hosts in vitro

Eimeria maxima
Long, P. L.; and Millard, B. J., 1979, Parasitology, v. 79 (3), 451-457
Eimeria maxima, immunological differences between laboratory strains and field isolates effect of mixed immunizing inoculum on heterologous challenge

Eimeria maxima
Long, P. L.; and Millard, B. J., 1979, Avian Path., v. 8 (3), 213-228
Eimeria spp., young chickens kept in litter pens, immunization, response to challenge with homologous and heterologous strains, effect of host age and of immunizing dose, timing of onset of immunity, longevity of immunity

Eimeria maxima
Long, P. L.; Millard, B. J.; and Smith, K. M., 1979, Avian Path., v. 8 (4), 453-467
Eimeria spp., chickens, effect of 4 anticoccidial drugs on development of immunity, field and laboratory conditions

Eimeria maxima
Long, P. L.; and Rowell, J. G., 1975, Brit. Poultry Sc., v. 16 (6), 583-592
Eimeria spp., method of sampling surface litter of commercial broiler houses for laboratory estimation of numbers of coccidial oocysts

Eimeria maxima
McDougald, L. R.; Karlsson, T.; and Reid, W. M., 1979, Avian Dis., v. 23 (4), 999-1005
coccidiosis, chickens (exper.), natural outbreak of infectious bursal disease (IBD) during comparison of anticoccidials for their effect on development of immunity, interaction between diseases, immunity to coccidiosis not blocked by IBD

Eimeria maxima
Major, J. R., jr.; and Ruff, M. D., 1978, Exper. Parasitol., v. 45 (2), 234-240
Eimeria spp., effects of coccidiosis on digestive capacity (amylase) of broiler chickens, changes in pancreatic, luminal, and surface-bound amylolytic activity, reduction in amylolytic activity as pH went below 5.0

Eimeria maxima
Major, J. R., jr.; and Ruff, M. D., 1978, J. Parasitol., v. 64 (4), 706-711
Eimeria spp.-infected broilers, reduced disaccharidase activity in region of intestine with maximum infection, this reduction is related to both time and severity of infection and can contribute to overall reduction in nutrient absorption

Eimeria maxima
Maxey, B. W.; and Page, R. K., 1977, Poultry Science, v. 56 (6), 1909-1913
Eimeria brunetti, E. maxima, chickens, lincomycin feed medication for control of necrotic enteritis

Eimeria maxima
Migaki, T. T.; Chappel, L. R.; and Babcock, W. E., 1979, Poultry Science, v. 58 (5), 1192-1196
Eimeria spp., chicks (exper.), salinomycin, monensin, lasalocid, efficacy in battery trials

Eimeria maxima
Morrison, W. D.; Ferguson, A. E.; and Leeson, S., 1979, Poultry Science, v. 58 (5), 1160-1166
Eimeria spp., chicks (exper.), salinomycin and stenorol compared with other anticoccidials, efficacy and effect on chick performance

Eimeria maxima, illus.
Norton, C. C.; and Joyner, L. P., 1978, Parasitology, v. 77 (3), 243-248
Eimeria maxima, development of resistance to Lerbek, appearance of and subsequent selection for abnormal bisporocystic oocysts

Eimeria maxima
Oikawa, H.; and Kawaguchi, H., 1975, Kiseichugaku Zasshi (Japan. J. Parasitol.), v. 24 (5), 276-283
Eimeria maxima, E. necatrix, chickens, effect of single vs. repeated vs. successive infections on manifestation of symptoms, food intake and body weight gain, and oocyst production

Eimeria maxima
Olson, G.; et al., 1978, Poultry Science, v. 57 (5), 1245-1250
Eimeria spp. field isolates, chickens (exper.), arprinocid in comparison trials with marketed drugs, effective against all isolates tested including those refractory to many of the other products

Eimeria maxima
Onaga, H.; et al., 1978, Nippon Zyuisi-Kai Zassi (J. Japan Vet. Med. Ass.), v. 31 (10), 592-596
Eimeria spp., chicks and cell cultures, monensin

Eimeria maxima, illus.
  Pittilo, R. M.; and Ball, S. J., 1979, Parasitology, v. 79 (2), 259-265
    Eimeria maxima, fine structure of developing macrogamete

E[imeria] maxima
  Pujic, P.; Krecov, M.; and Simic, V., 1977, Vet. Glasnik, v. 31 (4), 277-280
    E[imeria] spp., chickens, stenorol prevents coccidiosis under laboratory and field conditions, during fattening period

Eimeria maxima
  Rollinson, D.; Joyner, L. P.; and Norton, C. C., 1979, Parasitology, v. 78 (3), 361-367
    Eimeria maxima (Weybridge) and E. maxima (indentata) were distinguished by electrophoretic mobility of phosphoglucomutase, this enzyme was used as marker to detect genetic transfer of methyl benzoquate resistance between resistant and sensitive lines of these parasites

Eimeria maxima
  Rose, M. E.; and Hesketh, P., 1979, Infect. and Immun., v. 26 (2), 630-637
    Eimeria spp. infections in normal animals vs. in animals with functional deficiencies in either T-lymphocytes or B-lymphocytes

Eimeria maxima, illus.
  Rose, M. E.; Hesketh, P.; and Ogilvie, B. M., 1979, Immunology, v. 36 (1), 71-79
    Eimeria maxima, chickens, E. nieschulzi, rats, primary and secondary infections, E. maxima-immunized chickens challenged with E. acervulina: peripheral blood leucocyte response, correlation with resistance to reinfection

Eimeria maxima
  Ruff, M. D., 1978, Proc. 1978 Maryland Nutrition Conf. Feed Mfr. (University of Maryland, Mar. 16-17), 32-37
    Eimeria spp., chickens, anticoccidials, safe withdrawal times, effect on nutrient malabsorption

Eimeria maxima
  Ruff, M. D.; et al., 1978, Avian Dis., v. 22 (1), 32-41
    Eimeria spp., broilers (exper.), arprinocid, drug efficacy in both battery and floor-pen trials

Eimeria maxima
  Ruff, M. D.; et al., 1979, Poultry Science, v. 58 (2), 298-303
    Eimeria spp., battery raised broilers (exper.), narasin compared with monensin

Eimeria maxima
  Ruff, M. D.; Anderson, W. I.; and Reid, W. M., 1978, J. Parasitol., v. 64 (2), 306-311
    Eimeria spp. in broilers, arprinocid decreased number of oocysts produced, fewer of the oocysts sporulated, and those oocysts which did sporulate were less infective than those from unmedicated birds

Eimeria maxima
  Ruff, M. D.; Wyatt, R. D.; and Witlock, D. R., 1978, J. Parasitol., v. 64 (1), 23-26
    Eimeria spp. in broilers, severe infection increased prothrombin times compared with uninfected birds

Eimeria maxima
  Ryley, J. F.; and Hardman, L., 1978, Parasitology, v. 76 (1), 11-20
    Eimeria spp., chicks (exper.), effects of dietary vitamin K on severity of disease with particular attention to effects of vitamin K on response to anticoccidial drugs, concluded that use of vitamin K deficient diet for experimental work is quite justified

Eimeria maxima
  Ryley, N. G.; and Ryley, J. F., 1978, Parasitology, v. 77 (1), 33-39
    Eimeria, 10 spp., survival and viability of oocysts following prolonged exposure to saturated sodium chloride solution, additional observations on virulence of Eimeria tenella for chickens following such exposure, no evidence obtained to contraindicate use of salt-flotation methods for separation of oocysts from feces

Eimeria maxima
  Schindler, P.; et al., 1979, Poultry Science, v. 58 (1), 23-27
    Eimeria spp., broiler chicken pen trials, arprinocid in feed highly effective prophylaxis, comparison with halofuginone, monensin, nicarbazin, and pancoxin: England; France; Germany

Eimeria maxima
  Tamas, T.; et al., 1978, Poultry Science, v. 57 (2), 381-385
    Eimeria acervulina, E. maxima, E. necatrix, E. tenella, arprinocid and dichloro analog L-628,914 in feed, decrease in oocysts' capacity to sporulate

Eimeria maxima var. indentata
  Joyner, L. P.; et al., 1978, Parasitology, v. 77 (1), 27-31
    "The term 'variety' is considered unnecessary: it is proposed that the above parasites should be given the status of strains. Parasites previously described as E. maxima var. indentata for example, would be better referred to as E. maxima (indentata)."

Eimeria media Kessel, 1929, illus.
  Catchpole, J.; and Norton, C. C., 1979, Parasitology, v. 79 (2), 249-257
    Eimeria spp. in rabbits from 3 commercial rabbitries, species identification, level of infection, % occurrence and % predominance of different spp.: South East England

Eimeria media
  Danek, J.; et al., 1978, Biol. a Chem. Zivoc. Vyroby, Vet., v. 14 (v. 20) (2), 151-169
    Eimeria spp., rabbits (nat. and exper.), sulphacombine, controlled test, subacute toxicity, no negative effect on followed indicators

Eimeria media
  Gallazzi, D., 1977, Folia Vet. Latina, v. 7 (4), 371-380
    Eimeria spp., rabbits, cyclical variations in excretion of fecal oocysts, seasonal effects, effect of pregnancy, parturition, and lactation, removal of mother from litter, infection of litters, relationship between maternal coccidial levels and those in the young, performance

Eimeria media
Peeters, J. E.; Halen, P.; and Meulemans, G., 1979, Brit. Vet. J., v. 135 (4), 349-354
Eimeria spp., rabbits (nat. and exper.), efficacy of robenidine

Eimeria meleagrimitis Tyzzer, illus.
Augustine, P. C.; and Doran, D. J., 1978, J. Protozool., v. 25 (1), 82-86
Eimeria meleagrimitis, development from sporozoites and merozoites in turkey kidney cell cultures

Eimeria meleagrimitis
Augustine, P. C.; and Thomas, O. P., 1979, Avian Dis., v. 23 (4), 854-862
Eimeria meleagrimitis, turkeys (exper.), reduced feed consumption and weight gains, blood and organ changes

Eimeria meleagrimitis
Edgar, S. A.; and Flanagan, C., 1979, Poultry Science, v. 58 (6), 1483-1489
Eimeria spp., turkeys, halofuginone effective

Eimeria meleagrimitis
Hamet-Foure, N.; Macar, C.; and Robin, B., 1979, Avian Path., v. 8 (1), 107-113
Eimeria meleagrimitis, E. adenoeides, turkeys, activity of clopidol with methylbenzoquate and amprolium with ethopabate: France

Eimeria meleagrimitis
McDougald, L. R., 1979, Poultry Science, v. 58 (1), 76-80
coccidiosis, histomoniasis, turkeys, tests for efficacy and compatibility indicate that amprolium and carbarsone can be used in combination

Eimeria meleagrimitis
McDougald, L. R.; and Johnson, J. K., 1979, Poultry Science, v. 58 (1), 72-75
Eimeria spp., turkeys, efficacy of arprinocid compared with amprolium, floor pen studies

Eimeria meleagrimitis
McDougald, L. R.; and McQuistion, T. E., 1978, Avian Dis., v. 22 (4), 765-770
Eimeria spp., turkeys, coccidiosis management, innate (age) resistance and acquired immunity vs. anticoccidial medication

Eimeria meleagrimitis, illus.
Madden, P. A.; and Ruff, M. D., 1979, J. Parasitol., v. 65 (2), 234-242
Eimeria spp. in turkeys, effects on structural integrity of intestinal and cecal mucosa, scanning electron microscopy, comparison of damage with parasite distribution as seen by light microscopy

Eimeria meleagrimitis
Mitrovic, M.; Schildknecht, E. G.; and Marusich, W. L., 1979, Poultry Science, v. 58 (5), 1154-1159
Eimeria spp., turkey poults (exper.), single and mixed infections, lasalocid sodium, controlled battery trials

Eimeria meleagrimitis
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Eimeria meleagrimitis
Reid, W. M.; Anderson, W. I.; and McDougald, L. R., 1978, Avian Path., v. 7 (4), 569-576
Eimeria spp., turkey poults, anticoccidial protection and development of immunity while using monensin

Eimeria meleagrimitis
Weber, L. J., 1978, Canad. Vet. J., v. 19 (1), 21 [Letter]
Eimeria meleagrimitis, E. adenoides, turkeys, outbreak causing great economic loss despite anticoccidial drug in medicated feed; need for recognizing development of resistant strains of coccidia: Beamsville, Ontario

Eimeria merlangi sp. nov., illus.
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
Odontogadus merlangus euxinus (intestinal wall, gall bladder): Black Sea (region near Sevastopol)

E[imeria] mitis
Balauca, N., 1978, Arch. Vet., Inst. Cercet. Vet. si Bioprep. Pasteur, v. 13, 127-141
experimental reproduction of necrotic enteritis in chicks with mixed infections of Clostridium perfringens and coccidia

Eimeria mitis
Khovanskikh, A. E., 1977, Sel'skokhoz. Biol., v. 12 (2), 245-250
Eimeria tenella, E. mitis, biochemical characteristics of pathogenesis in chickens, review

Eimeria mitis
Kucera, J.; and Lom, J., 1979, Veterinarstvi, v. 29 (8), 351-353
Eimeria spp., Heterakis gallinae, incidence at large scale poultry farms using cage breeding methods: Cechach

Eimeria mitis
Latif, B. M. A.; Ali, S. R.; and Versenyi, L., 1974, Acta Vet. Beograd, v. 24 (3), 135-138
Eimeria spp., chickens (feces), rate of infection: Baghdad area, Iraq

Eimeria mitis
Musaev, M. A.; and Elchiev, Ia. Ia., 1970, Parazitologiia, Leningrad, v. 4 (5), 494-500
Eimeria tenella, E. mitis, chickens, serum protein changes over course of infection

Eimeria mitis
Musaev, M. A.; and Ibragimova, G. G., 1974, Parazitologiia, Leningrad, v. 8 (1), 70-73
Eimeria tenella, E. mitis, chickens (exper.), changes in lecithin content of blood serum at different stages of infection, extent of biochemical changes depends on pathogenicity of different species and host age

Eimeria mitis
Musaev, M. A.; and Surkova, A. M., 1972, Parazitologiia, Leningrad, v. 6 (1), 11-15
Eimeria tenella-, E. mitis-infected chickens (exper.), changes in activity of alkaline and acid phosphatases of small intestine depend on species of coccidia, age of host, and stage of infection

Eimeria mitis
  Oikawa, H.; and Kawaguchi, H., 1976, Kiseichugaku Zasshi (Japan. J. Parasitol.), v. 25 (3), 141-147
    Eimeria praecox, E. mitis, chickens, single, repeated or successive infection, effect on manifestation of symptoms and oocyst production

Eimeria mitis
  Shirley, M. W., 1979, Parasitology, v. 78 (2), 221-237
    Eimeria mivati, enzyme electrophoresis, cross-immunity, and other studies suggest that confusion over taxonomic status arose because some cultures were contaminated with E. acervulina; possibility exists that E. mivati and E. mitis are same species; E. diminuta or E. acervulina var. diminuta "should . . . be referred to as E. mivati (diminuta)."

Eimeria mitis Tyzzer, 1929
  Surkova, A. M., 1972, Parazitologiia, Leningrad, v. 6 (2), 171-175
    Eimeria tenella, E. mitis, chickens (exper.), changes in total, residual, and protein nitrogen content in liver, depends on stage of development of parasite, host age, and species of Coccidia

Eimeria mivati
  Chappel, L. R.; and Babcock, W. E., 1979, Poultry Science, v. 58 (2), 304-307
    Eimeria spp., broilers (exper.), salinomycin, monensin, lasalocid, drug toleration and anticoccidial efficacy compared in 5 field trials, commercial facilities

Eimeria mivati
  Edgar, S. A.; and Flanagan, C., 1979, Poultry Science, v. 58 (6), 1469-1475
    Eimeria spp. (recent field isolates resistant to various drugs), chickens, halofuginone, action cidal rather than static

Eimeria mivati
  Kellogg, F. E.; Doster, G. L.; and Johnson, J. K., 1971, J. Wildlife Dis., v. 7 (3), 186-187
    Indian red junglefowl, pen-raised (small intestine)

Eimeria mivati
  Kucera, J.; and Lom, J., 1979, Veterinarstvi, v. 29 (8), 351-353
    Eimeria spp., Heterakis gallinae, incidence at large scale poultry farms using cage breeding methods: Cechach

Eimeria mivati
  Long, P. L.; and Millard, B. J., 1979, Avian Path., v. 8 (3), 213-228
    Eimeria spp., young chickens kept in litter pens, immunization, response to challenge with homologous and heterologous strains, effect of host age and of immunizing dose, timing of onset of immunity, longevity of immunity

Eimeria mivati
  Long, P. L.; Millard, B. J.; and Smith, K. M., 1979, Avian Path., v. 8 (4), 453-467
    Eimeria spp., chickens, effect of 4 anticoccidial drugs on development of immunity, field and laboratory conditions

Eimeria mivati
  McDougald, L. R.; Karlsson, T.; and Reid, W. M., 1979, Avian Dis., v. 23 (4), 999-1005
    coccidiosis, chickens (exper.), natural outbreak of infectious bursal disease (IBD) during comparison of anticoccidials for their effect on development of immunity, interaction between diseases, immunity to coccidiosis not blocked by IBD

Eimeria mivati
  Migaki, T. T.; Chappel, L. R.; and Babcock, W. E., 1979, Poultry Science, v. 58 (5), 1192-1196
    Eimeria spp., chicks (exper.), salinomycin, monensin, lasalocid, efficacy in battery trials

Eimeria mivati, illus.
  Novilla, M. N.; and Medina, C. S., 1972, Philippine J. Vet. Med., v. 11 (2), 28-44
    Eimeria mivati, young chickens (exper.); pathology

Eimeria mivati
  Ruff, M. D.; et al., 1978, Avian Dis., v. 22 (1), 32-41
    Eimeria spp., broilers (exper.), arprinocid, drug efficacy in both battery and floor-pen trials

Eimeria mivati
  Ruff, M. D.; et al., 1979, Poultry Science, v. 58 (2), 298-303
    Eimeria spp., battery raised broilers (exper.), narasin compared with monensin

Eimeria mivati
  Ruff, M. D.; Anderson, W. I.; and Reid, W. M., 1978, J. Parasitol., v. 64 (2), 306-311
    Eimeria spp. in broilers, arprinocid decreased number of oocysts produced, fewer of the oocysts sporulated, and those oocysts which did sporulate were less infective than those from unmedicated birds

Eimeria mivati
  Ryley, J. F.; and Hardman, L., 1978, J. Parasitol., v. 64 (5), 878-881
    Eimeria acervulina, E. mivati, speciation studies (cross-immunity and drug resistance studies), some immunological relationship was demonstrated but the failure of the 2 organisms to interbreed in the drug resistance studies lends support to status of E. mivati as distinct species

Eimeria mivati
  Shirley, M. W., 1978, Ztschr. Parasitenk., v. 57 (1), 83-87
    Eimeria mivati and E. mivati var. diminuta strains differing in sensitivity to sulphaquinoxaline and electrophoretic mobility of lactate dehydrogenase crossed; electrophoretic variation of enzymes a further marker for genetic studies

Eimeria mivati
  Shirley, M. W., 1979, Avian Path., v. 8 (4), 469-475
    Eimeria mivati, chickens (exper.), 3 strains (2 chicken-maintained, 1 embryo-adapted), pathogenicity compared with E. acervulina, cross-protection between virulent and attenuated strains

Eimeria mivati
Shirley, M. W., 1979, Parasitology, v. 78 (2), 221-237
Eimeria mivati, enzyme electrophoresis, cross-immunity, and other studies suggest that confusion over taxonomic status arose because some cultures were contaminated with E. acervulina; possibility exists that E. mivati and E. mitis are same species; E. diminuta or E. acervulina var. diminuta "should . . . be referred to as E. mivati (diminuta)."

Eimeria mivati var. diminuta
Shirley, M. W., 1978, Ztschr. Parasitenk., v. 57 (1), 83-87
Eimeria mivati and E. mivati var. diminuta strains differing in sensitivity to sulphaquinoxaline and electrophoretic mobility of lactate dehydrogenase crossed; electrophoretic variation of enzymes a further marker for genetic studies

Eimeria (?) mucosa (Blanchard, 1885) nov. comb.
Levine, N. D., 1979, Ann. Trop. Med. and Parasitol., v. 73 (1), 91-92
Balbiania mucosa, descriptive history, erroneously indicated as Sarcocystis; "Until it is studied further, its correct name should be considered Eimeria (?) mucosa (Blanchard, 1885) nov. comb."

Eimeria mustelae
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Mustela nivalis (Jejunum, Rectum): Neusiedlerseegebiet, nordlichen Burgenland

Eimeria myotis
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Myotis myotis (Jejunum): Neusiedlerseegebiet, nordlichen Burgenland

E[imeria] necatrix
Balauca, N., 1978, Arch. Vet., Inst. Cercet. Vet. si Bioprep. Pasteur, v. 13, 127-141
experimental reproduction of necrotic enteritis in chicks with mixed infections of Clostridium perfringens and coccidia

E[imeria] necatrix
Bedrnik, P., 1977, Veterinarstvi, v. 27 (10), 458-459
coccidiosis, chickens, current prevalence, amprol plus treatment

imeria necatrix
Bedrnik, P.; et al., 1979, Arch. Geflugelk., v. 43 (1), 7-10
Eimeria spp., chickens (exper.), comparative efficiency of anticoccidials in combination with growth promotant nitrovin

imeria necatrix
Chappel, L. R.; and Babcock, W. E., 1979, Poultry Science, v. 58 (2), 304-307
Eimeria spp., broilers (exper.), salinomycin, monensin, lasalocid, drug toleration and anticoccidial efficacy compared in 5 field trials, commercial facilities

imeria necatrix
Cruthers, L. R.; et al., 1978, Poultry Science, v. 57 (5), 1227-1233
Eimeria spp., broiler chicks (exper.), lonomycin in feed, high degree of anticoccidial activity, comparison with lasalocid and monensin

Eimeria necatrix
Edgar, S. A.; and Flanagan, C., 1979, Poultry Science, v. 58 (6), 1469-1475
Eimeria spp. (recent field isolates resistant to various drugs), chickens, halofuginone, action cidal rather than static

Eimeria necatrix
Greuel, E.; and Kuehnhold, W., 1977, Prakt. Tierarzt, v. 58 (5), 338-341
Eimeria spp., chickens (exper.), synergistic effect of metichlorpindol and methylbenzoquate, rotation program with other coccidiostats discussed

Eimeria necatrix
Kilgore, R. L.; et al., 1978, Poultry Science, v. 57 (4), 907-911
Eimeria spp., broiler chickens (exper.), arprinocid, effects on weight, feed efficiency, and histological lesions

Eimeria necatrix
Kilgore, R. L.; et al., 1979, Poultry Science, v. 58 (1), 67-71
Eimeria spp., chickens, floor-pen trials evaluating 4 methods of induced exposure to coccidiosis suitable for use in drug research operations, laboratory sporulated oocysts spread over litter most satisfactory method

Eimeria necatrix
Kucera, J.; and Lom, J., 1979, Veterinarstvi, v. 29 (8), 351-353
Eimeria spp., Heterakis gallinae, incidence at large scale poultry farms using cage breeding methods: Cechach

Eimeria necatrix
Kutzer, E.; et al., 1979, Wien. Tierarztl. Monatsschr., v. 66 (6-7), 197-202
Eimeria spp., broiler chickens, arprinocid, drug efficacy compared with monensin

Eimeria necatrix
Latif, B. M. A.; Ali, S. R.; and Versenyi, L., 1974, Acta Vet. Beograd, v. 24 (3), 135-138
Eimeria spp., chickens (feces), rate of infection: Baghdad area, Iraq

Eimeria necatrix
Lee, E. H.; and Onderka, D. K., 1978, Canad. Vet. J., v. 19 (9), 237-243
Eimeria spp., chickens, incidence of clinical coccidiosis by month of year and age of host in the Ontario Veterinary Services Branch records 1973-1977, possible use of data to synchronize drug rotation with change in incidence

Eimeria necatrix
Long, P. L.; and Millard, B. J., 1979, Avian Path., v. 8 (3), 213-228
Eimeria spp., young chickens kept in litter pens, immunization, response to challenge with homologous and heterologous strains, effect of host age and of immunizing dose, timing of onset of immunity, longevity of immunity

Eimeria necatrix
Long, P. L.; Millard, B. J.; and Smith, K. M., 1979, Avian Path., v. 8 (4), 453-467
Eimeria spp., chickens, effect of 4 anticoccidial drugs on development of immunity, field and laboratory conditions

Eimeria necatrix
McDougald, L. R.; Karlsson, T.; and Reid, W. M., 1979, Avian Dis., v. 23 (4), 999-1005
coccidiosis, chickens (exper.), natural outbreak of infectious bursal disease (IBD) during comparison of anticoccidials for their effect on development of immunity, interaction between diseases, immunity to coccidiosis not blocked by IBD

Eimeria necatrix
Major, J. R., jr.; and Ruff, M. D., 1978, Exper. Parasitol., v. 45 (2), 234-240
Eimeria spp., effects of coccidiosis on digestive capacity (amylase) of broiler chickens, changes in pancreatic, luminal, and surface-bound amylolytic activity, reduction in amylolytic activity as pH went below 5.0

Eimeria necatrix
Major, J. R., jr.; and Ruff, M. D., 1978, J. Parasitol., v. 64 (4), 706-711
Eimeria spp.-infected broilers, reduced disaccharidase activity in region of intestine with maximum infection, this reduction is related to both time and severity of infection and can contribute to overall reduction in nutrient absorption

Eimeria necatrix
Migaki, T. T.; Chappel, L. R.; and Babcock, W. E., 1979, Poultry Science, v. 58 (5), 1192-1196
Eimeria spp., chicks (exper.), salinomycin, monensin, lasalocid, efficacy in battery trials

Eimeria necatrix
Mladenovic, Z.; Movsesijan, M.; and Borojevic, D., 1978, Vet. Glasnik, v. 32 (10), 829-834
Eimeria spp., chickens (exper.), mixed infections, cycostat, nitryl, and stenorol

Eimeria necatrix
Morrison, W. D.; Ferguson, A. E.; and Leeson, S., 1979, Poultry Science, v. 58 (5), 1160-1166
Eimeria spp., chicks (exper.), salinomycin and stenorol compared with other anticoccidials, efficacy and effect on chick performance

Eimeria necatrix
Noblet, G. P.; and Turk, D. E., 1979, Poultry Science, v. 58 (2), 392-403
Eimeria spp., chicks (exper.), no interference with overall intestinal absorption of ^{14}C-glucose, results suggest probable compensatory absorption by sections of intestine unaffected by a specific coccidial infection

Eimeria necatrix
Oikawa, H.; and Kawaguchi, H., 1975, Kiseichugaku Zasshi (Japan. J. Parasitol.), v. 24 (5), 276-283
Eimeria maxima, E. necatrix, chickens, effect of single vs. repeated vs. successive infections on manifestation of symptoms, food intake and body weight gain, and oocyst production

Eimeria necatrix
Olson, G.; et al., 1978, Poultry Science, v. 57 (5), 1245-1250
Eimeria spp. field isolates, chickens (exper.), arprinocid in comparison trials with marketed drugs, effective against all isolates tested including those refractory to many of the other products

Eimeria necatrix
Panitz, E., 1979, Parasitology, v. 78 (1), 33-40
Eimeria spp., chicks, anticoccidial efficacy and cross-resistance patterns of N,N'-bis (3,4 ditrifluoromethylphenyl) methylmalonamide compound have no practical application because of weight gain depression

E[imeria] necatrix
Pujic, P.; Krecov, M.; and Simic, V., 1977, Vet. Glasnik, v. 31 (4), 277-280
E[imeria] spp., chickens, stenorol prevents coccidiosis under laboratory and field conditions, during fattening period

Eimeria necatrix
Ruff, M. D., 1978, Proc. 1978 Maryland Nutrition Conf. Feed Mfr. (University of Maryland, Mar. 16-17), 32-37
Eimeria spp., chickens, anticoccidials, safe withdrawal times, effect on nutrient malabsorption

Eimeria necatrix
Ruff, M. D.; et al., 1978, Avian Dis., v. 22 (1), 32-41
Eimeria spp., broilers (exper.), arprinocid, drug efficacy in both battery and floor-pen trials

Eimeria necatrix
Ruff, M. D.; et al., 1979, Poultry Science, v. 58 (2), 298-303
Eimeria spp., battery raised broilers (exper.), narasin compared with monensin

Eimeria necatrix
Ruff, M. D.; Anderson, W. I.; and Reid, W. M., 1978, J. Parasitol., v. 64 (2), 306-311
Eimeria spp. in broilers, arprinocid decreased number of oocysts produced, fewer of the oocysts sporulated, and those oocysts which did sporulate were less infective than those from unmedicated birds

Eimeria necatrix
Ruff, M. D.; Wyatt, R. D.; and Witlock, D. R., 1978, J. Parasitol., v. 64 (1), 23-26
Eimeria spp. in broilers, severe infection increased prothrombin times compared with uninfected birds

Eimeria necatrix
Ryley, J. F.; and Hardman, L., 1978, Parasitology, v. 76 (1), 11-20
Eimeria spp., chicks (exper.), effects of dietary vitamin K on severity of disease with particular attention to effects of vitamin K on response to anticoccidial drugs, concluded that use of vitamin K deficient diet for experimental work is quite justified

Eimeria necatrix
Ryley, N. G.; and Ryley, J. F., 1978, Parasitology, v. 77 (1), 33-39
Eimeria, 10 spp., survival and viability of oocysts following prolonged exposure to saturated sodium chloride solution, additional observations on virulence of Eimeria tenella for chickens following such exposure, no evidence obtained to contraindicate use of salt-flotation methods for separation of oocysts from feces

Eimeria necatrix
Schindler, P.; et al., 1979, Poultry Science, v. 58 (1), 23-27
Eimeria spp., broiler chicken pen trials, arprinocid in feed highly effective prophylaxis, comparison with halofuginone, monensin, nicarbazin, and pancoxin: England; France; Germany

Eimeria necatrix
Sevcik, B.; et al., 1979, Biol. a Chem. Zivoc. Vyroby, Vet., v. 15 (v. 21), (4), 375-383
coccidiosis, broilers, field trials of anticoccidials: CSSR

Eimeria necatrix
Singh, J.; and Gill, B. S., 1976, Riv. Parassitol., Roma, v. 37 (1), 57-62
Eimeria necatrix, different levels of infection, chicks, activity of decoquinate used prophylactically and therapeutically, effect on development of immunity

Eimeria necatrix
Singh, J.; and Gill, B. S., 1976, Riv. Parassitol., Roma, v. 37 (1), 63-70
Eimeria necatrix, different levels of infection, chicks, activity of amprolium used prophylactically and therapeutically, effect on development of immunity

Eimeria necatrix
Tamas, T.; et al., 1978, Poultry Science, v. 57 (2), 381-385
Eimeria acervulina, E. maxima, E. necatrix, E. tenella, arprinocid and dichloro analog L-628,914 in feed, decrease in oocysts' capacity to sporulate

Eimeria necatrix
Thompson, J. E.; Fernando, M. A.; and Pasternak, J., 1979, Biochim. et Biophys. Acta, v. 555 (3), 472-484
Eimeria necatrix, induction of gel-phase lipid in plasma membrane of chick intestinal cells after infection, membrane lipid of developing parasites remains exclusively liquid crystalline at physiological temperature

Eimeria necatrix
Timchenko, A. D., 1972, Parazitologiia, Leningrad, v. 6 (6), 509-512
Eimeria spp., survival of non-sporulated and sporulated oocysts on soil surface and at various depths under conditions prevailing in southern Ukraine during winter to spring period, implications for prophylaxis and control of coccidiosis

Eimeria neodebliecki
Upadhyay, A. N.; and Ahluwalia, S. S., 1976, Indian Vet. J., v. 53 (12), 972-973
mixed coccidial infection, pig, first infection did not confer any immunity, pig became resistant to challenge after second infection

Eimeria neodebliecki
Upadhyay, A. N.; and Ahluwalia, S. S., 1978, Indian Vet. J., v. 55 (6), 458-461
coccidia, pigs (exper.), mixed infection with 4 spp. , macro- and microgametocytic stages, mostly not identified to species

Eimeria neodebliecki
Upadhyay, A. N.; and Ahluwalia, S. S., 1978, Indian Vet. J., v. 55 (10), 829-830
4 spp. of coccidia, pigs (exper.), only slightly pathogenic, no gross observable lesions

Eimeria neoirresidua
Wiggins, J. P.; and Rothenbacher, H., 1979, J. Parasitol., v. 65 (3), 393-394
Sylvilagus floridanus (feces): central Pennsylvania

Eimeria neoleporis
Wiggins, J. P.; and Rothenbacher, H., 1979, J. Parasitol., v. 65 (3), 393-394
Sylvilagus floridanus (feces): central Pennsylvania

Eimeria nieschulzi, illus.
Dubremetz, J. F.; and Torpier, G., 1978, J. Ultrastructure Research, v. 62 (2), 94-109
Eimeria nieschulzi, structure of three-membranous pellicle, freeze fracture technique

Eimeria nieschulzi
Duszynski, D. W.; et al., 1978, J. Parasitol., v. 64 (1), 83-88
Trichinella spiralis, suppressed rejection in immunized rats concurrently infected with Eimeria nieschulzi

Eimeria nieschulzi
Duszynski, D. W.; et al., 1978, J. Protozool., v. 25 (3, pt. 2), 370-374
Eimeria nieschulzi, rats, intestinal transit time during infection, on basis of findings it is difficult to implicate altered intestinal transit time in symptoms such as diarrhea which attend coccidiosis

Eimeria nieschulzi
Duszynski, D. W.; Roy, S. A.; and Castro, G. A., 1978, J. Protozool., v. 25 (2), 226-231
Eimeria nieschulzi, structural and functional changes in small intestine of infected rats (increase in intestinal mass; changes in mucosal structure especially increased crypt depth; decrease in peroxidase levels in lamina propria; reduction of brush border disaccharidase activity), intensity of all changes was directly dose-dependent

Eimeria nieschulzi
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Rattus norvegicus (Jejunum, Rectum)
R. rattus (Jejunum)
all from Neusiedlerseegebiet, nordlichen Burgenland

Eimeria nieschulzi
McQuistion, T. E.; and Schurr, K. M., 1978, J. Protozool., v. 25 (3, pt. 2), 374-377
Eimeria nieschulzi, rats, effect of infection on leukocyte levels

Eimeria nieschulzi, illus.
Marchiondo, A. A.; and Duszynski, D. W., 1978, J. Parasitol., v. 64 (1), 163-164
Eimeria nieschulzi, two methods for preparation of permanent light microscopy slides of oocysts

Eimeria nieschulzi, illus.
Marchiondo, A. A.; Duszynski, D. W.; and Speer, C. A., 1978, J. Protozool., v. 25 (4), 434-437
Eimeria nieschulzi, scanning electron micrographs of oocysts, transmission electron micrographs of oocyst wall and sporocyst wall

Eimeria nieschulzi
Rose, M. E.; et al., 1979, Parasite Immunol., v. 1 (2), 125-132
Eimeria nieschulzi, Nippostrongylus brasiliensis, failure of nude (athymic) rats to become resistant to reinfection

Eimeria nieschulzi, illus.
Rose, M. E.; and Hesketh, P., 1979, Infect. and Immun., v. 26 (2), 630-637
Eimeria spp. infections in normal animals vs. in animals with functional deficiencies in either T-lymphocytes or B-lymphocytes

Eimeria nieschulzi
Rose, M. E.; Hesketh, P.; and Ogilvie, B. M., 1979, Immunology, v. 36 (1), 71-79
Eimeria maxima, chickens, E. nieschulzi, rats, primary and secondary infections, E. maxima-immunized chickens challenged with E. acervulina: peripheral blood leucocyte response, correlation with resistance to reinfection

Eimeria ninakohlyakimovae
Ajayi, J. A.; and Todd, A. C., 1977, Bull. Animal Health and Prod. Africa, v. 25 (3), 257-261
prevalence, prepatent period
sheep (feces): Wisconsin

Eimeria ninae kohlyakimovi
Ermatova, D. U., 1978, Veterinariia, Moskva (3), 69-70
Eimeria spp., sheep and goats, occurrence, seasonality, host age: Kashkadar'insk oblast

Eimeria ninakohlyakimovae
Fitzgerald, P. R.; and Mansfield, M. E., 1978, Am. J. Vet. Research, v. 39 (1), 7-10
Eimeria spp., lambs (nat. and exper.), effect of monensin on oocyst discharge, feed consumption, fecal output, and weight gain (by sex of host): Illinois

Eimeria ninakohlyakimovae
Foreyt, W. J.; Gates, N. L.; and Wescott, R. B., 1979, Am. J. Vet. Research, v. 40 (1), 97-100
Eimeria spp., confinement-reared lambs (exper.) from weaning to market weight, monensin and lasalocid highly effective in eliminating oocysts, weight gains and feed conversion rates measured

Eimeria ninaekohljakimovi (Jakimoff, Rastegaeff, 1929), illus.
Gobzem, V. R., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 124-130
synonymy
[Ovis aries] (feces): Belorussia

Eimeria ninakohlyakimovi
Golemanski, V. G., 1977, Priroda, Sofiia, v. 26 (1), 71-74
Ovis musimon: Bulgaria

Eimeria ninakohlyakimovae
Horton, G. M. J.; and Stockdale, P. H. G., 1979, Am. J. Vet. Research, v. 40 (7), 966-970
Eimeria spp., lambs, amprolium and monensin, oocyst discharge, feed utilization, and ruminal metabolism

Eimeria ninakohlyakimovae
Krishnamurthy, R.; and Kshirsagar, H. S., 1976, Marathwada Univ. J. Sc. (Nat. Sc.), v. 15 (8), 153-156
Eimeria spp., goats (feces), prevalence and seasonal incidence: slaughter house at Parbhani, Mahrasthra state

Eimeria ninakohlyakimovae Yakimoff & Rastegaieff, illus.
McDougald, L. R., 1979, J. Protozool., v. 26 (1), 109-113
Eimeria christenseni oocysts from goat, failure to infect sheep; E. ninakohlyakimovae-like oocysts from sheep, failure to infect goats, E. ninakohlyakimovae-like coccidia of the 2 hosts belong to separate species, E. ovinoidalis sp. n. proposed for species from sheep, E. ninakohlyakimovae retained for species from goats

Eimeria ninakohlyakimovae
McDougald, L. R.; and Dunn, W. J., 1978, Am. J. Vet. Research, v. 39 (9), 1459-1462
Eimeria spp., lambs (exper.), efficacy of monensin in feed

Eimeria ninakohlyakimovae
Mason, P., 1977, N. Zealand Vet. J., v. 25 (1-2), 30-33
Eimeria spp., lambs, development of naturally acquired infection: Otago, New Zealand

Eimeria ninakohlyakimovae
Rama, S. P.; et al., 1978, Indian Vet. Med. J., v. 2 (4), 197-199
Eimeria parva, E. ninakohlyakimovae, sheep (exper.), blood values 11th day post-infection

Eimeria ninakohlyakimovae
Samizadeh-Yazd, A.; et al., 1979, Am. J. Vet. Research, v. 40 (8), 1107-1109
Eimeria spp., lambs, efficacy of monensin and aureomycin separately and combined

Eimeria nina-kohl-yakimovae
Santiago, M. A. M.; da Costa, U. C.; and da Silva, O. L., 1977, Rev. Centro Cien. Rurais, v. 7 (3), 297-301
Eimeria spp., sheep, natural infection, amprolium, furoxone, not sufficient control for sheep grazing on pasture

Eimeria ninakohlyakimovae
Seidel, B., 1979, Zool. Garten, n. F., v. 49 (2), 131-160
Ovibos moschatus, management in zoological park, includes information on parasites and their treatment: Tierpark Berlin

Eimeria ninaėkohlyakimovae
Tarazona, J. M.; and Fernandez, J. M., 1974, An. Inst. Nac. Invest. Agrar., s. Hig. y San. Animal (1), 55-62
Eimeria spp., sheep, incidence, distribution, review of clinical aspects, pathology, epizootiology: Spain

Eimeria nocens
  Golovneva, L. F., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 116-119
    helminths and protozoa of domestic waterfowl, prevalence under intensive rearing conditions

Eimeria nutriae, illus.
  Nukerbaeva, K. K., 1978, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (3), 16-21
    Myocastor coypus (feces): Alma-Ata zoopark

Eimeria odocoilei
  Woolf, A.; and Harder, J. D., 1979, Wildlife Monogr. (67), 53 pp.
    Odocoileus virginianus: Rachelwood Wildlife Research Preserve, New Florence, Pennsylvania

Eimeria ondatrazibethicae
  Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
    Ondatra zibethica (Jejunum, Rectum): Neusiedlerseegebiet, nordlichen Burgenland

Eimeria oreoecetes sp. n., illus.
  Stabler, R. M.; et al., 1979, J. Parasitol., v. 65 (2), 272-274
    Lagopus leucurus: Alpine meadows
    Dendragapus obscurus: forested mountains
    (feces of all): all from Colorado Rockies

Eimeria ovina
  Fitzgerald, P. R.; and Mansfield, M. E., 1978, Am. J. Vet. Research, v. 39 (1), 7-10
    Eimeria spp., lambs (nat. and exper.), effect of monensin on oocyst discharge, feed consumption, fecal output, and weight gain (by sex of host): Illinois

Eimeria ovina
  Foreyt, W. J.; Gates, N. L.; and Wescott, R. B., 1979, Am. J. Vet. Research, v. 40 (1), 97-100
    Eimeria spp., confinement-reared lambs (exper.) from weaning to market weight, monensin and lasalocid highly effective in eliminating oocysts, weight gains and feed conversion rates measured

Eimeria ovina, illus.
  Golemanski, V. G., 1977, Priroda, Sofiia, v. 26 (1), 71-74
    Ovis musimon: Bulgaria

Eimeria ovina
  Horton, G. M. J.; and Stockdale, P. H. G., 1979, Am. J. Vet. Research, v. 40 (7), 966-970
    Eimeria spp., lambs, amprolium and monensin, oocyst discharge, feed utilization, and ruminal metabolism

Eimeria ovina
  McDougald, L. R.; and Dunn, W. J., 1978, Am. J. Vet. Research, v. 39 (9), 1459-1462
    Eimeria spp., lambs (exper.), efficacy of monensin in feed

Eimeria ovina
  Mason, P., 1977, N. Zealand Vet. J., v. 25 (1-2), 30-33
    Eimeria spp., lambs, development of naturally acquired infection: Otago, New Zealand

Eimeria ovina
  Samizadeh-Yazd, A.; et al., 1979, Am. J. Vet. Research, v. 40 (8), 1107-1109
    Eimeria spp., lambs, efficacy of monensin and aureomycin separately and combined

Eimeria ovinoidalis sp. n., illus.
  McDougald, L. R., 1979, J. Protozool., v. 26 (1), 109-113
    Eimeria christenseni oocysts from goat, failure to infect sheep; E. ninakohlyakimovae-like oocysts from sheep, failure to infect goats, E. ninakohlyakimovae-like coccidia of the 2 hosts belong to separate species, E. ovinoidalis sp. n. proposed for species from sheep, E. ninakohlyakimovae retained for species from goats

Eimeria pallida
  Ajayi, J. A.; and Todd, A. C., 1977, Bull. Animal Health and Prod. Africa, v. 25 (3), 257-261
    prevalence, prepatent period
    sheep (feces): Wisconsin

Eimeria pallida
  Fitzgerald, P. R.; and Mansfield, M. E., 1978, Am. J. Vet. Research, v. 39 (1), 7-10
    Eimeria spp., lambs (nat. and exper.), effect of monensin on oocyst discharge, feed consumption, fecal output, and weight gain (by sex of host): Illinois

Eimeria pallida
  Foreyt, W. J.; Gates, N. L.; and Wescott, R. B., 1979, Am. J. Vet. Research, v. 40 (1), 97-100
    Eimeria spp., confinement-reared lambs (exper.) from weaning to market weight, monensin and lasalocid highly effective in eliminating oocysts, weight gains and feed conversion rates measured

Eimeria pallida
  Samizadeh-Yazd, A.; et al., 1979, Am. J. Vet. Research, v. 40 (8), 1107-1109
    Eimeria spp., lambs, efficacy of monensin and aureomycin separately and combined

Eimeria pallida
  Tarazona, J. M.; and Fernandez, J. M., 1974, An. Inst. Nac. Invest. Agrar., s. Hig. y San. Animal (1), 55-62
    Eimeria spp., sheep, incidence, distribution, review of clinical aspects, pathology, epizootiology: Spain

Eimeria parva
  Ajayi, J. A.; and Todd, A. C., 1977, Bull. Animal Health and Prod. Africa, v. 25 (3), 257-261
    prevalence, prepatent period
    sheep (feces): Wisconsin

Eimeria parva
  Ermatova, D. U., 1978, Veterinariia, Moskva (3), 69-70
    Eimeria spp., sheep and goats, occurrence, seasonality, host age: Kashkadar'insk oblast

Eimeria parva
  Fitzgerald, P. R.; and Mansfield, M. E., 1978, Am. J. Vet. Research, v. 39 (1), 7-10
    Eimeria spp., lambs (nat. and exper.), effect of monensin on oocyst discharge, feed consumption, fecal output, and weight gain (by sex of host): Illinois

Eimeria parva
Foreyt, W. J.; Gates, N. L.; and Wescott, R. B., 1979, Am. J. Vet. Research, v. 40 (1), 97-100
Eimeria spp., confinement-reared lambs (exper.) from weaning to market weight, monensin and lasalocid highly effective in eliminating oocysts, weight gains and feed conversion rates measured

Eimeria parva (Kotlan, Mocsu, & Vajda, 1929), illus.
Gobzem, V. R., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 124-130
[Ovis aries] (feces): Belorussia

Eimeria parva, illus.
Golemanski, V. G., 1977, Priroda, Sofiia, v. 26 (1), 71-74
Ovis musimon: Bulgaria

Eimeria parva
Krishnamurthy, R.; and Kshirsagar, H. S., 1976, Marathwada Univ. J. Sc. (Nat. Sc.), v. 15 (8), 153-156
Eimeria spp., goats (feces), prevalence and seasonal incidence: slaughter house at Parbhani, Mahrasthra state

Eimeria parva
McDougald, L. R.; and Dunn, W. J., 1978, Am. J. Vet. Research, v. 39 (9), 1459-1462
Eimeria spp., lambs (exper.), efficacy of monensin in feed

Eimeria parva
Mason, P., 1977, N. Zealand Vet. J., v. 25 (1-2), 30-33
Eimeria spp., lambs, development of naturally acquired infection: Otago, New Zealand

Eimeria parva Mocsy, Vajda, 1929, illus.
Nukerbaeva, K. K., 1978, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (3), 16-21
Capra ibex sibirica (feces): Alma-Ata zoopark

Eimeria parva
Rama, S. P.; et al., 1978, Indian Vet. Med. J., v. 2 (4), 197-199
Eimeria parva, E. ninakohlyakimovae, sheep (exper.), blood values 11th day post-infection

Eimeria parva, illus.
Rama, S.; Singh, C. D. N.; and Sinha, B. K., 1977, Kerala J. Vet. Sc., v. 8 (2), 243-246
Eimeria parva, lambs, symptoms, pathology

Eimeria parva
Samizadeh-Yazd, A.; et al., 1979, Am. J. Vet. Research, v. 40 (8), 1107-1109
Eimeria spp., lambs, efficacy of monensin and aureomycin separately and combined

Eimeria parva
Santiago, M. A. M.; da Costa, U. C.; and da Silva, O. L., 1977, Rev. Centro Cien. Rurais, v. 7 (3), 297-301
Eimeria spp., sheep, natural infection, amprolium, furoxone, not sufficient control for sheep grazing on pasture

Eimeria parva
Seidel, B., 1979, Zool. Garten, n. F., v. 49 (2), 131-160
Ovibos moschatus, management in zoological park, includes information on parasites and their treatment: Tierpark Berlin

Eimeria parva
Tarazona, J. M.; and Fernandez, J. M., 1974, An. Inst. Nac. Invest. Agrar., s. Hig. y San. Animal (1), 55-62
Eimeria spp., sheep, incidence, distribution, review of clinical aspects, pathology, epizootiology: Spain

Eimeria parvula
Golovneva, L. F., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 116-119
helminths and protozoa of domestic waterfowl, prevalence under intensive rearing conditions

Eimeria pellerdyi nouvelle espece, illus.
Coudert, P., 1977, Compt. Rend. Acad. Sc., Paris, v. 285, s. D, Sc. Nat. (8), 885-887
Eimeria pellerdyi n. sp., E. intestinalis, pure isolates distinguished in coccidia-free rabbits on basis of duration of life cycle, site of development, and strict specificity of acquired immunity; pathogenicity
Oryctolagus cuniculus (caecum, colon) (exper.)

E[imeria] pellerdyi
Coudert, P.; Vaissaire, J.; and Licois, D., 1978, Rec. Med. Vet., v. 154 (5), 437-440
E[imeria] pellerdyi, E[imeria] intestinalis, young rabbits, hematological changes

Eimeria pellerdyi (Coudert, 1977)
Licois, D.; Coudert, P.; and Mongin, P., 1978, Ann. Recherches Vet., v. 9 (1), 1-10
Eimeria intestinalis, E. pellerdyi, rabbits (exper.), changes in water metabolism in diarrhoeic hosts

Eimeria pellita Supperer 1952, illus.
Ernst, J. V.; and Todd, K. S., jr., 1977, Proc. Helminth. Soc. Washington, v. 44 (2), 221-223
redescription of sporulated oocyst
Holstein-Friesian calves (feces): eastern Alabama

Eimeria perforans (Leuckart, 1879) Sluiter & Swellengrebel, 1912, illus.
Catchpole, J.; and Norton, C. C., 1979, Parasitology, v. 79 (2), 249-257
Eimeria spp. in rabbits from 3 commercial rabbitries, species identification, level of infection, % occurrence and % predominance of different spp.: South East England

Eimeria perforans (Leuckart, 1879), illus.
Coudert, P.; Licois, D.; and Streun, A., 1979, Ztschr. Parasitenk., v. 59 (3), 227-234
Eimeria perforans, isolation of pure strain using specific-pathogen free rabbits, measurements, sporulation time, pathogenicity

Eimeria perforans
Danek, J.; et al., 1978, Biol. a Chem. Zivoc. Vyroby, Vet., v. 14 (v. 20) (2), 151-169
Eimeria spp., rabbits (nat. and exper.), sulphacombine, controlled test, subacute toxicity, no negative effect on followed indicators

Eimeria perforans
Gallazzi, D., 1977, Folia Vet. Latina, v. 7 (4), 371-380
Eimeria spp., rabbits, cyclical variations in excretion of fecal oocysts, seasonal effects, effect of pregnancy, parturition, and lactation, removal of mother from litter, infection of litters, relationship between maternal coccidial levels and those in the young, performance

Eimeria perforans
Pav, J., 1978, Veterinarstvi, v. 28 (2), 84-86
coccidia, cestodes, prevalence in hares: Czechoslovakia

Eimeria perforans
Peeters, J. E.; Halen, P.; and Meulemans, G., 1979, Brit. Vet. J., v. 135 (4), 349-354
Eimeria spp., rabbits (nat. and exper.), efficacy of robenidine

Eimeria perminuta
Napravnik, J.; and Fajkos, A., 1978, Sborn. Vysoke Skoly Zemedelske Praze, Fak. Agron., Rada B, Zivoc. Vyroba (1), 3-19
Eimeria spp., nematodes, pigs, soft runs as source of contamination, weather and climatic conditions

Eimeria perminuta
Upadhyay, A. N.; and Ahluwalia, S. S., 1976, Indian Vet. J., v. 53 (12), 972-973
mixed coccidial infection, pig, first infection did not confer any immunity, pig became resistant to challenge after second infection

Eimeria perminuta
Upadhyay, A. N.; and Ahluwalia, S. S., 1978, Indian Vet. J., v. 55 (6), 458-461
coccidia, pigs (exper.), mixed infection with 4 spp. , macro- and microgametocytic stages, mostly not identified to species

Eimeria perminuta
Upadhyay, A. N.; and Ahluwalia, S. S., 1978, Indian Vet. J., v. 55 (10), 829-830
4 spp. of coccidia, pigs (exper.), only slightly pathogenic, no gross observable lesions

Eimeria persica Levine and Becker, 1933, illus.
Ovezmukhammedov, A., 1978, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (5), 85-87
description
Natrix natrix persa (intestine): lake Maloe Delili, Western Turkmenia

Eimeria peruviana Yakimoff, 1934, illus.
Nukerbaeva, K. K., 1978, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (3), 16-21
Lama glamma (feces): Alma-Ata zoopark

Eimeria phasiani
Bejsovec, J., 1978, Ang. Parasitol., v. 19 (2), 76-85
Eimeria phasiani and E. colchici in Phasianus colchicus, dynamics of incidence dependent upon host biotope, host movements, season, temperature, and humidity: Mittelbohmen

Eimeria phasiani
Williams, R. B., 1978, Vet. Parasitol., v. 4 (2), 193-197
Pavo cristatus: Essex, Great Britain

Eimeria pileata sp. n., illus.
Straneva, J. E.; and Kelley, G. L., 1979, J. Protozool., v. 26 (4), 530-532
Clethrionomys gapperi (feces): Brush Valley, Indiana County, Pennsylvania

Eimeria piriformis Kotlan & Pospesch, 1934, illus.
Catchpole, J.; and Norton, C. C., 1979, Parasitology, v. 79 (2), 249-257
Eimeria spp. in rabbits from 3 commercial rabbitries, species identification, level of infection, % occurrence and % predominance of different spp.: South East England

Eimeria ponderosa Wetzel, 1942, illus.
Brglez, J.; and Bidovec, A., 1978, Acta Parasitol. Iugoslavica, v. 9 (2), 71-73
Capreolus capreolus: hunting district, and Zoological Garden in Ljubljana, SR Sloveniji, Jugoslaviji

Eimeria ponderosa Wetzel, 1942, illus.
Nukerbaeva, K. K., 1978, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (3), 16-21
Capreolus capreolus (feces): Alma-Ata zoopark

Eimeria praecox
Chapman, H. D., 1978, Ztschr. Parasitenk., v. 56 (2), 115-121
Eimeria spp., excystation, roles of various enzymes

Eimeria praecox
Kucera, J.; and Lom, J., 1979, Veterinarstvi, v. 29 (8), 351-353
Eimeria spp., Heterakis gallinae, incidence at large scale poultry farms using cage breeding methods: Cechach

Eimeria praecox
Latif, B. M. A.; Ali, S. R.; and Versenyi, L., 1974, Acta Vet. Beograd, v. 24 (3), 135-138
Eimeria spp., chickens (feces), rate of infection: Baghdad area, Iraq

Eimeria praecox
Oikawa, H.; and Kawaguchi, H., 1976, Kiseichugaku Zasshi (Japan. J. Parasitol.), v. 25 (3), 141-147
Eimeria praecox, E. mitis, chickens, single, repeated or successive infection, effect on manifestation of symptoms and oocyst production

Eimeria punctata (Landers, 1955)
Gobzem, V. R., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 124-130
as syn. of Eimeria crandallis (Honess, 1942)

Eimeria punctata Landers, 1955, illus.
Nukerbaeva, K. K., 1978, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (3), 16-21
Ovis canadensis canadensis (feces): Alma-Ata zoopark

Eimeria punctata
Samizadeh-Yazd, A.; et al., 1979, Am. J. Vet. Research, v. 40 (8), 1107-1109
Eimeria spp., lambs, efficacy of monensin and aureomycin separately and combined

Eimeria punctata
Tarazona, J. M.; and Fernandez, J. M., 1974, An. Inst. Nac. Invest. Agrar., s. Hig. y San. Animal (1), 55-62
Eimeria spp., sheep, incidence, distribution, review of clinical aspects, pathology, epizootiology: Spain

Eimeria riedmuelleri, "resembled"
Desser, S. S., 1978, J. Parasitol., v. 64 (5), 933-935
Rupicapra rupicapra (feces): Westland game farm

Eimeria robertsoni
Gottschalk, C., 1973, Ang. Parasitol., v. 14 (1), 44-54
endo-parasites of Lepus europaeus, seasonal dynamics, distribution according to locality, sex and age of host, economic importance of parasitism for regional hunting: Ostthuringen, DDR

Eimeria robertsoni (Madsen 1938) Carvalho 1943, illus.
Samoil, H. P.; and Samuel, W. M., 1977, Canad. J. Zool., v. 55 (10), 1671-1683
synonymy, redescription, key
Lepus americanus (feces): near Rochester, north-central Alberta

Eimeria robusta Supperer and Kutzer, 1961, illus.
Brglez, J.; and Bidovec, A., 1978, Acta Parasitol. Iugoslavica, v. 9 (2), 71-73
Cervus elaphus: hunting districts of Snezniku, SR Sloveniji, Jugoslaviji

Eimeria rochesterensis sp. n., illus.
Samoil, H. P.; and Samuel, W. M., 1977, Canad. J. Zool., v. 55 (10), 1671-1683
key
Lepus americanus (feces): near Rochester, north-central Alberta

Eimeria rowani sp. n., illus.
Samoil, H. P.; and Samuel, W. M., 1977, Canad. J. Zool., v. 55 (10), 1671-1683
key
Lepus americanus (feces): near Rochester, north-central Alberta

Eimeria ruficaudati Gill and Ray 1960, illus.
Samoil, H. P.; and Samuel, W. M., 1977, Canad. J. Zool., v. 55 (10), 1671-1683
redescription, key
Lepus americanus (feces): near Rochester, north-central Alberta

Eimeria rutili Dogiel et Bychowsky
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[Rutilus rutilus]: Upper Kama

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Eimeria spp., nematodes, pigs, soft runs as source of contamination, weather and climatic conditions

Eimeria schamchorica
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Meriones erythrourus: Uzbek SSR

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Lepus europaeus: Bulgaria

Eimeria semisculpta
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endo-parasites of Lepus europaeus, seasonal dynamics, distribution according to locality, sex and age of host, economic importance of parasitism for regional hunting: Ostthuringen, DDR

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Lochkarev, V. A.; and Evpak, I. D., 1977, Veterinariia, Moskva (6), 64-65
E[imeria] zurni, E. smithi, calves, novarsenol effective

Eimeria smithi Yakimoff et Galouzo, 1927, illus.
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Eimeria smithi, valid species, not a synonym of Eimeria bovis

Eimeria solipedum, illus.
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morphology, biology
horse: Yugoslavia

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Sorex araneus (Jejunum): Neusiedlerseegebiet, nordlichen Burgenland

Eimeria spleni, illus.
Lom, J., 1971, Folia Parasitol., v. 18 (4), 289-293
Eimeria subepithelialis, E. spleni, fish, morphology, walls of oocysts and spores, spore envelopes, electron microscopy
Campostoma anomalum: Flemings Creek, Washtenaw County, Michigan

Eimeria stiedae
Barriga, O. O.; and Arnoni, J. V., 1979, Exper. Parasitol., v. 48 (3), 407-414
Eimeria stiedae in Oryctolagus cuniculus, pathological effects produced by graded infections: body weight, oocyst output, serum glutamic pyruvic and serum oxalacetic transaminases, bilirubinemia, lipemia, glycemia, proteinemia, mortality, carcass and liver weights

Eimeria stiedai, illus.
Cerna, Z.; and Senaud, J., 1971, Folia Parasitol., v. 18 (2), 177-178
Eimeria stiedai, peculiarities of fine structure of merozoites

Eimeria stiedai
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Frandsen, J. C., 1978, Comp. Biochem. and Physiol., v. 60B (3), 303-307
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Eimeria stiedai
Hein, B.; and Laemmler, G., 1978, Ztschr. Parasitenk., v. 57 (3), 199-211
Eimeria stiedai, rabbits (exper.), alteration of serum enzyme activities

Eimeria stiedai, illus.
Jolley, W. R.; Allen, J. V.; and Nyberg, P. A., 1979, Internat. J. Parasitol., v. 9 (3), 199-204
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Eimeria stiedae
McCallister, G. L., 1979, J. Parasitol., v. 65 (1), 24
Eimeria, use of commercial soft drinks as source of carbon dioxide for excystation of oocysts

Eimeria stiedae
Owen, D., 1978, Lab. Animals, v. 12 (2), 49-50
Eimeria stiedae, E. intestinalis, standard rabbit diet heavily contaminated with oocysts, sterilization by autoclaving and irradiation was satisfactory for elimination of oocysts from diet but pelleting even at 68° was unsatisfactory

Eimeria stiedae
Pav, J., 1978, Veterinarstvi, v. 28 (2), 84-86
coccidia, cestodes, prevalence in hares: Czechoslovakia

Eimeria stiedai
Peeters, J. E.; Halen, P.; and Meulemans, G., 1979, Brit. Vet. J., v. 135 (4), 349-354
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Eimeria stiedai
Ryley, N. G.; and Ryley, J. F., 1978, Parasitology, v. 77 (1), 33-39
Eimeria, 10 spp., survival and viability of oocysts following prolonged exposure to saturated sodium chloride solution, additional observations on virulence of Eimeria tenella for chickens following such exposure, no evidence obtained to contraindicate use of salt-flotation methods for separation of oocysts from feces

Eimeria stiedai, illus.
Scholtyseck, E.; and Ratanavichien, A., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 553-556
Eimeria stiedai, rabbits, endodyogeny observed, confirms theory that endodyogeny is primary and fundamental process of asexual reproduction in the Coccidia and that schizogony is a secondary process which has developed from endodyogeny

Eimeria subepithelialis, illus.
Koerting, W., 1977, Fisch u. Umwelt (4), 37-48
fish parasites, histopathological changes

Eimeria subepithelialis, illus.
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Eimeria subepithelialis, E. spleni, fish, morphology, walls of oocysts and spores, spore envelopes, electron microscopy
Cyprinus carpio

Eimeria subepithelialis
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Evplov, N. N.; and Nazarov, V. G., 1977, Veterinariia, Moskva (6), 65-66
Eimeria spp., calves, chemococcide effective; compared with biomycin and norsulfazol: Belgorodsk oblast

imeria subsphaerica
Graefner, G.; Graubmann, H. D.; and Kron, A., 1978, Monatsh. Vet.-Med., v. 33 (23), 910-912
Eimeria spp., cattle on rearing and fattening farms, intensity and extensity of infection in relation to certain environmental conditions: DDR

Eimeria subsphaerica
Hiepe, T.; Romeyke, D.; and Jungmann, R., 1978, Monatsh. Vet.-Med., v. 33 (23), 904-910
Eimeria spp., calves reared under conditions of industrialized cattle farming, course of infection, clinical symptoms, monthly distribution, control measures: DDR

Eimeria subspherica
Ward, J. K.; Ferguson, D. L.; and Parkhurst, A. M., 1979, J. Animal Sc., v. 49 (2), 306-309
gastrointestinal parasites, beef cows (feces), level of infection, effect of animal age and season of year: Mead, Nebraska

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Davronov, O., 1973, Parazitologiia, Leningrad, v. 7 (1), 79-86
Rhombomys opimus: Zadar'insk, Alty-Aryksk, and Karshinsk regions, Uzbek SSR

Eimeria sylvilagi
Wiggins, J. P.; and Rothenbacher, H., 1979, J. Parasitol., v. 65 (3), 393-394
Sylvilagus floridanus (feces): central Pennsylvania

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as syn. of Eimeria wombati (Gilruth and Bull, 1912) comb. nov.
"probability of misidentification of the type host"

Eimeria tenella
Augustine, P. C.; Vetterling, J. M.; and Doran, D. J., 1977, Proc. Helminth. Soc. Washington, v. 44 (2), 147-149
Eimeria tenella, comparison of amprolium- and buquinolate-resistant strains to a drug-sensitive strain with respect to (1) oocyst production in chicks and (2) infectivity, rate of development, and oocyst production in primary chick kidney cell cultures

Eimeria tenella
Aver'ianov, A. V.; et al., 1978, Molek. Biol., v. 12 (3), 646-653
Eimeria tenella-infected chicks, relationship between RNA and protein biosynthesis in liver

Eimeria tenella
Bajwa, R. S.; and Gill, B. S., 1975, Ann. Soc. Belge Med. Trop., v. 55 (1), 31-36
Eimeria tenella, irradiation of oocysts with gamma-rays reduced number of viable sporozoites rather than attenuating the parasite

Eimeria tenella
Bajwa, R. S.; and Gill, B. S., 1977, Acta Vet. Brno, v. 46 (1-2), 149-158
Eimeria tenella, chickens (exper.), decoquinate vs. amprolium used prophylactically vs. therapeutically, anticoccidial activity against different levels of infection, effect on development of immunity

E[imeria] tenella
Bedrnik, P., 1977, Veterinarstvi, v. 27 (10), 458-459
coccidiosis, chickens, current prevalence, amprol plus treatment

Eimeria tenella
Bedrnik, P.; et al., 1979, Arch. Geflugelk., v. 43 (1), 7-10
Eimeria spp., chickens (exper.), comparative efficiency of anticoccidials in combination with growth promotant nitrovin

Eimeria tenella, illus.
Beier, T. V.; and Shibalova, T. A., 1974, Parazitologiia, Leningrad, v. 8 (5), 449-455
Eimeria tenella, increase in DNA in nuclei of chicken caecal cells infected with 2nd generation schizonts

Eimeria tenella, illus.
Bemrick, W. J.; and Hammer, R. F., 1978, Avian Dis., v. 22 (1), 86-94
Eimeria tenella, chickens (exper.), cecal lesions, scanning electron microscopy

Eimeria tenella
Brown, G. R.; and Foubister, A. J., 1979, J. Med. Chem., v. 22 (8), 997-999
benzo-15-crown-5 polyethers, synthesis, in vivo and in vitro tests against Eimeria tenella

Eimeria tenella
Chapman, H. D., 1978, Avian Path., v. 7 (2), 269-277
Eimeria maxima, E. brunetti, E. tenella, chickens, effect of monensin on development of immunity acquired by repeated low-level infections

Eimeria tenella
Chapman, H. D., 1978, Parasitology, v. 76 (2), 177-183
Eimeria tenella, Houghton strain, experimental development of resistance to amprolium, clopidol, and methyl benzoquate

Eimeria tenella
Chapman, H. D., 1978, Ztschr. Parasitenk., v. 56 (2), 115-121
Eimeria spp., excystation, roles of various enzymes

Eimeria tenella, illus.
Chappel, L. R., 1979, J. Parasitol., v. 65 (1), 137-143
Eimeria spp., chicks, site of action of salinomycin

Eimeria tenella
Chappel, L. R.; and Babcock, W. E., 1979, Poultry Science, v. 58 (2), 304-307
Eimeria spp., broilers (exper.), salinomycin, monensin, lasalocid, drug toleration and anticoccidial efficacy compared in 5 field trials, commercial facilities

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Eimeria tenella-infected chicks (exper.), periodicity of caecal defaecation

Eimeria tenella
Cruthers, L. R.; et al., 1978, Poultry Science, v. 57 (5), 1227-1233
Eimeria spp., broiler chicks (exper.), lonomycin in feed, high degree of anticoccidial activity, comparison with lasalocid and monensin

Eimeria tenella
Davis, P. J.; Parry, S. H.; and Porter, P., 1978, Immunology, v. 34 (5), 879-888
Eimeria tenella, chickens, serological and secretory immune responses evaluated in terms of various anti-coccidial activities, results suggest that intestinal secretory IgA system plays essential role in protective immune response

Eimeria tenella
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Eimeria tenella
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Eimeria tenella, 13 strains, intraspecific variability with respect to virulence, reproductive capability, and immunogenic properties: USSR

Eimeria tenella
Doran, D. J.; and Augustine, P. C., 1978, J. Protozool., v. 25 (4), 544-546
Eimeria tenella, vitamin requirements for development in primary cultures of chicken kidney cells

Eimeria tenella
Dykstra, D. D.; and Reid, W. M., 1978, Poultry Science, v. 57 (1), 85-89
Eimeria tenella in conventional, bacteria-free, and monofloral (Escherichia coli and Bacteroides sp.) chicks (exper.), comparison of cecal lesions, weight gain, clinical signs, and mortality

Eimeria tenella
Dykstra, D. D.; and Reid, W. M., 1978, Poultry Science, v. 57 (2), 398-402
Eimeria tenella, gnotobiotic chickens, monensin suppressed bacterial numbers in absence of Eimeria, prevented increase in bacterial numbers in presence of Eimeria

Eimeria tenella
Edgar, S. A.; and Flanagan, C., 1979, Poultry Science, v. 58 (6), 1469-1475
Eimeria spp. (recent field isolates resistant to various drugs), chickens, halofuginone, action cidal rather than static

Eimeria tenella
Edgar, S. A.; and Flanagan, C., 1979, Poultry Science, v. 58 (6), 1476-1482
Eimeria spp. (recent field isolates resistant to various drugs), halofuginone with roxarsone and/or bacitracin MD

Eimeria tenella
Greuel, E.; and Kuehnhold, W., 1977, Prakt. Tierarzt, v. 58 (5), 338-341
Eimeria spp., chickens (exper.), synergistic effect of metichlorpindol and methylbenzoquate, rotation program with other coccidiostats discussed

Eimeria tenella
Grier, N., 1979, J. Pharm. Sc., v. 68 (4), 407-411
Eimeria tenella, trichostrongyle, synthesis, in vitro and in vivo evaluation of quaternary salts of 4-Phenyl-1,2,3,6-tetrahydropyridine and 3,6-Dimethyl-6-phenyltetrahydro-2H-1,3-oxazine, none effective

Eimeria tenella
Hayat, B., 1976, Pakistan J. Scient. Research, Lahore, v. 28, 38-40
Eimeria tenella, chickens, immunization with gamma-irradiated oocysts

Eimeria tenella
Hildebrandt, J.; Meingassner, J. G.; and Mieth, H., 1978, Zentralbl. Vet.-Med., Reihe B., v. 25 (3), 186-193
Eimeria tenella, kidney cell cultures, chickens (exper.), septamycin, activity largely confined to first generation schizont

Eimeria tenella
Ishii, T.; and Onaga, H., 1978, Nippon Zyuisi-Kai Zassi (J. Japan Vet. Med. Ass.), v. 31 (9), 534-538
Eimeria spp., chicks (exper.) and cell cultures, activity of chlorobromofebrifugine

Eimeria tenella
Jeffers, T. K., 1978, Avian Dis., v. 22 (1), 157-161
Eimeria tenella isolants obtained from monensin-treated flocks, monensin sensitivity tests revealed no monensin-resistant isolants

Eimeria tenella
Johnson, J.; Reid, W. M.; and Jeffers, T. K., 1979, Poultry Science, v. 58 (1), 37-41
Eimeria tenella, chickens, floor-pen experiments comparing immunogenicity of 2 non-pathogenic parasite strains with control strain used in a commercial planned immunozation program, strain Wis-F-125 gave flock immunity equivalent to control strain

Eimeria tenella, illus.
Jolley, W. R.; Allen, J. V.; and Nyberg, P. A., 1979, Internat. J. Parasitol., v. 9 (3), 199-204
Eimeria stiedai, Eimeria tenella, micropyle and oocyst wall changes associated with chemically-mediated in vitro excystation

Eimeria tenella
Karlsson, T.; and Reid, W. M., 1978, Avian Dis., v. 22 (3), 487-495
Eimeria tenella, broiler chicks, effect of anticoccidials in feed on development of immunity to coccidiosis

Eimeria tenella
Kellogg, F. E.; Doster, G. L.; and Johnson, J. K., 1971, J. Wildlife Dis., v. 7 (3), 186-187
Indian red junglefowl, pen-raised (ceca)

Eimeria tenella
Khovanskikh, A. E., 1974, Parazitologiia, Leningrad, v. 8 (5), 456-459
Eimeria tenella, use of amino acids from host's cell for protein synthesis

Eimeria tenella
Khovanskikh, A. E., 1977, Sel'skokhoz. Biol., v. 12 (2), 245-250
Eimeria tenella, E. mitis, biochemical characteristics of pathogenesis in chickens, review

Eimeria tenella
Khovanskikh, A. E.; Krylov, M. V.; and Krylov, V. F., 1972, Parazitologiia, Leningrad, v. 6 (1), 8-10
Eimeria tenella-infected chickens (exper.), zinc concentration in various tissues and organs at different stages of infection

Eimeria tenella
Khovanskikh, A. E.; Krylov, M. V.; and Ludinova, I. C., 1974, Parazitologiia, Leningrad, v. 8 (2), 164-169
Eimeria tenella, chickens (exper.), absorption of iron in small intestine, concentration of iron in tissues and organs

Eimeria tenella
Khovanskikh, A. E.; and Kuznetsova, N. A., 1975, Parazitologiia, Leningrad, v. 9 (1), 77-81
Eimeria tenella-infected chickens, intensity of C^{14}-glycine inclusion into proteins of various organs, changes in total proteins and gamma-globulin in blood serum, correlation between increased biosynthesis of proteins in immunocompetent organs and increase in gamma-globulin in blood serum

Eimeria tenella
Kilgore, R. L.; et al., 1978, Poultry Science, v. 57 (4), 907-911
Eimeria spp., broiler chickens (exper.), arprinocid, effects on weight, feed efficiency, and histological lesions

Eimeria tenella
Kilgore, R. L.; et al., 1979, Poultry Science, v. 58 (1), 67-71
Eimeria spp., chickens, floor-pen trials evaluating 4 methods of induced exposure to coccidiosis suitable for use in drug research operations, laboratory sporulated oocysts spread over litter most satisfactory method

Eimeria tenella
Krylov, M. V.; et al., 1975, Parazitologiia, Leningrad, v. 9 (1), 82-91
Eimeria tenella, mechanisms of resistance to glycamide

Eimeria tenella
Krylov, V. F., 1978, Veterinariia, Moskva (10), 68-69
Eimeria tenella strain resistant to pharmcoccid after 35 laboratory passages in chickens, cross-resistance only to rigecoccin

Eimeria tenella
Kucera, J.; and Lom, J., 1979, Veterinarstvi, v. 29 (8), 351-353
Eimeria spp., Heterakis gallinae, incidence at large scale poultry farms using cage breeding methods: Cechach

Eimeria tenella
Kutzer, E.; et al., 1979, Wien. Tierarztl. Monatsschr., v. 66 (6-7), 197-202
Eimeria spp., broiler chickens, arprinocid, drug efficacy compared with monensin

Eimeria tenella
Latif, B. M. A.; Ali, S. R.; and Versenyi, L., 1974, Acta Vet. Beograd, v. 24 (3), 135-138
Eimeria spp., chickens (feces), rate of infection: Baghdad area, Iraq

Eimeria tenella
Latter, V. S.; and Wilson, R. G., 1979, Parasitology, v. 79 (1), 169-175
Eimeria tenella, factors influencing assessment of anticoccidial activity in cell culture

Eimeria tenella
Lazdynia, M. A.; et al., 1976, Med. Parazitol. i Parazitar. Bolezni, v. 45 (2), 189-192
Ascaridia galli, volatile fatty acids, composition, effects on Eimeria tenella infection in chickens

Eimeria tenella
Lee, E. H., 1979, Canad. Vet. J., v. 20 (4), 102-104
Eimeria tenella, drug-resistant field strains, White Leghorn chickens, single and low-level oocyst infections, treatment with robenidine or decoquinate

Eimeria tenella
Lee, E. H.; and Onderka, D. K., 1978, Canad. Vet. J., v. 19 (9), 237-243
Eimeria spp., chickens, incidence of clinical coccidiosis by month of year and age of host in the Ontario Veterinary Services Branch records 1973-1977, possible use of data to synchronize drug rotation with change in incidence

Eimeria tenella
Long, P. L.; and Millard, B. J., 1979, Avian Path., v. 8 (3), 213-228
Eimeria spp., young chickens kept in litter pens, immunization, response to challenge with homologous and heterologous strains, effect of host age and of immunizing dose, timing of onset of immunity, longevity of immunity

Eimeria tenella
Long, P. L.; and Millard, B. J., 1979, Parasitology, v. 78 (2), 239-247
Eimeria maxima, rejection by 'foreign' host (Numida meleagris); E. tenella, E. grenieri, survival of sporozoites in peritoneal macrophages from 'foreign' vs. normal hosts in vitro

Eimeria tenella
Long, P. L.; Millard, B. J.; and Smith, K. M., 1979, Avian Path., v. 8 (4), 453-467
Eimeria spp., chickens, effect of 4 anticoccidial drugs on development of immunity, field and laboratory conditions

Eimeria tenella
Long, P. L.; and Rowell, J. G., 1975, Brit. Poultry Sc., v. 16 (6), 583-592
Eimeria spp., method of sampling surface litter of commercial broiler houses for laboratory estimation of numbers of coccidial oocysts

Eimeria tenella, illus.
Luecht, I. G.; Millard, B. J.; and Scholtyseck, E. O., 1978, Parasitology, v. 76 (2), 185-191
Eimeria tenella, embryo-adapted strain, fine structure and development in chicken embryos, complete endogenous cycle is restricted to epithelial cells of chorio-allantoic membrane, no major ultrastructual changes have occurred as result of repeated embryo passage

Eimeria tenella
McCallister, G.; and Cowgill, L. M., 1977, Proc. Helminth. Soc. Washington, v. 44 (2), 218-219
Eimeria tenella, efficiency of Seinhorst filter for recovery of oocysts from large quantities of feces as compared to 3 previously applied methods

Eimeria tenella
McDougald, L. R.; and Galloway, R. B., 1977, Ztschr. Parasitenk., v. 54 (1), 95-100
Eimeria tenella in vitro, development inhibited by serum from chickens fed anticoccidial drugs, technique to assay drug activity and to characterize and quantitate therapeutic effect

Eimeria tenella
McDougald, L. R.; Karlsson, T.; and Reid, W. M., 1979, Avian Dis., v. 23 (4), 999-1005
coccidiosis, chickens (exper.), natural outbreak of infectious bursal disease (IBD) during comparison of anticoccidials for their effect on development of immunity, interaction between diseases, immunity to coccidiosis not blocked by IBD

Eimeria tenella
McLoughlin, D. K.; and Chute, M. B., 1978, J. Parasitol., v. 64 (5), 874-877
Eimeria tenella, chickens, robenidine protected against cecal coccidiosis initiated by parasite strain with no previous drug exposure, no cross resistance found with 13 strains resistant to other anticoccidials, when the sensitive strain was serially propagated in chickens medicated with robenidine it became resistant, no cross resistance found when this experimental strain was tested against 12 other anticoccidials

Eimeria tenella
McManus, E. C.; et al., 1979, Exper. Parasitol., v. 47 (1), 13-23
Eimeria tenella, chickens (exper.), t-butylaminoethanol alone or in synergistic combination with sulfaquinoxaline and pyrimethamine, anticoccidial efficacy, specific reversal of toxicity for parasite and host by choline and dimethylaminoethanol

Eimeria tenella
McManus, E. C.; and Rogers, E. F., 1979, Exper. Parasitol., v. 48 (2), 235-238
Eimeria tenella, chickens, synergistic interaction of sulfaquinoxaline and t-butylaminoethanol

Eimeria tenella
McQuistion, T. E.; and McDougald, L. R., 1979, Ztschr. Parasitenk., v. 59 (2), 107-113
Eimeria tenella, surgical ligation of chick ceca used to study role of absorption and extraintestinal transport in action of anticoccidial drugs

Eimeria tenella, illus.
Madden, P. A.; and Vetterling, J. M., 1978, J. Protozool., v. 25 (3, pt. 2), 298-301
Eimeria tenella, schizogony, scanning electron microscopy

Eimeria tenella
Major, J. R., jr.; and Ruff, M. D., 1978, Exper. Parasitol., v. 45 (2), 234-240
Eimeria spp., effects of coccidiosis on digestive capacity (amylase) of broiler chickens, changes in pancreatic, luminal, and surface-bound amylolytic activity, reduction in amylolytic activity as pH went below 5.0

Eimeria tenella
Manuel, M. F., 1972, Philippine J. Vet. Med., v. 11 (2), 92-100
Eimeria tenella, broilers (exper,), evaluation of amprol plus and coyden-25 with or without 3-nitro-50 (roxarsone)

Eimeria tenella
Manuel, M. F.; and de Leon, M. L., 1978, Philippine J. Vet. Med., v. 17 (1-2), 129-141
Eimeria tenella, White Leghorn chicks (exper.), comparative efficacy of Esb$_3$ and Abi-Zet$_{50}$

Eimeria tenella
Manuel, M. F.; and Neri, R. A., 1975, Philippine J. Vet. Med., v. 14 (1), 106-116
Eimeria tenella, White Leghorn cockerels, efficacy of 7 water-soluble coccidiostats

Eimeria tenella
Matsuzawa, T., 1978, Parasitology, v. 77 (2), 235-241
Eimeria tenella, chickens, beclotiamine, mode of action studies; attempts to potentiate or antagonize its activity revealed that pyrithiamine and 2,4-dinitrophenol also showed slight anticoccidial activity and that a combination of 2,4-DNP and beclotiamine was effective but weight gain was not as good as with beclotiamine alone

Eimeria tenella
Matsuzawa, T., 1979, Japan. Poultry Sc., v. 16 (3), 137-140
Eimeria tenella, chicks, carbohydrate in host diet, effect on infection

Eimeria tenella, illus.
Matsuzawa, T., 1979, Poultry Science, v. 58 (4), 1007-1008
Eimeria tenella in chicken ceca, incorporation of H^3-glucose, electron microscope autoradiography

Eimeria tenella
Meingassner, J. G.; et al., 1979, Poultry Science, v. 58 (2), 308-313
Eimeria tenella, broilers (exper.), monensin, lasalocid, anticoccidial activity enhanced when combined with tiamulin, tiamulin alone was inactive; metabolic fate of monensin and tiamulin separately or in combination monitored using isolated, perfused rat liver

Eimeria tenella
Migaki, T. T.; Chappel, L. R.; and Babcock, W. E., 1979, Poultry Science, v. 58 (5), 1192-1196
Eimeria spp., chicks (exper.), salinomycin, monensin, lasalocid, efficacy in battery trials

Eimeria tenella
Milushev, I., 1978, Vet. Med. Nauki, v. 15 (8), 26-29
Eimeria tenella oocysts remain viable in digestive tract of Musca domestica and when excreted; flies containing E. tenella oocysts produce infection when fed to chicks

Eimeria tenella
Mladenovic, Z.; Movsesijan, M.; and Borojevic, D., 1978, Vet. Glasnik, v. 32 (10), 829-834
Eimeria spp., chickens (exper.), mixed infections, cycostat, nitryl, and stenorol

Eimeria tenella
Mørch, J., 1978, Nord. Vet. Med., v. 30 (6), 253-259
Eimeria spp., chickens (exper.), efficacy of coccidiostats in feed, better production efficiency of medicated groups

Eimeria tenella, illus.
Mogollon, J. D.; and Molina, T. de P., 1977, Rev. Inst. Colomb. Agropec, v. 14 (4), 473-488
Eimeria tenella field strain, sensitivity against 3 anticoccidial drugs

Eimeria tenella
Morisawa, Y.; et al., 1978, J. Med. Chem., v. 21 (2), 194-199
Eimeria tenella, synthesis and anticoccidial activity of methyl-2(6)-nitro- and -3(5)-nitropyridinecarboxamides

Eimeria tenella
Morozova, T. I.; and Kravtsov, V. A., 1978, Tsitologiia, v. 20 (7), 817-819
Eimeria tenella, E. acervulina, sporozoites, ratio of rhoptry-microneme elements to anterior pole cytoplasm, such ratios may be species specific

Eimeria tenella
Morrison, W. D.; Ferguson, A. E.; and Leeson, S., 1979, Poultry Science, v. 58 (5), 1160-1166
Eimeria spp., chicks (exper.), salinomycin and stenorol compared with other anticoccidials, efficacy and effect on chick performance

Eimeria tenella
Musaev, M. A.; and Elchiev, Ia. Ia., 1970, Parazitologiia, Leningrad, v. 4 (5), 494-500
Eimeria tenella, E. mitis, chickens, serum protein changes over course of infection

Eimeria tenella
Musaev, M. A.; and Ibragimova, G. G., 1974, Parazitologiia, Leningrad, v. 8 (1), 70-73
Eimeria tenella, E. mitis, chickens (exper.), changes in lecithin content of blood serum at different stages of infection, extent of biochemical changes depends on pathogenicity of different species and host age

Eimeria tenella
Musaev, M. A.; and Surkova, A. M., 1972, Parazitologiia, Leningrad, v. 6 (1), 11-15
Eimeria tenella-, E. mitis-infected chickens (exper.), changes in activity of alkaline and acid phosphatases of small intestine depend on species of coccidia, age of host, and stage of infection

Eimeria tenella
Musaev, M. A.; and Surkova, A. M., 1974, Parazitologiia, Leningrad, v. 8 (2), 170-174
Eimeria tenella, chickens (exper.), acid and alkaline phosphatase activity of small intestinal mucosa, comparison of one infection (non-immune) vs. 3 successive infections (immune)

Eimeria tenella
Noblet, G. P.; and Turk, D. E., 1979, Poultry Science, v. 58 (2), 392-403
Eimeria spp., chicks (exper.), no interference with overall intestinal absorption of ^{14}C-glucose, results suggest probable compensatory absorption by sections of intestine unaffected by a specific coccidial infection

Eimeria tenella
Oikawa, H.; and Kawaguchi, H., 1975, Kiseichugaku Zasshi (Japan. J. Parasitol.), v. 24 (3), 99-106
Eimeria acervulina, E. tenella, chickens, effect of single vs. repeated vs. successive infections of mixed species on manifestation of symptoms, food intake and body weight gain, and oocyst production

Eimeria tenella
Olson, G.; et al., 1978, Poultry Science, v. 57 (5), 1245-1250
Eimeria spp. field isolates, chickens (exper.), arprinocid in comparison trials with marketed drugs, effective against all isolates tested including those refractory to many of the other products

Eimeria tenella
Onaga, H.; et al., 1978, Nippon Zyuisi-Kai Zassi (J. Japan Vet. Med. Ass.), v. 31 (10), 592-596
Eimeria spp., chicks and cell cultures, monensin

[Eimeria tenella] E. tenella
Orekhova, M. M., 1973, Vet. Nauka--Proizvod., Trudy, Minsk, v. 11, 156-159
[Eimeria tenella], chickens, 3,5-dinitrobenzamide having negative effect on schizonts and gamonts, but no effect on sporozoites; no effect on nucleic acids of endogenous stages shown

E[imeria] tenella
Orekhova, M. M., 1974, Vet. Nauka--Proizvod., Trudy, Minsk, v. 12, 139-140
E[imeria] tenella, chicks, coccidiostatic activity of iramin in combination with metacyl

Eimeria tenella
Panitz, E., 1979, Parasitology, v. 78 (1), 33-40
Eimeria spp., chicks, anticoccidial efficacy and cross-resistance patterns of N,N'-bis (3,4 ditrifluoromethylphenyl) methylmalonamide compound have no practical application because of weight gain depression

Eimeria tenella
Parre, J.; Hussar, U.; and Schattschneider, T., 1977, Eesti Pollumaj. Akad. Teadusl. Toode Kogum. (104), 81-89
Eimeria tenella, immunization and subsequent invasion, chicks, mitotic activity of thymus lymphocytes depressed, number of degenerative cell forms in thymus raised, changes correlated with increasing host age

Eimeria tenella
Parre, J.; and Olkonen, E., 1977, Eesti Pollumaj. Akad. Teadusl. Toode Kogum. (104), 100-108
Eimeria tenella, E. acervulina, E. brunetti, polyvaccine tested, chicks maintained in battery cages or deep litter; simultaneous zoalene treatment efficacious in deep litter maintenance

Eimeria tenella
Parre, J.; and Suigusaar, M., 1977, Eesti Pollumaj. Akad. Teadusl. Toode Kogum. (104), 90-99
Eimeria tenella sporozoites in vitro, neutralized by immune serum and immune gammaglobulin, proof of specific antibodies in serum of immune birds

Eimeria tenella
Pastuszko, J., 1976, Acta Parasitol. Polon., v. 24 (11-19), 103-117
Eimeria tenella, chickens, immunization using X-ray attenuated oocysts

Eimeria tenella
Patton, W. H.; and Brigman, G. P., 1979, J. Parasitol., v. 65 (4), 526-530
Eimeria tenella, use of sodium taurodeoxycholate for excystation of sporozoites, far more effective than other bile salts or extracts or pooled chicken bile

Eimeria tenella
Perov, M. F., 1974, Parazitologiia, Leningrad, v. 8 (5), 460-462
Eimeria tenella, fatty acid composition of lipids of oocysts and oocyst membranes

Eimeria tenella
Perov, M. F.; and Tal'drik, A. A., 1974, Parazitologiia, Leningrad, v. 8 (4), 365-367
Eimeria tenella, amino acid content of oocysts and oocyst membranes

Eimeria tenella
Platz, S., 1977, Deutsche Tierarztl. Wchnschr., v. 84 (5), 178-180
Eimeria tenella, infectivity after short-time composting of poultry manure, chicks (exper.)

E[imeria] tenella
Pujic, P.; Krecov, M.; and Simic, V., 1977, Vet. Glasnik, v. 31 (4), 277-280
E[imeria] spp., chickens, stenorol prevents coccidiosis under laboratory and field conditions, during fattening period

Eimeria tenella
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Eimeria tenella, illus.
Rose, M. E.; and Lee, D. L., 1977, Ztschr. Parasitenk., v. 54 (1), 1-7
Eimeria tenella, in vitro uptake of sporozoites by peritoneal exudate cells (macrophages and heterophils) from normal and immunized chickens, electron microscopy, greater uptake by cells from immunized birds but no difference in appearance of sporozoites; entry by phagocytosis

Eimeria tenella
Ruff, M. D., 1978, Proc. 1978 Maryland Nutrition Conf. Feed Mfr. (University of Maryland, Mar. 16-17), 32-37
Eimeria spp., chickens, anticoccidials, safe withdrawal times, effect on nutrient malabsorption

Eimeria tenella
Ruff, M. D.; et al., 1978, Avian Dis., v. 22 (1), 32-41
Eimeria spp., broilers (exper.), arprinocid, drug efficacy in both battery and floor-pen trials

Eimeria tenella
Ruff, M. D.; et al., 1979, Poultry Science, v. 58 (2), 298-303
Eimeria spp., battery raised broilers (exper.), narasin compared with monensin

Eimeria tenella
Ruff, M. D.; Anderson, W. I.; and Reid, W. M., 1978, J. Parasitol., v. 64 (2), 306-311
Eimeria spp. in broilers, arprinocid decreased number of oocysts produced, fewer of the oocysts sporulated, and those oocysts which did sporulate were less infective than those from unmedicated birds

Eimeria tenella
Ruff, M. D.; Wyatt, R. D.; and Witlock, D. R., 1978, J. Parasitol., v. 64 (1), 23-26
Eimeria spp. in broilers, severe infection increased prothrombin times compared with uninfected birds

Eimeria tenella
Ryley, J. F.; and Hardman, L., 1978, Parasitology, v. 76 (1), 11-20
Eimeria spp., chicks (exper.), effects of dietary vitamin K on severity of disease with particular attention to effects of vitamin K on response to anticoccidial drugs, concluded that use of vitamin K deficient diet for experimental work is quite justified

Eimeria tenella
Ryley, J. F.; and Wilson, R. G., 1978, Methods Cultiv. Parasites in Vitro, 111-128
cell and tissue culture of protozoa, review

Eimeria tenella
Ryley, N. G.; and Ryley, J. F., 1978, Parasitology, v. 77 (1), 33-39
Eimeria, 10 spp., survival and viability of oocysts following prolonged exposure to saturated sodium chloride solution, additional observations on virulence of Eimeria tenella for chickens following such exposure, no evidence obtained to contraindicate use of salt-flotation methods for separation of oocysts from feces

Eimeria tenella
Saad, F. E.; et al., 1974, J. Egypt. Vet. Med. Ass., v. 34 (1-2), 81-84
Eimeria tenella, chicks (exper.), blood changes during course of infection

Eimeria tenella
Sanda, A., 1977, Acta Vet. Brno, v. 46 (3-4), 311-314
Eimeria tenella, one-day-old chicks, reliable immunity within 5-6 days after low initial dose of oocysts

Eimeria tenella
Schindler, P.; et al., 1979, Poultry Science, v. 58 (1), 23-27
Eimeria spp., broiler chicken pen trials, arprinocid in feed highly effective prophylaxis, comparison with halofuginone, monensin, nicarbazin, and pancoxin: England; France; Germany

Eimeria tenella
Sevcik, B.; et al., 1974, Veterinaria, Praha, v. 16 (5-6), 421-588
Eimeria tenella, chickens, 613 substances screened as coccidiostats, extensive detailed statistical results

Eimeria tenella
Sevcik, B.; et al., 1979, Biol. a Chem. Zivoc. Vyroby, Vet., v. 15 (v. 21), (4), 375-383
coccidiosis, broilers, field trials of anticoccidials: CSSR

Eimeria tenella
Sherkov, S., 1977, Vet. Med. Nauki, v. 14 (2), 37-44
Eimeria tenella, chickens (exper.), amprolium alone and with additional amounts of thiamine in feed mixture, evaluation of prophylactic use, thiamine contributes to lowered activity of amprolium

E[imeria] tenella
Sherkov, Sh., 1977, Vet. Sbirka, v. 75 (3), 35-38
E[imeria] tenella, pancoxin, chickens raised under conditions of high and low temperatures; influence of thiamine on development of coccidiosis

Eimeria tenella, illus.
Shibalova, T. A.; Morozova, T. I.; and Kravtsov, V. A., 1974, Parazitologiia, Leningrad, v. 8 (3), 266-271
Eimeria tenella, sporozoites, merozoites, ultrastructure

Eimeria tenella
Singh, J.; and Hussain, O., 1978, Indian Vet. J., v. 55 (1), 56-60
Eimeria tenella, chicks (exper.), amprolium provided better protection than codrinal, both drugs interfered to some extent with development of immunity

Eimeria tenella
Smith, C. K. II; and Strout, R. G., 1979, Exper. Parasitol., v. 48 (3), 325-330
Eimeria tenella, accumulation and retention of lasalocid and narasin by extracellular sporozoites

E[imeria] tenella
Stoianov, P.; et al., 1978, Vet. Med. Nauki, v. 15 (8), 105-114
E[imeria] tenella, broiler chickens, varied temperature and moisture regimes, blood biochemistry, host resistance, efficacy of pancoxin plus

Eimeria tenella, illus.
Stotish, R. L.; Wang, C. C.; and Meyenhofer, M., [1979], J. Parasitol., v. 64 (6), 1978, 1074-1081
Eimeria tenella, oocyst wall, lipid composition, carbohydrate composition, protein content, amino acid analysis, proposed organization of cell wall, results suggest explanation for physical and mechanical resistance of oocyst wall as well as possible mechanisms for excystation of sporulated oocysts

Eimeria tenella (Railliet et Lucet, 1891)
Surkova, A. M., 1972, Parazitologiia, Leningrad, v. 6 (2), 171-175
Eimeria tenella, E. mitis, chickens (exper.), changes in total, residual, and protein nitrogen content in liver, depends on stage of development of parasite, host age, and species of Coccidia

Eimeria tenella
  Takhar, B. S.; and Farrell, D. J., 1979, Brit. Poultry Sc., v. 20 (2), 197-211
    Eimeria acervulina- or Eimeria tenella-infected chickens, energy and nitrogen metabolism

Eimeria tenella
  Tamas, T.; et al., 1978, Poultry Science, v. 57 (2), 381-385
    Eimeria acervulina, E. maxima, E. necatrix, E. tenella, arprinocid and dichloro analog L-628,914 in feed, decrease in oocysts' capacity to sporulate

Eimeria tenella
  Timchenko, A. D., 1972, Parazitologiia, Leningrad, v. 6 (6), 509-512
    Eimeria spp., survival of non-sporulated and sporulated oocysts on soil surface and at various depths under conditions prevailing in southern Ukraine during winter to spring period, implications for prophylaxis and control of coccidiosis

Eimeria tenella
  Tsang, C. L., 1977, J. Agric. and Forest., v. 26, 1-11
    Eimeria tenella, chickens, immunization using oocysts attenuated by chilling method

Eimeria tenella
  Tsang, C. L.; and Chen, K. H., 1977, J. Chinese Soc. Vet. Sc., v. 3 (2), 9-13
    Eimeria tenella, several breeds of chickens, resistance and susceptibility correlated with heredity: Taiwan

Eimeria tenella, illus.
  Wang, C. C., 1978, Comp. Biochem. and Physiol., v. 61B (4), 571-579
    Eimeria tenella, ribosomes, purification from oocysts, characterization, biochemical activity, pharmacological properties, all evidence demonstrates prokaryotic characteristics of these ribosomes and suggests that the parasite may be one of the most primitive eukaryotes

Eimeria tenella
  Wang, C. C.; et al., 1979, Biochem. Pharmacol., v. 28 (15), 2249-2260
    arprinocid inhibits hypoxanthine-guanine transport, may be mode of anticoccidial action

Eimeria tenella
  Wang, C. C.; Simashkevich, P. M.; and Stotish, R. L., 1979, Biochem. Pharmacol., v. 28 (15), 2241-2248
    Eimeria tenella, mode of anticoccidial action of arprinocid

Eimeria tenella
  Wang, C. C.; and Stotish, R. L., 1978, Comp. Biochem. and Physiol., v. 61B (2), 307-313
    Eimeria tenella, multiple leucine aminopeptidases in oocysts, changes during sporulation

Eimeria tenella
  Welliver, R. C.; and Ogra, P. L., 1978, J. Am. Vet. Med. Ass., v. 173 (5, pt. 2), 560-564
    importance of local immunity in enteric infection, colloquium presentation with brief mention of Eimeria tenella

Eimeria tenella
  Williams, R. B., 1978, Vet. Parasitol., v. 4 (2), 193-197
    Eimeria tenella, failure to experimentally infect Pavo cristatus

Eimeria tenella
  Witlock, D. R.; and Wyatt, R. D., 1978, Avian Dis., v. 22 (3), 481-486
    Eimeria tenella, dietary aflatoxin, and combination of both, broiler chicks, prolongation of prothrombin time

Eimeria townsendi
  Gottschalk, C., 1973, Ang. Parasitol., v. 14 (1), 44-54
    endo-parasites of Lepus europaeus, seasonal dynamics, distribution according to locality, sex and age of host, economic importance of parasitism for regional hunting: Ostthuringen, DDR

Eimeria townsendi (Carvalho 1943) Pellerdy 1956, illus.
  Samoil, H. P.; and Samuel, W. M., 1977, Canad. J. Zool., v. 55 (10), 1671-1683
    synonymy, redescription, key
    Lepus americanus (feces): near Rochester, north-central Alberta

Eimeria uniungulati, illus.
  Tomanovic, B., 1977, Vet. Glasnik, v. 31 (4), 281-285
    morphology, biology
    horse: Yugoslavia

Eimeria ursini Supperer 1957
  Barker, I. K.; Munday, B. L.; and Presidente, P. J. A., 1979, J. Parasitol., v. 65 (3), 451-456
    "probability of misidentification of the type host"
    measurements
    Lasiorhinus latifrons (feces): South Australia

Eimeria utahensis, illus.
  Ernst, J. V.; and Chobotar, B., 1978, J. Parasitol., v. 64 (1), 27-34
    Eimeria utahensis, endogenous life cycle in Dipodomys ordii (exper.)

Eimeria uzbekistanica sp. n., illus.
  Davronov, O., 1973, Parazitologiia, Leningrad, v. 7 (1), 79-86
    Meriones meridianus: Karshinsk and Ul'ianovsk regions, Uzbek SSR

Eimeria variabilis (Thelohan, 1893) Reichenow, 1921, illus.
  Davies, A. J., 1978, J. Protozool., v. 25 (1), 15-21
    synonymy, redescription, electron microscopy, cytochemistry
    Cottus bubalis (pyloric ceca and rectum): Aberystwyth and Porth Cwyfan, Wales, U. K.

Eimeria vermiformis Ernst, Chobotar & Hammond, 1971, illus.
  Kelley, G. L.; and Youssef, N. N., 1977, Ztschr. Parasitenk., v. 53 (1), 23-29
    Eimeria vermiformis, development from sporozoite to mature first-generation schizonts in cell cultures

Eimeria virginianus
Woolf, A.; and Harder, J. D., 1979, Wildlife Monogr. (67), 53 pp.
Odocoileus virginianus: Rachelwood Wildlife Research Preserve, New Florence, Pennsylvania

Eimeria vulpis
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Vulpes vulpes (mittleres Jejunum, Ileum, Rectum): Neusiedlerseegebiet, nordlichen Burgenland

Eimeria waiganiensis sp. n., illus.
Varghese, T., 1978, J. Parasitol., v. 64 (2), 312-314
Chalcophaps indica
Otidiphaps nobilis
(feces of both): both from Waigani area, University of Papua New Guinea

Eimeria weybridgensis
Mason, P., 1977, N. Zealand Vet. J., v. 25 (1-2), 30-33
Eimeria spp., lambs, development of naturally acquired infection: Otago, New Zealand

Eimeria wombati (Gilruth and Bull, 1912) comb. nov., illus.
Barker, I. K.; Munday, B. L.; and Presidente, P. J. A., 1979, J. Parasitol., v. 65 (3), 451-456
measurements, redescription
Syns.: Eimeria tasmaniae (Supperer, 1957); Ileocystis wombati Gilruth and Bull, 1912
Lasiorhinus latifrons (feces, duodenum, lower small intestine): South Australia

Eimeria wyomingensis
Cotteleer, C.; and Fameree, L., 1978, Schweiz. Arch. Tierh., v. 120 (3), 149-156
Eimeria spp., cattle, rate of infestation: Belgium

Eimeria wyomingensis
Hiepe, T.; Romeyke, D.; and Jungmann, R., 1978, Monatsh. Vet.-Med., v. 33 (23), 904-910
Eimeria spp., calves reared under conditions of industrialized cattle farming, course of infection, clinical symptoms, monthly distribution, control measures: DDR

Eimeria yakimoffmatschoulskyi
Restani, R., 1974, Parassitologia, v. 16 (1), 83-86
capra (exper.)
agnello (exper.)
(feces of all)

Eimeria zurnabadensis Yakimoff, 1931, illus.
Musaev, M. A.; and Manafova, Sh. G., 1971, Parazitologiia, Leningrad, v. 5 (4), 310-315
Eimeria zurnabadensis, valid species, not a synonym of Eimeria canadensis

Eimeria zuernii
Cotteleer, C.; and Fameree, L., 1978, Schweiz. Arch. Tierh., v. 120 (3), 149-156
Eimeria spp., cattle, rate of infestation: Belgium

Eimeria zurni
Evplov, N. N.; and Nazarov, V. G., 1977, Veterinariia, Moskva (6), 65-66
Eimeria spp., calves, chemococcide effective; compared with biomycin and norsulfazol: Belgorodsk oblast

Eimeria zuernii
Fitzgerald, P. R.; and Mansfield, M. E., 1979, J. Parasitol., v. 65 (5), 824-825
Eimeria spp., calves (exper.), lasalocid

Eimeria zuernii, illus.
Fox, J. E., 1978, Mod. Vet. Pract., v. 59 (8), 599-603
bovine coccidiosis, review, emphasis on prevention and control; field tests, decoquinate against Eimeria bovis and E. zuernii prevented clinical signs of disease with no observable signs of toxicity

Eimeria zuerni
Gobzem, V. R.; and Nazarov, V. G., 1978, Veterinariia, Moskva (3), 67-69
Eimeria spp., calves, diagnostic difficulties, clinical symptoms, chemoprophylactic substances tested at various dosages and in various combinations

Eimeria zuerni
Graefner, G.; Graubmann, H. D.; and Kron, A., 1978, Monatsh. Vet.-Med., v. 33 (23), 910-912
Eimeria spp., cattle on rearing and fattening farms, intensity and extensity of infection in relation to certain environmental conditions: DDR

Eimeria zuerni
Hiepe, T.; Romeyke, D.; and Jungmann, R., 1978, Monatsh. Vet.-Med., v. 33 (23), 904-910
Eimeria spp., calves reared under conditions of industrialized cattle farming, course of infection, clinical symptoms, monthly distribution, control measures: DDR

E[imeria] zurni
Lochkarev, V. A.; and Evpak, I. D., 1977, Veterinariia, Moskva (6), 64-65
E[imeria] zurni, E. smithi, calves, novarsenol effective

Eimeria zuernii
Ryley, N. G.; and Ryley, J. F., 1978, Parasitology, v. 77 (1), 33-39
Eimeria, 10 spp., survival and viability of oocysts following prolonged exposure to saturated sodium chloride solution, additional observations on virulence of Eimeria tenella for chickens following such exposure, no evidence obtained to contraindicate use of salt-flotation methods for separation of oocysts from feces

Eimeria zuernii
Stockdale, P. H. G.; and Yates, W. D. G., 1978, Vet. Parasitol., v. 4 (3), 209-214
Eimeria zuernii, calves (exper.), successful chemotherapy with amprolium or monensin, resistance to reinfection after chemotherapy

Eimeria zuerni, illus.
Supperer, R., 1973, Tieraerztl. Prax., v. 1 (4), 403-412
parasites of calves, diagnosis, pathology, therapy, review

Eimeria zuernii
Ward, J. K.; Ferguson, D. L.; and Parkhurst, A. M., 1979, J. Animal Sc., v. 49 (2), 306-309
gastrointestinal parasites, beef cows (feces), level of infection, effect of animal age and season of year: Mead, Nebraska

Eimeria zuernii
Zajicek, D.; et al., 1977, Veterinarstvi, v. 27 (11), 507-508
Eimeria spp., calves, prevalence in large-scale breeding

Eimeridae [sp.]
Byman, D.; et al., 1977, Canad. J. Zool., v. 55 (2), 376-380
Canis lupus (feces): northeastern Minnesota

Eimeriidae
Cotteleer, C.; and Fameree, L., 1976, Rev. Med. Liege, v. 31 (23), 729-734
Eimeriidae, incidence in cats: Belgique

Embryocola sp., illus.
Michajlow, W., 1970, Acta Parasitol. Polon., v. 18 (42-50), 473-482
Acanthocyclops vernalis

Embryocola sp.
Michajlow, W., 1978, Bull. Acad. Polon. Sc., Cl. V, s. Sc. Biol., v. 26 (1), 51-54
Pseudoboeckella silvestri: reservoir on King George Island, Southern Shetland Islands, Antarctica

Embryocola diflagellatus Mich. et Wita, illus.
Michajlow, W., 1977, Zhurnal Obsh. Biol., v. 38 (5), 657-675
Microcyclops varicans

Embryocola sulcatus Mich.
Michajlow, W., 1978, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 26 (4), 269-272
Mesocyclops leuckarti: water body of Botanical Gardens, Brisbane and Melbourne, Australia

Encephalitozoon
Pletcher, J. M., 1978, J. Am. Vet. Med. Ass., v. 172 (3), 216, 219 [Letter]
". . .continued use of 'nosema' as a synonym for 'encephalitozoon' is not in keeping with current knowledge"; morphologic differences and differing host specificities separate the two genera

Encephalitozoon [sp.]
Botha, W. S.; Van Dellen, A. F.; and Stewart, C. G., 1979, J. South African Vet. Ass., v. 50 (2), 135-144
Encephalitozoon [sp.], dogs (nat. and exper.), clinical, clinicopathological, macroscopic, and histopathological findings, electron microscopic and cultural confirmation, transmission experiment: South Africa

Encephalitozoon sp.
Carpenter, J. W.; and Hillman, C. N., 1979, Ann. Proc., Am. Ass. Zoo Vet., 36-47
Hepatozoon sp., Encephalitozoon sp., mixed infection in Siberian ferret kits
Mustela eversmanni: Patuxent Wildlife Research Center, Laurel, Maryland

Encephalitozoon cuniculi
Bywater, J. E. C., 1979, Lab. Animals, v. 13 (2), 149-151
encephalitozoonosis, discussion of reported human cases, doubt that is is zoonotic

Encephalitozoon cuniculi
Bywater, J. E.; and Kellett, B. S., 1978, Infect. and Immun., v. 21 (2), 360-364
Encephalitozoon cuniculi, existence in specific-pathogen-free rabbit colony, small-sized samples failed to reveal presence of infection with low prevalence, organism probably present in original stock of unit, possibility of establishing Encephalitozoon-free colony by culling all positive reactors using India ink immunoreaction test, incidence (familial, sexual, and age-related) and possible routes of transmission

Encephalitozoon cuniculi
Bywater, J. E. C.; and Kellett, B. S., 1978, Lab. Animal Sc., v. 28 (4), 402-404
Encephalitozoon cuniculi, specific pathogen-free rabbit colony, diagnosis by modified India-ink immunoreaction test, eradication by culling of seropositive animals

Encephalitozoon cuniculi
Bywater, J. E. C.; and Kellett, B. S., 1979, Lab. Animals, v. 13 (4), 293-297
Encephalitozoon cuniculi, 4 generations of a family of rabbits tested at different ages, antibody titres, distribution of histopathological lesions

Encephalitozoon cuniculi
Cox, J. C.; and Gallichio, H. A., 1978, Research Vet. Sc., v. 24 (2), 260-261
Encephalitozoon cuniculi in rabbits, histopathology, value of serology (indirect immunofluorescence) in early diagnosis

Encephalitozoon cuniculi Levaditi, Nicolau & Schoen, illus.
Cox, J. C.; Hamilton, R. C.; and Attwood, H. D., 1979, J. Protozool., v. 26 (2), 260-265
Encephalitozoon cuniculi, rabbits infected orally, intratracheally, or intravenously, serum antibody levels, excretion of spores in urine, frequency of organisms in several organs and severity of lesions

Encephalitozoon cuniculi, illus.
Gannon, J., 1978, Lab. Animals, v. 12 (3), 125-127
Encephalitozoon cuniculi, rabbits (nat. and exper.), diagnosis, immunoperoxidase test, comparison with immunofluorescence test

Encephalitozoon cuniculi, illus.
Hamilton, R. C.; Cox, J. C.; and Pye, D., 1977, J. Gen. Microbiol., v. 98 (1), 305-307
Encephalitozoon cuniculi, wall structure of sporonts grown in human fibroblasts

Encephalitozoon cuniculi Levaditi, Nicolau, Schoen, 1923
Kaliakin, V. N.; and Slepchenko, A. R., 1971, Parazitologiia, Leningrad, v. 5 (6), 559-562
Encephalitozoon cuniculi, mice of several strains and substrains, no natural infections found, susceptibility to experimental infection, parasite virulence increases with mouse passage

Encephalitozoon cuniculi
Kellett, B. S.; and Bywater, J. E. C., 1978, Lab. Animals, v. 12 (2), 59-60
Encephalitozoon cuniculi, detection of antibodies in rabbit sera, modified india-ink immuno-technique

ncephalitozoon cuniculi
Kellett, B. S.; and Bywater, J. E. C., 1979, Lab. Animals, v. 13 (3), 197-198
Encephalitozoon cuniculi, technique for photographing india-ink immunoreaction

ncephalitozoon cuniculi
Pye, D.; and Cox, J. C., 1979, Lab. Animals, v. 13 (3), 193-195
Encephalitozoon cuniculi, in vitro infectivity assay based on enumeration of lesions which appear as macroscopically distinct foci in cell cultures

ncephalitozoon cuniculi
Shadduck, J. A.; et al., 1979, J. Parasitol., v. 65 (1), 123-129
Encephalitozoon cuniculi, infectivity of rabbit isolate for rabbits, mice, rats, and rhesus monkeys, macroscopic and microscopic lesions; infectivity of rabbit, mouse, and hamster isolates in cell culture and in mice

ncephalitozoon cuniculi, illus.
Shadduck, J. A.; Bendele, R.; and Robinson, G. T., 1978, Vet. Path., v. 15 (4), 449-460
Encephalitozoon cuniculi in puppies (renal tubule cells, endothelial cells, brain), clinical signs, lesions, description of organism, immunofluorescence serology: Texas

cephalitozoon cuniculi, illus.
Shadduck, J. A.; and Geroulo, M. J., 1979, Lab. Animal Sc., v. 29 (3), 330-334
Encephalitozoon cuniculi, rabbits, diagnosis, indirect microagglutination test

cephalitozoon cuniculi
Shadduck, J. A.; Kelsoe, G.; and Helmke, R. J., 1979, J. Parasitol., v. 65 (1), 185-188
microsporida spontaneously contaminating cell culture prepared from baboon placental cells, ultrastructurally identical to Encephalitozoon cuniculi and different from Nosema connori

ephalitozoon cuniculi
Shadduck, J. A.; and Polley, M. B., 1978, J. Protozool., v. 25 (4), 491-496
Encephalitozoon cuniculi, propagation in vitro using rabbit choroid plexus (RCP) cells, some factors influencing infectivity and replication (passage level of organisms; passage level, age, and source of RCP cells; antibiotics; storage time and temperature including freezing; elevated temperature; chemical disinfectants; centrifugation; physical and chemical treatments)

ephalitozoon cuniculi
Waller, T., 1979, Lab. Animals, v. 13 (3), 227-230
Encephalitozoon cuniculi, survival of spores after exposure to various temperatures and disinfectants; growth-inhibition effect of drugs in cell cultures

cephalitozoon (Nosema) cuniculi
Waller, T.; Lyngset, A.; and Morein, B., 1979, Lab. Animals, v. 13 (3), 207-208
Encephalitozoon cuniculi, rabbits, diagnosis, simple method for collection of blood on filter paper for india-ink immunoreaction

Encephalitozoon cuniculi
Waller, T.; Morein, B.; and Fabiansson, E., 1978, Lab. Animals, v. 12 (3), 145-148
Encephalitozoon cuniculi, rabbits, humoral immune response following different routes of infection, india-ink immunoreaction test, indirect immunofluorescent antibody test, and immunodiffusion test, immunoglobulin classes involved, possible use of results in eradication program

Encephalitozoon cuniculi
Wilson, J. M., 1979, Research Vet. Sc., v. 26 (1), 114
Encephalitozoon cuniculi, presence of antibodies in blood of Oryctolagus cuniculus and Vulpes vulpes, india ink immunoreaction Vulpes vulpes (brain, ileum smooth muscle tissue): Chipping Norton in Oxfordshire

Endamoeba histolytica. See Entamoeba histolytica.

Endolimax sp.
Marzochi, M. C. de A., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (4), 249-256
human intestinal parasites, eggs and cysts from water used to irrigate vegetable gardens, increased risk of crop contamination in dry season: Ribeirao Preto, Sao Paulo, Brazil

E[ndolimax] nana
Apt, W.; et al., 1977, Rev. Med. Chile, v. 105 (6), 388-389
human intestinal protozoa, modified Teleman and PAFS methods compared for diagnostic purposes

Endolimax nana
Bartlett, M. S.; et al., 1978, J. Clin. Microbiol., v. 7 (6), 524-528
modified zinc sulfate flotation technique evaluated in comparison with formalin-ether concentration method for recovery of protozoan cysts and helminth eggs and larvae from feces preserved in formalin less than and longer than 1 month, results suggest that (except for schistosomes) F-$ZnSO_4$ compares favorably to FE method for detecting infections of clinical significance

Endolimax nana
Blecka, L. J., 1978, J. Parasitol., v. 64 (2), 362-363
patients infected with intestinal parasites, attempt to quantitate immunoglobulin levels in fecal extracts with radial immunodiffusion, mean IgA levels higher than in controls, other immunoglobulin classes rarely detectable

Endolimax nana
Echeverria, P.; and Cross, J. H., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (4), 476-479
Giardia lamblia, Entamoeba spp., Endolimax nana, study of American college students who had had high incidence of diarrheal disease while traveling in the Orient, findings suggest that G. lamblia and Entamoeba histolytica should be considered in differential diagnosis of gastrointestinal disease of travellers to the Orient

E[ndolimax] nana
Okelo, G. B. A.; Mukiibi, J. M.; and Kyobe, J., 1979, East African Med. J., v. 56 (6), 266-269
intestinal parasites, humans, mean eosinophil count higher in infected patients vs. controls, no correlation between eosinophil levels and different intestinal helminths: Nairobi, Kenya

Endolimax nana
Pasricha, A.; and Narang, P., 1978, Indian J. Med. Research, v. 67, 934-936
human intestinal parasites, evaluation of kerosene as substitute for ether in the formol-ether concentration diagnostic technique, morphology of ova and cysts equally well preserved by both techniques

Endolimax nana
Sargeaunt, P. G.; and Williams, J. E., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (2), 225-227
pathogenic and non-pathogenic intestinal amoeba of man, all species can easily be distinguished by characteristic electrophoretic isoenzyme patterns

Endolimax nana
Soto Travieso, R.; et al., 1977, Rev. Cubana Med. Trop., v. 29 (3), 115-120
human intestinal parasites, diagnosis, modified Baermann's technique compared with standard diagnostic methods

Endolimax nana
William, D. C.; et al., 1978, Sex. Transmit. Dis., v. 5 (4), 155-157
enteric protozoa, Enterobius vermicularis, high rate of infection in homosexual men who practice anilingus: New York City

Endotrypanum
Williams, P.; and Coelho, M. de V., 1978, Advances Parasitol., v. 16, 1-42
Leishmania, taxonomy and nomenclature, Phlebotominae as hosts for Trypanosomatidae, review

Endotrypanum sp.
Zeledon, R.; Ponce, C.; and Murillo, J., 1979, J. Parasitol., v. 65 (2), 275-279
Bradypus griseus
Choloepus hoffmanni (spleen)
all from Costa Rica

Endotrypanum schaudinni Mesnil & Brimont, 1908, illus.
Christensen, H. A.; and Herrer, A., 1976, J. Med. Entom., v. 13 (3), 299-303
Endotrypanum schaudinni in Choloepus hoffmanni, infection acquired in laboratory-reared Lutzomyia sanguinaria, L. gomezi, and L. trapidoi during xenodiagnostic trials, infection rates and localization of flagellates in sand fly gut

Entamoeba
Cavier, R.; and Cenac, J., 1972, Therapie, v. 27 (4), 733-742
Trichomonas vaginalis, Entamoeba, in vitro and in vivo (rats, hamsters), efficacy of flunidazole compared with metronidazole

Entamoeba
Dutta, G. P., 1977, J. Scient. and Indust. Research, India, v. 36 (5), 226-240
Entamoeba histolytica and other anaerobic amoebae, cytoplasmic inclusions, recent advances in cytochemistry and ultrastructure role in physiology and pathogenesis, review

Entamoeba
Gutteridge, W. E.; and Coombs, G. H., 1977, Biochemistry of parasitic protozoa, 172 pp., illus.
biochemistry of parasitic protozoa, textbook: methodology; catabolism and generation of energy; nucleic acid metabolism; protein metabolism; lipid metabolism; biochemical mechanism of drug action; isolation of parasitic protozoa from infected animals; culture of parasitic protozoa

Entamoeba
Lumsden, W. H. R., 1978, Handb. Exper. Immunol. (Weir), 3. ed., 42.1-42.47
demonstration of antibodies to Protozoa, extensive review

Entamoeba
Meerovitch, E., 1978, Methods Cultiv. Parasites in Vitro, 19-37
Entamoeba, Giardia lamblia, Trichomonas vaginalis, cultivation, review

Entamoeba
Sebesteny, A., 1979, Lab. Animals, v. 13 (3), 189-191
intestinal protozoa, successful transmission to and establishment in specific-pathogen-free mice exposed to intestinal contents of infected hamsters

Entamoeba sp.
Amaro, A.; Sena, S.; and dos Santos, E., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 123
Atelopus moreirae: Veu da Noiva, Maromba, em Itatiaia, no Estado do Rio de Janeiro, Brasil

Entamoeba sp.
Goldsmid, J. M.; and Rogers, S., 1978, J. South African Vet. Ass., v. 49 (2), 109-111
Papio ursinus (feces): Rhodesia, imported from Northern Transvaal

Entamoeba sp. (Laredo isolate), illus.
Injeyan, H.; Huebner, E.; and Meerovitch, E., 1979, J. Protozool., v. 26 (2), 253-259
Entamoeba sp. (Laredo isolate), morphologically distinct colchicine-resistant variant, properties compared to those of parent strain

Entamoeba sp.
Marzochi, M. C. de A., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (4), 249-256
human intestinal parasites, eggs and cysts from water used to irrigate vegetable gardens, increased risk of crop contamination in dry season: Ribeirao Preto, Sao Paulo, Brazil

Entamoeba sp.
Marzochi, M. C. de A., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (3), 148-155
enteroparasitic cysts and eggs, contamination of green vegetables and kitchen garden soils, epidemiological survey, most commonly found during dry-season when fecal polluted brooks were used for irrigation: Ribeirao Preto, Sao Paulo, Brasil

Entamoeba barreti
Diamond, L. S.; Harlow, D. R.; and Cunnick, C. C., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (4), 431-432
Entamoeba spp., new medium for axenic cultivation

Entamoeba barreti
Gillin, F. D.; and Diamond, L. S., 1978, J. Protozool., v. 25 (4), 539-543
Entamoeba histolytica, other Entamoeba spp., technique for clonal growth in agar, possible use in drug testing

Entamoeba bovis, illus.
Kingston, N.; and Stabler, R. M., 1978, J. Parasitol., v. 64 (1), 14-16
brief description
captive Odocoileus virginianus (feces): Georgia

Entamoeba coli
Akhtaruzzaman, K. M.; et al., 1978, Tropenmed. u. Parasitol., v. 29 (4), 427-431
comparison of different methods for detection of intestinal protozoa and helminths in human stool

Entamoeba coli
Apt, W.; et al., 1977, Rev. Med. Chile, v. 105 (6), 388-389
human intestinal protozoa, modified Teleman and PAFS methods compared for diagnostic purposes

Entamoeba coli
Bartlett, M. S.; et al., 1978, J. Clin. Microbiol., v. 7 (6), 524-528
modified zinc sulfate flotation technique evaluated in comparison with formalin-ether concentration method for recovery of protozoan cysts and helminth eggs and larvae from feces preserved in formalin less than and longer than 1 month, results suggest that (except for schistosomes) F-$ZnSO_4$ compares favorably to FE method for detecting infections of clinical significance

ntamoeba coli
Blecka, L. J., 1978, J. Parasitol., v. 64 (2), 362-363
patients infected with intestinal parasites, attempt to quantitate immunoglobulin levels in fecal extracts with radial immunodiffusion, mean IgA levels higher than in controls, other immunoglobulin classes rarely detectable

tamoeba coli
de Carneri, I.; and Grassi, L., 1974, Parassitologia, v. 16 (1), 79-81
Dientamoeba fragilis, Entamoeba histolytica, E. coli, prevalence in 500 individuals, taxonomic implications of differential drug sensitivity

ntamoeba coli
Chu, J. K., 1972, Taehan Uihak Hyophoe Chi (J. Korean Med. Ass.), v. 15 (8), 685-690
human parasites, differential diagnosis

Entamoeba coli
Echeverria, P.; and Cross, J. H., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (4), 476-479
Giardia lamblia, Entamoeba spp., Endolimax nana, study of American college students who had had high incidence of diarrheal disease while traveling in the Orient, findings suggest that G. lamblia and Entamoeba histolytica should be considered in differential diagnosis of gastrointestinal disease of travellers to the Orient

Entamoeba coli
Engbaek, K., 1978, Ugeskr. Laeger, v. 140 (1), 14-17
Giardia lamblia and other intestinal parasites of children in day-nurseries and kindergartens, incidence, clinical symptoms, management: Denmark

Entamoeba coli
Engbaek, K.; and Larsen, S. O., 1979, Ugeskr. Laeger, v. 141 (17), 1128-1131
Giardia lamblia, Entamoeba coli, Trichuris trichiura, socio-epidemiological study of infected families, emphasis on living conditions, social standards, and clinical symptoms: Denmark

Entamoeba coli, illus.
Goldsmid, J. M.; and Rogers, S., 1978, J. South African Vet. Ass., v. 49 (2), 109-111
Papio ursinus (feces): Rhodesia, imported from Northern Transvaal

Entamoeba coli, illus.
Kingston, N.; and Stabler, R. M., 1978, J. Parasitol., v. 64 (1), 14-16
brief description
captive Odocoileus virginianus (feces): Georgia

Entamoeba coli
Mirza, M. Y.; and al-Rawas, A. Y., 1978, J. Protozool., v. 25 (3, pt. 2), 285-286
Nesokia indica: Baghdad district, Iraq

Entamoeba coli
Molina, J. G., 1971, Rev. Soc. Brasil. Med. Trop., v. 5 (1), 17-20
human intestinal parasites, comparison of Kato and Hoffman diagnostic methods; Kato method clearly superior only in detecting Trichocephalus trichiurus

Entamoeba coli
Owen, D., 1978, Lab. Animals, v. 12 (2), 79-80
Entamoeba coli of human and primate origin, attempted transmission to specified-pathogen-free rodents, results indicate that rodent-human contact is probably not responsible for infiltration of SPF barriers
mice (exper.)
rats (exper.)

Entamoeba coli
Pasricha, A.; and Narang, P., 1978, Indian J. Med. Research, v. 67, 934-936
human intestinal parasites, evaluation of kerosene as substitute for ether in the formol-ether concentration diagnostic technique, morphology of ova and cysts equally well preserved by both techniques

Entamoeba coli, illus.
Proctor, E. M., 1977, Ztschr. Parasitenk., v. 52 (1), 113-115
Entamoeba coli, filamentous virus-like material in nucleus of trophozoites from human colon and from culture

Entamoeba coli
Rajasekaran, P., 1978, Madras Agric. J., v. 65 (9), 613-616
intestinal parasites, children, infection rates in relation to sources of drinking water (open dug wells, water taps in street, water tap within household): Athur block, Madurai district, Tamil Nadu

Entamoeba coli
Rondanelli, E. G.; et al., 1974, Parassitologia, v. 16 (1), 89-91
Entamoeba coli cysts, ultrastructure, comparison with E. histolytica encysted in same culture conditions

Entamoeba coli
Sargeaunt, P. G.; and Williams, J. E., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (2), 164-166
Entamoeba histolytica (14 stocks), E. coli (1 stock), electrophoretic isoenzyme patterns

Entamoeba coli
Scaglia, M.; et al., 1979, Internat. Arch. Allergy and Applied Immunol., v. 59 (4), 465-468
intestinal parasite load in relation to serum IgE levels in individuals from 2 ethnic groups from North and South Rwanda

Entamoeba coli
Soto Travieso, R.; et al., 1977, Rev. Cubana Med. Trop., v. 29 (3), 115-120
human intestinal parasites, diagnosis, modified Baermann's technique compared with standard diagnostic methods

Entamoeba coli
William, D. C.; et al., 1978, Sex. Transmit. Dis., v. 5 (4), 155-157
enteric protozoa, Enterobius vermicularis, high rate of infection in homosexual men who practice anilingus: New York City

Entamoeba gingivalis
Dzhemileva, T.; Stoianova, O.; and Popov, K., 1976, Stomatologiia, Sofiia, v. 58 (3), 165-169
Entamoeba gingivalis, incidence in young people with normal gingiva, or with chronic catarrhal or experimental gingivitis, possible significance of parasite enzymes in pathogenicity

Entamoeba gingivalis
Stoianova, O.; and Dzhemileva, T., 1976, Stomatologiia, Sofiia, v. 58 (2), 147-152
Entamoeba gingivalis, humans, etiology in peridontal diseases

Entamoeba hartmanni
Bartlett, M. S.; et al., 1978, J. Clin. Microbiol., v. 7 (6), 524-528
modified zinc sulfate flotation technique evaluated in comparison with formalin-ether concentration method for recovery of protozoan cysts and helminth eggs and larvae from feces preserved in formalin less than and longer than 1 month, results suggest that (except for schistosomes) F-$ZnSO_4$ compares favorably to FE method for detecting infections of clinical significance

Entamoeba hartmanni
Blecka, L. J., 1978, J. Parasitol., v. 64 (2), 362-363
patients infected with intestinal parasites, attempt to quantitate immunoglobulin levels i fecal extracts with radial immunodiffusion, mean IgA levels higher than in controls, other immunoglobulin classes rarely detectabl

Entamoeba hartmanni
Echeverria, P.; and Cross, J. H., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (4), 476-479
Giardia lamblia, Entamoeba spp., Endolimax nana, study of American college students who had had high incidence of diarrheal disease while traveling in the Orient, findings suggest that G. lamblia and Entamoeba histolytica should be considered in differential diagnosis of gastrointestinal disease of travellers to the Orient

Entamoeba hartmanni
Pasricha, A.; and Narang, P., 1978, Indian J. Med. Research, v. 67, 934-936
human intestinal parasites, evaluation of kerosene as substitute for ether in the formol-ether concentration diagnostic technique, morphology of ova and cysts equally well preserved by both techniques

Entamoeba hartmanni
Sargeaunt, P. G.; and Williams, J. E., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (2), 225-227
pathogenic and non-pathogenic intestinal amoeba of man, all species can easily be distinguished by characteristic electrophoretic isoenzyme patterns

Entamoeba histolytica
Abdus Sattar, A. E. M., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 201-202
Entamoeba histolytica, 35 year-old male, case report, cutaneous amoebic ulcer on right leg: Bangladesh

Entamoeba histolytica
Ailiarova, S. T., 1972, Parazitologiia, Leningrad, v. 6 (1), 19-21
Entamoeba histolytica, hyaluronidase activity of 10 strains compared, activity reduced but not lost by long laboratory cultivation, does not depend on associated microflora but on amoebae themselves

Entamoeba histolytica, illus.
Ailiarova, S. T., 1973, Parazitologiia, Leningrad, v. 7 (2), 169-174
Entamoeba histolytica strains from patients and from carriers, increase in infection rate and extent of pathological changes in irradiated vs. nonirradiated rats

Entamoeba histolytica
Akhtaruzzaman, K. M.; et al., 1978, Tropenmed. u. Parasitol., v. 29 (4), 427-431
comparison of different methods for detection of intestinal protozoa and helminths in human stool

Entamoeba histolytica
Alam, M.; and Ahmad, S., 1979, Indian J. Exper. Biol., v. 17 (10), 1152-1153
Entamoeba histolytica, prolonged cultivation without frequent subculturing

E[ntamoeba] histolytica
Albarran Trevino, C.; and Valle Gonzalez, A., 1978, Semana Med. Mexico (1167), an. 24, v. 52 (11), 309-312
amoebiasis, human intestinal, surgical complications (fulminant infection, ameboma, appendicitis), clinical review

E[ntamoeba] histolytica
Alonso, R. E., 1976, Semana Med. (4956), an. 83, v. 149 (7), 234-237
intestinal parasites, school children, clinical studies with tinidazole and mebendazole: Institute Aguirre 'Patronato de la Infancia'

Entamoeba histolytica
Alvarez-Cordero, R., 1973, Rev. Brasil. Med., v. 30 (11), 688-691
Entamoeba histolytica, human infections in presence of intestinal complications as appendicitis, colitis, amebomas, clinical management

Entamoeba histolytica
Ambroise-Thomas, P.; Desgeorges, P. T.; and Monget, D., 1978, Bull. World Health Organ., v. 56 (5), 797-804
Toxoplasma gondii, Entamoeba histolytica, Trichinella spiralis, Echinococcus granulosus, human, diagnosis by enzyme-linked immunosorbent assay with a modified micromethod, parallel study by comparative serological tests

ntamoeba histolytica, illus.
Amyx, H. L.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (5), 888-891
Entamoeba histolytica, natural outbreak of amebic dysentery in colony of Ateles spider monkeys with severe hepatic abscesses in many, may provide valuable model host: primate colony, Patuxent Wildlife Research Center, Laurel, Maryland

ntamoeba histolytica
Apt, W.; et al., 1976, Rev. Med. Chile, v. 104 (11), 791-793
Entamoeba histolytica, human intestinal amoebiasis, successful treatment with nimorazole

ntamoeba histolytica
Apt, W.; et al., 1977, Rev. Med. Chile, v. 105 (6), 388-389
human intestinal protozoa, modified Teleman and PAFS methods compared for diagnostic purposes

tamoeba histolytica
Apt, W.; and Morales, A., 1977, Rev. Med. Chile, v. 105 (7), 452-455
human amoebic hepatic abscess, analysis of 7 cases, clinical features, diagnosis, therapy: Santiago, Chile

ntamoeba histolytica
Archampong, E. Q.; and Clark, C. G., 1973, Ann. Roy. Coll. Surg. England, v. 52 (1), 36-48
Entamoeba histolytica, humans, pathology, complications, surgical management, review

ntamoeba histolytica
Argiolas, N.; et al., 1974, Rassegna Med. Sarda, v. 77, n.s., v. 26 (3), 221-260
Entamoeba histolytica, human, extensive historical and diagnostic review, clinical aspects, epidemiology, pathology

Entamoeba histolytica
Aswapokee, N.; et al., 1974, Siriraj Hosp. Gaz., v. 26 (3), 371-376
Entamoeba histolytica, human hepatic abscess, single course of metronidazole plus closed aspiration of abscess, clinical trials, treatment well tolerated without side effects: Siriraj Hospital

Entamoeba histolytica
Atias, A.; et al., 1977, Rev. Med. Chile, v. 105 (7), 456-459
Entamoeba histolytica, invasive intestinal infection in nursing infants, statistics of 30 cases, metronidazole therapy: Chile

Entamoeba histolytica, illus.
Aust-Kettis, A.; and Sundqvist, K. G., 1978, Scand. J. Immunol., v. 7 (1), 35-44
Entamoeba histolytica, distribution and redistribution of antigen determinants and Con A receptors on surface, reappearance of antigen, effect of metabolic inhibitors and pH on ligand induced redistribution, capping and endocytosis in phagocytizing amoebae and influence of inhibitory compounds, variation of expression of surface antigens

Entameba histolytica
Awadalla, M. A.; Hussein, L. A.; and Salem, M., 1979, Nutrition Rep. Internat., v. 19 (5), 627-633
parasitic infestations in children, effect on intestinal absorption as determined by assay of fasting serum carotene and vitamin A levels and by vitamin A tolerance tests: Orphanage Institute of Giza and El-Zeitoun, Egypt

Entamoeba histolytica, illus.
Axton, J. H. M., 1972, South African Med. J., v. 46 (10), 258-259
Entamoeba histolytica, 21-day-old African female infant, case report, amoebic proctocolitis and liver abscess, probable source of infection was fecal contamination at birth, successfully treated with metronidazole

Entamoeba histolytica
Bailenger, J.; et al., 1972, Therapeutique, v. 48 (2), 117-128
dangers of cortisone therapy in the presence of human parasitic pathology, clinical review

Entamoeba histolytica
Band, R. N.; and Cirrito, H., 1979, J. Protozool., v. 26 (2), 282-286
Entamoeba histolytica in axenic culture, growth response to hydrogen, carbon dioxide, and oxygen

Entamoeba histolytica
Bank, S.; et al., 1971, South African Med. J., v. 45 (9), 219-225
Entamoeba histolytica, human, clinical classification of amoebic colitis, case reports, diagnosis, therapy

Entamoeba histolytica
Baranski, M. C.; and da Silva, A. F., 1976, Rev. Brasil. Clin. e Terap., v. 5 (11), 417-420
human intestinal amoebiasis, clinical trials testing the therapeutic effectiveness of tinidazole, recommended for individual and mass therapy because of ease of administration and virtual absence of side effects

Entamoeba histolytica
Barkin, R. M.; et al., 1975, Rocky Mountain Med. J., v. 72 (8), 340-344
medical screening of recently adopted Vietnamese orphans, scabies, Entamoeba histolytica, Giardia lamblia, and Pneumocystis carinii among medical problems encountered: Denver area, Colorado

Entamoeba histolytica
Bartlett, M. S.; et al., 1978, J. Clin. Microbiol., v. 7 (6), 524-528
modified zinc sulfate flotation technique evaluated in comparison with formalin-ether concentration method for recovery of protozoan cysts and helminth eggs and larvae from feces preserved in formalin less than and longer than 1 month, results suggest that (except for schistosomes) F-$ZnSO_4$ compares favorably to FE method for detecting infections of clinical significance

Entamoeba histolytica
Basit, N.; Siddiquiz, M. A.; and Ahmed, Z., [1978], Riv. Parassitol., Roma, v. 38 (2-3), 1977, 259-262
Entamoeba histolytica, in vitro effect of Euphorbia hirta extracts, good prospects

Entamoeba histolytica, illus.
Beaver, P. C.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (6), 1133-1136
Entamoeba histolytica, 4-month-old girl, infection of skin of eyelid with extension into orbit, case report: Colombia

Entamoeba histolytica
Belli, E.; and Palenzona, C., 1976, Rassegna Patol. Appar. Respir., v. 26 (2), 105-109
Entamoeba histolytica, humans, clinical analysis of secondary pulmonary infections of amoebiasis, diagnosis, case reports

Entamoeba histolytica
Benazet, F.; et al., 1970, Scand. J. Infect. Dis., v. 2 (2), 139-143
intestinal and hepatic parasites, nitroheterocyclic antiparasitics, laboratory studies of chemotherapeutic activity and toxicity in exper. animals

Endamoeba histolytica, illus.
Benitez Soto, L., 1974, Semana Med. Mexico (1034), an. 21, v. 81 (10), 295-305
Endamoeba histolytica, human, epidemiology, pathology, diagnosis, review

Entamoeba histolytica
Berkovits, L., 1975, Magy. Radiol., v. 27 (5), 283-289
Entamoeba histolytica, colitis, humans, pathologic changes determined by radiologic means

Entamoeba histolytica
Bertrand, J.; Carli-Basset, C.; and Jankowski, J. M., 1975, Rev. Franc. Gastro-Enterol. (108), 45-49
Entamoeba histolytica, woman with hemorrhagic recto-colitis and ichthyosis associated with intestinal amoebiasis, additional severe resistance to various amoebicidal drugs, case report

Entamoeba histolytica
Beurier, J., 1974, Rev. Infirm. Afrique Noire (24), 5-15
Entamoeba histolytica, human amoebic dysentery, brief clinical review

Entamoeba histolytica
Bhandari, B.; and Sankhla, K., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (3), 345-346 [Letter]
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tourist diarrhea of parasitic and bacterial origins, clinical review

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probable hepatic amoebiasis, young man of African origin, therapy with amoebicides resulted in relief of symptoms: Italy

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parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

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parasitic infections of human liver and bile ducts, radiologic diagnosis, general review

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[Entamoeba] histolytica
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Entamoeba histolytica, humans, pathology associated with various forms of infection (colitis, ameboma, hepatic abscess, genital and cutaneous amoebiasis)

Entamoeba histolytica
Richle, R.; et al., 1978, Arzneimittel-Forsch., v. 28 (4), 612-625
trichomoniasis, amoebiasis, lambliasis, extensive in vitro and in vivo trials (humans, domestic animals, laboratory animals) with ornidazole to establish chemotherapeutic properties, efficacy slightly superior to metronidazole in comparative trials

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Entamoeba histolytica, cutaneous infection of genital area of 14-month-old child, clinical case report

E[ntamoeba] histolytica
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E[ntamoeba] histolytica, human hepatic abscess, diagnosis using the latex agglutination test gave some false positive reactions in normal controls, recommendations therefore that the quantitative indirect agglutination test be used for diagnostic purposes

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Said-Fernandez, S.; and Lopez-Revilla, R., 1978, Ztschr. Parasitenk., v. 56 (3), 219-225
Entamoeba histolytica (two strains), E. invadens, E. moshkovskii, proteins analyzed by disc electrophoresis and immunodiffusion, suggests use as taxonomic criterion

E[ntamoeba] histolytica
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E[ntamoeba] histolytica, humans with chronic intestinal amoebiasis, clinical trials with ornidazole vs. metronidazole

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Entamoeba histolytica
Sargeaunt, P. G.; and Williams, J. E., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (2), 164-166
Entamoeba histolytica (14 stocks), E. coli (1 stock), electrophoretic isoenzyme patterns

Entamoeba histolytica
Sargeaunt, P. G.; and Williams, J. E., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (2), 225-227
pathogenic and non-pathogenic intestinal amoeba of man, all species can easily be distinguished by characteristic electrophoretic isoenzyme patterns

Entamoeba histolytica
Sargeaunt, P. G.; Williams, J. E.; and Grene, J. D., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (5), 519-521
Entamoeba histolytica, differentiation of invasive and non-invasive forms by isoenzyme electrophoresis

Entamoeba [histolytica]
Scaffidi, V., 1973, Riforma Med., v. 87 (19), 743-760
Entamoeba [histolytica], human, amoeboma and ulcerative amoebic colitis, clinical variables, diagnosis, therapeutics

Entamoeba histolytica
Scaglia, M.; et al., 1979, Internat. Arch. Allergy and Applied Immunol., v. 59 (4), 465-468
intestinal parasite load in relation to serum IgE levels in individuals from 2 ethnic groups from North and South Rwanda

Entamoeba histolytica
Scheff, G. J., 1978, Zentralbl. Bakteriol., 1. Abt. Orig., Reihe A, v. 242 (3), 419-421
Entamoeba histolytica, monoxenic cultures, growth enhanced with 6 Fusobacteria strains while Bacteroides strains did not possess this property

Entamoeba histolytica
Schenone, H.; et al., 1979, Bol. Chileno Parasitol., v. 34 (1-2), 2-6
Entamoeba histolytica and/or Giardia lambiia, children, clinical trials with varying doses of tinidazole: ciudad de Santiago

Entamoeba histolytica, illus.
Schiefer, B.; and Loew, F. M., 1978, Vet. Path., v. 15 (3), 428-431
Entamoeba histolytica in pet Lagothrix (ileum), mixed infection with Salmonella, pathology: Saskatoon, Canada

Entamoeba histolytica, illus.
Sepulveda, B.; et al., 1975, Rev. Fac. Med., Univ. Nac. Auton. Mexico, an. 18, v. 18 (1), 5-31
Entamoeba histolytica, extensive round table discussion on diagnosis, clinical management, treatment, immunology

Entamoeba histolytica
Shabot, J. M.; and Patterson, M., 1978, Am. J. Digest. Dis., n.s., v. 23 (2), 110-118
Entamoeba histolytica, 10-year (1966-1976) retrospective analysis of 15 patients with amoebic liver abscess: clinical findings, laboratory findings, management, case reports: Texas

Entamoeba histolytica
Sheehan, D. J.; et al., 1979, J. Clin. Microbiol., v. 10 (2), 128-133
Entamoeba histolytica, human, diagnosis, comparison of microscopic, cultural, counterimmunoelectrophoresis, and indirect hemagglutination techniques

E[ntamoeba] histolytica, illus.
Silard, R.; et al., 1977, Arch. Roumaines Path. Exper. et Microbiol., v. 36 (1), 55-60
E[ntamoeba] histolytica, intrahepatic inoculation in Cricetus auratus of human strain associated with Blastocystis hominis, liver abscess, diagnosis, results demonstrate the tissue adaptability of B. hominis and its potential as a conditioned pathogen

Entamoeba histolytica
Skromne-Kadlubik, G.; Medina, J.; and Rauda, L., 1976, SPM Salud Pub. Mexico, v. 18 (2), 395-398
Entamoeba histolytica, human, comparative survey, conventional medications vs. radiation therapy

Entamoeba histolytica
Smith, M. A. P., 1976, South African Med. J., v. 50 (15), 592-594
amoebiasis, humans, case report of multiple colonic strictures resulting from infection, clinical management

Entamoeba histolytica
Sobrevilla Cruz, R., 1977, Semana Med. Mexico (1172), an. 24. v. 93 (4), 83-86
Entamoeba histolytica, children, prevalence survey, clinical management: Mexico

Entamoeba histolytica
Sobrinho, J. B.; Lima, I. A.; and Brito, E. S. (filho), 1976, Rev. Brasil. Cirug., v. 66 (1-2), 19-24
Entamoeba histolytica, human hepatic abscess, hospital case survey, clinical and surgical treatment review: Manaus, Brazil

E[ntamoeba] histolytica
Solano, J. N. de R.; Merino, C. G.; and Barrera, R. R., 1976, Rev. Brasil. Clin. e Terap., v. 5 (9), 353-365
human intestinal amoebiasis, successful clinical trials using tinidazole

Entamoeba histolytica
Soto Travieso, R.; et al., 1977, Rev. Cubana Med. Trop., v. 29 (3), 115-120
human intestinal parasites, diagnosis, modified Baermann's technique compared with standard diagnostic methods

E[ntamoeba] histolytica
Stemberger, H., 1978, Immun. u. Infekt., v. 6 (2), 71-78
E[ntamoeba] histolytica, investigation of cytolytic action of antibody, complement, and normal human peripheral blood lymphocytes as well as action of peripheral blood lymphocytes from donor with amoebic liver abscess

Entamoeba histolytica
Stierli, J.; and Steffen, R., 1978, Schweiz. Med. Wchnschr., v. 108 (34), 1310-1317
Entamoeba histolytica, retrospective study of 453 persons who became infected through travel or living abroad, symptoms, pathology, therapy: Zurich region of Switzerland

Entamoeba histolytica
Stuiver, P. C.; and Goud, T. J. L. M., 1978, Brit. Med. J. (6134), v. 2, 394-395
Entamoeba histolytica, possible hazards of administering corticosteroids to persons who may have amoebiasis, steroid therapy may result in acute amoebic dysentery or exacerbation of amoebiasis, case reports

Entamoeba histolytica
Takeuchi, T.; et al., 1979, Comp. Biochem. and Physiol., v. 62B (3), 281-285
Entamoeba histolytica, axenically cultivated trophozoites, enzymatic mechanism of L-serine oxidation

Entamoeba histolytica
Thacker, S. B.; et al., 1979, Am. J. Pub. Health, v. 69 (12), 1279-1281
Entamoeba histolytica, Giardia lamblia, control attempts in a residential facility for mentally retarded persons: Washington, D. C.

E[ntamoeba] histolytica
Timmermans, P. M., 1979, East African Med. J., v. 56 (4), 166-169
E[ntamoeba] histolytica, humans, intestinal tissue amoebiasis, treatment with tissue plus luminal amoebicides, 7 case reports; differentiation of tissue vs. uncomplicated amoebic dysenteries

Entamoeba histolytica
Triger, D. R., 1978, J. Trop. Med. and Hyg., v. 81 (4), 54-59
Entamoeba histolytica, human, liver abscess, clinical features, response to therapy, review of 24 cases: Wessex region, Great Britain

Entamoeba histolytica
Trissl, D.; et al., 1978, J. Exper. Med., v. 148 (5), 1137-1145
Entamoeba, several species and strains, comparison of in vitro ingestion of human erythrocytes (HRBC), E. histolytica isolated from cases of human dysentery show significantly higher phagocytic rate of HRBC ingestion than nonpathogenic strains and than other Entamoeba not pathogenic for mammals, however all Entamoeba tested are able to ingest HRBC

Entamoeba histolytica-like ameba, Laredo strain
Trissl, D.; et al., 1978, J. Exper. Med., v. 148 (5), 1137-1145
Entamoeba, several species and strains, comparison of in vitro ingestion of human erythrocytes (HRBC), E. histolytica isolated from cases of human dysentery show significantly higher phagocytic rate of HRBC ingestion than nonpathogenic strains and than other Entamoeba not pathogenic for mammals, however all Entamoeba tested are able to ingest HRBC

Entamoeba histolytica
Trzaska, B.; and Leinweber-Noiszewska, H., 1974, Ann. Acad. Med. Stetinensis, suppl. 10, 19-25
Entamoeba histolytica, acute infection in Polish sailor who acquired disease in West Africa, therapy with bemarsal, mexaform and spiramycin resulted in relapse and chronic infection, apparent cure with metronidazole; pathology and clinical aspects of amoebiasis

Entamoeba histolytica
Unger, K. W., 1978, N. England J. Med., v. 298 (20), 1148 [Letter]
Entamoeba histolytica, increasing incidence of intestinal infections in male homosexuals, importance of inclusion in differential diagnosis in cases with persistent intestinal symptoms: New York

Entamoeba histolytica
de la Vega, J. M., 1973, Rev. Cir. Hosp. Juarez, Mexico (187-188), v. 44, 1972-1973, 22-30
human intestinal amoebiasis, clinical manifestations, review of presenting forms (ameboma, fulminating forms, acute appendicitis), differential diagnosis, pathology: Mexico

Entamoeba histolytica
del Villar Ponce, J. P.; and Alvarez Chacon, R., 1975, Semana Med. Mexico (1071), an. 22, v. 84 (11), 325-327
Entamoeba histolytica, human cutaneous, case report, clinical aspects

Entamoeba histolytica
del Villar Ponce, J. P.; and Alvarez Chacon, R., 1976, Semana Med. Centroamerica y Panama (541), an. 11, v. 27 (6), 148-150
amoebiasis, human cutaneous, case report, clinical aspects: Mexico D.F.

Entamoeba histolytica
Vinayak, V. K.; et al., 1979, J. Parasitol., v. 65 (3), 407
Macaca mulatta naturally infected with ameba morphologically indistinguishable from Entamoeba histolytica, indirect hemagglutination titers to antigens from Entamoeba histolytica and from ameba of Macaca mulatta

Entamoeba histolytica
Vinayak, V. K.; Chitkara, N. L.; and Chhuttani, P. N., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (3), 266-268
Entamoeba histolytica-infected rats pretreated with corticosteroids, irradiation or both, exacerbation of amoebic pathology, corticosteroid therapy possibly aggravates otherwise sub-clinical infection

Entamoeba histolytica
Vinayak, V. K.; and Chopra, A. K., 1978, Ann. Trop. Med. and Parasitol., v. 72 (6), 549-551
Syphacia obvelata-infected mice challenged intracaecally with Entamoeba histolytica trophozoites showed higher level of amoebic infection than mice without helminths, mice with heaviest helminth infection were most susceptible to amoebic infection

Entamoeba histolytica
Vullo, V.; et al., 1979, Boll. Ist. Sieroterap. Milanese, v. 58 (2), 141-147
hydatidosis, amebiasis, urinary schistosomiasis, human, diagnosis, immunoperoxidase and immunofluorescence techniques compared

Entamoeba histolytica
Wang, L. T., 1973, Taiwan i Hsueh Hui Tsa Chih (J. Formosan Med. Ass.), v. 72 (11), 619-623
Entamoeba histolytica, humans, diagnosis by indirect haemagglutination test using antigen from axenic culture, more specific for amoebic liver abscess than for intestinal infection

Entamoeba histolytica
Wang, L. T., 1974, Taiwan i Hsueh Hui Tsa Chih (J. Formosan Med. Ass.), v. 73 (4), 196-202
Entamoeba histolytica, 38 strains cultured from infected stool and liver aspirates, growth and maintenance in various media

Entamoeba histolytica
Wang, L. T.; and Yang, S. P., 1971, Taiwan i Hsueh Hui Tsa Chih (J. Formosan Med. Ass.), v. 70 (3), 131-134
Entamoeba histolytica, human, oxytetracycline-resistant amoebic dysentery: Taiwan

Entamoeba histolytica
Wery, M.; et al., 1976, Ann. Soc. Belge Med. Trop., v. 56 (3), 169-182
Entamoeba histolytica, human, epidemiologic survey using immunofluorescence and fecal smears: Zaire

Entamoeba histolytica
Wery-Paskoff, S.; et al., 1971, Ann. Soc. Belges Med. Trop. Parasitol., v. 51 (2), 221-227
Entamoeba histolytica, human hepatic abscess, diagnosis, indirect fluorescent antibody test

Entamoeba histolytica
William, D. C.; et al., 1978, Sex. Transmit. Dis., v. 5 (4), 155-157
enteric protozoa, Enterobius vermicularis, high rate of infection in homosexual men who practice anilingus: New York City

Entamoeba histolytica
Winkelmann, E.; Raether, W.; and Gebert, U., 1978, Arzneimittel-Forsch., v. 28 (10), 1682-1684
activity of 16 novel 5-nitroimidazoles against protozoa in mice and golden hamsters, compared with metronidazole and tinidazole, structure-activity relationships

Entamoeba histolytica
Wissler, K.; and Eckert, J., 1979, Therap. Umschau, v. 36 (3), 233-240
Entamoeba histolytica, human, seroimmunologic survey comparing indirect immunofluorescent, latex agglutination, and indirect hemagglutination tests

Entamoeba histolytica
Wu, T. P.; et al., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (4), 467-468
Entamoeba histolytica, 49-year-old male, left lung, medical and surgical treatment of 2 abscesses, case report: Sining, China

Entamoeba histolytica
Yang, J.; and Kennedy, M. T., 1979, J. Clin. Microbiol., v. 10 (6), 778-785
Entamoeba histolytica, human, diagnosis, development and evaluation of enzyme-linked immunosorbent assay, compared with indirect fluorescent antibody and indirect hemagglutination techniques

Entamoeba histolytica
Yap, E. H.; et al., 1975, Singapore Med. J., v. 16 (1), 31-38
Entamoeba histolytica, evaluation of indirect haemagglutination test for diagnosis, trophozoites grown in bacteria-free cultures used as antigen, merits of technique discussed

Entamoeba histolytica
Young, K. H.; et al., 1979, J. Clin. Microbiol., v. 10 (6), 852-853
parasites, ethyl acetate as satisfactory substitute solvent in formalin-ether sedimentation technique for fecal specimens

Endamoeba histolytica
Yu, M.; and Kin, T. H., 1970, Singapore Med. J., v. 11 (1), 55-56
Endamoeba histolytica, Balantidium, Trichomonas, culture, new simple method

Endamoeba histolytica
Yu, M.; and Loo, L. A., 1970, Singapore Med. J., v. 11 (1), 59-61
Endamoeba histolytica, diagnosis, indirect fluorescent antibody test

Entamoeba histolytica
Zafar, M. H.; et al., 1976, Rawal Med. J., v. 5 (4), 140-142
Entamoeba histolytica, human, amoebic dysentery, clinical trials with teclozan, 94% cure rate with few side effects

Entamoeba histolytica
Zwierz, C.; et al., 1974, Ann. Acad. Med. Stetinensis, suppl. 10, 27-32
Entamoeba histolytica, human, description of post-parasitic syndrome which results in continued intestinal complaints after parasitic cure, attributed to disturbances in biological equilibrium of alimentary tract

Entamoeba histolytica
Zwierz, C.; et al., 1975, Ann. Soc. Belge Med. Trop., v. 55 (5), 403-413
amoebiasis and other human intestinal parasites, persons returning from tropical countries, laboratory diagnostic problems, test recommendations: Poland

Entamoeba invadens
Bos, H. J., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (1), 120 [Letter]
Entamoeba histolytica, E. invadens, correlation between cytopathogenic effects of trophozoites and their soluble extracts, presence of fetal calf serum largely inhibited cell damage

Entamoeba invadens
Bos, H. J.; et al., 1978, Acta Leidensia, v. 46, 17-30
cryopreservation of parasitic protozoa

Entamoeba invadens
Diamond, L. S.; Harlow, D. R.; and Cunnick, C. C., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (4), 431-432
Entamoeba spp., new medium for axenic cultivation

Entamoeba invadens
Frank, W., 1974, Ann. Soc. Belge Med. Trop., v. 54 (4-5), 343-349
Morelia sp.
Kinosternon leucostomum
Ophiosaurus apodus
(intestine and liver of all)

Entamoeba invadens
Frank, W.; Sachsse, W.; and Winkelstracter, K. H., 1976, Salamandra, v. 12 (3), 120-126
Entamoeba invadens in Chelonia mydas and Caretta caretta, fatal infections, pathology, Cuora amboinensis found to be source of infection for Chelonia mydas: Saarbruckener Zoologischen Garten

Entamoeba invadens
Gillin, F. D.; and Diamond, L. S., 1978, J. Protozool., v. 25 (4), 539-543
Entamoeba histolytica, other Entamoeba spp., technique for clonal growth in agar, possible use in drug testing

Entamoeba invadens, illus.
Ishii, A., 1973, Snake, v. 5 (1-2), 133-140

Entamoeba invadens
Ishii, A.; et al., 1971, Snake, v. 3 (1), 30-34
Entamoeba invadens, oral inoculation in snakes of the Amami Islands, pathology
Trimeresurus flavoviridis
T. okinavensis
Dinodon semicarinatus
Opheodrys semicarinata
Ampiesma pryeri
(all exper.)

Entamoeba invadens
Ishii, A.; et al., 1976, Snake, v. 8 (1), 64-68
Entamoeba invadens, oral inoculation of some amphibians and reptiles, possible biological control of the habu snake
Takydromus tachydromoides
Japalura polygonata
Eumeces marginatus
Pseudemys scripta
Geoclemys reevesii
Trionyx sinensis
Trionyx sp.
(all exper.)

Entamoeba invadens
Ishii, A.; and Hayashi, Y., 1978, Snake, v. 10 (1), 16-17
Entamoeba invadens, unsuccessful attempt to transmit parasitic infection from exper. infected Geoclemys reevesii to Trimeresurus flavoviridis for purposes of biological control

Entamoeba invadens
Ishii, A.; and Noboru, H., 1971, Snake, v. 3 (1), 24-29
Entamoeba invadens, exper. infection of Trimeresurus flavoviridis and Ahaetulla nusutus, pathology; attempts to infect laboratory rats and mice unsuccessful

Entamoeba invadens
Lopez-Revilla, R.; and Gomez, R., 1978, Exper. Parasitol., v. 44 (2), 243-248
Entamoeba histolytica strains, E. invadens, E. moshkovskii, fluctuations of DNA content of axenic trophozoites, variations generally but not always occurred within a range characteristic of each species, possible causes of variation, estimation of DNA content could help to distinguish these species despite variation

Entamoeba invadens, illus.
McCaul, T. F.; and Bird, R. G., 1978, Internat. J. Parasitol., v. 8 (6), 501-506
Entamoeba histolytica, E. invadens, trophozoites, localization of thiamine pyrophosphatase activity within cytoplasmic fine structure

Entamoeba invadens, illus.
Mishra, G. S.; and Gonzalez, J. P., 1978, Arch. Inst. Pasteur Tunis, v. 55 (3), 303-326
Clemmys caspica leprosa: Tunisie

Entamoeba invadens
Mueller, M.; et al., 1979, Comp. Biochem. and Physiol., v. 64B (1), 97-100
Tritrichomonas foetus, Trichomonas vaginalis, Entamoeba invadens, effects of 2,4-dinitrophenol (including effect on accumulation of metronidazole)

Entamoeba invadens
Nseka, K.; and Mueller, M., 1978, Compt. Rend. Soc. Biol., Paris, v. 172 (6), 1094-1098
Tritrichomonas foetus, Entamoeba invadens, effect of glycolysis inhibitors on uptake of metronidazole

ntamoeba invadens
Pennisi, L.; Mento, G.; and Faraone, U., 1976, Riv. Parassitol., Roma, v. 37 (1), 105-111
Entamoeba, isolation of 5 strains from sewage of Messina, characterization by thermo-resistance and behavior in hypotonic solutions, comparison with known Entamoeba strains

ntamoeba invadens
Said-Fernandez, S.; and Lopez-Revilla, R., 1978, Ztschr. Parasitenk., v. 56 (3), 219-225
Entamoeba histolytica (two strains), E. invadens, E. moshkovskii, proteins analyzed by disc electrophoresis and immunodiffusion, suggests use as taxonomic criterion

ntamoeba invadens
Trissl, D.; et al., 1978, J. Exper. Med., v. 148 (5), 1137-1145
Entamoeba, several species and strains, comparison of in vitro ingestion of human erythrocytes (HRBC), E. histolytica isolated from cases of human dysentery show significantly higher phagocytic rate of HRBC ingestion than nonpathogenic strains and than other Entamoeba not pathogenic for mammals, however all Entamoeba tested are able to ingest HRBC

ntamoeba moshkovskii, illus.
Carosi, G., 1974, Ann. Soc. Belge Med. Trop., v. 54 (4-5), 265-277
Entamoeba moshkovskii and free-living amoeba of Hartmannella-Naegleria group, ultrastructural comparisons

ntamoeba moshkovskii
Diamond, L. S.; Harlow, D. R.; and Cunnick, C. C., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (4), 431-432
Entamoeba spp., new medium for axenic cultivation

ntamoeba moshkovskii
Felix-Silva, E.; and Mayrink, W., 1974, Rev. Inst. Med. Trop. S. Paulo, v. 16 (4), 203-221
Emtamoeba moshkovskii, distribution in mud and water, new habitat types, new foci: Brazil; Uruguay

ntamoeba moshkovskii
Gillin, F. D.; and Diamond, L. S., 1978, J. Protozool., v. 25 (4), 539-543
Entamoeba histolytica, other Entamoeba spp., technique for clonal growth in agar, possible use in drug testing

tamoeba moshkovskii
Lopez-Revilla, R.; and Gomez, R., 1978, Exper. Parasitol., v. 44 (2), 243-248
Entamoeba histolytica strains, E. invadens, E. moshkovskii, fluctuations of DNA content of axenic trophozoites, variations generally but not always occurred within a range characteristic of each species, possible causes of variation, estimation of DNA content could help to distinguish these species despite variation

tamoeba moshkovskii
Pennisi, L.; Mento, G.; and Faraone, U., 1976, Riv. Parassitol., Roma, v. 37 (1), 105-111
Entamoeba, isolation of 5 strains from sewage of Messina, characterization by thermo-resistance and behavior in hypotonic solutions, comparison with known Entamoeba strains

Entamoeba moshkovskii
Said-Fernandez, S.; and Lopez-Revilla, R., 1978, Ztschr. Parasitenk., v. 56 (3), 219-225
Entamoeba histolytica (two strains), E. invadens, E. moshkovskii, proteins analyzed by disc electrophoresis and immunodiffusion, suggests use as taxonomic criterion

Entamoeba moshkovskii
Trissl, D.; et al., 1978, J. Exper. Med., v. 148 (5), 1137-1145
Entamoeba, several species and strains, comparison of in vitro ingestion of human erythrocytes (HRBC), E. histolytica isolated from cases of human dysentery show significantly higher phagocytic rate of HRBC ingestion than nonpathogenic strains and than other Entamoeba not pathogenic for mammals, however all Entamoeba tested are able to ingest HRBC

Entamoeba muris
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Rattus norvegicus (Jejunum, Ileum)
Mus musculus (Jejunum, Rectum)
Apodemus flavicollis (Jejunum, Ileum)
A. sylvaticus (Jejunum, Ileum)
Ondatra zibethica (Jejunum, Ileum)
Microtus arvalis (Ileum)
all from Neusiedlerseegebiet, nordlichen Burgenland

Entamoeba nana
Akhtaruzzaman, K. M.; et al., 1978, Tropenmed. u. Parasitol., v. 29 (4), 427-431
comparison of different methods for detection of intestinal protozoa and helminths in human stool

Entamoeba polecki, illus.
Salaki, J. S.; Shirey, J. L.; and Strickland, G.T., 1979, Am. J. Trop. Med. and Hyg., v. 28 (2), 190-193
Entamoeba polecki, 24-year-old Peace Corps volunteer (feces), symptomatic intestinal infection cured with diloxanide furoate and metronidazole: United States (previously in Upper Volta)

Entamoeba terrapinae
Diamond, L. S.; Harlow, D. R.; and Cunnick, C. C., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (4), 431-432
Entamoeba spp., new medium for axenic cultivation

Entamoeba terrapinae
Gillin, F. D.; and Diamond, L. S., 1978, J. Protozool., v. 25 (4), 539-543
Entamoeba histolytica, other Entamoeba spp., technique for clonal growth in agar, possible use in drug testing

Enteromonas hominis, illus.
Harvey, J., 1975, Saguenay Med., v. 20 (5), 146-156
parasitic flagellates, life cycle, infection in man, clinical signs, therapy, brief review

Entodiscidae fam. n.
Iankovskii, A. V., 1973, Parazitologiia, Leningrad, v. 7 (3), 214-219
Hymenostomatida, Tetrahymenina
includes: Entodiscus (type genus); Cryptochilum; Biggaria; Madsenia; Ptyssostoma; Andreula; Symbionecta gen. n.; Pectenita gen. n.; Gullmarella

Entodiscus Madsen, 1931 (type genus)
Iankovskii, A. V., 1973, Parazitologiia, Leningrad, v. 7 (3), 214-219
Hymenostomatida, Tetrahymenina, Entodiscidae fam. n.

Entopolypoides species, illus.
Wolf, R. E.; et al., 1978, Ann. Int. Med., v. 88 (6), 769-773
Entopolypoides sp. intraerythrocytic parasitosis in 2 humans with hepatic dysfunction, description of serum factors inhibiting phytohemagglutinin response by normal lymphocytes suggests that hepatic dysfunction induces serum factors that diminish cellular immunity and allow patent parasitemia to develop: United States (one patient was born in Turkey; the other was a merchant seaman who traveled extensively)

Entopolypoides macaci
Kaschula, V. R.; Van Dellen, A. F.; and de Vos, V., 1978, J. South African Vet. Ass., v. 49 (3), 223-227
Cercopithecus aethiops pygerythrus: South Africa

Eperythrozoon sp.
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Sorex araneus (Lunge)
S. minutus (Blut, Lunge, Milz)
Rattus norvegicus (Blut, Herz, Lunge, Milz, Placenta)
R. rattus (Herz, Lunge, Leber, Milz)
Mus musculus (Blut, Herz, Lunge, Leber, Milz)
Cricetus cricetus (Leber, Milz)
all from Neusiedlerseegebiet, nordlichen Burgenland

Eperythrozoon ovis
Ilemobade, A. A.; and Blotkamp, C., 1978, Tropenmed. u. Parasitol., v. 29 (3), 307-310
Eperythrozoon ovis, sheep, indirect immunofluorescent antibody test, evaluation for diagnostic purposes, recommended for use in epidemiological studies

Eperythrozoon ovis
Ilemobade, A. A.; and Blotkamp, C., 1978, Tropenmed. u. Parasitol., v. 29 (3), 311-314
Eperythrozoon ovis, sheep, incidence survey using the indirect immunofluorescent antibody test, 36% were sero-positive but only 12 had positive smears for parasites indicating endemic stability of area: Nigeria

Eperythrozoon ovis
Ilemobade, A. A.; and Blotkamp, C., 1978, Tropenmed. u. Parasitol., v. 29 (4), 443-450
Eperythrozoon ovis, sheep (exper.), pathologic changes in blood picture, concurrent or superimposed infection with Trypanosoma vivax did not affect growth of either parasite

Eperythrozoon ovis
Sonoda, M.; et al., 1977, Nippon Zyuisi-Kai Zassi (J. Japan Vet. Med. Ass.), v. 30 (7), 374-379
Eperythrozoon ovis, sheep (nat. and exper.), clinical and hematological observations: Japan

Eperythrozoon ovis
Sutton, R. H., 1978, N. Zealand Vet. J., v. 26 (9), 224, 229-230
Eperythrozoon ovis, sheep (exper.), body and organ weight measurements, pathology

Eperythrozoon ovis
Sutton, R. H., 1979, Vet. Parasitol., v. 5 (1
11-15
Eperythrozoon ovis, effect of infection on reductive potential of sheep erythrocytes

Eperythrozoon suis
Henry, S. C., 1979, J. Am. Vet. Med. Ass., v. 174 (6), 601-603
Eperythrozoon suis, swine, clinical signs of infection confirmed by indirect hemagglutination and measuring packed cell volume; oxytetracycline and arsanilic acid combined with lice control, arsenic toxicosis

Eperythrozoon suis, illus.
Hoffmann, R.; and Saalfeld, K., 1977, Deutsche Tierarztl. Wchnschr., v. 84 (1), 7-9
Eperythrozoon suis, outbreak in a fattening pig farm, pathogenesis, therapeutic possibilities discussed: Germany

Eperythrozoon wenyonii
Minami, T., 1977, Japan Agric. Research Quart. v. 11 (4), 234-238
Japanese Babesia sp., cattle (exper.), capil lary-tube agglutination test, discrimination from B. bigemina and B. argentina was almost possible, no cross reactions with other protozoa

Eperythrozoon wenyoni, illus.
Sutton, R. H.; Charleston, W. A. G.; and Collins, G. H., 1977, N. Zealand Vet. J., v. 2 (1-2), 8-9
Jersey cow (blood) (nat. and exper.): Manawatu, New Zealand

Eperythrozoonosis
Leman, A. D.; Cropper, M.; and Rodeffer, H. E., 1974, Theriogenology, v. 2 (6), 149-160
infectious swine reproductive diseases, includes brief discussions on eperythrozoonosis and toxoplasmosis

Epistylis sp.
Arthur, J. R.; Margolis, L.; and Arai, H. P., 1976, J. Fish. Research Bd. Canada, v. 33 (11), 2489-2499
Cottus cognatus (fins): Aishihik Lake, Yukon Territory

Epistylis sp.
Esch, G.; et al., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 687-693
Epistylis-Aeromonas complex, centrarchid fish, incidence, spatial distribution of lesions, host size class (age), body condition, seasonal periodicity, influence of thermal effluent on disease
Micropterus salmoides
Lepomis gulosus
L. auritus
L. macrochirus
all from Par Pond reservoir, Savannah River Plant near Aiken, South Carolina

Epistylis sp., illus.
Hazen, T. C.; et al., 1978, J. Protozool., v. 25 (3, pt. 2), 351-355
". . . it is concluded that Epistylis sp. is a benign ectocommensal and that A[eromonas] hydrophila is the primary etiologic agent of red-sore disease [of Micropterus salmoides]."

pistylis [sp.]
Lewis, R. E.; Siler, J. R.; and Olmsted, L. L., 1977, Proc. 31. Ann. Conf. Southeast. Ass. Fish and Wildlife Agencies, 372-379
Epistylis [sp.], fishes, host specificity, intensity of infestation, attachment site, factors affecting prevalence (host length, water quality, season)
Dorosoma cepedianum
Notemigonus crysoleucas
Notropis niveus
Moxostoma macrolepidotum
Ictalurus catus
I. platycephalus
Morone chrysops
M. saxatilis
Lepomis auritus
L. gulosus
L. macrochirus
L. microlophus
Micropterus salmoides
Pomoxis nigromaculatus
Etheostoma fusiforme
Perca flavescens
all from North Carolina

pistylis sp.
Rogers, W. A.; and Hawke, J. P., 1978, Tr. Am. Micr. Soc., v. 97 (2), 244
Dorosoma cepedianum
Pomoxis nigromaculatus
Micropterus salmoides
(skin of all): all from Alabama

istylis sp., illus.
Turner, J. T.; Postek, M. T.; and Collard, S. B., 1979, Tr. Am. Micr. Soc., v. 98 (1), 136-138
Epistylis sp. on Acartia tonsa, bacterial colonization near ciliate-produced lesions in exoskeleton suggests that bacteria may utilize dissolved copepod body contents: upper Escambia Bay, Florida

Epistylis lwoffi
Iziumova, N. A.; Mashtakov, A. V.; and Kashkovskii, V. V., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36) 64-68
[Lucioperca lucioperca] (gills): Kamsk reservoir

Epistylis lwoffi Faure-Fremiet
Lester, R. J. G., 1974, Syesis, v. 7, 195-200
Gasterosteus aculeatus (gills, body surface): near Vancouver, British Columbia

Euglenoidina
Michajlow, W. K., 1972, Parazitologiia, Leningrad, v. 6 (1), 3-7
Euglenoidina of Copepoda, stages of adaptation to parasitism

Eugregarines, acephaline, illus.
Hembree, S. C., 1979, Mosquito News, v. 39 (3), 575-582
Aedes aegypti
A. albopictus
A. chrysolineatus
Armigeres subalbatus
all from Thailand

Eutreptia parasitica sp. n., illus.
Michajlow, W., 1978, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 26 (4), 269-272
Eutreptia parasitica sp. n., development cycle
Microcyclops varicans (eyes, limbs and body cavity of cephalothorax region): water body of Botanical Garden, Brisbane, Australia

Fabespora vermicola, illus.
Weidner, E.; and Overstreet, R. M., 1979, Cell
and Tissue Research, v. 201 (3), 331-342
Fabespora vermicola, sporogenesis

Facieplatycauda gen. n.
Wyatt, E. J., 1979, J. Protozool., v. 26 (1),
47-51
Myxobolidae
tod: F. pratti sp. n.

Facieplatycauda pratti sp. n. (tod), illus.
Wyatt, E. J., 1979, J. Protozool., v. 26 (1),
47-51
Catostomus luxatus (kidneys): Klamath
County, Oregon

Fallisia simplex
Telford, S. R., jr., 1979, Ann. Parasitol.,
v. 54 (2), 129-144
reported earlier as Plasmodium tropiduri
Plica umbra: Guyana

Farinocystis tenebrioides n. sp., illus.
Purrini, K.; and Ormieres, R., 1979, Zool.
Anz., Jena, v. 202 (5-6), 437-443
Farinocystis tenebrioides n. sp., life cycle,
pathogenicity
Tenebrioides mauretanicus (Fettkorper):
Kosova-Gebietes, Jugoslawien

Farinocystis tribolii
Purrini, K.; and Ormieres, R., 1979, Zool.
Anz., Jena, v. 202 (5-6), 437-443
Tenebrioides mauretanicus: Kosova-Gebietes,
Jugoslawien

Farinocystis tribolii Weiser, 1953, illus.
Zizka, Z., 1977, Ztschr. Parasitenk., v. 54
(3), 217-228
Farinocystis tribolii in Tribolium castaneum,
fine structure, developmental stages in
sporogony, parasite-host relations (mitochondria
of host concentrated around schizonts,
consumption of host fat body by
parasites, host development stopped)

Farinocystis tribolii Weiser, 1953, illus.
Zizka, Z., 1978, J. Protozool., v. 25 (1),
50-56
Farinocystis tribolii, fine structure of
schizonts and free merozoites and their
development in fat body of larval Tribolium
castaneum

Filiformis subgen. n.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro,
v. 10 (2), 23-24
subgen. of Protoopalina
type sp.: Protoopalina (Filiformis) filiformis
Metcalf, 1923

Filiformis
Amaro, A., 1966, Atas Soc. Biol. Rio de
Janeiro, v. 10 (3), 49-52
subgen. of Protoopalina, key

Filiformis Amaro, 1966, preoccupied by Filiformis
Hirshfield, Isquith & Bhandary, 1965
Amaro, A., 1969, Atas Soc. Biol. Rio de
Janeiro, v. 12 (5-6), 237-243
subgen. of Protoopalina
renamed: Longicorpus subgen. n.

Flagellates
Arias, J. R.; and de Freitas, R. A., 1978,
Acta Amazonica, v. 8 (3), 387-396
Lutzomyia maripaensis
L. spinosa
L. (longipalpis) sp.
L. (shannoni) sp.
all from regiao de Manaus, Amazonas, Brasi

Flagellate
Boghen, A. D., 1978, Canad. J. Zool., v. 56
(11), 2460-2462
prevalence in different host size classes
Homarus americanus (gill filaments): Nort
umberland strait, southern Gulf of St.
Lawrence

Flagellates
Holt, P. E.; Brown, A.; and Brown, B., 1978,
Vet. Rec., v. 102 (18), 404-405
Strongyloides [sp.] in Lampropeltis getulus
holbrooki, clinical symptoms, unsuccessful
treatment with thiabendazole, mixed infection
with flagellates and Ochetosomatidae
sp., treatment for flagellates with metron
dazole also unsuccessful, case report
Lampropeltis getulus holbrooki (feces)

Fr[ancaiella] colchica
Kolev, V.; et al., 1977, Vet. Med. Nauki, v.
(3), 54-60
Protozoa as cause of abortions in cows: dis
trict of Rousse

Francaiellosis
Rakhimov, T. Kh.; et al., 1977, Veterinariia,
Moskva (10), 75-77
piroplasmosis, francaiellosis, cattle,
dimidine and imidocarb tested in various
doses, recommended for control

Frenkelia
Beier, T. V.; Shibalova, T. A.; and Kostenko,
L. A., 1978, [Cytology of coccidia], 185 pp.,
illus.

Frenkelia, illus.
Goebel, E.; Rommel, M.; and Krampitz, H. E.,
1978, Ztschr. Parasitenk., v. 55 (1), 29-42
Frenkelia, schizonts and merozoites, electron
microscopy

Frenkelia
Rommel, M., 1978, Ztschr. Parasitenk., v. 57
(3), 269-283
cyst-forming coccidia, life cycle, taxonomy,
comparative review

Frenkelia sp.
Frank, C., 1978, Ang. Parasitol., v. 19 (3),
137-154
Clethrionomys glareolus (Gehirn): Neusiedlerseegebiet,
nordlichen Burgenland

Frenkelia [sp.], illus.
Tadros, W.; and Laarman, J. J., 1978, Ztschr.
Parasitenk., v. 58 (1), 41-46
Frenkelia [sp.] in Clethrionomys glareolus
(brain, spinal chord), apparent congenital
transmission, epizootiological significance

Frenkelia buteonis
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk.,
v. 58 (2), 115-139
nomen dubium

Frenkelia clethrionomyobuteonis, illus.
  Geisel, O.; et al., 1978, Vet. Path., v. 15 (5), 621-630
    Frenkelia clethrionomyobuteonis in Clethrionomys glareolus (exper.), pathomorphology

Frenkelia glareoli (Erhardova, 1955) Biocca, 1968
  Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
    definition, intermediate and final hosts, synonymy

Frenkelia microti (Findlay and Middleton, 1934) Biocca, 1968
  Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
    definition, intermediate and final hosts, synonymy

Frenkelia microti, illus.
  Rommel, M.; and Krampitz, H. E., 1978, Zentralbl. Vet.-Med., Reihe B, v. 25 (4), 273-281
    Frenkelia microti, life cycle
    Microtus agrestis (exper.)
    M. arvalis (nat. and exper.): Grafrath, westlich Munchens
    Apodemus sylvaticus (exper.)
    A. flavicollis (exper.)
    A. agrarius (exper.)
    Mus musculus (exper.)
    Mastomys natalensis (exper.)
    Rattus norvegicus (exper.)
    Mesocricetus auratus (exper.)
    Cricetus cricetus (exper.)
    Chinchilla laniger (exper.)
    Buteo buteo (exper.)
    Oryctolagus cuniculus (exper.)

Ganymedes apsteini n. sp. [nom. nud.]
Theodorides, J.; and Desportes, I., 1972, Compt. Rend. Acad. Sc., Paris, v. 274, s. D, Sc. Nat. (24), 3251-3253 [For complete author reference see Supplement 20, Part 1]

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Desportes, I.; Vivares, C. P.; and Theodorides, J., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (3), 261-277
Ganymedes, Porospora, Thiriotia, epicytic ultrastructure, taxonomic significance
Eucopia hanseni: Villefranche-sur-Mer (Alpes-Maritimes)

Ganymedes haeckeli (Mingazzini) [n. comb.?]
Theodorides, J.; and Desportes, I., 1972, Compt. Rend. Acad. Sc., Paris, v. 274, s. D, Sc. Nat. (24), 3251-3253 [For complete author reference see Supplement 20, Part 1]

Ganymedes vibiliae Theod. et Desp. (1972), illus.
Desportes, I.; Vivares, C. P.; and Theodorides, J., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (3), 261-277
Ganymedes, Porospora, Thiriotia, epicytic ultrastructure, taxonomic significance
Vibilia armata
V. propinqua
all from Villefranche-sur-Mer (Alpes-Maritimes)

Giardia
Bingham, A. K.; and Meyer, E. A., 1979, Nature, London (5694), v. 277, 301-302
Giardia, excystation in acidic solutions, pattern of trophozoite emergence during excystation, method for routine in vitro induction of excystation, establishment of in vitro axenic cultures from excysted trophozoites

Giardia
Feldman, R., 1977, Acta Bioquim. Clin. Latinoam., v. 11 (2), 179-180
protozoa, techniques for microscopical diagnosis

Giardia
Fernandes, J. L.; and Garcia, F., 1971, Rev. Soc. Brasil. Med. Trop., v. 5 (3), 155-158
human intestinal parasites, clinical trials with combined piperazine and thiabendazole: Sao Paulo

Giardia
Kavousi, S., 1979, Am. J. Trop. Med. and Hyg., v. 28 (1), 19-23
giardiasis, infants and children, short and long-term followup after treatment with quinacrine vs. metronidazole

Giardia
Lakhonina, G., 1978, Eesti NSV Teaduste Akad. Toimetised, v. 27, Biol. ser. (1), 47-54
Giardia, culture in solid medium; saccharolytic activity

Giardia, illus.
Meyer, E. A., 1979, Environment. Protect. Technol. Ser. (EPA-600/2-79-063), 28 pp.
Giardia, concentration and purification of cysts from feces, induction of and determination of factors involved in excystation, effect of various storage temperatures on survival as determined by cultural excystation method

Giardia, illus.
Moffatt, R. E.; and Olfert, E., 1976, Proc. 19. Ann. Meet., Am. Ass. Vet. Lab. Diagn., 171-178
Giardia, Trichomonas, Wistar rats (intestine), diarrhea, histopathology

Giardia
Nesvadba, J., 1979, Kleintier-Praxis, v. 24 (4), 177-179
Giardia, cat, ornidazole, metronidazole, case report

Giardia
Prakash, P.; and Saxena, S., 1976, Rajasthan Med. J., v. 15 (4), 240-244
giardiasis in children, evaluation of metronidazole using varying dosage schedules, recommendations for therapy

Giardia
Sebesteny, A., 1979, Lab. Animals, v. 13 (3), 189-191
intestinal protozoa, successful transmissio to and establishment in specific-pathogen-free mice exposed to intestinal contents of infected hamsters

Giardia, illus.
Sugano, H.; and Ando, K., 1978, Nippon Zyuisi-Kai Zassi (J. Japan Vet. Med. Ass.), v. 31 (11), 635-638
Giardia, dogs (nat. and exper.), diarrhea, metronidazole

Giardia
Zierdt, W. S., 1978, Am. J. Clin. Path., v. 70 (1), 89-93
human intestinal parasites, description of simple method for recovery of eggs, larva an cysts from feces by concentration device, comparison with other methods of parasite re covery

Giardia sp.
Bingham, A. K.; et al., 1979, Exper. Parasitol v. 47 (2), 284-291
Giardia sp., excystation in vitro, effects o temperature, pH, time, and incubation medium eosin exclusion and excystation compared as methods of determining cyst viability, effec of temperature on cyst viability

Giardia sp.
Marzochi, M. C. de A., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (4), 249-256
human intestinal parasites, eggs and cysts from water used to irrigate vegetable gardens, increased risk of crop contamination ir dry season: Ribeirao Preto, Sao Paulo, Brazil

Giardia sp.
Marzochi, M. C. de A., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (3), 148-155
enteroparasitic cysts and eggs, contamination of green vegetables and kitchen garden soils, epidemiological survey, most commonly found during dry-season when fecal polluted brooks were used for irrigation: Ribeirao Preto, Sao Paulo, Brasil

Giardia sp.
Mirza, M. Y.; and al-Rawas, A. Y., 1978, J. Protozool., v. 25 (3, pt. 2), 285-286
Nesokia indica: Baghdad district, Iraq

Giardia sp., illus.
Panigrahy, B.; et al., 1978, Avian Dis., v. 22 (4), 815-818
Giardia sp., parakeets (intestine), case reports, successful treatment with dimetridazole: commercial aviaries in Texas

Giardia sp.
Rajasekaran, P., 1978, Madras Agric. J., v. 65 (9), 613-616
intestinal parasites, children, infection rates in relation to sources of drinking water (open dug wells, water taps in street, water tap within household): Athur block, Madurai district, Tamil Nadu

Giardia agilis Kunstler, 1882, illus.
Machado, D. A. (filho), 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (4), 91-94
description
Elosiidae
Bufonidae
(intestino delgado of all): all from Brasil

Giardia agilis Kuenstler, 1882, illus.
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
description
Rana temporaria (strevo (az zaludek), kloaka): CSSR

Giardia canis
Barlough, J. E., 1979, J. Small Animal Practice, v. 20 (10), 613-623
Giardia canis, morphology and diagnosis, transmission, prevalence, clinical signs, pathogenesis, treatment, review

iardia caprae
Padmavathi, P.; et al., 1978, Indian Vet. J., v. 55 (11), 917
Giardia caprae, lambs (feces), possible cause of diarrhea

iardia duodenalis race peromysci Filice, 1952
Grant, D. R.; and Woo, P. T. K., 1978, Canad. J. Zool., v. 56 (6), 1348-1359
as syn. of Giardia peromysci Filice, 1952 emend.

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Cabrera M., G.; et al., 1977, Rev. Med. Chile, v. 105 (5), 297-303
Entamoeba histolytica, Giardia intestinalis, Ascaris lumbricoides, Hymenolepis nana, listed as etiologic agents of human diarrheal syndrome

iardia intestinalis
Danö, P.; Olsen, J.; and Vejlsgaard, R., 1973, Scand. J. Gastroenterol., v. 8 (2), 151-154
Giardia intestinalis, humans, prevalence survey of patients with gastrointestinal symptoms, 1 positive case out of 190 surveyed: Denmark

iardia intestinalis, illus.
Harvey, J., 1973, Saguenay Med., v. 20 (3), 146-156
parasitic flagellates, life cycle, infection in man, clinical signs, therapy, brief review

Giardia intestinalis
Merdivenci, A.; Baydemir, M.; and Sengul, M., 1976, Turk Hijiy. ve Deneysel Biyol. Dergisi, v. 36 (2), 238-247
Giardia intestinalis, children, tinidazole and nitrimidazine compared, both successful

Giardia intestinalis
Warhurst, D. C.; and Wright, S. G., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (5), 601 [Letter]
Giardia intestinalis, cryopreservation

Giardia lamblia
Akhtaruzzaman, K. M.; et al., 1978, Tropenmed. u. Parasitol., v. 29 (4), 427-431
comparison of different methods for detection of intestinal protozoa and helminths in human stool

G[iardia] lamblia
Alonso, R. E., 1976, Semana Med. (4956), an. 83, v. 149 (7), 234-237
intestinal parasites, school children, clinical studies with tinidazole and mebendazole: Institute Aguirre 'Patronato de la Infancia'

Giardia lamblia
Alvarez Gomez, L. de las N., 1977, Rev. Cubana Med. Trop., v. 29 (2), 61-65
Giardia lamblia, technique for the preparation of antigen extract from trophozoites and cysts, study of immunoallergic characteristics in experimental animals and normal human controls

Giardia lamblia
Alvarez Gomez, L. de las N.; and Sordo Gonzalez, M., 1977, Rev. Cubana Med. Trop., v. 29 (3), 129-133
Giardia lamblia, method for obtaining specific antiserum in rabbits

Giardia lamblia
Amato Neto, V.; et al., 1972, Rev. Soc. Brasil. Med. Trop., v. 6 (3), 147-149
Giardia lamblia, humans, therapeutic trials showed tinidazole to be useful antiparasitic treatment: Brazil

Giardia lamblia, illus.
Amin, N., 1979, Postgrad. Med., v. 66 (5), 151-156, 158
Giardia lamblia, human, incidence and epidemiology, pathophysiology, pathogenesis, clinical manifestations, diagnosis, treatment, prevention, review

Giardia lamblia
Ansdell, V. E.; and Common, J. D. A., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 206-207
Giardia lamblia, 21-year-old Kenyan Asian, corneal damage after therapy with mepacrine: London

G[iardia] lamblia
Apt, W.; et al., 1977, Rev. Med. Chile, v. 105 (6), 388-389
human intestinal protozoa, modified Teleman and PAFS methods compared for diagnostic purposes

Giardia lamblia
Aust-Kettis, A.; and Magnius, L., 1973, Scand. J. Infect. Dis., v. 5 (4), 289-292
Giardia lamblia, infection in group of students after a visit to Leningrad: Stockholm, Sweden

Giardia lamblia
Aust-Kettis, A.; and Thoren, G., 1974, Scand. J. Infect. Dis., v. 6 (4), 349-353
Giardia lamblia, inter- and intrafamilial infection involving 5 families, case reports, epidemiology, index case thought to be 3-year-old child from Israel that had recently been adopted by family members: Lindesberg, Sweden

Giardia lamblia
Awadalla, M. A.; Hussein, L. A.; and Salem, M., 1979, Nutrition Rep. Internat., v. 19 (5), 627-633
parasitic infestations in children, effect on intestinal absorption as determined by assay of fasting serum carotene and vitamin A levels and by vitamin A tolerance tests: Orphanage Institute of Giza and El-Zeitoun, Egypt

Giardia lamblia
Baranski, M. C.; et al., 1975, Rev. Soc. Brasil. Méd. Trop., v. 9 (1), 15-18
Giardia lamblia, trials testing the therapeutic efficacy of tinidazole, adults and children both achieved cure rate of over 96%: Brasil

Giardia lamblia
Barkin, R. M.; et al., 1975, Rocky Mountain Med. J., v. 72 (8), 340-344
medical screening of recently adopted Vietnamese orphans, scabies, Entamoeba histolytica, Giardia lamblia, and Pneumocystis carinii among medical problems encountered: Denver area, Colorado

Giardia lamblia
Bartlett, M. S.; et al., 1978, J. Clin. Microbiol., v. 7 (6), 524-528
modified zinc sulfate flotation technique evaluated in comparison with formalin-ether concentration method for recovery of protozoan cysts and helminth eggs and larvae from feces preserved in formalin less than and longer than 1 month, results suggest that (except for schistosomes) F-$ZnSO_4$ compares favorably to FE method for detecting infections of clinical significance

Giardia lamblia
Bassett, M. L.; Danta, G.; and Cook, T. A., 1978, Brit. Med. J. (6129), v. 2, 19
Giardia lamblia, humans with associated peripheral neuropathy of unknown etiology, 2 case reports, resolution of giardiasis and neurological symptoms after metronidazole therapy

Giardia lamblia
Blanco Rabassa, E.; et al., 1975, Rev. Cubana Pediat., v. 47 (2), 247-263
Giardia lamblia resulting in malabsorption syndrome in children, clinical features of syndrome and pathologic changes in intestine

Giardia lamblia
Blas Bava, A.; and Nino, R. F., 1974, Semana Med. (4876), an. 81, v. 145 (54), 2534-2539
Giardia lamblia, humans, clinical trials testing 2 dosage schedules of tinidazole, clinically and parasitologically effective with only mild side effects

Giardia lamblia
Blecka, L. J., 1978, J. Parasitol., v. 64 (2), 362-363
patients infected with intestinal parasites, attempt to quantitate immunoglobulin levels fecal extracts with radial immunodiffusion, mean IgA levels higher than in controls, other immunoglobulin classes rarely detectab

Giardia lamblia
Blenkinsopp, W. K.; Gibson, J. A.; and Haffenden, G. P., 1978, Lancet, London (8071), v. 1, 994 [Letter]
Giardia lamblia, case report of infection in elderly woman resulting in severe atrophy of jejunal mucosa with dense plasma-cell infiltrate, complete recovery after therapy with metronidazole: England (had traveled to Far East)

Giardia lamblia
Blom, M.; Prag, J. B.; and Nørredam, K., 1979, Am. J. Trop. Med. and Hyg., v. 28 (1), 76-83
intestinal parasites, children from Vietnam-Cambodia at and after arrival in Denmark, α_1-acid glycoprotein, α_1-antitrypsin, and ceruloplasmin concentrations, eosinophilia, sedimentation rates, effect of T.A.B.-choler vaccination

Giardia lamblia
Botero R., D., 1978, Ann. Rev. Pharmacol. and Toxicol., v. 18, 1-15
chemotherapy of common intestinal protozoan and helminth infections in humans, review of antiparasitic drugs in current use

Giardia lamblia, illus.
Brandborg, L. L., 1979, Am. J. Med., v. 67 (6), 999-1006
diseases of malabsorption, histologic diagnosis

Giardia lamblia, illus.
Brasitus, T. A., 1979, Am. J. Med., v. 67 (6), 1058-1065
parasitic infections, association with malabsorption in man

Giardia lamblia
Buck, A. A.; Anderson, R. I.; and MacRae, A. A., 1978, Tropenmed. u. Parasitol., v. 29 (3), 253-268
polyparasitism, humans, epidemiology, assessment of combined effects of multiple infections on an individual's state of health, comparative analysis of field data obtained from several tropical villages

Giardia lamblia
Canzonieri, C. J.; et al., 1975, Rev. Brasil. Clin. e Terap., v. 4 (12), 479-482
Trichomonas vaginalis, Giardia lamblia, Entamoeba histolytica, humans, clinical trials testing efficacy of tinidazole therapy

Giardia lamblia
Chavalittamrong, B.; et al., 1978, Southeast Asian J. Trop. Med. and Pub. Health, v. 9 (1), 51-54
Giardia lamblia, children, prevalence, associated symptoms: Out-patient Department, Siriraj Hospital, Thailand

iardia lamblia
Chu, J. K., 1972, Taehan Uihak Hyophoe Chi (J. Korean Med. Ass.), v. 15 (8), 685-690
human parasites, differential diagnosis

iardia lamblia, illus.
Collins, J. P.; Keller, K. F.; and Brown, L., 1978, Am. J. Trop. Med. and Hyg., v. 27 (4), 835-836
Giardia lamblia, 'ghost' forms of cysts in human fecal sample initially misdiagnosed as Isospora oocysts

iardia lamblia
Corral Carranceja, I.; et al., 1975, Rev. San. e Hig. Pub., v. 49 (2), 159-178
intestinal parasites, epidemiologic survey of children on admission to Sanatorio Marino de Gorliz

ardia lamblia
Craun, G. F., 1979, Am. J. Pub. Health, v. 69 (8), 817-819
Giardia lamblia, humans, waterborne outbreaks occurring from contaminated drinking water, seasonal distribution, data indicate that disinfection as only treatment for surface water sources is ineffective, review: United States

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Ondatra zibethica (Duodenum, Jejunum, Rectum)
Microtus agrestis (Jejunum)
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allergic form of myocarditis, 26-year-old
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differential diagnosis of chronic malabsorption and maldigestion syndromes in children
mentions lambliasis and ascaridiasis

Giardiasis
Nitzulescu, V.; Gherman, I.; and Popescu, A.,
1977, Rev. Pediat. (Pediat.), Bucuresti, v. 2
(2), 189-192
giardiasis, human, therapy with tinidazole,
good results

Giardiasis
Novis, B. H.; Bank, S.; and Marks, I. N., 197
South African Med. J., v. 46 (16), 455-462
small intestinal malabsorption, humans, giardiasis among causes: South-western Cap

Giardiasis
Novis, B. H.; Bank, S.; and Marks, I. N., 1973, South African Med. J., v. 47 (26), 1139-1142
gastric acid secretions in intestinal malabsorption conditions including giardiasis, humans

Giardiasis
Okelo, G. B. A.; Mukiibi, J. M.; and Kyobe, J.
1979, East African Med. J., v. 56 (6), 266-269
intestinal parasites, humans, mean eosinophil count higher in infected patients vs. controls, no correlation between eosinophil levels and different intestinal helminths: Nairobi, Kenya

Giardiasis
Panigrahy, B.; et al., 1979, J. Am. Vet. Med. Ass., v. 175 (4), 359-361
parakeets from 6 aviaries

rdiasis
ichle, R.; et al., 1978, Arzneimittel-Forsch., . 28 (4), 612-625
trichomoniasis, amoebiasis, lambliasis, extensive in vitro and in vivo trials (humans, domestic animals, laboratory animals) with ornidazole to establish chemotherapeutic properties, efficacy slightly superior to metronidazole in comparative trials

rdiasis
aha, K.; et al., 1979, Indian J. Med. Research, . 70, 22-32
parasitic diseases, human, serum immunoglobulin and complement profile: India

rdiasis
pech, H. J., 1978, Deutsche Med. Wchnschr., . 103 (50), 2008-2012
lambliasis, epidemiology, pathogenesis, clinical aspects, diagnosis, treatment, review

rdiasis
ulaiman, A.; and Gips, C., 1974, Tijdschr. astro-enterol., v. 17 (6), 373-382
identification of gallbladder bile during duodenal intubation, includes information on diagnosis of human lambliasis

vannolaia
arnham, P. C. C., 1979, J. Trop. Med. and yg., v. 82 (9-10), 178-182
avian Plasmodium spp. of 4 subgenera, list of type material in Garnham Collection of Wellcome Museum of Medical Science; neotypes designated

bidian cyst-like bodies
ilali, M.; and Scholtyseck, E., 1979, Acta et. Scand., v. 20 (3), 321-328
globidian cyst-like bodies containing multinucleate schizonts or merozoites, sheep (abomasum), ultrastructure, merozoite formation, host cell confirmed as being alive and intact: Bonn

bidium [sp.], illus.
ineme, C. N.; and Njoku, C. O., 1978, Vet. ec., v. 103 (19), 418-420
Globidium [sp.], Yankasa sheep, clinical symptoms, gross and histopathological lesions, importance of differentiating from intestinal coccidiosis: Nigeria

oides subgen. n.
aro, A., 1966, Atas Soc. Biol. Rio de Jairo, v. 10 (2), 29-31
subgen. of Cepedea
type sp.: Cepedea (Globoides) globosa Metcalf, 1923

oides
naro, A., 1966, Atas Soc. Biol. Rio de aneiro, v. 10 (3), 49-52
subgen. of Cepedea, key

ulocephalus, nov. gen.
audoin, J., 1965, Arch. Zool. Exper. et Gen., . 105 (2), 245-258
Actinocephalidae
tod: G. hydropsyches, n. g., n. sp.

ulocephalus hydropsyches, n. g., n. sp.
d), illus.
udoin, J., 1965, Arch. Zool. Exper. et Gen., 105 (2), 245-258
Hydropsyche sp.: torrents d'Auvergne

Glossatella piscicola
Rydlo, M., 1977, Wien. Tierarztl. Monatschr., v. 64 (2), 58-60
Glossatella piscicola, possible pathogenicity discussed
Regenbogenforelle (Korperoberflache)

Glugea sp.
Chernyshenko, A. S., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 105-113
ichthyoparasite fauna, extensity and intensity of invasion, species composition
[Pisces] bychok-zelenchak
[Pleuronectes flesus]
all from 4 estuaries, Black Sea (northern coastal region)

Glugea anomala (Moniez, 1887), illus.
Dykova, I.; and Lom, J., 1978, J. Fish Dis., v. 1 (1), 83-90
Glugea anomala, Gasterosteus aculeatus (exper.) (stomach), xenoparasitic formations typical of intracellular development responsible for marked tissue reaction in host, histopathology

Glugea anomala (Moniez)
Lester, R. J. G., 1974, Syesis, v. 7, 195-200
Gasterosteus aculeatus (body cavity): near Vancouver, British Columbia

Glugea anomala (Moniez, 1887) Gurley, 1893
Lom, J.; and Laird, M., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 569-580
brief description
Gasterosteus aculeatus (branchial cavity, subcutaneous connective tissue): Bay of Fundy, New Brunswick

Glugea atherinae n. sp., illus.
Berrebi, P., 1979, Ztschr. Parasitenk., v. 60 (2), 105-122
Glugea atherinae n. sp., ultrastructure, life cycle
Atherina boyeri (tube digestif, cavite generale): Lagunes du Languedoc et de Provence

Glugea berglax n. sp., illus.
Lom, J.; and Laird, M., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 569-580
Macrourus berglax (muscular layer of intestine): northern edge of Grand Banks of Newfoundland

Glugea bracteata Strickland, 1913
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora bracteata (Strickland, 1913) comb. n.

Glugea branchiale (Nemeczek, 1911) emend., illus.
Lom, J.; and Laird, M., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 569-580
description
Melanogrammus aeglefinus (gill filaments): Passamaquoddy Bay at St. Andrews, Bay of Fundy, New Brunswick, Canada

Glugea habrodesmi Loubes, Maurand, Gasc & Bouix, illus.
Loubes, C., 1979, J. Protozool., v. 26 (2), 200-208
Microsporida, synaptonemal complexes demonstrated in 6 genera but not in Nosema, implications for life cycles
Habrodesmus falx
Oxydesmus granulosus
all from Republique Populaire de Benin (ex-Dahomey)

Glugea heraldi n. sp., illus.
Blasiola, G. C., jr., 1979, J. Fish Dis., v. 2 (6), 493-500
Glugea heraldi n. sp., histopathology
Hippocampus erectus (subcutaneous connective tissue): Florida

Glugea hertwigi, illus.
Scarborough, A.; and Weidner, E., 1979, Biol. Bull., v. 157 (2), 334-343
Glugea hertwigi in Osmerus mordax, prevalence in ovaries of spawning female hosts, transmission to young smelt by direct ingestion of spores or by ingestion of spore-carrying zooplankton, parasite development and xenoma growth
Osmerus mordax (intestine, ovaries) (nat. and exper.): Wheatley, Ontario, on Lake Erie; Jones River, near Plymouth, Cape Cod, Massachusetts
cladocerans (digestive tract) (exper.)
copepods (digestive tract) (exper.)

Glugea hertwigi, illus.
Sherburne, S. W.; and Bean, Laurie L., 1979, Fish. Bull., National Oceanic and Atmos. Admin., v. 77 (2), 503-509
Glugea hertwigi and piscine erythrocytic necrosis (PEN) in Osmerus mordax, incidence and geographic distribution, no apparent relation between G. hertwigi and PEN: North Atlantic coast from Massachusetts to the Canadian Maritimes; inland lakes of Maine

Glugea hertwigi Weissenberg
Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
parasites of fishes: China

Goussia variabilis Labbe, 1896
Davies, A. J., 1978, J. Protozool., v. 25 (1), 15-21
as syn. of Eimeria variabilis (Thelohan, 1893) Reichenow, 1921

Grahamella sp.
Abranches, P.; and Magalhaes, C., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (6), 361-365
Rhinolophus ferrumequinum: Olhos de Agua, near Santarem, Portugal

Grahamella [sp.]
Butabaeva, M., 1979, Uzbek. Biol. Zhurnal (2), 50-53
[Citellus fulvus]
[Allactaga severtzovi]
[Allactaga elater]
all from Kashkadar'insk oblast

Grahamella sp.
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Sorex araneus (Blut, Herz, Lunge)
S. minutus (Blut, Lunge, Milz)
Myotis myotis (Lunge)
Rattus norvegicus (Blut, Herz, Lunge, Leber, Milz)
R. rattus (Blut, Herz, Lunge, Leber)
Mus musculus (Blut, Herz, Lunge, Leber)
Apodemus flavicollis (Blut, Herz, Lunge, Milz)
A. sylvaticus (Blut, Herz, Lunge, Leber, Milz, Muskulatur)
Microtus oeconomus (Blut, Herz, Lunge)
M. arvalis (Herz, Lunge)
M. agrestis (Blut, Herz, Lunge)
all from Neusiedlerseegebiet, nordlichen Burgenland

Grahamella [sp.], illus.
Lage, H. A.; and Lage, S. A., 1971, Atas Soc. Biol. Rio de Janeiro, v. 14 (5-6), 125-127
Rattus norvegicus: Rio de Janeiro, Guanabar

Grahamella sp.
Leong, T. S.; et al., 1979, Southeast Asian J. Trop. Med. and Pub. Health, v. 10 (1), 122-126
parasites of Rattus r. diardii, influence of human habitats on rat parasite fauna
Rattus rattus diardii (blood): Kuala Lumpur and nearby villages

Grahamella peromysci
Anderson, J. F.; Magnarelli, L. A.; and Kurz, J., 1979, J. Parasitol., v. 65 (4), 599-604
Peromyscus leucopus (blood): Connecticut

Grahamia [sp.]
Berdyev, A. S., 1977, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (1), 74-79
Hemiechinus hypomelas: southern Turkmenista

Grahamia alactagae Tartakowskyi, 1917
Berdyev, A. S., 1977, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (1), 74-79
Alactaga elater: southern Turkmenistan

Grahamia gerbilli Zasukhin, 1937
Berdyev, A. S., 1977, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (1), 74-79
Rhombomys opimus: southern Turkmenistan

Grahamia sylvaticus Parswanidse, 1925
Berdyev, A. S., 1977, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (1), 74-79
Mus musculus severtzovi: southern Turkmenistan

Grebnickiella gracilis (Grebnecki), illus.
Molon-Noblot, S., 1978, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 20 (2), 79-97
Grebnickiella gracilis, gametogenesis, electron microscopy

Grebnickiella gracilis (Grebnicki), illus.
Molon-Noblot, S.; and Desportes, I., 1977, Compt. Rend. Acad. Sc., Paris, v. 285, s. D, Sc. Nat. (3), 217-219
Grebnickiella gracilis, first description of synaptonemal complexes in meiotic nucleus of sporozoon

Grebneckiella pixellae Misra
Dass, C. M. S.; and Singotam, L., 1978, Electron Micr., v. 2, Biol., 446-447
Grebneckiella pixellae, ultrastructure of early developmental stages

regarina Dufour, 1828
Levine, N. D., 1979, J. Protozool., v. 26 (4),
532-536
Gregarinidae
diagnosis

regarina sp., illus.
Lage, H. A.; and Lage, S. A., 1971, Atas Soc.
Biol. Rio de Janeiro, v. 14 (5-6), 169-172
description
Cycloneda sanguinea: Manguinhos

egarina sp.
Massot, N.; and Ormieres, R., 1979, Ann. Parasitol.,
v. 54 (3), 267-275
Nebria lafresnayei

egarina sp.
Purrini, K.; Bukva, V.; and Baeumler, W.,
1979, Pedobiologia, v. 19 (5), 329-339
Eupelops subuliger
E. torulosus
Rhysotritia ardua
all from Nadelwaldbestanden in Suddeutschland

egarina sp.
Purrini, K.; and Ormieres, R., 1979, Zool.
Anz., Jena, v. 202 (5-6), 437-443
Tenebrioides mauretanicus: Kosova-Gebietes,
Jugoslawien

egarina ambigua sp. nov., illus.
Amoji, S. D.; and Rodgi, S. S., 1976, Riv.
Parassitol., Roma, v. 37 (1), 43-56
Forficula ambigua (mid-intestine): Karnatak
University Campus, Dharwar, India

garina blaberae, illus.
Philippe, M.; et al., 1979, Biol. Cell., v. 35
(2), 165-174
Gregarina blaberae, glycoconjugates of cell
surface studied by carbohydrate cytochemistry
and biochemistry on isolated 'ghost' fraction

garina blaberae Frenzel, illus.
zet, O.; Ormieres, R.; and Bergoin, M.,
965, Arch. Zool. Exper. et Gen., v. 105 (2),
99-303
Gregarina blaberae, life cycle, development
Blabera fusca (caecums intestinaux, tube
digestif anterieur): Amerique du Sud

garina brasiliensis Pinto, 1918
evine, N. D., 1979, J. Protozool., v. 26 (4),
32-536
as syn. of Chilogregarina brasiliensis (Pinto,
1918) comb. n.

arina coccinellae Lipa, 1967, illus.
pa, J. J.; Pruszynski, S.; and Bartkowski,
, 1975, Acta Parasitol. Polon., v. 23 (41-
1), 453-461
role of pathogens and parasites in survival
of Coccinellidae spp. during winter
Exochomus quadripustulatus
Harmonia quadripustulata
Myrrha octodecimguttata
all from Poland

garina cuneata
urrini, K.; and Ormieres, R., 1979, Zool.
nz., Jena, v. 202 (5-6), 437-443
Tenebrioides mauretanicus: Kosova-Gebietes,
Jugoslawien

Gregarina dichirotrichi n. sp., illus.
Massot, N.; and Ormieres, R., 1979, Ann. Parasitol.,
v. 54 (3), 267-275
Dichirotrichus pallidus: bords de l'Etang
de Thau (Sete)

Gregarina echiuri
Greeff, R., 1879, Nova Acta Acad. Nat. Curios.,
v. 41, pt. 2 (1), 1-172
as syn. of Conorhynchus gibbosus nov. gen. et
nov. spec.

Gregarina fuscozetis n. sp., illus.
Purrini, K.; Bukva, V.; and Baeumler, W.,
1979, Pedobiologia, v. 19 (5), 329-339
Fuscozetes setosus: Nadelwaldbestanden in
Suddeutschland

Gregarina garnhami, illus.
Mackenzie, C.; and Walker, M. H., 1979, Cell
and Tissue Research, v. 202 (1), 33-39
Gregarina garnhami, bacteria-like structures
in endoplasm, light and electron microscopy

Gregarina garnhami, illus.
Walker, M. H.; et al., 1979, J. Protozool., v.
26 (4), 566-574
Gregarina garnhami, structure and gliding
movement, light microscopy, scanning and
transmission electron microscopy

Gregarina hyalella Batten & DeGiusti, 1949
Levine, N. D., 1979, J. Protozool., v. 26 (4),
532-536
as syn. of Degiustia hyalella (Batten &
DeGiusti, 1949) comb. nov.

Gregarina kamenote Hoshide, 1951
Levine, N. D., 1979, J. Protozool., v. 26 (4),
532-536
as syn. of Cirrigregarina kamenote (Hoshide,
1951) comb. n.

Gregarina longa
Sherlock, P. L., 1979, Parasitology, v. 78 (2),
207-220
Tipula paludosa (intestine)

Gregarina macrotermitis n. sp., illus.
Kalavati, C.; and Narasimhamurti, C. C.,
1978, Proc. Indian Acad. Sc., Sect. B,
Animal Sc., v. 87 (12), 397-404
Macrotermes estherae (foregut, lumen of midgut,
hindgut): Andhra University at
Waltair

Gregarina megaspora sp. nov., illus.
Amoji, S. D.; and Rodgi, S. S., 1976, Riv.
Parassitol., Roma, v. 37 (1), 43-56
Forficula ambigua (mid-intestine): Karnatak
University Campus, Dharwar, India

Gregarina nitida Geus, 1969
Levine, N. D., 1979, J. Protozool., v. 26 (4),
532-536
as syn. of Arachnocystis nitida (Geus, 1969)
comb. n.

Gregarina ophoni Tuzet et Ormieres, 1956, illus.
Massot, N.; and Ormieres, R., 1979, Ann. Parasitol.,
v. 54 (3), 267-275
Ophonus similis: region de Sete

Gregarina oribatae Geus, 1969
Levine, N. D., 1979, J. Protozool., v. 26 (4),
532-536
as syn. of Arachnocystis oribatae (Geus,
1969) comb. n.

Gregarina peloris n. sp., illus.
  Massot, N.; and Ormieres, R., 1979, Ann. Parasitol., v. 54 (3), 267-275
    Pelor obesus: Lac d'Oncet (Hautes-Pyrenees)

Gregarina postneri n. sp., illus.
  Purrini, K.; Bukva, V.; and Baeumler, W., 1979, Pedobiologia, v. 19 (5), 329-339
    Hermannia gibba (Caeca, Magen): Nadelwaldbestanden in Suddeutschland

Gregarina pterotracheae (Stuart, 1971) Labbe (1899)
  Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
    as syn. of Molluskocystis pterotracheae (Stuart, 1871) comb. n.

Gregarina ruszkowski Lipa, 1967, illus.
  Lipa, J. J.; Pruszynski, S.; and Bartkowski, J., 1975, Acta Parasitol. Polon., v. 23 (41-51), 453-461
    role of pathogens and parasites in survival of Coccinellidae spp. during winter
    Adalia bipunctata
    Exochomus quadripustulatus
    (gut lumen of all): all from Poland

Gregarina scutovertexi Erhardova, 1955
  Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
    as syn. of Arachnocystis scutovertexi (Erhardova, 1955) comb. n.

Gregarina spissa Henry, 1938
  Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
    as syn. of Cirrigregarina spissa (Henry, 1938) comb. n.

Gregarina steini
  Purrini, K.; and Ormieres, R., 1979, Zool. Anz., Jena, v. 202 (5-6), 437-443
    Tenebrioides mauretanicus: Kosova-Gebietes, Jugoslawien

Gregarines, possibly Nematopsis-Porospora group, illus.
  Sawyer, T. K.; Newman, M. W.; and Otto, S. V., 1975, Proc. National Shellfish Ass., v. 65, 1974, 15-19
    gregarines, possibly Nematopsis-Porospora group in Crassostrea virginica, seasonal pathology suggests that parasites overwinter in hibernating oysters, undergo vegetative growth in the spring, and then perish or undergo further development in an unknown host
    Crassostrea virginica (digestive tract): New Haven Harbor, Connecticut; Chesapeake Bay tributaries, Maryland

Gregarines?
  Smit, F. G. A. M., 1978, J. Nat. Hist., v. 11 (6), 693-702
    Palaeopsylla t. tauberi (stomach): Dhaulagiri region of western Nepal

Gregarinidae Labbe, 1899
  Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
    diagnosis, includes: Gregarina; Degiustia gen. n.; Cirrigregarina gen. n.; Molluskocystis gen. n.

Gullmarella Fenchel, 1964
  Iankovskii, A. V., 1973, Parazitologiia, Leningrad, v. 7 (3), 214-219
    Hymenostomatida, Tetrahymenina, Entodiscidae fam. n.

Gurleya chironomi Loubes et Maurand, 1975, illus.
  Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135

Gurleya chironomi Loubes & Maurand, illus.
  Loubes, C., 1979, J. Protozool., v. 26 (2), 200-208
    Microsporida, synaptonemal complexes demonstrated in 6 genera but not in Nosema, implications for life cycles
    Orthocladius sp.: environs de Montpellier

Gurleya corynoneurae sp. n., illus.
  Voronin, V. N., 1979, Zool. Zhurnal, v. 58 (4), 592-594
    Corynoneura sp. (fat body): lake Vrevo, Leningrad oblast and lake Uchenoe, Pskovsk oblast

Haemamoeba
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
avian Plasmodium spp. of 4 subgenera, list of type material in Garnham Collection of Wellcome Museum of Medical Science; neotypes designated

Haematoxenus Uilenberg, 1964
van Vorstenbosch, C. J. A. H. V.; Uilenberg, G.; and van Dijk, J. E., 1978, Research Vet. Sc., v. 24 (2), 214-221
as syn. of Theileria

Haematoxenus sp., illus.
Young, A. S.; et al., 1978, Tropenmed. u. Parasitol., v. 29 (3), 281-288
theilerial parasites, incidence in Syncerus caffer, role as reservoir of cattle pathogenic theilerioses in East Africa

Haematoxenus [sp.]
Young, A. S.; Burridge, M. J.; and Payne, R. C., 1977, Trop. Animal Health and Prod., v. 9 (1), 37-34
Syncerus caffer (blood): Mara region, Kenya

Haematoxenus separatus Uilenberg and Andreasen, 1974
van Vorstenbosch, C. J. A. H. V.; Uilenberg, G.; and van Dijk, J. E., 1978, Research Vet. Sc., v. 24 (2), 214-221
as syn. of Theileria separata (Uilenberg and Andreasen 1974) [n. comb.]

Haematoxenus veliferus Uilenberg, 1964
van Vorstenbosch, C. J. A. H. V.; Uilenberg, G.; and van Dijk, J. E., 1978, Research Vet. Sc., v. 24 (2), 214-221
as syn. of Theileria velifera (Uilenberg 1964) [n. comb.]

Haemobartonella sp.
Gretillat, S., 1978, Bull. Acad. Vet. France, n.s., v. 51 (3), 351-358
Haemobartonella sp., horses, symptoms, diagnosis, pathology, treatment with chlorpromazine + berenil: Niger

aemobartonella sp.
North, D. C., 1978, J. Small Animal Practice, v. 19 (12), 769-773
Haemobartonella sp., non-splenectomized dog (blood), case report, haemolytic anaemia, post-mortem examination: Kent, Great Britain

aemobartonella bovis, illus.
Rodriguez, O. N.; et al., 1975, Folia Vet., v. 19 (1-2), 221-231
ganado vacuno (eritrocitos): Cuba

aemobartonella bovis
Rodriguez, O. N.; et al., 1975, Folia Vet., v. 19 (1-2), 233-242
isolation of bacteria resembling Rickettsia from Haemobartonella bovis cultures

aemobartonella canis, illus.
Bellamy, J. E. C.; MacWilliams, P. S.; and Searcy, G. P., 1978, J. Am. Vet. Med. Ass., v. 173 (4), 397-401
cold-agglutinin hemolytic anemia complicated by Haemobartonella canis, dog, case report

Haemobartonella canis, illus.
Sonoda, M.; et al., 1978, Japan. J. Vet. Sc., v. 40 (3), 335-341
Haemobartonella canis, presence in blood of intact and splenectomized dogs experimentally infected with Babesia gibsoni, scanning and transmission electron microscopy: Japan

Haemobartonella canis, illus.
West, H. J., 1979, J. Small Animal Practice, v. 20 (9), 543-549
Haemobartonella canis, dog, case report, oxytetracycline hydrochloride

Haemobartonella felis
Guelfi, J. F., 1978, Comp. Immunol., Microbiol. and Infect. Dis., v. 1 (1-2), 77-82
Haemobartonella felis in cats (nat. and exper.), value of examining blood smear for diagnosis and prognosis: region of Toulouse

Haemobartonella felis, illus.
Krieger-Huber, S., 1974, Tieraerztl. Prax., v. 2 (2), 221-224
Haemobartonella felis, infectious anemia in cats, clinical aspects, diagnosis, treatment

Haemobartonella felis, illus.
Lundborg, L. E.; Karlbom, I.; and Christensson, D., 1978, Svensk Vet.-Tidn., v. 30 (10), 417-419
Haemobartonella felis, infectious anemia, case report; review of diagnosis and treatment
katt: Sweden

Haemobartonella felis
Maede, Y., 1978, Japan. J. Vet. Sc., v. 40 (2), 141-146
Haemobartonella felis in splenectomized vs. non-splenectomized cats, relations between parasitemia and hematocrit value, erythrocyte osmotic fragility and direct Coombs tests, organ distribution of ^{51}Cr-labeled erythrocytes

Haemobartonella felis, illus.
Maede, Y.; and Murata, H., 1978, Japan. J. Vet. Sc., v. 40 (2), 203-205
Haemobartonella felis, detachment of organisms from parasitized erythrocytes by reticular cells in spleen of cat, ultrastructural observations

Haemobartonella felis, illus.
Manuel, M. F.; and Abalos, H. P., 1975, Philippine J. Vet. Med., v. 14 (1), 125-136
incidence survey
domestic cats: Philippines

Haemobartonella felis, illus.
Simpson, C. F.; Gaskin, J. M.; and Harvey, J. W., 1978, J. Parasitol., v. 64 (3), 504-511
Haemobartonella felis-parasitized cat erythrocytes, ultrastructural characteristics of damage caused by parasitism, presence of crystalloid inclusions, implications for pathogenesis of anemia associated with this infection

Haemobartonella felis, illus.
Tury, E.; Papp, L.; and Horvath, Z., 1977, Magy. Allat. Lapja, v. 99, v. 32 (4), 249-250
Haemobartonella felis, cats (nat. and exper.), severe anemia: Hungary

Haemobartonella felis
Watson, A. D. J.; Farrow, B. R. H.; and Hoskins, L. P., 1978, Austral. Vet. Practitioner, v. 8 (3), 129-132
Haemobartonella felis, cats (nat. and exper.), thiacetarsamide sodium therapy; prednisolone, tetracycline, and chloramphenicol given sequentially

Haemobartonella muris, illus.
Petrelli, F.; and Marsili, G., 1970, Boll. Soc. Ital. Biol. Sper., v. 46 (2), 71-72
Haemobartonella muris, splenectomized rats, course of infection not altered by administration of biotin

Haemobartonella muris
Schottelius, J., 1977, Tropenmed. u. Parasitol., v. 28 (4), 533-538
Trypanosoma cruzi, course of infection in non-splenectomized SPF rats with and without Haemobartonella muris infection

Haemobartonella muris
Sogandares-Bernal, F.; and Chandler, J., 1978, J. Parasitol., v. 64 (3), 547-548
Haemobartonella muris infections of laboratory mice as problem in routine passage of trypanosomes, method devised to remove most of Haemobartonella from donor blood sample to be transferred to recipient

Haemobartonella procyoni, illus.
Schaffer, G. D.; et al., 1978, J. Am. Vet. Med. Ass., v. 173 (9), 1148-1151
hematotropic parasites of Procyon lotor, carrier potential as related to translocation and release for hunting purposes, practice is considered biologically hazardous: southeastern United States
Procyon lotor: Hillsborough County, Florida

Haemobartonella ranarum Cunha & Muniz, 1926
da Costa, S. C. G.; et al., 1970, Atas Soc. Biol. Rio de Janeiro, v. 12, Suppl., 7-13
Leptodactylus ocellatus: Manguinhos, Guanabara

Haemobartonella ranarum, illus.
da Costa, S. C. G.; and Martinez B., N., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (5-6), 269-271
morphology, diagnostic differentiation
Leptodactylus ocellatus

Haemobartonellosis
Austerman, J. W., 1979, Vet. Med. and Small Animal Clin., v. 74 (7), 954
haemobartonellosis, dog, case report, tetracycline

Haemogregarina sp.
da Costa, S. C. G.; and Lins, D. C., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 39-40
sporozoans, staining for nucleic acids
Bothrops jararacusu

Haemogregarina sensu lato [sp.], illus.
da Costa, S. C. G.; Pereira, N. M.; and Gomes, D. C., 1971, Atas Soc. Biol. Rio de Janeiro, v. 14 (5-6), 113-115
Geochelone denticulata (sangue): Serra do Navio no Km 190 da Estrada de Ferro Amapa, Amapa

Haemogregarina sp.
Glassman, A. B.; Holbrook, T. W.; and Bennett, C. E., 1979, J. Parasitol., v. 65 (2), 323-324
Placobdella multilineata-infested Alligator mississippiensis (dorsal and lateral body surfaces, within oral cavity), increased eosinophilia; Haemogregarina sp. in alligators (blood) and in histologic sections of leeches probable vectorial relationship

Haemogregarina [sp.], illus.
Guerrero, S.; and Ayala, S. C., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (5), 283-288
Dendrobates trivittatus (sangre): selva amazonica, Huanuco, Peru

Haemogregarina sp., illus.
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102
Pholis gunnellus: Kent Island, New Brunswick
Spheroides maculatus: Woods Hole, Massachusetts
Liparis atlanticus: Kent Island, New Brunswick
(blood of all)

Haemogregarina [sp.], illus.
Paperna, I., 1979, Ann. Parasitol., v. 54 (4), 385-392
Sparus aurata (kidneys, liver and gut wall): cultured at Israel Oceanographic and Limnologic Research Mariculture Laboratory at Elat, gulf of Aqaba, Red Sea; collected in Bardawil lagoon on Mediterranean coast of Sinai
Siganus luridus (liver): northern gulf of Aqaba, Red Sea

Haemogregarina achiri Saunders (1955)
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102
". . . it is submitted that the name H. achiri may have to be discarded as a synonym of H. platessae."

Haemogregarina aeglefini Henry, 1913, illus.
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102
description
Syn.: H. urophysis Fantham et al.
Melanogrammus aeglefinus: St. Andrews, New Brunswick
Pollachius virens: St. Andrews, New Brunswick
Urophycis tenuis: St. Andrews, New Brunswick; Kittery, Me.
(blood of all)

Haemogregarina bigemina Laveran and Mesnil, 1901, illus.
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102
brief description
Centropristis striatus (blood): Woods Hole, Massachusetts

Haemogregarina bigemina
Ollenschlaeger, B., 1975, Tieraerztl. Prax., v. 3 (1), 99-107
blood and other parasites of commercial fish, pathology, transmission, therapy, clinical review

aemogregarina catostomi
Ollenschlaeger, B., 1975, Tieraerztl. Prax., v. 3 (1), 99-107
blood and other parasites of commercial fish, pathology, transmission, therapy, clinical review

aemogregarina cyprini, illus.
Kudriashova, Iu. V.; and Naumova, A. M., 1978, Veterinariia, Moskva (4), 76-78
Haemogregarina cyprini, carp, schizogonial stages in blood plasma and kidney smears, host blood values

aemogregarina cyprini
Ollenschlaeger, B., 1975, Tieraerztl. Prax., v. 3 (1), 99-107
blood and other parasites of commercial fish, pathology, transmission, therapy, clinical review

aemogregarina delagei Laveran & Mesnil, 1902
Becker, C. D.; and Overstreet, R. M., 1979, J. Fish Dis., v. 2 (6), 469-479
Raja eglanteria (blood): northern Gulf of Mexico

aemogregarina delagei
Daily, D. D., 1978, J. Parasitol., v. 64 (2), 361-362
Raja erinacea: Whiting Bay, Maine

aemogregarina delagei Laveran and Mesnil, 902, illus.
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102
description
Squalus acanthias
Raja erinacea
R. radiata
(blood of all): all from St. Andrews, New Brunswick

aemogregarina gadi pollachii Henry
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102
discarded as nomen nudum

aemogregarina irkulakpiki Laird, 1961
Arthur, J. R.; Margolis, L.; and Arai, H. P., 1976, J. Fish. Research Bd. Canada, v. 33 (11), 2489-2499
Prosopium cylindraceum (erythrocytes): Aishihik Lake, Yukon Territory
Coregonus clupeaformis (erythrocytes): Aishihik Lake and Stevens Lake, Yukon Territory

emogregarina leptodactyli Lesage, 1908
da Costa, S. C. G.; et al., 1970, Atas Soc. Biol. Rio de Janeiro, v. 12, Suppl., 7-13
Leptodactylus ocellatus: Manguinhos, Guanabara

emogregarina leptodactyli
da Costa, S. C. G.; and Lins, D. C., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 39-40
sporozoans, staining for nucleic acids

emogregarina leptodactyli Lesage, 1908, illus.
da Costa, S. C. G.; da Silva, A. M.; and Martinez Bernaola, N., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (1-2), 55-58
Haemogregarina leptodactyli, cytochemical studies
Leptodactylus ocellatus: Brazil

Haemogregarina mavori n. sp., illus.
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102
Macrozoarces americanus (blood): St. Andrews, New Brunswick

Haemogregarina mugili Carini, 1932
Becker, C. D.; and Overstreet, R. M., 1979, J. Fish Dis., v. 2 (6), 469-479
Mugil cephalus (blood): northern Gulf of Mexico

Haemogregarina myoxocephali
Daily, D. D., 1978, J. Parasitol., v. 64 (2), 361-362
Myoxocephalus octodecemspinosus: Whiting Bay, Maine

Haemogregarina myoxocephali Fantham, Porter and Richardson, 1942, illus.
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102
description
Myoxocephalus octodecemspinosus (blood) St. Andrews, New Brunswick; Great Bay, N.H.

Haemogregarina platessae Lebailly, 1904
Becker, C. D.; and Overstreet, R. M., 1979, J. Fish Dis., v. 2 (6), 469-479
Paralichthys lethostigma
Trinectes maculatus
Achirus lineatus
Symphurus plagiusa
(blood of all): all from northern Gulf of Mexico

Haemogregarina platessae Lebailly, 1904, illus.
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102
description; ". . . it is submitted that the name H. achiri may have to be discarded as a synonym of H. platessae."
Paralichthys dentatus: Woods Hole, Massachusetts
Pseudopleuronectes americanus: St. Andrews, New Brunswick
(blood of all)

Haemogregarina pollachii Henry
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102
discarded as nomen nudum

Haemogregarina sachai n. sp., illus.
Kirmse, P. D., 1978, J. Fish Dis., v. 1 (4), 337-342
Haemogregarina sachai n. sp., life cycle
Scophthalmus maximus (blood, haemopoietic tissue and other organs): fish farms, Hunterston, Ayrshire, Scotland

Haemogregarina sachai, illus.
Kirmse, P., 1979, Ztschr. Parasitenk., v. 58 (3), 201-210
Haemogregarina sachai from Scophthalmus maximus, fine structure of intracellular and extracellular stages

Haemogregarina simondi, illus.
Kirmse, P., 1979, Ztschr. Parasitenk., v. 59 (2), 131-140
Haemogregarina simondi, ultrastructure of developing stages

Haemogregarina simondi (Laveran and Mesnil, 1901), illus.
Kirmse, P., 1979, Ztschr. Parasitenk., v. 59 (2), 141-150
Haemogregarina simondi, redescription of life cycle
Solea solea (blood): White Fish Authority fish farm, Hunterston, Scotland

Haemogregarina stepanowi, probably
Keymer, I. F., 1978, Vet. Rec., v. 103 (26-27), 577-582
parasitic and other diseases of terrapins and turtles, necropsy survey
Emys orbicularis

Hemogregarina stepanovi (Dan., 1886), illus.
Mishra, G. S.; and Gonzalez, J. P., 1978, Arch. Inst. Pasteur Tunis, v. 55 (3), 303-326
Clemmys caspica leprosa: Tunisie

Haemogregarina uncinata sp. n., illus.
Khan, R. A., 1978, J. Parasitol., v. 64 (1), 35-44
Haemogregarina uncinata sp. n., life cycle, leech (Johanssonia sp.) as probable vector
Lycodes lavalaei (erythrocytes): Grand Banks, northwestern Atlantic Ocean, off Newfoundland
L. vahlii (erythrocytes): Grand Banks, northwestern Atlantic Ocean, off Newfoundland
Johanssonia sp. (stomach ceca, intestinal lumina, intestinal epithelium) (exper.)

Haemogregarina urophysis Fantham et al.
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102
as syn. of H. aeglefini Henry, 1913

Haemogregarine
Becker, C. D.; and Overstreet, R. M., 1979, J. Fish Dis., v. 2 (6), 469-479
Dasyatis sabina (blood): northern Gulf of Mexico

Haemogregarines
Beier, T. V.; and Sidorenko, N. V., 1972, Parazitologiia, Leningrad, v. 6 (4), 385-390
haemogregarine-infected erythrocytes of Lacerta armeniaca and L. saxicola nairensis, changes in hemoglobin and total protein content

Haemogregarine, similar to Haemogregarine E-type, illus.
Ishii, A., 1973, Snake, v. 5 (1-2), 133-140
Trimeresurus flavoviridis (blood, liver): Amami-oshima, southern Japan

Haemogregarines
Keymer, I. F., 1978, Vet. Rec., v. 103 (26-27), 577-582
Emydoidea blandingi

Hemogregarines
Moreno, E.; and Bolanos, R., 1977, Rev. Biol. Trop., v. 25 (1), 47-57
Lachesis muta (pulmon, sangre, higado)
Crotalus durissus (sangre)
Bothrops asper (sangre)
B. picadoi (sangre)
B. nummifer (sangre)
B. godmani (sangre)
Micrurus nigrocinctus (sangre)
Clelia clelia (sangre)
Boa constrictor (sangre)
Amblyomma dissimile (cavidad general, ampolla rectal)
all from Costa Rica

Haemogregarines
Peirce, M. A.; and Mead, C. J., 1977, J. Nat. Hist., v. 11 (6), 597-600
Troglodytes troglodytes
Sylvia curruca
Phylloscopus trochilis
Parus caeruleus
P. major
all from near Tring, Hertfordshire

Haemogregarine A-type, illus.
Toshioka, S., 1970, Snake, v. 2 (1), 53-56
Agkistrodon halys: Japan

Haemogregarine B-type, illus.
Toshioka, S., 1970, Snake, v. 2 (1), 53-56
Agkistrodon halys: Korea

Haemogregarine C-type, illus.
Toshioka, S., 1970, Snake, v. 2 (1), 53-56
Rhabdophis t. tigrinus

Haemogregarine D-type, illus.
Toshioka, S., 1970, Snake, v. 2 (1), 53-56
Dinodon semicarinatus: Japan Snake Institut

Haemogregarine E-type, illus.
Toshioka, S., 1970, Snake, v. 2 (1), 53-56
Trimeresurus okinavensis: Japan Snake Institute

Haemogregarine F-type, illus.
Toshioka, S., 1970, Snake, v. 2 (1), 53-56
Trimeresurus tokarensis: Japan Snake Institute

Haemogregarine G-type, illus.
Toshioka, S., 1970, Snake, v. 2 (1), 53-56
Elaphe quadrivirgata: Japan Snake Institute

Haemohormidium Henry
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102
Syn.: Babesiosoma Jakowska and Nigrelli

Haemohormidium sp., illus.
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102
description
Hemitripterus americanus
Hippoglossoides platessoides
(blood of all): all from St. Andrews, New Brunswick

Haemohormidium aulopi (Mackerras and Mackerras, 1925) [n. comb.]
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102

Haemohormidium jahni (Nigrelli, 1929) [n. comb.]
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102

Haemohormidium mariae (Hoare, 1930) [n. comb.]
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102

Haemohormidium rubrimarensis (Saunders, 1960) [n. comb.]
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102

Haemohormidium stableri (Schmittner and McGhee, 1961) [n. comb.]
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102

aemohormidium tetragonis (Becker and Katz, 965) [n. comb.]
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102

aemoproteus
Bennett, G. F., 1978, Canad. J. Zool., v. 56 (8), 1721-1725
key to species of Haemoproteus parasitic in Meropidae

aemoproteus
Greiner, E. C.; Eveleigh, E. S.; and Boone, W. M., 1978, J. Med. Entom., v. 14 (6), 701-704
collection of Culicoides spp. in bird-baited traps, relative abundance and seasonal occurrence in relation to concurrent avian Haemoproteus infections: Tantramar Marshes, New Brunswick

emoproteus (or Parahaemoproteus) [sp.]
Baker, J. R., 1974, J. Zool., London, v. 172 (2), 169-190
Fringilla coelebs
Garrulus glandarius
Muscicapa striata
Passer domesticus
Sturnus vulgaris
Turdus ericetorum
T. merula
Athene noctua
Columba palumbus
Lagopus scoticus
all from Britain

emoproteus [sp.]
Bennett, G. F.; Cameron, M.; and White, E., 1975, Canad. J. Zool., v. 53 (10), 1432-1442
hematozoa of passeriforms, prevalence, effect of climate, application of insecticide, and large-scale environmental alteration
Carpodacus purpureus
Junco hyemalis
Melospiza georgiana
M. lincolnii
M. melodia
Passerculus sandwichensis
Pheucticus ludovicianus
Pooecetes gramineus
Spinus tristis
Zonotrichia albicollis
sparrows, unknown
Agelaius phoeniceus
Dolichonyx oryzivoros
Molothrus ater
Quiscalus quiscula
Dumetella carolinensis
Dendroica coronata
D. fusca
D. magnolia
D. palmarum
D. pensylvanica
D. petechia
D. striata
D. tigrina
Geothlypis trichas
Seiurus novaboracensis
Setophaga ruticilla
Vermivora peregrina
V. ruficapilla
Wilsonia pusilla
Catharus guttatus
C. ustulatus
Turdus migratorius

Haemoproteus [sp.].-- Continued.
Bennett, G. F.; Cameron, M.; and White, E., 1975, Canad. J. Zool., v. 53 (10), 1432-1442
Empidonax trailii
Empidonax sp.
Tyrannus tyrannus
Vireo olivaceous
Piranga olivacea
Catharus fuscescens
all from New Brunswick

Haemoproteus sp.
Bennett, G. F.; White, E. M.; and Williams, N. A., 1977, J. Wildlife Dis., v. 13 (3), 251-257
Pogoniulus bilineatus
Chrysococcyx caprius
Hirundo abyssinica
H. griseopyga
Aerops albicollis
Melittophagus variegatus
Anthus leucophrys
Macronyx croceus
Motacilla alba
M. flava
Dyaphorophyia castanea
Parisoma plumbeum
Tchitrea nigriceps
Cinnyris chloropygius
Nectarinia senegaiensis
N. verticalis
Plocepasser mahali
P. aurantius
P. cucullatus
P. luteolus
P. nigerrimus
P. nigricollis
P. ocularis
P. weynsi
Quelea erythrops
Pycnonotus virens
Camaroptera brevicaudata
Sylvia borin
Trichastoma fulvescens
Zosterops senegalensis
(blood of all): all from Uganda

Haemoproteus sp.
Kigaye, M. K., 1977, Bull. Animal Health and Prod. Africa, v. 25 (3), 295-298
Leptoptilos crumeniferus (blood): Mukuno abattoir, about 16 km from Kampala, Uganda

Haemoproteus [sp.]
Kocan, A. A.; Snelling, J.; and Greiner, E. C., 1977, J. Wildlife Dis., v. 13 (3), 304-306
Strix varia
Bubo virginianus
Otus asio
Buteo jamaicensis
Falco sparverius
Accipiter cooperii
Buteo lagopus
all from Oklahoma

Haemoproteus [sp.]
Peirce, M. A.; and Mead, C. J., 1977, J. Nat. Hist., v. 11 (6), 597-600
Muscicapa striata: near Tring, Hertfordshire

Haemoproteus [sp.]
White, E. M.; and Bennett, G. F., 1978, Canad. J. Zool., v. 56 (10), 2110-2116
Hirundo senegalensis
H. tahitica
Ptyonoprogne fuligula
Hirundo cucullata

Haemoproteus [sp.]
Williams, N. A., 1978, J. Parasitol., v. 64 (3), 556-558
Junco hyemalis
Melospiza melodia
Spinus pinus
Zonotrichia leucophrys
Vermivora celata
Catharus guttatus
C. ustalatus
all from 16 km west of Courtenay, British Columbia

Haemoproteus [spp.]
Williams, N. A.; and Bennett, G. F., 1978, Canad. J. Zool., v. 56 (4, pt. 1), 596-607
Accipiter cooperii: Maryland
Buteo jamaicensis: Maryland
B. platypterus: "
Anas rubripes: "
Olor columbianus: "
Oxyura jamaicensis: "
Egretta thula: Maryland or New Jersey
Nycticorax nycticorax: New Jersey
Cathartes aura: Maryland, and/or New Jersey
Vireo flavifrons: Maryland
Corvus brachyrhynchos: "
Cyanocitta cristata: Maryland, and/or New Jersey
Cardinalis cardinalis: Maryland, and/or New Jersey
Junco hyemalis: Maryland, and/or New Jersey
Melospiza melodia: Maryland, and/or New Jersey
Pheucticus ludovicianus: Maryland or New Jersey
Pipilo erythrophthalmus: Maryland, and/or New Jersey
Spinus tristis: Maryland or New Jersey
Spizella passerina: Maryland, and/or New Jersey
S. pusilla: Maryland, and/or New Jersey
Zonotrichia albicollis: Maryland, and/or New Jersey
Agelaius phoeniceus: Maryland, and/or New Jersey
Icterus galbula: Maryland and/or New Jersey
Molothrus ater: Maryland, and/or New Jersey
Quiscalus quiscula: Maryland, and/or New Jersey
Dumetella carolinensis: Maryland, and/or New Jersey
Mimus polyglottus: Maryland or New Jersey
Toxostoma rufum: Maryland, and/or New Jersey
Parus bicolor: Maryland, and/or New Jersey
Dendroica coronata: Maryland, and/or New Jersey
D. petechia: Maryland
Geothlypis trichas: Maryland
Icteria virens: Maryland, and/or New Jersey
Mniotilta varia: Maryland, and/or New Jersey
Seiurus aurocapillus: Maryland, and/or New Jersey
Wilsonia citrina: Maryland
Colinus virginianus: Maryland, and/or New Jersey
Passer domesticus: Maryland, and/or New Jersey
Porzana carolinensis: Maryland
Calidris minutilla: "
C. pusilla: Maryland
Otus asio: "
Strix varia: Maryland, and/or New Jersey
Sturnus vulgaris: Maryland, and/or New Jersey
Piranga olivacea: Maryland

Haemoproteus [spp.].-- Continued.
Williams, N. A.; and Bennett, G. F., 1978, Canad J. Zool., v. 56 (4, pt. 1), 596-607
Hylocichla mustelina: Maryland or New Jersey
Turdus migratorius: Maryland, and/or New Jersey
Myiarchis crinitus: Maryland
Sayornis phoebe: Maryland, and/or New Jerse
Tyrannus tyrannus: Maryland
Vireo olivaceus: Maryland, and/or New Jersey

Haemoproteus [sp.]
Winchell, E. J., 1978, J. Parasitol., v. 64 (3), 558-559
Piranga ludoviciana: El Salvador

Haemoproteus spp.
Wink, M.; and Bennett, G. F., 1976, J. Wildlife Dis., v. 12 (4), 587-590
Estrildidae
Passer griseus
Ploceus cucullatus
Ploceus nigerrimus
Ploceus atrogularis
Hippolais polyglotta
all from Ghana

Haemoproteus acanthis Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus accipiter Zeiniev, 1975; Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus archilochus Coatney and West 1938
Williams, N. A., 1978, J. Parasitol., v. 64 (3), 556-558
Selasphorus rufus: 16 km west of Courtenay, British Columbia

Haemoproteus asio Zeiniev, 1975; Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus brodkorbi sp. nov., illus.
Forrester, D. J.; et al., 1977, Canad. J. Zool., v. 55 (8), 1268-1274
Leptoptilos crumeniferus: Mukono, 10 mi. from Kampala, Uganda
Mycteria americana: Fisheating Creek Wildlife Refuge, Glades County, Palmdale, Florida
(blood of all)

Haemoproteus caprimulgi Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
homonym of H. caprimulgi Williams, Bennett & Mahrt, 1975

Haemoproteus caprimulgi Williams, Bennett, and Mahrt 1975
Williams, N. A., 1978, J. Parasitol., v. 64 (3), 556-558
Chordeiles minor: 16 km west of Courtenay, British Columbia

Haemoproteus chelidonis Franchini, 1922
White, E. M.; and Bennett, G. F., 1978, Canad. J. Zool., v. 56 (10), 2110-2116
taxonomic review, species inquirenda

Haemoproteus chukari Tartakovsky, 1913
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus columbae Kruse, illus.
Ahmed, F. E.; and Mohammed, A. H. H., 1978, J. Protozool., v. 25 (2), 174-177
Haemoproteus columbae in pigeons, growth and development of gametocytes, effect on host cell, multiple infection of erythrocytes, sex ratio

aemoproteus columbae Kruse
Ahmed, F. E.; and Mohammed, A. H. H., 1978, Ztschr. Parasitenk., v. 57 (3), 229-236
Haemoproteus columbae in Columba livia (exper.), course of infection, relapse, and immunity to reinfection

Haemoproteus columbae
da Costa, S. C. G.; and Lins, D. C., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 39-40
sporozoans, staining for nucleic acids

Haemoproteus columbae
Qureshi, M. I.; and Sheikh, A. H., 1978, Pakistan J. Sc., v. 30 (1-6), 165-167
chickens, Desi, white leghorns: Lahore district
pigeons: Lahore Zoo

Haemoproteus columbae Kruse
Winchell, E. J., 1978, J. Parasitol., v. 64 (3), 558-559
Columbina talpacoti: El Salvador

Haemoproteus coraciatis Tartakovsky, 1913
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus coturnix Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus crumenium (Hirst, 1905)
Forrester, D. J.; et al., 1977, Canad. J. Zool., v. 55 (8), 1268-1274
nomen dubium

Haemoproteus cuculus Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus danilewskyi Kruse, 1890
Kairullaev, K. K., 1979, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (1), 19-21
Phoenicurus phoenicurus
Sylvia borin
S. curruca
S. atricapilla
Parus major
P. coeruleus
(blood of all): all from central course of Ural river, Priural region, Ural oblast

aemoproteus emberiza Berson, 1964
Kairullaev, K. K., 1979, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (1), 19-21
Emberiza citrinella (blood): central course of Ural river, Priural region, Ural oblast

aemoproteus emberiza Zeiniev, 1975, quotes Ber-
on, 1964, as authority
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus enucleator
Bennett, G. F.; White, E. M.; and Williams, N. A., 1977, J. Wildlife Dis., v. 13 (3), 251-257
Ispidina picta (blood): Uganda

Haemoproteus erythropi Tartakovsky, 1913
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus fallisi
Bennett, G. F.; White, E. M.; and Williams, N. A., 1977, J. Wildlife Dis., v. 13 (3), 251-257
Cossypha natalensis (blood): Uganda

Haemoproteus fallisi
Peirce, M. A.; and Mead, C. J., 1977, J. Nat. Hist., v. 11 (6), 597-600
Turdus merula: near Tring, Hertfordshire

Haemoproteus fallisi Bennett and Campbell
Winchell, E. J., 1978, J. Parasitol., v. 64 (3), 558-559
Turdus grayi: El Salvador

Haemoproteus fringillae/orizivorae
Bennett, G. F.; White, E. M.; and Williams, N. A., 1977, J. Wildlife Dis., v. 13 (3), 251-257
Lagonosticta rubricata (blood): Uganda

Haemoproteus fringilla Labbe, 1894
Kairullaev, K. K., 1979, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (1), 19-21
Fringilla montifringilla
F. coelebs
(blood of all): all from central course ot Ural river, Priural region, Ural oblast

Haemoproteus fringillae Labbe/orizivora Anschutz complex
Winchell, E. J., 1978, J. Parasitol., v. 64 (3), 558-559
Vermivora peregrina
Vireo gilvus
all from El Salvador

Haemoproteus galatheae Laird and Laird, 1959
Bennett, G. F.; Greiner, E. C.; and Campbell, A. G., 1975, Canad. J. Zool., v. 53 (3), 634-638
as syn. of H. plataleae de Mello, 1935

Haemoproteus garrulus Zeiniev, 1975; Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus halcyonis
Bennett, G. F.; White, E. M.; and Williams, N. A., 1977, J. Wildlife Dis., v. 13 (3), 251-257
Alcedo cristata
Halcyon senegalensis
(blood of all): all from Uganda

Haemoproteus hirundinis (Sergent & Sergent, 1905) emend Levine & Campbell, 1971
White, E. M.; and Bennett, G. F., 1978, Canad. J. Zool., v. 56 (10), 2110-2116
taxonomic review, valid species

Haemoproteus janovyi sp. n., illus.
Greiner, E. C.; and Mundy, P. J., 1979, J. Parasitol., v. 65 (1), 147-153
hematozoa from southern African vultures, prevalence, host age class, seasonal variation
Gyps africanus: Sengwa Wildlife Research Area, Rhodesia
Trigonoceps occipitalis: Sengwa Wildlife Research Area, Rhodesia
Torgos tracheliotus: Sengwa Wildlife Research Area, Rhodesia
Necrosyrtes monachus: Rhodesia [and/or] South Africa

Haemoproteus lairdi sp. n., illus.
Bennett, G. F., 1978, Canad. J. Zool., v. 56 (8), 1721-1725
key
Melittophagus variegatus (blood): Entebbe, Uganda

Haemoproteus lanii
Bennett, G. F.; White, E. M.; and Williams, N. A., 1977, J. Wildlife Dis., v. 13 (3), 251-257
Telophorus sulfureopectus (blood): Uganda

Haemoproteus lanii
Wink, M.; and Bennett, G. F., 1976, J. Wildlife Dis., v. 12 (4), 587-590
Lanius senator: Ghana

Haemoproteus lanuidae Yakunin, 1976
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus loxiae Tartakovsky, 1913
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus maccallumi, illus.
Ayala, S. C.; et al., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (6), 411-416
Haemoproteus, Plasmodium, and hippoboscid ectoparasites of Zenaida auriculata caucae, infection patterns and dove population dynamics, seasonal prevalence
Zenaida auriculata caucae (blood)
Columbina talpacoti
domestic pigeons
all from Cauca River valley, Colombia

Haemoproteus maccallumi Novy and MacNeal, 1904
Williams, N. A.; and Bennett, G. F., 1978, Canad. J. Zool., v. 56 (4, pt. 1), 596-607
Zenaidura macroura: Maryland

Haemoproteus majoris Zeiniev, 1975; Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
homonym of Haemoproteus majoris (Laveran)

Haemoproteus manwelli sp. n., illus.
Bennett, G. F., 1978, Canad. J. Zool., v. 56 (8), 1721-1725
key
Merops orientalis (blood): Maharashtra, India; Chiengmai, Thailand
M. viridis (blood): Subang, Malaysia

Haemoproteus meropi Zargar, 1945
Bennett, G. F., 1978, Canad. J. Zool., v. 56 (8), 1721-1725
emended to: Haemoproteus meropis

Haemoproteus meropis Tartakovskii, 1913
Bennett, G. F., 1978, Canad. J. Zool., v. 56 (8), 1721-1725
nomen nudum

Haemoproteus meropis (Zargar, 1945) emend. Bennett, 1978, illus.
Bennett, G. F., 1978, Canad. J. Zool., v. 56 (8), 1721-1725
redescription, key
emendation of: Haemoproteus meropi Zargar, 1945
Merops viridis: Subang, Malaysia
M. leschnaulti
Aerops albicollis: Masaka, Uganda
(blood of all)

Haemoproteus meropis Tartakovsky, 1913
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus merulla Zeiniev, 1975; Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus monticola Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus motasillae Zeiniev, 1975
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus muscicapa Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus myophonus Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus nettionis
Greiner, E. C.; Eveleigh, E. S.; and Boone, W. M., 1978, J. Med. Entom., v. 14 (6), 701-704
collection of Culicoides spp. in bird-baited traps, relative abundance and seasonal occurrence in relation to concurrent avian Haemoproteus infections: Tantramar Marshes, New Brunswick

Haemoproteus nettionis
Williams, N. A.; Calverley, B. K.; and Mahrt, J. L., 1977, J. Wildlife Dis., v. 13 (3), 226-229
Anas platyrhynchos (blood): Central Alberta and Mackenzie Delta, Northwest Territories
Anas acuta (blood): Mackenzie Delta, Northwest Territories

Haemoproteus oenante Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus orizivorae/fringillae
Bennett, G. F.; White, E. M.; and Williams, N. A., 1977, J. Wildlife Dis., v. 13 (3), 251-257
Lagonosticta rubricata (blood): Uganda

Haemoproteus orizivorae
Peirce, M. A.; and Mead, C. J., 1977, J. Nat. Hist., v. 11 (6), 597-600
Sylvia atricapilla: near Tring, Hertfordshire

emoproteus orizivora
Winchell, E. J., 1978, J. Parasitol., v. 64 (3), 558-559
Dendroica magnolia
Polioptila albiloris
Pheucticus ludovicianus
all from El Salvador

emoproteus orizivora Anschutz/fringillae
bbe complex
Winchell, E. J., 1978, J. Parasitol., v. 64 (3), 558-559
Vermivora peregrina
Vireo gilvus
all from El Salvador

emoproteus otus Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

emoproteus palumbis
Peirce, M. A.; and Mead, C. J., 1977, J. Nat. Hist., v. 11 (6), 597-600
Columba palumbus: near Tring, Hertfordshire

emoproteus passeris Kruse, 1890
Kairullaev, K. K., 1979, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (1), 19-21
Passer montanus (blood): central course of Ural river, Priural region, Ural oblast

emoproteus pastor Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

emoproteus peircei sp. nov., illus.
Forrester, D. J.; et al., 1977, Canad. J. Zool., v. 55 (8), 1268-1274
Ephippiorhynchus senegalensis: Sudan
Leptoptilos sp.: Southeast Asia
Pseudotantalus ibis: Zaire

moproteus peircei sp. nov., possibly
Forrester, D. J.; et al., 1977, Canad. J. Zool., v. 55 (8), 1268-1274
Pseudotantalus ibis: Mali

emoproteus pelouroi Tendeiro, 1947, illus.
Bennett, G. F.; Greiner, E. C.; and Campbell, A. G., 1975, Canad. J. Zool., v. 53 (3), 634-638
valid species, redescription

moproteus perdix Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

moproteus phoenicurus Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

emoproteus plataleae de Mello, 1935, illus.
Bennett, G. F.; Greiner, E. C.; and Campbell, A. G., 1975, Canad. J. Zool., v. 53 (3), 634-638
valid species, redescription
Syn.: Haemoproteus galatheae Laird and Laird, 1959
Eudocimus albus (blood): southern Florida
Threskiornis molucca pygmaeus: Rennell Island, British Solomon Islands

emoproteus pratasi
Wink, M.; and Bennett, G. F., 1976, J. Wildlife Dis., v. 12 (4), 587-590
Francolinus ahantensis: Ghana

Haemoproteus prognei Coatney and Roudabush, 1937, illus.
White, E. M.; and Bennett, G. F., 1978, Canad. J. Zool., v. 56 (10), 2110-2116
redescription
Hirundo abyssinica: Entebbe, Uganda
H. daurica: central Thailand
H. rustica: Entebbe, Uganda
Progne subis: Dennisville, New Jersey

Haemoproteus prognei Coatney and Roudabush, 1937
Williams, N. A.; and Bennett, G. F., 1978, Canad. J. Zool., v. 56 (4, pt. 1), 596-607
Progne subis: Maryland, and/or New Jersey

Haemoproteus raymundi
Wink, M.; and Bennett, G. F., 1976, J. Wildlife Dis., v. 12 (4), 587-590
Cyanomitra olivacea: Ghana

Haemoproteus sanguinus
Wink, M.; and Bennett, G. F., 1976, J. Wildlife Dis., v. 12 (4), 587-590
Pycnonotus barbatus
Eurillas virens
all from Ghana

Haemoproteus sittae Zeiniev, 1975; Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus stellaris n. sp., illus.
White, E. M.; and Bennett, G. F., 1978, Canad. J. Zool., v. 56 (10), 2110-2116
Hirundo griseopyga (blood): Kasenyi, Uganda

Haemoproteus sternae Zeiniev, 1975; Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus sylvia Musaev & Zeiniev, 1977
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Haemoproteus tinnunculus
Peirce, M. A.; and Mead, C. J., 1977, J. Nat. Hist., v. 11 (6), 597-600
Falco subbuteo: near Tring, Hertfordshire

Haemoproteus velans
Bennett, G. F.; Cameron, M.; and White, E., 1975, Canad. J. Zool., v. 53 (10), 1432-1442
hematozoa of passeriforms, prevalence, effect of climate, application of insecticide, and large-scale environmental alteration
Colaptes auratus: New Brunswick

Haemosporidian, undetermined species, illus.
Telford, S. R., jr., 1978, Internat. J. Parasitol., v. 8 (5), 341-353
description of gametocytes
Thecadactylus rapicaudus (proerythrocytes, erythrocytes, macrophage, monocyte): Village of Tierra Caliente, Municipio Manrique, Estado Cojedes, Venezuela; Portuguesa, Venezuela; San Blas Territory, Panama

Hammondia
Beier, T. V.; Shibalova, T. A.; and Kostenko, L. A., 1978, [Cytology of coccidia], 185 pp., illus.

Hammondia
Gill, H. S.; et al., 1978, J. Parasitol., v. 64 (3), 549-551
dogs fed Bubalus bubalis diaphragm muscles with visible cysts of Sarcocystis fusiformis shed unsporulated oocysts belonging either to Hammondia or Isospora

Hammondia
Heydorn, A. O., 1979, Berl. u. Munchen. Tierarztl. Wchnschr., v. 92 (11), 214-220
coccidia of cats, life cycle, epidemiology, review

Hammondia
Rommel, M., 1978, Ztschr. Parasitenk., v. 57 (3), 269-283
cyst-forming coccidia, life cycle, taxonomy, comparative review

Hammondia hammondi
Dubey, J. P., 1978, J. Protozool., v. 25 (3, pt. 2), 382-384
Hammondia hammondi, Besnoitia jellisoni, Toxoplasma gondii, BCG, comparison of cross-protection in hamsters

Hammondia hammondi Frenkel and Dubey 1975
Dubey, J. P.; and Wong, M. M., 1978, J. Parasitol., v. 64 (3), 551-552
Hammondia hammondi, experimentally infectious to only 1 of 4 monkey species (Saguinus nigricollis), all 4 species developed low and transitory antibody titers to Toxoplasma gondii antigen after inoculation with Hammondia oocysts

Hammondia hammondi, illus.
Entzeroth, R.; Scholtyseck, E.; and Greuel, E., 1978, Naturwissenschaften, v. 65 (7), 395
"The application of different techniques led to the conclusion that, besides three ultramorphologically different sarcosporidian cysts, the roe-deer [Capreolus capreolus] muscles contain coccidia of the genera Toxoplasma and Hammondia."

Hammondia hammondi
Fayer, R.; and Frenkel, J. K., 1979, J. Parasitol., v. 65 (5), 756-762
6 spp. of feline coccidia, oocysts fed to calves, calf tissue then fed to cats: neither Besnoitia, Hammondia, nor Sarcocystis were pathogenic for calves nor did they establish patent infections which could be transmitted back to cats; Cystoisospora spp. were not pathogenic for calves but could be transmitted back to cats; Toxoplasma strains were slightly to moderately pathogenic for calves and could be transmitted back to cats

Hammondia hammondi
Markus, M. B., 1979, Ann. Trop. Med. and Parasitol., v. 73 (4), 393-394
Hammondia hammondi Frenkel, 1974 is correct authorship rather than Frenkel and Dubey, 1975 as previously published

Hammondia hammondi, illus.
Mason, R. W., 1978, Ztschr. Parasitenk., v. 57 (2), 101-106
Hammondia hammondi, first detection in Australia and first record from natural intermediate host
cats (exper.)
Rattus rattus: Australia
Mus musculus (nat. and exper.): Australia

Hammondia hammondi
Perrotin, C.; and Graber, M., 1977, Rev. Elevage et Med. Vet. Pays Trop., n.s., v. 30 (4), 377-382
Sarcocystis spp., domestic animals, review of life cycle and differential characters from Toxoplasma gondii and Hammondia hammond

Hammondia hammondi
Weiland, G.; Rommel, M.; and von Seyerl, F., 1979, Berl. u. Munchen. Tierarztl. Wchnschr., v. 92 (2), 30-32
Toxoplasma gondii, Hammondia hammondi, mice, serological cross-reactions

Hammondia heydorni
Boch, J.; Boehm, A.; and Weiland, G., 1979, Berl. u. Munchen. Tierarztl. Wchnschr., v. 92 (12), 240-243
coccidia, dogs (feces), coprological and serological survey: South Germany

Hammondia heydorni-like oocysts
Dubey, J. P.; Fayer, R.; and Seesee, F. M., 1978, J. Am. Vet. Med. Ass., v. 173 (9), 1167-1170
coyotes (feces): Montana

Hammondia pardalis sp. n., illus.
Hendricks, L. D.; et al., 1979, J. Protozool., v. 26 (1), 39-43
Felis pardalis (nat. and exper.) (intestinal contents): Albrook Air Force Station, Panama Canal Zone
F. catus (exper.)
F. yagouaroundi (exper.)
Mus musculus (exper.) (mesenteric lymph nodes, lungs, and intestinal submucosa)

Haplosporidan, illus.
Armstrong, D. A.; and Armstrong, J. L., 1974, Proc. National Shellfish Ass., v. 64, 68-72
haplosporidan in Tresus capax, incidence of infection observed by the presence of cysts in host tissue, age of host, pathology: Yaquina Bay, Oregon

Haplosporidans, resembling Minchinia costalis, illus.
Newman, M. W., 1971, Proc. 1970 National Shellfish. Ass., v. 61, 59-63
Crassostrea virginica (mantle): New Haven Harbor, Connecticut

Haplosporidans, resembling Minchinia nelsoni
Newman, M. W., 1971, Proc. 1970 National Shellfish. Ass., v. 61, 59-63
Crassostrea virginica: New Haven Harbor, Connecticut

Haptophrya Stein
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key

Haptophryidae
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key

Hartmannella
Botto, C.; and Rodriguez, N., 1975, Rev. Uruguaya Patol. Clin. y Microbiol., v. 13 (1), 3-16
human amoebic meningoencephalitis, etiology, epidemiology, pathology, diagnosis, therapy, review

rtmannella-Naegleria free-living amoebae, illus.
Carosi, G., 1974, Ann. Soc. Belge Med. Trop., v. 54 (4-5), 265-277
Entamoeba moshkovskii and free-living amoeba of Hartmannella-Naegleria group, ultrastructural comparisons

rtmannella-Naegleria groups
Das, S. R., 1975, New Approaches Ident. Microorgan., 451-458
Naegleria-Hartmannella groups, agar plate method for isolation and cultivation of amoebae from infected humans and animals, soil, sewage sludge, fresh water, and other substrates

rtmanella, illus.
Fleischer, N. K. F., 1979, Med. Welt., v. 30 (44), 1625-1630
parasitic tropical diseases, humans, central nervous system involvement, clinical review

rtmannella
Gottlieb, B.; and Reyes, H., 1977, Rev. Med. Chile, v. 105 (7), 467-471
Naegleria, Acanthamoeba, Hartmannella, free-living amoeba now thought to be agents responsible for human meningoencephalitis, diagnosis, pathology, therapy, review

rtmanella
Shirkina, N. I., 1975, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (27), 15-17
Hartmanella, amebas of Limax group, strains N and A-1, morphology, pathogenicity to mice

artmanella-Acanthamoeba group amoeba, illus.
Stibbs, H. H.; et al., 1979, J. Invert. Path., v. 33 (2), 159-170
Hartmanella-Acanthamoeba group amoeba isolated from Biomphalaria glabrata (Schistosoma mansoni susceptible and resistant strains) (pericardium, mantle), in vitro killing of S. mansoni sporocysts by amoebae; whether amoebae contribute to snail resistance is unknown

artmannella sp.
Pernin, P.; and Riany, A., 1978, Ann. Parasitol., v. 53 (4), 333-344
survey of swimming pools for presence of free-living amebae, potential danger for swimmers: Lyon

artmanella sp., illus.
Scholten, T., 1979, Canad. Med. Ass. J., v. 120 (3), 264, 267-268 [Letter]
soil amoebae potentially pathogenic to man, existence in Canada, results of survey from various areas of Ontario, experimental infections in mice

artmannella [sp.]
Watson, P. G., 1975, Tr. Ophth. Soc. United Kingdom, v. 95 (2), 204-206
Hartmannella infection of eye, woman, pathology, case history

artmannella astronyxis new species, illus.
Ray, D. L.; and Hayes, R. E., 1954, J. Morphol., v. 95 (1), 159-188
free-living: contents of tube of water placed in laboratory termite culture

Hartmannella castellani, illus.
Carosi, G., 1974, Ann. Soc. Belge Med. Trop., v. 54 (4-5), 265-277
Entamoeba moshkovskii and free-living amoeba of Hartmannella-Naegleria group, ultrastructural comparisons

Hartmanella culbertsoni
Lal, A. A.; and Garg, N. K., 1979, Exper. Parasitol., v. 48 (3), 331-336
Hartmanella culbertsoni, meningoencephalitic mice, biochemical changes in brain

Hartmannella culbertsoni
Rai, R. D.; and Mohan Rao, V. K., 1978, Indian J. Exper. Biol., v. 16 (8), 903-905
Hartmannella culbertsoni, mice, experimental amoebic meningoencephalitis, gross biochemical changes in brain

Hartmannella culbertsoni
Singh, U. S.; Raizada, M. K.; and Rao, V. K. M., 1978, Zentralbl. Bakteriol., 1 Abt. Orig., Reihe A, v. 241 (3), 358-367
Hartmannella culbertsoni, axenically grown, purification and properties of L-histidine ammonia-lyase, marked inhibitory effect of certain amoebicidal drugs and divalent cations

Hartmannella vermiformis n. sp., illus.
Page, F. C., 1967, J. Protozool., v. 14 (3), 499-521
Pigeon Lake, Wisconsin; Kankakee River at Schneider, Indiana

Hartmannella vermiformis Page, 1967
Derr-Harf, C.; et al., 1978, Ann. Parasitol., v. 53 (5), 467-477
isolation of 164 strains of free-living amoebae from public water supplies and swimming pools, some strains of Acanthamoeba polyphaga, A. castellanii, A. lenticulata, and A. comandoni were pathogenic for mice, none of the Naegleria isolated were pathogenic for mice, levels of bromine and chlorine used were insufficient to eliminate these amoebae, potential danger to users and suggested control measures: Strasbourg

Hartmannella vermiformis
Kadlec, V., 1978, J. Protozool., v. 25 (2), 235-237
15 strains of free-living amebae found in variety of domestic animals, 3 isolates tested were not pathogenic for laboratory animals, some features of isolates differed from those previously known for members of these genera
dog (bronchi)
turkey (trachea)

Hartmannella vermiformis
Simitzis, A. M.; Le Goff, F.; and L'Azou, M. T., 1979, Ann. Parasitol., v. 54 (2), 121-127
free-living amebae, 9 strains isolated from nasal mucosa of 1039 healthy humans sampled, pathogenicity for mice and in cell culture tested

Hartmannellidae
Rivera, F.; et al., 1979, Tr. Am. Micr. Soc., v. 98 (3), 465-469
protozoa polluting tap water, concentration and identification in culture: Federal District, Mexico City

Hegneriella Earl, 1971
Earl, P. R., 1979, Tr. Am. Micr. Soc., v. 98 (4), 549-557
Hegneriellina [i.e.? Hegneriellinae]

Hegneriellina [i.e.? Hegneriellinae] Earl, 1971
Earl, P. R., 1979, Tr. Am. Micr. Soc., v. 98 (4), 549-557
Opalinidae
includes: Hegneriella; Zelleriella

Helcosoma tropicum Wright, 1903
Gardener, P. J., 1977, Trop. Dis. Bull., v. 74 (12), 1069-1088
as syn. of Leishmania tropica (Wright, 1903) Luehe, 1906

Helicosporidium sp., illus.
Hembree, S. C., 1979, Mosquito News, v. 39 (3), 575-582
Aedes aegypti (nat. and exper.)
Culex quinquefasciatus (nat. and exper.)
(fat body of all): all from Thailand

Helicosporidium sp., illus.
Sayre, R. M.; and Clark, T. B., 1978, J. Invert. Path., v. 31 (2), 260-261
Daphnia magna: laboratory cultures
Aedes aegypti (exper.)

Hematodinium perezi Chatton and Poisson 1931, illus.
MacLean, S. A.; and Ruddell, C. L., 1978, J. Parasitol., v. 64 (1), 158-160
Cancer irroratus (hepatopancreas): Mid-Atlantic Bight, New York Bight Apex near Ambrose Light, and continental shelf and slope waters off New Jersey coast
C. borealis: Mid-Atlantic Bight, continental shelf and slope waters off New Jersey coast
Ovalipes ocellatus: Mid-Atlantic Bight, New York Bight Apex near Ambrose Light

Hemiophrys macrostoma Chen
Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
parasites of fishes: China

Hemogregarina. See Haemogregarina.

Hemogregarines. See Haemogregarines.

Henneguya sp.
Herman, R. L.; and Putz, R. E., 1970, J. Wildlife Dis., v. 6 (3), 173
Ictalurus punctatus (gill tissue): Maryland

Henneguya sp.
Meyers, T. R., 1978, Proc. Helminth. Soc. Washington, v. 45 (1), 120-128
Cynoscion regalis (pelvic fin, anal fin, caudal fin, pectoral fin)
Pomatomus saltatrix (bulbus arteriosus)
all from Raritan Bay, New Jersey

Henneguya adiposa Minchew, illus.
Current, W. L., 1979, J. Protozool., v. 26 (2), 209-217
Henneguya adiposa, ultrastructure of plasmodium wall and sporogenesis
Ictalurus punctatus (among connective tissue bands of adipose fin): Fish Farming Experimental Station, Stuttgart, Arkansas

Henneguya creplini
Dykova, I.; and Lom, J., 1978, J. Fish Biol., v. 12 (3), 197-202
Henneguya spp., developmental stages in hosts, histopathological changes in fish gill, inflammatory reaction may be a temperature dependent immune response
Acerina cernua (gills): South Bohemia

Henneguya creplini (Gurley, 1894)
Wierzbicki, K., 1970, Acta Parasitol. Polon., v. 18 (1-12), 45-55
brief description
Perca fluviatilis (gill filaments): Lake Dargin, Mazurian Lakeland, Poland

Henneguya exilis Kudo, illus.
Current, W. L.; and Janovy, J., jr., 1978, J. Protozool., v. 25 (1), 56-65
Henneguya exilis, comparative ultrastructure of interlamellar and intralamellar types of infection in Ictalurus punctatus, demonstrate that plasmodia of the 2 clinical types differ with respect to arrangement within gill filament, that interlamellar form causes extensive hyperplasia of host basal epithelium while intralamellar form does not, that clinical types differ with respect to ultrastructure of plasmodium walls, and that there is structural difference in surface coat covering plasmodia of the 2 clinical types

Henneguya exilis
Markiw, M. E.; and Wolf, K., 1978, J. Fish. Research Bd. Canada, v. 35 (6), 828-832
Myxosoma cerebralis, rabbits immunized with antigens extracted from mature spores or prespore stages, antisera and globulins used in fluorescent antibody techniques, direct fluorescent antibody test showed higher specificity than indirect FAT in cross reactions with other species of myxosporidians

Henneguya exilis Kudo, 1934
Mitchell, L. G., 1978, J. Protozool., v. 25 (1), 100-105
Ictalurus punctatus (gills, skin, gall bladder): Iowa

Henneguya psorospermica Thelohan, 1895
Andrews, C., 1979, J. Fish. Biol., v. 15 (2), 195-209
parasite fauna of Perca fluviatilis, host specificity, comparison with different localities in British Isles, factors affecting composition
Perca fluviatilis (gill filaments): Llyn Tegid, Wales

Henneguya psorospermica, illus.
Dykova, I.; and Lom, J., 1978, J. Fish Biol., v. 12 (3), 197-202
Henneguya spp., developmental stages in hosts, histopathological changes in fish gill, inflammatory reaction may be a temperature dependent immune response
Perca fluviatilis (blood vessels at bases of fine respiratory lamellae)
Esox lucius (afferent filament artery)
all from South Bohemia

Henneguya psorospermica, illus.
Podlipaev, S. A., 1972, Parazitologiia, Leningrad, v. 6 (6), 506-508
Myxosporidia, 6 spp., lipid inclusions in spores and cysts

Henneguya psorospermica, illus.
Podlipaev, S. A., 1974, Parazitologiia, Leningrad, v. 8 (6), 535-542
Myxobolidae spp., polysaccharides in spores, nature of iodinophilous vacuole

Henneguya salminicola
Olson, R. E., 1978, Calif. Fish and Game, v. 64 (2), 117-120
Oncorhynchus kisutch (musculature): Pacific Ocean off Newport, Oregon

Henneguya zschokkei (Gurley, 1893)
Arthur, J. R.; Margolis, L.; and Arai, H. P., 1976, J. Fish. Research Bd. Canada, v. 33 (11), 2489-2499
Coregonus clupeaformis (musculature, pelvic fin)
Prosopium cylindraceum (caudal musculature)
all from Aishihik Lake, Yukon Territory

Henneguya zschokkei (Gurley)
Iziumova, N. A.; Mashtakov, A. V.; and Timoshechkina, L. G., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36), 71-76
[Abramis brama]: Chusovaia river

Henneguya zschokkei
Makhovenko, E. T., 1972, Parazitologiia, Leningrad, v. 6 (4), 369-375
Salvelinus alpinus inhabiting different ecological niches, possible use of differences in parasite fauna between groups as biological tags: Lake Azabach'e, Kamchatka

Henneguya zschokkei
Markiw, M. E.; and Wolf, K., 1978, J. Fish. Research Bd. Canada, v. 35 (6), 828-832
Myxosoma cerebralis, rabbits immunized with antigens extracted from mature spores or prespore stages, antisera and globulins used in fluorescent antibody techniques, direct fluorescent antibody test showed higher specificity than indirect FAT in cross reactions with other species of myxosporidians

epatocystis
Garnham, P. C. C., 1977, Protozoology, v. 3, 135-142
taxonomic characters of species discussed

epatocystis sp., illus.
Landau, I., 1973, Mem. Mus. National Hist. Nat., Paris, n. s., s. A, Zool., v. 77, 1-62
Coccidiomorpha, life cycles, mechanisms assuring continuation of transmission, thesis

epatocystis sp. Landau, 1973 pro parte
Rosin, G.; Landau, I.; and Hugot, J. P., 1978, Ann. Parasitol., v. 53 (5), 447-459
as syn. of: Nycteria krampitzi n. sp.

epatocystis sp.
Stafford, E. E.; Galdikas-Brindamour, B.; and Beaudoin, R. L., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (1), 106-107 [Letter]
Pongo pygmaeus (blood): Southern Kalimantan (Indonesian Borneo)

patocystis sp.
Yap, L. F.; et al., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (3), 345-353
Ratufa bicolor: Jenderak Utara, Malaysia

Hepatocystis fieldi sp. n., illus.
Garnham, P. C. C.; and Edeson, J. F. B., 1962, Riv. Malariol., v. 41 (1-3), 1-8
Tragulus javanicus (liver, erythrocytes of peripheral blood): Malaya

Hepatocystis oriheli sp. nov., illus.
Garnham, P. C. C., 1977, Protozoology, v. 3, 135-142
Miopithecus talapoin (blood, liver): Rio Muni, West Africa, imported to United States of America

Hepatocystis perronae n. sp., illus.
Landau, I.; and Adam, J. P., 1971, Cahiers O. R. S. T. O. M., s. Entom. Med., v. 9 (4), 373-378 [Correction for 1971 a as published in Supplement 20]
[lapsus p. 373 as H. perronnae]
Myonycteris torquata (liver)
Lissonycteris angolensis (liver)
all from Congo-Brazzaville

Hepatocystis perronae
Landau, I., 1973, Mem. Mus. National Hist. Nat., Paris, n. s., s. A, Zool., v. 77, 1-62
Coccidiomorpha, life cycles, mechanisms assuring continuation of transmission, thesis
Myonycteris torquata: Maboke, Republique Centrafricaine

Hepatocystis perronnae n. sp. [lapsus p. 373 for H. perronae n. sp.]
Landau, I.; and Adam, J. P., 1971, Cahiers O. R. S. T. O. M., s. Entom. Med., v. 9 (4), 373-378 [Correction for 1971 a as published in Supplement 20]

Hepatocystis vassali yokogawai Wu
Garnham, P. C. C., 1977, Protozoology, v. 3, 135-142
as syn. of Hepatocystis yokogawai (Wu) [n. rank]

Hepatocystis yokogawai (Wu) [n. rank]
Garnham, P. C. C., 1977, Protozoology, v. 3, 135-142
Syn.: Hepatocystis vassali yokogawai Wu

Hepatozoon
Landau, I., 1973, Mem. Mus. National Hist. Nat., Paris, n. s., s. A, Zool., v. 77, 1-62
Coccidiomorpha, life cycles, mechanisms assuring continuation of transmission, thesis

Hepatozoon-like hemogregarines, illus.
Ayala, S. C., 1975, Rev. Inst. Med. Trop. S. Paulo, v. 17 (4), 218-224
Anolis pinchoti
Ameiva ameiva
Cnemidophorus lemniscatus
Mabuya mabuya
Aristelliger georgeensis
(blood of all): all from Providencia Island, western Caribbean

Hepatozoon [sp.]
Baker, J. R., 1974, J. Zool., London, v. 172 (2), 169-190
Apodemus sylvaticus
Clethrionomys glareolus
Microtus agrestis
Rattus norvegicus
R. rattus
Sciurus carolinensis
S. vulgaris
all from Britain

Hepatozoon [sp.], presumably H. griseisciuri
Britt, D.; and Molyneux, D. H., 1979, J. Parasitol., v. 65 (3), 408
Sciurus carolinensis (blood): Delamere Forest, Cheshire, England

Hepatozoon sp.
Carpenter, J. W.; and Hillman, C. N., 1979, Ann. Proc., Am. Ass. Zoo Vet., 36-47
Hepatozoon sp., Encephalitozoon sp., mixed infection in Siberian ferret kits
Mustela eversmanni: Patuxent Wildlife Research Center, Laurel, Maryland

Hepatozoon sp.
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Sorex araneus (Milz)
Microtus oeconomus (Herz, Leber, Milz)
all from Neusiedlerseegebiet, nordlichen Burgenland

Hepatozoon sp., illus.
Klopfer, U.; Nobel, T. A.; and Neumann, F., 1973, Vet. Path., v. 10 (3), 185-190
Hepatozoon sp., cats, prevalence of schizonts in myocardium: Israel

Hepatozoon sp.
Lainson, R., 1977, Protozoology, v. 3, 87-93
Caiman crocodilus crocodilus (blood): Barcarena, north Para State, Brazil

Hepatozoon sp., illus.
Pessoa, S. B.; and de Biasi, P., 1974, Rev. Patol. Trop., v. 3 (2), 221-224
Leimadophis poecilogyrus (blood): Santa Catarina
Haementeria gracilis (exper.)

Hepatozoon-type hemogregarine
Telford, S. R., jr., [1979], J. Parasitol., v. 64 (6), 1978, 1126-1127
Sceloporus undulatus
Eumeces laticeps
E. inexpectatus
all from Florida

Hepatozoon alactagae Zasukhin, 1936, illus.
Berdyev, A. S., 1977, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (1), 74-79
Alactaga elater
Alactagulus acontion
all from southern Turkmenistan

Hepatozoon canis, illus.
Craig, T. M.; et al., 1978, J. Am. Vet. Med. Ass., v. 173 (8), 967-972
Hepatozoon canis, dogs (blood), clinical, radiographic, and hematologic findings: Texas

Hepatozoon domerguei Landau, Chabaud, Michel et Brygoo 1970, illus.
Landau, I., 1973, Mem. Mus. National Hist. Nat., Paris, n. s., s. A, Zool., v. 77, 1-62
Coccidiomorpha, life cycles, mechanisms assuring continuation of transmission, thesis
Madagascarophis colubrina (nat. and exper.)
Culex pipiens fatigans (exper.)
C. p. pipiens (exper.)
Anopheles stephensi (exper.)
Lioheterodon modestus (exper.)
Python sebae (exper.)
Oplurus sebae (exper.)

Hepatozoon erhardovae Krampitz 1964
Frank, C., 1977, Ztschr. Parasitenk., v. 53 (3), 251-254
Hepatozoon erhardovae, incidence in Clethrionomys glareolus (blood), distribution of schizonts in lungs; accumulation of gametocytes in Ixodes ricinus (nat. and exper.) or Neotrombicula zachvatkini (nat. and exper.) after biting heavily infected C. glareolus; occasionally transmitted to non-specific host, Apodemus flavicollis (exper.), but only by specific vector, Megabothris turbidus (exper.): South-Western Styria and areas around Neusiedlersee

Hepatozoon erhardovae, illus.
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Clethrionomys glareolus (Herz, Lunge, Leber, Milz): Neusiedlerseegebiet, nordlichen Burgenland

Hepatozoon gerbilli Christophers, 1905, illus.
Berdyev, A. S., 1977, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (1), 74-79
Rhombomys opimus: southern Turkmenistan

Hepatozoon lavieri
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Microtus arvalis (Leber): Neusiedlerseegebiet, nordlichen Burgenland

Hepatozoon microti Coles, 1914
Butabaeva, M., 1979, Uzbek. Biol. Zhurnal (2), 50-53
[Allactaga severtzovi]
[Allactaga elater]
all from Kashkadar'insk oblast

Hepatozoon microti
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Microtus agrestis (Leber): Neusiedlerseegebiet, nordlichen Burgenland

Hepatozoon muris (Balfour, 1905), illus.
Lage, H. A.; and Lage, S. A., 1971, Atas Soc. Biol. Rio de Janeiro, v. 14 (5-6), 109-111
incidence, description
Rattus norvegicus (figado, pulmao)
R. frugivorus
all from Rio de Janeiro, Guanabara

Hepatozoon procyonis, illus.
Schaffer, G. D.; et al., 1978, J. Am. Vet. Med. Ass., v. 173 (9), 1148-1151
hematotropic parasites of Procyon lotor, carrier potential as related to translocation and release for hunting purposes, practice is considered biologically hazardous: southeastern United States
Procyon lotor: Glades County, Florida; Liberty County, Georgia; Brown County, Texas

patozoon procyonis Richards, 1961, illus.
Schneider, C. R., 1967, Rev. Biol. Trop., v. 15
(1), 123-135
supplemented description
Procyon cancrivorus panamensis (myocardium,
blood): near town of Pacora, east of Panama
City

patozoon prodhoni n. sp., illus.
Landau, I., 1973, Mem. Mus. National Hist.
Nat., Paris, n. s., s. A, Zool., v. 77, 1-62
Coccidiomorpha, life cycles, mechanisms
assuring continuation of transmission,
thesis
Oplurus quadrimaculatus (sang): Baie de
Loukaro

patozoon sylvatici Coles 1914
Frank, C., 1977, Ztschr. Parasitenk., v. 53
(3), 307-310
Hepatozoon sylvatici transmitted from naturally to experimentally infected Apodemus
sylvaticus and A. flavicollis by Laelaps
agilis; L. agilis transmission of H. sylvatici to non-specific host, Clethrionomys
glareolus; gonads of female mites possibly
have stimulating effect on protozoan development; schizonts from bone marrow and liver
of Apodemus flavicollis differ in morphology

atozoon sylvatici, illus.
Frank, C., 1978, Ang. Parasitol., v. 19 (3),
137-154
Apodemus flavicollis (Herz, Lunge)
A. sylvaticus (Blut, Herz, Lunge, Milz)
all from Neusiedlerseegebiet, nordlichen
Burgenland

rpetomonas sp.
Camargo, E. P.; Itow, S.; and Alfieri, S. C.,
[1979], J. Parasitol., v. 64 (6), 1978, 1120-
1121
Trypanosomatidae, 18 spp. of 6 genera, proteolytic activities in cell extracts

petomonas-like
sh, W. R.; Holz, G. G., jr.; and Beach, D.
H., 1978, J. Parasitol., v. 64 (3), 546-547
26 trypanosomatid species, cultivation in new
chemically-defined medium RE III

petomonas sp.
alinari, S.; and Camargo, E. P., 1978, Exper.
arasitol., v. 46 (2), 277-282
trypanosomatid protozoa, 16 spp., survey for
acetylornithinase and ornithine acetyltransferase, metabolic and nutritional implications

petomonas sp.
endes, N. F.; et al., 1979, Transplant. Proc.,
. 11 (2), 1304-1305
cross-reactions between Trypanosomatidae
cell extracts and HLA antigens

petomonas, sp., illus.
einman, D.; and Cheong, W. H., 1978, J. Proto-
ool., v. 25 (2), 167-169
Herpetomonas sp. infections in laboratory-reared Aedes aegypti and A. albopictus (Malphighian tubules of both), 30 to 40% of the
flagellates contained intracytoplasmic rod-shaped structures strongly resembling bacteria: Institute for Medical Research, Kuala
Lumpur, Malaysia

Herpetomonas sp.
Yoshida, N.; et al., 1978, J. Protozool., v. 25
(4), 550-555
Herpetomonas spp., enzymes of ornithine-arginine metabolism

Herpetomonas [sp.]
Yoshida, N.; Freymueller, E.; and Wallace,
F. G., 1978, J. Protozool., v. 25 (4), 421-425
Musca (intestine)
Sarcophaga (intestine)
all from Sao Paulo, State of Sao Paulo,
Brazil

Herpetomonas ampelophilae, illus.
Rowton, E. D.; and McGhee, R. B., 1978, J.
Protozool., v. 25 (2), 232-235
Herpetomonas in Drosophila melanogaster, introduction of 10 infected flies into stabilized population of 10,000 flies resulted in
increase of infection from 0.01% to 90% in
14 days, infection fluctuated between 5% and
100% during 250 days, parasites were found
predominantly endotrophically but also peritrophically and in Malpighian tubules, it
is suggested that all Herpetomonas in D.
melanogaster be referred to as H. ampelophilae

Herpetomonas donovani (Laveran & Mesnil, 1903)
Mesnil, 1906
Gardener, P. J., 1977, Trop. Dis. Bull., v. 74
(12), 1069-1088
as syn. of Leishmania donovani (Laveran &
Mesnil, 1903) Ross, 1903

Herpetomonas farunculosa (Firth, 1891) Patton,
1922
Gardener, P. J., 1977, Trop. Dis. Bull., v. 74
(12), 1069-1088
as syn. of Leishmania tropica (Wright, 1903)
Luehe, 1906

Herpetomonas infantum (Nicolle, 1908) Patton,
1909
Gardener, P. J., 1977, Trop. Dis. Bull., v. 74
(12), 1069-1088
as syn. of Leishmania donovani (Laveran &
Mesnil, 1903) Ross, 1903

Herpetomonas mariadeanei sp. n. [? nomen nudum]
Yoshida, N.; Freymuller, E.; and Wallace, F.
G., 1978, Rev. Inst. Med. Trop. S. Paulo, v.
20 (4), 229 [Abstract]
Muscina stabulans

Herpetomonas mariadeanei sp. n., illus.
Yoshida, N.; Freymueller, E.; and Wallace,
F. G., 1978, J. Protozool., v. 25 (4), 421-425
Muscina stabulans (intestine): Sao Paulo,
State of Sao Paulo, Brazil

Herpetomonas megaseliae
Camargo, E. P.; Itow, S.; and Alfieri, S. C.,
[1979], J. Parasitol., v. 64 (6), 1978, 1120-
1121
Trypanosomatidae, 18 spp. of 6 genera, proteolytic activities in cell extracts

Herpetomonas megaseliae
Daggett, P. M.; Decker, J. E.; and Janovy, J.,
1978, Comp. Biochem. and Physiol., v. 59A (4),
363-366
6 insect trypanosomatids, attempted adaptation to mice, alteration of component elements of excreted factors (EF) produced by
the 3 adapted species, EF components of
Leishmania donovani isolates differ with
varying infectivity for hamsters

Herpetomonas megaseliae
Fish, W. R.; Holz, G. G., jr.; and Beach, D. H., 1978, J. Parasitol., v. 64 (3), 546-547
26 trypanosomatid species, cultivation in new chemically-defined medium RE III

Herpetomonas megaseliae
Galinari, S.; and Camargo, E. P., 1978, Exper. Parasitol., v. 46 (2), 277-282
trypanosomatid protozoa, 16 spp., survey for acetylornithinase and ornithine acetyltransferase, metabolic and nutritional implications

Herpetomonas megaseliae
Mendes, N. F.; et al., 1979, Transplant. Proc., v. 11 (2), 1304-1305
cross-reactions between Trypanosomatidae cell extracts and HLA antigens

Herpetomonas megaseliae
Yoshida, N.; et al., 1978, J. Protozool., v. 25 (4), 550-555
Herpetomonas spp., enzymes of ornithine-arginine metabolism

Herpetomonas megaseliae
Yoshida, N.; and Camargo, E. P., 1978, J. Bacteriol., v. 136 (3), 1184-1186
trypanosomatids, excretion of urea or ammonia or both, varies according to genus, may be of taxonomic use

Herpetomonas muscarum (Leidy, 1856) Kent, 1880, illus.
Carvalho, A. L. de M., 1973, Rev. Patol. Trop., v. 2 (2), 223-274
Zelus leucogrammus (tubo digestivo)
Musca domestica
all from Brasil

Herpetomonas muscarum
Daggett, P. M.; Decker, J. E.; and Janovy, J., 1978, Comp. Biochem. and Physiol., v. 59A (4), 363-366
6 insect trypanosomatids, attempted adaptation to mice, alteration of component elements of excreted factors (EF) produced by the 3 adapted species, EF components of Leishmania donovani isolates differ with varying infectivity for hamsters

Herpetomonas muscarum
Yoshida, N.; and Camargo, E. P., 1978, J. Bacteriol., v. 136 (3), 1184-1186
trypanosomatids, excretion of urea or ammonia or both, varies according to genus, may be of taxonomic use

Herpetomonas muscarum ingenoplastis
Yoshida, N.; et al., 1978, J. Protozool., v. 25 (4), 550-555
Herpetomonas spp., enzymes of ornithine-arginine metabolism

Herpetomonas muscarum muscarum
Camargo, E. P.; Itow, S.; and Alfieri, S. C., [1979], J. Parasitol., v. 64 (6), 1978, 1120-1121
Trypanosomatidae, 18 spp. of 6 genera, proteolytic activities in cell extracts

Herpetomonas muscarum muscarum
Fish, W. R.; Holz, G. G., jr.; and Beach, D. H., 1978, J. Parasitol., v. 64 (3), 546-547
26 trypanosomatid species, cultivation in n chemically-defined medium RE III

Herpetomonas muscarum muscarum
Galinari, S.; and Camargo, E. P., 1978, Exper Parasitol., v. 46 (2), 277-282
trypanosomatid protozoa, 16 spp., survey fo acetylornithinase and ornithine acetyltransferase, metabolic and nutritional implications

Herpetomonas muscarum muscarum
Yoshida, N.; et al., 1978, J. Protozool., v. (4), 550-555
Herpetomonas spp., enzymes of ornithine-arg nine metabolism

Herpetomonas myoxi Laveran & Franchini, 1921 = Trypanosoma myoxi Blanchard, 1904
Gardener, P. J., 1977, Trop. Dis. Bull., v. 7 (12), 1069-1088

Herpetomonas pessoai (Galvao, Oliveira, Carvalho and Veiga 1970) Roitman, Brener, Roitman, and Kitajima 1976 [i.e., n. comb.]
Levine, N. D., 1978, J. Parasitol., v. 64 (4), 668
"correct name"
Syns.: Leptomonas pessoai; Herpetomonas samuelpessoai

Herpetomonas pessoai [(Galvao et al., 1970) Levine, 1978]
Levine, N. D., 1979, J. Parasitol., v. 65 (1), 116
preoccupied by H. pessoai Ayroza and Coutinho, 1941, "the name Herpetomonas samuelpessoai Roitman, Brener, Roitman, and Kitajima 1976 is the correct one for the species from Zelus."

Herpetomonas samuelpessoai, illus.
de Almeida, D. F.; and de Souza, W., 1978, J. Parasitol., v. 64 (1), 17-22
Herpetomonas samuelpessoai in basic medium or in complex media obtained through addition of various concentrations of KCl, NaCl, or glycerol, growth curves, changes in cell volume as function of osmolarity of medium, corresponding morphological changes

Herpetomonas samuelpessoai, illus.
Bunn, M. M.; et al., 1977, Ztschr. Parasitenk., v. 52 (3), 245-256
Herpetomonas samuelpessoai in vitro, 2-deoxy-D-glucose (2-DG) inhibits growth and respiration, modifies ultrastructure of cells; some carbohydrates decrease effect of 2-DG

Herpetomonas samuelpessoai
Camargo, E. P.; Itow, S.; and Alfieri, S. C., [1979], J. Parasitol., v. 64 (6), 1978, 1120-1121
Trypanosomatidae, 18 spp. of 6 genera, proteolytic activities in cell extracts

rpetomonas samuelpessoai, illus.
de Carvalho, T. U.; Souto-Padron, T.; and de Souza, W., 1979, Exper. Parasitol., v. 47 (3), 297-304
Herpetomonas samuelpessoai, electron-dense granules, electron microscopy and cytochemistry (peroxidase activity, basic proteins, acid phosphatase)

rpetomonas samuelpessoai
Daggett, P. M.; Decker, J. E.; and Janovy, J., 1978, Comp. Biochem. and Physiol., v. 59A (4), 363-366
6 insect trypanosomatids, attempted adaptation to mice, alteration of component elements of excreted factors (EF) produced by the 3 adapted species, EF components of Leishmania donovani isolates differ with varying infectivity for hamsters

rpetomonas samuelpessoai, illus.
Esteves, M. J. G.; et al., 1979, European J. Cell Biol., v. 20 (1), 113-115
Herpetomonas samuelpessoai, membrane-associated polysaccharides, influence of growth conditions

rpetomonas samuelpessoai
Fish, W. R.; Holz, G. G., jr.; and Beach, D. H., 1978, J. Parasitol., v. 64 (3), 546-547
26 trypanosomatid species, cultivation in new chemically-defined medium RE III

rpetomonas samuelpessoai
Galinari, S.; and Camargo, E. P., 1978, Exper. Parasitol., v. 46 (2), 277-282
trypanosomatid protozoa, 16 spp., survey for acetylornithinase and ornithine acetyltransferase, metabolic and nutritional implications

rpetomonas samuelpessoai
Goncalves de Lima, V. M. Q.; Roitman, I.; and ilgour, V., 1979, J. Protozool., v. 26 (4), 648-652
trypanosomatids, 7 species distinguished by electrophoretic mobilities of some isoenzymes

rpetomonas samuelpessoai Roitman et al. 1976
Levine, N. D., 1978, J. Parasitol., v. 64 (4), 658
as syn. of H. pessoai (Galvao, Oliveira, Carvalho, and Veiga 1970) Roitman, Brener, Roitman, and Kitajima 1976 [i.e., n. comb.]

petomonas samuelpessoai
Levine, N. D., 1979, J. Parasitol., v. 65 (1), 116
H. pessoai [(Galvao et al., 1970) Levine, 1978] is preoccupied by H. pessoai Ayroza and Coutinho, 1941, "the name Herpetomonas samuelpessoai Roitman, Brener, Roitman, and Kitajima 1976 is the correct one for the species from Zelus."

petomonas samuelpessoai
Mendonca-Previato, L.; Gorin, P. A. J.; and Previato, J. O., 1979, Biochemistry, Washington, v. 18 (1), 149-154
Herpetomonas samuelpessoai, polysaccharide components of cells grown on various culture media, possible role of certain polysaccharides in immunogenic terms

Herpetomonas samuelpessoai, illus.
Soares, T. C. B.; and de Souza, W., 1977, Ztschr. Parasitenk., v. 53 (2), 149-154
Trypanosoma cruzi, Herpetomonas samuelpessoai, fixation with glutaraldehyde-tannic acid for electron microscopy; structural details

Herpetomonas samuelpessoai, illus.
Souto-Padron, T.; and de Souza, W., 1979, J. Protozool., v. 26 (4), 551-557
trypanosomatids, various developmental stages, basic proteins, localization at fine-structural level with ethanolic phosphotungstic acid technique

Herpetomonas samuelpessoai, illus.
de Souza, W.; Chavez, B.; and Martinez-Palomo, A., 1979, J. Parasitol., v. 65 (1), 109-116
Herpetomonas samuelpessoai, cell membrane, freeze-fracture study

Herpetomonas samuelpessoai
Yoshida, N.; et al., 1978, J. Protozool., v. 25 (4), 550-555
Herpetomonas spp., enzymes of ornithine-arginine metabolism

Herpetomonas samuelpessoai
Yoshida, N.; and Camargo, E. P., 1978, J. Bacteriol., v. 136 (3), 1184-1186
trypanosomatids, excretion of urea or ammonia or both, varies according to genus, may be of taxonomic use

Herpetomonas tropica (Wright, 1903) Patton, 1909
Gardener, P. J., 1977, Trop. Dis. Bull., v. 74 (12), 1069-1088
as syn. of Leishmania tropica (Wright, 1903) Luehe, 1906

Herpetophrya Siedlecki
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key

Herpetophryinae
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key

Herpetosoma, subgenus
Godfrey, D. G., 1977, Protozoology, v. 3, 33-49
identification of morphologically similar trypanosomes of mammals

Herpetosoma
Molyneux, D. H., 1970, Ann. Soc. Belges Med. Trop. Parasitol., v. 50 (2), 229-237
Trypanosoma (Herpetosoma) spp., reproduction in vertebrate host, course of infection, transmission and development in fleas, review

Heteromita pusilla Perty, 1852
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
as syn. of Hexamita intestinalis Dujardin, 1841

Hexamastix polyphagae n. sp., illus.
Sultana, T., 1976, Riv. Parassitol., Roma, v. 37 (1), 1-5
Polyphaga indica (hindgut): Aurangabad, Maharashtra, India

Hexamita intestinalis
Ollenschlaeger, B., 1975, Tieraerztl. Prax., v. 3 (1), 99-107
blood and other parasites of commercial fish, pathology, transmission, therapy, clinical review

Hexamita intestinalis Dujardin, 1841, illus.
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
synonymy, description
Bombina bombina
B. variegata
Bufo bufo
B. viridis
B. calamita
Pelobates fuscus
Rana esculenta
R. temporaria
R. dalmatina
all from CSSR

Hexamita muris, illus.
Flatt, R. E.; Halvorsen, J. A.; and Kemp, R. L., 1978, Lab. Animal Sc., v. 28 (1), 62-65
Hexamita muris in laboratory mice (small intestine, between epithelial cells, lamina propria), case history, dimetridazole controlled clinical disease but did not eliminate the infection

Hexamita muris
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Rattus norvegicus (Jejunum)
Mus musculus (Jejunum)
Apodemus sylvaticus (Jejunum, Ileum, Rectum)
Ondatra zibethica (Duodenum, Jejunum)
Microtus agrestis (Jejunum)
all from Neusiedlerseegebiet, nordlichen Burgenland

Hexamita muris
MacDonald, T. T.; and Ferguson, A., 1978, Gastroenterology, v. 74 (3), 496-500
Giardia muris, Hexamita muris, mice (exper.), effects of chronic infection on small intestinal epithelial cell kinetics

Hexamita muris, illus.
Owen, R. L.; Nemanic, P. C.; and Stevens, D. P., 1979, Gastroenterology, v. 76 (4), 757-769
Giardia muris in immunocompetent mice, intestinal distribution of trophozoites, attachment and relationships to intestinal mucosa (particularly Peyer's patches), normal reaction of intestine and intestinal immune organs; includes some incidental observations on Hexamita muris

Hexamita muris, illus.
Van Kruiningen, H. J.; Knibbs, D. R.; and Burke, C. N., 1978, J. Am. Vet. Med. Ass., v. 173 (9), 1202-1204
Hexamita muris in laboratory mice (intestines), disease picture somewhat different than previously reported

Hexamitus murthii n. sp., illus.
Sultana, T., 1976, Riv. Parassitol., Roma, v. 37 (1), 7-11
Periplaneta americana (hindgut): Aurangabad, Maharashtra, India

Hexamita salmonis (Moore, 1923)
Arthur, J. R.; Margolis, L.; and Arai, H. P. 1976, J. Fish. Research Bd. Canada, v. 33 (1
2489-2499
Coregonus clupeaformis
Prosopium cylindraceum
Salvelinus namaycush
Thymallus arcticus
Lota lota
(gall bladder of all): all from Aishihik Lake, Yukon Territory

Hexamita salmonis (Moore)
Lester, R. J. G., 1974, Syesis, v. 7, 195-200
Gasterosteus aculeatus (intestine): near Vancouver, British Columbia

Hexamita salmonis
Ollenschlaeger, B., 1975, Tieraerztl. Prax., v. 3 (1), 99-107
blood and other parasites of commercial fis
pathology, transmission, therapy, clinical review

Hexamitus intestinalis Buetschli (1878)
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-
209
as syn. of Hexamita intestinalis Dujardin, 1841

Hexamitus intestinalis (Dujardin, 1841)
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-
209
as syn. of Hexamita intestinalis Dujardin, 1841

Hirmocystidae Grasse, 1953
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
diagnosis, includes: Arachnocystis gen. n.

Hirmocystis bengalensis n. sp., illus.
Haldar, D. P.; and Chakraborty, N., 1979, Ztschr. Parasitenk., v. 59 (2), 121-130
Hirmocystis bengalensis n. sp., life history
Myllocerus sp. 1 (midgut): Kalyani, West Bengal, India

Hirmocystis pitcharis n. sp., illus.
Haldar, D. P.; and Chakraborty, N., 1979, Ztschr. Parasitenk., v. 59 (2), 121-130
Hirmocystis pitcharis n. sp., life history
Xanthoprochilus sp. (midgut): Kalyani, West Bengal, India

Hirmocystis pseudoductis n. sp., illus.
Haldar, D. P.; and Chakraborty, N., 1979, Ztschr. Parasitenk., v. 59 (2), 121-130
Hirmocystis pseudoductis n. sp., life histor
Myllocerus sp. 2 (midgut): Kalyani, West Bengal, India

Hirmocystis ventricosa
Sherlock, P. L., 1979, Parasitology, v. 78 (2)
207-220
Tipula paludosa (intestine)

Histomonas meleagridis
Augustine, P. C.; and Chute, A. M., 1978, J. Parasitol., v. 64 (3), 425
Histomonas meleagridis, in vitro-adapted strain, inhibition of growth in conventional or gnotobiotic turkeys inoculated with 5 spp. of bacteria used for in vitro cultivation, supports hypothesis that in vitro culturing decreased in vivo reproductive capability by gradual elimination of more virulent organism

stomonas meleagridis
Chute, M. B.; Chute, A. M.; and Wilkins, G. C., 1978, Parasitology, v. 77 (1), 41-48
Histomonas meleagridis, therapeutic treatment of chickens with dimetridazole may reduce but not eliminate transmission of this protozoan by eggs of Heterakis gallinarum from medicated birds

stomonas meleagridis
Chute, A. M.; Lund, E. E.; and Wilkins, G. C., 1977, Proc. Helminth. Soc. Washington, v. 44 (2), 126-130
Histomonas meleagridis, Heterakis gallinarum, experimental infections in Indian Red Jungle Fowl and Light Brahma chickens compared with pheasants, turkeys, and New Hampshire chickens, susceptibility and potential for parasite dissemination
Gallus gallus murghi
Phasianus colchicus
Beltsville Small White turkeys
Gallus gallus
(all exper.)

istomonas] meleagridis, illus.
Damodaran, S.; and Viswanathan, S., 1978, Indian Vet. J., v. 55 (11), 922
Pavo cristata (liver): Zoological Gardens, Madras

stomonas meleagridis
Davidson, W. R.; Doster, G. L.; and McGhee, M. B., 1978, Avian Dis., v. 22 (4), 627-632
Heterakis bonasae, failure to transmit Histomonas meleagridis from infected bobwhite quails to turkeys

stomonas meleagridis
Davies, D. H.; et al., 1978, J. Med. Chem., v. 21 (2), 189-193
Trichomonas vaginalis, Entamoeba histolytica, 6'-hydroxy analogue of sisomicin, antiprotozoal activity demonstrated in laboratory trials; no activity against Histomonas meleagridis and no anthelmintic activity observed

tomonas meleagridis
irsch, R. P., 1979, Internat. J. Parasitol., . 9 (5), 395-399
Histomonas meleagridis-infected turkeys, dynamics of protozoan population density, plasma glutamic oxalacetic transaminase, plasma bilirubin concentration, relationship to clinical symptoms

tomonas meleagridis
Kellogg, F. E.; Doster, G. L.; and Johnson, . K., 1971, J. Wildlife Dis., v. 7 (3), 186-187
Indian red junglefowl, pen-raised (ceca, liver)

stomonas meleagridis
Kocan, A. A.; Hannon, L.; and Eve, J. H., 1979, Proc. Oklahoma Acad. Sc., v. 59, 20-22
Colinus virginianus (intestine): Oklahoma

tomonas meleagridis
McDougald, L. R., 1979, Poultry Science, . 58 (1), 76-80
coccidiosis, histomoniasis, turkeys, tests for efficacy and compatibility indicate that amprolium and carbarsone can be used in combination

Histomonas meleagridis
Manuel, M. F.; and Guerzon, V. A., 1978, Philippine J. Vet. Med., v. 17 (1-2), 121-128
Histomonas meleagridis, absence in ceca of Coturnix coturnix japonicum; incidental finding of developmental stages of Eimeria sp.: Philippines

Histomonas meleagridis
Ridala, V., 1977, Eesti Pollumaj. Akad. Teadusl. Toode Kogum. (104), 109-113
Histomonas meleagridis, turkeys, histopathology

Histomonas meleagridis
Schildknecht, E. G.; and Squibb, R. L., 1979, Parasitology, v. 78 (1), 19-31
Histomonas meleagridis in turkeys, effects of vitamins A, E, and K (alone and in combination with ipronidazole) on performance and on plasma enzymes, plasma enzyme levels correlated well with progressive pathological changes

Histomoniasis
Parre, J.; and Simovart, H. E., 1977, Eesti Pollumaj. Akad. Teadusl. Toode Kogum. (104), 114-118
histomoniasis, turkeys, age and seasonal dynamics in relation to epizootiology; disease outbreaks in young birds under stress conditions; nitazol satisfactory, trichopol good prophylactic and therapeutic effect

Hoplitophrya Stein
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key

Hoplitophryidae
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key

Hoplitophryinae
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key

Huffia
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
avian Plasmodium spp. of 4 subgenera, list of type material in Garnham Collection of Wellcome Museum of Medical Science; neotypes designated

Hyalinocysta gen. n.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Thelohaniidae fam. n., key
tod: H. chapmani sp. n.

Hyalinocysta chapmani sp. n. (tod), illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Culiseta melanura: near Kinder, Lousiana, U.S.A.

Hypotrichomonas acosta (Moskowitz), illus.
Warton, A.; and Honigberg, B. M., 1979, J. Protozool., v. 26 (1), 56-62
Hypotrichomonas acosta, Trichomonas vaginalis, Pentatrichomonas hominis, Tritrichomonas foetus, scanning electron microscopy

Ichtyobodo necator, illus.
Ellis, A. E.; and Wootten, R., 1978, J. Fish Dis., v. 1 (4), 389-393
Ichtyobodo necator in Salmo salar (gills), high mortality following transfer to seawater cages probably due to prior infestation in freshwater, pathology: farm in Scotland

Ichtyobodo necator (= Costia necatrix)
Robertson, D. A., 1979, J. Fish Dis., v. 2 (6), 481-491
Ichtyobodo necator on farmed salmonids, prevalence and intensity in relation to time, temperature, and host age; suggested that some form of host defense mechanism is operating
Salmo salar (gills)
S. gairdneri (gills, skin)
all from central Scotland

Ichthyophthiriasis
Bachinskii, V. P., 1969, Rybn. Khoziaist., Kiev (8), 104-106
parasites as possible causes of epizootics in fish: Kremenchugsk reservoir

Ichthyophthirius [sp.]
Voelker, F. A.; et al., 1978, Vet. Path., v. 15 (1), 40-48
Chaetodon (surface of integument)

Ichthyophthirius multifiliis Fouquet, 1876, illus.
Abe, I., 1978, Nippon Zyuisi-Kai Zassi (J. Japan Vet. Med. Ass.), v. 31 (6), 360-364
white-spot disease, reared fish, treatment and control

I[chthyophthirius] multifiliis
Abe, I., 1978, Nippon Zyuisi-Kai Zassi (J. Japan Vet. Med. Ass.), v. 31 (7), 417-420
white-spot disease, cultured fish, treatment and control

Ichthyophthirius multifiliis Fouquet
Armas, G., 1979, J. Fish Dis., v. 2 (6), 543-547
Mugil cephalus (body surface, gills): Rio Moche coastal lagoon, northern Peru

Ichthyophthirius multifiliis
Astakhova, T. V.; and Stepanova, G. A., 1972, Parazitologiia, Leningrad, v. 6 (4), 364-368
Ctenopharyngodon idella (skin, gills): pond and spawning-nursery fisheries, Volga delta

Ichthyophthirius multifiliis, illus.
Bremer, H., 1972, Ang. Parasitol., v. 13 (4), 247-248
Ichthyophthirius multifiliis, cell division

Ichthyophthirius multifiliis
Jackson, P. B. N., 1978, J. South African Vet. Ass., v. 49 (1), 57-59
parasites of Anguilla mossambica, importance of disease control in aquaculture
Anguilla mossambica (skin, gills): South Africa

Ichthyophthirius multifiliis Fouquet
Lester, R. J. G., 1974, Syesis, v. 7, 195-200
Gasterosteus aculeatus (between epidermis and dermis): near Vancouver, British Columbia

Ichthyophthirius multifiliis, illus.
Nigrelli, R. F.; Pokorny, K. S.; and Ruggieri, G. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 607-613
Ichthyophthirius multifiliis and other 'Ich-like' parasites of freshwater fishes appear to be limited in distribution and infectivity by temperature tolerances of their hosts, possible existence of multiple physiological races or even species possibly in more than one genus

Ichthyophthirius multifiliis
Vladimirov, V. L.; and Flerov, B. A., 1975, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (25), 35-37
Ichthyophthirius multifiliis, fish more susceptible when exposed to phenol or polychlorpinene (toxic water pollutants)

Ichthyophthirius multifiliis (Foquet, 1876)
Vlasenko, M. I.; and Meshcheriakova, A. A., 1977, Veterinariia, Moskva (4), 75-78
Chilodonella cyprini, Ichthyophthirius multifiliis, [Costia], fish, formalin solution for control, toxicity tested and safe levels established

Ileocystis
Barker, I. K.; Munday, B. L.; and Presidente, P. J. A., 1979, J. Parasitol., v. 65 (3), 451-456
as syn. of Eimeria

Ileocystis wombati Gilruth and Bull, 1912
Barker, I. K.; Munday, B. L.; and Presidente, P. J. A., 1979, J. Parasitol., v. 65 (3), 451-456
as syn. of Eimeria wombati (Gilruth and Bull, 1912) comb. nov.

Inodosporus Overstreet and Weidner, 1974
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Thelohaniidae fam. n., key

Inodosporus octosporus (Henneguy, 1892) Overstreet and Weidner, 1974
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
synonymy

Iodamoeba
Dutta, G. P., 1977, J. Scient. and Indust. Research, India, v. 36 (5), 226-240
Entamoeba histolytica and other anaerobic amoebae, cytoplasmic inclusions, recent advances in cytochemistry and ultrastructure, role in physiology and pathogenesis, review

Iodamoeba sp.
Marzochi, M. C. de A., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (4), 249-256
human intestinal parasites, eggs and cysts from water used to irrigate vegetable gardens, increased risk of crop contamination in dry season: Ribeirao Preto, Sao Paulo, Brazil

Iodamoeba sp.
Marzochi, M. C. de A., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (3), 148-155
enteroparasitic cysts and eggs, contamination of green vegetables and kitchen garden soils, epidemiological survey, most commonly found during dry-season when fecal polluted brooks were used for irrigation: Ribeirao Preto, Sao Paulo, Brasil

damoeba buetschlii
Akhtaruzzaman, K. M.; et al., 1978, Tropenmed. u. Parasitol., v. 29 (4), 427-431
comparison of different methods for detection of intestinal protozoa and helminths in human stool

damoeba butschlii
Bartlett, M. S.; et al., 1978, J. Clin. Microbiol., v. 7 (6), 524-528
modified zinc sulfate flotation technique evaluated in comparison with formalin-ether concentration method for recovery of protozoan cysts and helminth eggs and larvae from feces preserved in formalin less than and longer than 1 month, results suggest that (except for schistosomes) F-$ZnSO_4$ compares favorably to FE method for detecting infections of clinical significance

damoeba buetschlii
lecka, L. J., 1978, J. Parasitol., v. 64 (2), 62-363
patients infected with intestinal parasites, attempt to quantitate immunoglobulin levels in fecal extracts with radial immunodiffusion, mean IgA levels higher than in controls, other immunoglobulin classes rarely detectable

damoeba buetschlii
oldsmid, J. M.; and Rogers, S., 1978, J. South African Vet. Ass., v. 49 (2), 109-111
Papio ursinus (feces): Rhodesia, imported from Northern Transvaal

moeba butschlii
an, B. H.; William, D. C.; and Luminais, S. ., 1979, Brit. J. Vener. Dis., v. 55 (5), 375-78
amoebiasis, giardiasis, Iodamoeba butschlii, epidemic in a homosexual population: New York City

amoeba buetschlii
sricha, A.; and Narang, P., 1978, Indian J. d. Research, v. 67, 934-936
human intestinal parasites, evaluation of kerosene as substitute for ether in the formol-ether concentration diagnostic technique, morphology of ova and cysts equally well preserved by both techniques

moeba buetschlii
geaunt, P. G.; and Williams, J. E., 1979, Roy. Soc. Trop. Med. and Hyg., v. 73 (2), 5-227
pathogenic and non-pathogenic intestinal amoeba of man, all species can easily be distinguished by characteristic electrophoretic isoenzyme patterns

amoeba buetschlii
illiam, D. C.; et al., 1978, Sex. Transmit. is., v. 5 (4), 155-157
enteric protozoa, Enterobius vermicularis, high rate of infection in homosexual men who practice anilingus: New York City

pora
ier, T. V.; Shibalova, T. A.; and Kostenko, A., 1978, [Cytology of coccidia], 185 pp., lus.

Isospora
Collins, J. P.; Keller, K. F.; and Brown, L., 1978, Am. J. Trop. Med. and Hyg., v. 27 (4), 835-836
Giardia lamblia, 'ghost' forms of cysts in human fecal sample initially misdiagnosed as Isospora oocysts

Isospora
Gill, H. S.; et al., 1978, J. Parasitol., v. 64 (3), 549-551
dogs fed Bubalus bubalis diaphragm muscles with visible cysts of Sarcocystis fusiformis shed unsporulated oocysts belonging either to Hammondia or Isospora

Isospora
Heydorn, A. O., 1979, Berl. u. Munchen. Tierarztl. Wchnschr., v. 92 (11), 214-220
coccidia of cats, life cycle, epidemiology, review

Isospora
Rommel, M., 1978, Ztschr. Parasitenk., v. 57 (3), 269-283
cyst-forming coccidia, life cycle, taxonomy, comparative review

Isospora sp.
Byman, D.; et al., 1977, Canad. J. Zool., v. 55 (2), 376-380
Canis lupus (feces): northeastern Minnesota

Isospora [sp.]
Dissanaike, A. S.; and Poopalachelvam, M., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (3), 420
coccidian oocysts from stools of crows examined, "After sporulation, however, it was clear that they were oocysts of Isospora and not Eimeria ..." as reported by Bisseru and Lim in 1971: Klang, Malaysia

Isospora sp.
Farmer, J. N.; et al., 1978, Vet. Rec., v. 102 (4), 78-80
sheepdogs (feces): Gwyned, North Wales
greyhounds (feces): London area
Vulpes vulpes (feces): North Wales

Isospora spp.
Malloy, W. F.; and Embil, J. A., 1978, Canad. J. Comp. Med., v. 42 (1), 29-31
Toxocara canis, T. cati, Toxascaris leonina, and other parasites, prevalence in dogs or cats, percentages by sex and age, survey initiated because of cases of human visceral larva migrans, public health implications
dogs
cats
all from Halifax, Nova Scotia

Isospora [spp.], illus.
Schramlova, J.; and Blazek, K., 1978, Ztschr. Parasitenk., v. 55 (1), 43-48
Isospora [spp.], ultrastructure of cyst wall of two types of sarcocysts, "Entwicklungsstadien wahrscheinlich zweier Kokzidienarten der Gattung Isospora parasitieren."
Capreolus capreolus (Herz-, Skelett-, und Schlundmuskulatur)

Isospora belli, illus.
Bassleer, J.; Nautet, M.; and Dargent, C.,
1973, Rev. Med. Liege, v. 28 (8), 270-272
Isospora hominis, clinical observations on
5 cases, morphologic differentiation from
I. belli: residents of Belgium of various
nationalities

Isospora belli, illus.
Brandborg, L. L., 1979, Am. J. Med., v. 67
(6), 999-1006
diseases of malabsorption, histologic diagnosis

Isospora belli, illus.
Goldsmid, J. M.; Rogers, S.; and Mahomed, K.,
1976, South African Med. J., v. 50 (40), 1547-1550

Isospora belli, illus.
McCracken, A. W., 1972, South. Med. J., v. 65
(7), 800, 818
Isospora belli, natural infection in 8-year-
old Mexican-American girl and in technician
who diagnosed child's infection, case reports:
San Antonio, Texas

Isospora belli, illus.
Ohtaki, M.; et al., 1976, Tohoku J. Exper. Med.,
v. 120 (1), 43-51
human malignant lymphoma initiated with malabsorption
syndrome due to Isospora belli infection
and lymphocytosis, case report

Isospora belli
de Oliveira, G. S. C.; Barbosa, W.; and da
Silva, A. L., 1973, Rev. Patol. Trop., v. 2
(4), 387-395
Isospora hominis, I. belli, humans, epidemiologic
survey, clinical aspects, immunodiagnostic
comparisons with Toxoplasma gondii
(immunofluorescence, protein electrophoresis,
Sabin-Feldman dye test) resulted in frequent
cross-reactions: Goias, Brazil

Isospora belli
Sagua, H.; et al., 1978, Bol. Chileno Parasitol.,
v. 33 (1-2), 8-12
Isospora belli, resulting in human enteritis
outbreak, epidemiologic study, diagnosis by
identification of cysts in stools, verification
that water from industrial sewage treatment
plant was used locally to irrigate vegetables:
Antofagasta City, Chile

Isospora belli, illus.
Westerman, E. L.; and Christensen, R. P., 1979,
Ann. Int. Med., v. 91 (3), 413-414
Isospora belli, immunosuppressed woman concurrently
infected with Giardia lamblia,
severe diarrhea, rapid remission with co-trimoxazole,
case report

Isospora bigemina Stiles, 1891, illus.
Carneiro, J. R.; et al., 1974, Rev. Patol.
Trop., v. 3 (2), 217-219
Canis familiaris (fezes): Goiania-Go, Brasil

Isospora bigemina
Cotteleer, C.; and Fameree, L., 1976, Rev.
Med. Liege, v. 31 (23), 729-734
Eimeriidae, incidence in cats: Belgique

Isospora bigemina
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk.,
v. 58 (2), 115-139
nomen dubium

Isospora bigemina, illus.
Shibalova, T. A.; and Petrenko, V. I., 1972,
Parazitologiia, Leningrad, v. 6 (3), 201-205
Isospora bigemina, I. felis, I. rivolta,
sporozoites inoculated into various tissue
cultures, all 3 species invaded cultured
cells but only I. bigemina developed furth
resemblance of cultured parasites to Toxoplasma
gondii, culture material fed to mic
with necropsy and serological tests for
Toxoplasma negative

Isospora bigemina (Stiles, 1891) Luehe, 1906
Tadros, W.; and Laarman, J. J., 1978, Proc.
K. Nederl. Akad. Wetensch., s. C, Biol. and
Med. Sc., v. 81 (4), 466-468
as syn. of Sarcocystis miescheriana (Kuehn
1865) Lankester, 1882

Isospora boughtoni Volk, 1938
Duszynski, D. W.; and Box, E. D., 1978, J.
Parasitol., v. 64 (2), 326-329
as syn. of Sarcocystis debonei Vogelsang,
1929

Isospora burrowsi n. sp., illus.
Trayser, C. V.; and Todd, K. S., jr., 1978,
J. Vet. Research, v. 39 (1), 95-98
Isospora burrowsi n. sp., life cycle
Canis familiaris (nat. and exper.) (caudal
three-fifths of small intestine, cecum,
feces)

Isospora burrowsi
Boch, J.; Boehm, A.; and Weiland, G., 1979,
Berl. u. Munchen. Tierarztl. Wchnschr., v. 9
(12), 240-243
coccidia, dogs (feces), coprological and
serological survey: South Germany

Isospora buteonis Henry, 1932
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk.,
v. 58 (2), 115-139
nomen dubium

Isospora canis-like oocysts
Dubey, J. P.; Fayer, R.; and Seesee, F. M.,
1978, J. Am. Vet. Med. Ass., v. 173 (9), 1167
1170
coyotes (feces): Montana

Isospora canis, illus.
Franken, A. M. J.; de Figueiredo, M. J.; and
Coutinho, S. G., 1975, Rev. Soc. Brasil. Med.
Trop., v. 9 (1), 42-51
Isospora rivolta, I. canis, morphologic comparisons,
incidence in dogs (feces) from
various areas of city of Rio de Janeiro

Isospora canis, illus.
Hilali, M.; Ghaffar, F. A.; and Scholtyseck,
E., 1979, Acta Vet., Budapest, v. 27 (3), 233
243
Isospora canis, fine structure of endogenou
stages in dog small intestine

Isospora canis, illus.
Jensen, J. B.; and Edgar, S. A., 1978, J.
Protozool., v. 25 (2), 169-173
Isospora canis, fine structure of penetration
of cultured cells by sporozoites

Isospora canis
Oduye, O. O.; and Bobade, P. A., 1979, J. Smal
Animal Practice, v. 20 (3), 181-184
Isospora canis, kennelled dogs, outbreak of
haemorrhagic diarrhoea, sulphadimidine treat
ment

spora canivelocis
Frank, C., 1978, Ang. Parasitol., v. 19 (3),
37-154
  Vulpes vulpes (mittleres Jejunum, Ileum, Rectum): Neusiedlerseegebiet, nordlichen Burgenland

ospora capreoli Svanbaev, 1958, illus.
Nukerbaeva, K. K., 1978, Izvest. Akad. Nauk
Kazakhsk. SSR, s. Biol. (3), 16-21
  Capreolus capreolus (feces): Alma-Ata zoopark

spora eversmanni
Frank, C., 1978, Ang. Parasitol., v. 19 (3),
37-154
  Mustela eversmanni (letztes Drittel des Jejunum): Neusiedlerseegebiet, nordlichen Burgenland

ospora felis
Cotteleer, C.; and Fameree, L., 1976, Rev.
Med. Liege, v. 31 (23), 729-734
  Eimeriidae, incidence in cats: Belgique

spora felis
Dubey, J. P., 1978, J. Protozool., v. 25 (3,
t. 2), 380-382
  Toxoplasma gondii, chronically infected cats re-excreted T. gondii oocyts after superinfection with Isospora felis, this re-excretion was prevented in cats infected with I. felis before T. gondii infection, administration of BCG before Toxoplasma infection had no apparent effect on outcome of infection

spora felis
Dubey, J. P.; and Frenkel, J. K., 1974, Vet.
ath., v. 11 (4), 350-379
  Toxoplasma gondii, cats (exper.), immunity, effects of host age and corticosteroid administration; excretion of T. gondii, Isospora felis, and I. rivolta oocysts from cats previously infected and challenged with all three coccidia

spora felis
ast, M. J.; and Powell, E. C., 1978, J. Para-
tol., v. 64 (1), 162-163
  mice used by Powell, E. C.; and McCarley, J. B., 1975, J. Parasitol., v. 61 (5), 928-931, were infected with both Sarcocystis muris and Isospora felis

spora felis
Shibalova, T. A.; and Petrenko, V. I., 1972,
arazitologiia, Leningrad, v. 6 (3), 201-205
  Isospora bigemina, I. felis, I. rivolta, sporozoites inoculated into various tissue cultures, all 3 species invaded cultured cells but only I. bigemina developed further, resemblance of cultured parasites to Toxoplasma gondii, culture material fed to mice with necropsy and serological tests for Toxoplasma negative

spora felis
mith, D. D.; and Frenkel, J. K., 1978, J. Para-
itol., v. 64 (2), 315-319
  Sarcocystis muris, Toxoplasma gondii, Isospora spp., experimental transmission to mice by ingestion of Blatella germanica and Periplaneta americana exposed to cat feces containing these coccidia or by direct ingestion of the feces, results indicate possible role of cockroaches as transport hosts and sources of cross-infection in laboratories, need for stringent control

Isospora frenkeli sp. nov. [nomen nudum]
  Arcay-de-Peraza, L., 1976, Resum. Trab. Libres, Fed. Latinoam. Parasitol. (San Jose, Dec. 7-11), 26 [Abstract]
    gato (intestino) (nat. and exper.)
    ratones blancos (epitelio intestinal) (exper.)

Isospora gallicolumbae sp. n., illus.
  Varghese, T., 1978, J. Protozool., v. 25 (4), 425-426
    Gallicolumba beccarii (feces): Waigani area, University, Port Moresby, Papua New Guinea

Isospora hominis, illus.
  Bassleer, J.; Nautet, M.; and Dargent, C., 1973, Rev. Med. Liege, v. 28 (8), 270-272
    Isospora hominis, clinical observations on 5 cases, morphologic differentiation from I. belli: residents of Belgium of various nationalities

Isospora hominis
  Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
    nomen dubium

Isospora hominis
  de Oliveira, G. S. C.; Barbosa, W.; and da Silva, A. L., 1973, Rev. Patol. Trop., v. 2 (4), 387-395
    Isospora hominis, I. belli, humans, epidemiologic survey, clinical aspects, immunodiagnostic comparisons with Toxoplasma gondii (immunofluorescence, protein electrophoresis, Sabin-Feldman dye test) resulted in frequent cross-reactions: Goias, Brazil

Isospora lacazei Labbe 1893, illus.
  Hernandez Rodriguez, S.; et al., 1976, Rev. Iber. Parasitol., v. 36 (1-2), 67-79
    Isospora lacazei, Carduelis carduelis (exper.), life cycle, ultrastructure of intestinal phases

Isospora lacazei Labbe 1893
  Hernandez Rodriguez, S.; et al., 1976, Rev. Iber. Parasitol., v. 36 (1-2), 81-88
    Passer domesticus
    Serinus canarius
    Passer hispaniolensis
    Carduelis carduelis (nat. and exper.)
    all from provincia de Cordoba, Espana

Isospora lacazei, illus.
  Speer, C. A.; et al., 1979, Ztschr. Parasitenk., v. 59 (3), 219-225
    Isospora lacazei, oocyst wall, scanning and transmission electron microscopy

Isospora lieberkuehni (Labbe, 1896) Laveran et Mensil, 1902, illus.
  Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
    description
    Rana esculenta (ledviny): CSSR

Isospora mayuri
  Williams, R. B., 1978, Vet. Parasitol., v. 4 (2), 193-197
    Isospora mayuri, first British record, oocyst morphology, failure to infect chickens, degree of pathogenicity uncertain, treatment with sulphaquinoxaline and diaveridine (Saquadil) in drinking water
    Pavo cristatus: Essex, Great Britain

Isospora neorivolta sp. n., illus.
Dubey, J. P.; and Mahrt, J. L., [1979], J. Parasitol., v. 64 (6), 1978, 1067-1073
Isospora neorivolta sp. n. proposed for I. rivolta of Mahrt, 1967; endogenous development compared with I. ohioensis
domestic dog (lamina propria of posterior half of small intestine) (exper.)

Isospora ohioensis, illus.
Dubey, J. P., 1978, J. Am. Vet. Med. Ass., v. 173 (2), 192-197
Isospora ohioensis, dogs (epithelium of small intestine, cecum, and colon) (exper.), pathology in young pups, pathogenicity was greatest in newborn and suckling pups whereas older pups (40-384 days at first inoculation) acquired immunity within 1 week

Isospora ohioensis
Dubey, J. P., 1978, Parasitology, v. 77 (1), 1-11
Isospora ohioensis, life cycle, endogenous stages in dog following oocyst-induced and mouse-induced infections, occurrence throughout small intestine, caecum, and colon, extraintestinal stages not found but biological evidence indicated invasion of spleens and mesenteric lymph nodes of dogs fed oocysts

Isospora ohioensis-like oocysts
Dubey, J. P.; Fayer, R.; and Seesee, F. M., 1978, J. Am. Vet. Med. Ass., v. 173 (9), 1167-1170
coyotes (feces): Montana

Isospora ohioensis
Dubey, J. P.; and Mahrt, J. L., [1979], J. Parasitol., v. 64 (6), 1978, 1067-1073
Isospora neorivolta sp. n. proposed for I. rivolta of Mahrt, 1967; endogenous development compared with I. ohioensis
dog (epithelium, throughout small intestine) (exper.)

Isospora ohioensis, illus.
Dubey, J. P.; and Mehlhorn, H., 1978, J. Parasitol., v. 64 (4), 689-695
Isospora ohioensis, persistence and structure of extraintestinal stages in tissues of mice inoculated with oocysts, these give rise to stages in dogs that are different from oocyst-induced infection in dogs, discussion of mice as transport vs. intermediate host

Isospora ohioensis-like organism, illus.
Dubey, J. P.; Weisbrode, S. E.; and Rogers, W. A., 1978, J. Am. Vet. Med. Ass., v. 173 (2), 185-191
Isospora ohioensis-like organism, dog (small intestine, cecum, colon, terminal ileum, villous epithelium, lamina propria, intestinal glands), description of asexual and sexual stages, pathology, attempted treatment with sulfaguanidine unsuccessful, case report: Ohio

Isospora putorii
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Mustela putorius (Jejunum): Neusiedlerseegebiet, nordlichen Burgenland

Isospora putorii (Railliet and Lucet, 1891) Becker, 1934
Tadros, W.; and Laarman, J. J., 1978, Proc. K. Nederl. Akad. Wetensch., s. C, Biol. and Med. Sc., v. 81 (4), 466-468
as syn. of Sarcocystis putorii (Railliet and Lucet, 1891) comb. nov.

Isospora rivolta
Cotteleer, C.; and Fameree, L., 1976, Rev. Med. Liege, v. 31 (23), 729-734
Eimeriidae, incidence in cats: Belgique

Isospora rivolta (Grassi, 1879), illus.
Dubey, J. P., 1979, J. Protozool., v. 26 (3), 433-443
Isospora rivolta, life cycle in cats and mice pathogenicity for newborn but not for weaned cats
cats (small intestine, in epithelial cells of villi and glands of Lieberkuhn) (exper.)
mice (mesenteric lymph nodes, spleen, liver, skeletal muscles) (exper.)

Isospora rivolta
Dubey, J. P.; and Frenkel, J. K., 1974, Vet. Path., v. 11 (4), 350-379
Toxoplasma gondii, cats (exper.), immunity, effects of host age and corticosteroid administration; excretion of T. gondii, Isospora felis, and I. rivolta oocysts from cats previously infected and challenged with all three coccidia

Isospora rivolta (Grassi 1879) of Mahrt, 1967
Dubey, J. P.; and Mahrt, J. L., [1979], J. Parasitol., v. 64 (6), 1978, 1067-1073
Isospora neorivolta sp. n. proposed for I. rivolta of Mahrt, 1967; endogenous development compared with I. ohioensis

Isospora rivolta, illus.
Franken, A. M. J.; de Figueiredo, M. J.; and Coutinho, S. G., 1975, Rev. Soc. Brasil. Med. Trop., v. 9 (1), 42-51
Isospora rivolta, I. canis, morphologic comparisons, incidence in dogs (feces) from various areas of city of Rio de Janeiro

Isospora rivolta
Matsui, T.; et al., 1977, Kiseichugaku Zasshi (Japan. J. Parasitol.), v. 26 (4), 235-239
Isospora rivolta, cats, mice (all exper.), sulfamonomethoxine

Isospora rivolta
Shibalova, T. A.; and Petrenko, V. I., 1972, Parazitologiia, Leningrad, v. 6 (3), 201-205
Isospora bigemina, I. felis, I. rivolta, sporozoites inoculated into various tissue cultures, all 3 species invaded cultured cells but only I. bigemina developed further, resemblance of cultured parasites to Toxoplasma gondii, culture material fed to mice with necropsy and serological tests for Toxoplasma negative

Isospora rivolta
Smith, D. D.; and Frenkel, J. K., 1978, J. Parasitol., v. 64 (2), 315-319
Sarcocystis muris, Toxoplasma gondii, Isospora spp., experimental transmission to mice by ingestion of Blatella germanica and Periplaneta americana exposed to cat feces containing these coccidia or by direct ingestion of the feces, results indicate possible role of cockroaches as transport hosts and sources of cross-infection in laboratories, need for stringent control

sospora rustamovi sp. n., illus.
Ovezmukhammedov, A., 1977, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (6), 67-68
Phrynocephalus reticulatus bannicovi (posterior section of intestine): western Turkmenia

sospora suis
Stuart, B. P.; and Lindsay, D., 1979, J. Am. Vet. Med. Ass., v. 175 (4), 328-329
Isospora suis, baby pigs (nat. and exper.), clinical disease

sospora suis
Upadhyay, A. N.; and Ahluwalia, S. S., 1976, Indian Vet. J., v. 53 (12), 972-973
mixed coccidial infection, pig, first infection did not confer any immunity, pig became resistant to challenge after second infection

Isospora suis
Upadhyay, A. N.; and Ahluwalia, S. S., 1978, Indian Vet. J., v. 55 (6), 458-461
coccidia, pigs (exper.), mixed infection with 4 spp. , macro- and microgametocytic stages, mostly not identified to species

Isospora suis
Upadhyay, A. N.; and Ahluwalia, S. S., 1978, Indian Vet. J., v. 55 (10), 829-830
4 spp. of coccidia, pigs (exper.), only slightly pathogenic, no gross observable lesions

Ithania Ludwig, 1947
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Ithania wenrichi Ludwig, 1947
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Kala azar. [See also Leishmaniasis]

Kala-azar
Bada, J. L.; et al., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (3), 347-348 [Letter]
kala-azar, longstanding evolution in asplenic male patient, case report: Barcelona, Spain

Kala azar
Bertrand, A.; Jourdan, J.; and Prefaut, C., 1975, Semaine Hop. Paris, v. 51 (10), 667-676
granulopenia of infectious origins, humans, kala azar and malaria listed among causes

Kala-azar
Camacho-Sanchez, M., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (5), 544-545
kala-azar, human, distribution in Colombia, South America

Kala-azar
Dar, F. K., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (5), 555-556 [Letter]
kala-azar, 2 1/2-year-old boy, first reported autochthonous case in Libya

Kala-azar
Fuller, G. K.; et al., 1979, Ann. Trop. Med. and Parasitol., v. 73 (5), 417-431
kala-azar, humans, extensive epidemiologic survey using the leishmanin skin test: south-west Ethiopia

Kala azar
Rassadi, P., 1979, Nouv. Presse Med., v. 8 (40), 2367-2368 [Letter]
kala azar, children, incidence from 1967-1975, pediatric hospital: Iran

Kala azar
Zylberait, D.; et al., 1979, Ann. Med. Int., Paris, v. 130 (10), 437-442
kala azar, woman, hemolytic anemia, possible abnormal sensitivity of erythrocyte membrane to complement

Karotomorpha ambystomae (Das Gupta 1935)
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
as syn. of Karotomorpha bufonis Dobell (1909)

Karotomorpha bufonis Dobell (1909), illus.
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
synonymy, description
Bufo bufo
B. viridis
Rana temporaria
all from CSSR

Karotomorpha swezei (Grasse 1926)
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
as syn. of Karotomorpha bufonis Dobell (1909)

Karyolysus
Landau, I., 1973, Mem. Mus. National Hist. Nat., Paris, n. s., s. A, Zool., v. 77, 1-62
Coccidiomorpha, life cycles, mechanisms assuring continuation of transmission, thesis

Karyolysus sp., illus.
Beier, T. V., 1979, Tsitologiia, v. 21 (3), 295-299
Karyolysus sp. trophozoites, interaction with lizard liver cells during host hibernation
Lacerta saxicola nairensis (liver): lake Sevan, Armenia

Klossia Schneider, 1875
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Klossia sp. Chatton, 1939
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Klossia sp. Mullin and Colley, 1972
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Klossia bigemina (Labbe, 1896) Labbe, 1899
Levine, N. D., 1977, Protozoology, v. 3, 173-182
Syn.: Eimeria bigemina Labbe, 1896

Klossia dimidiata Schneider, 1885
Levine, N. D., 1977, Protozoology, v. 3, 173-182
as syn. of Adelea dimidiata (Schneider, 1885) Labbe, 1896

Klossia helicina Schneider, 1875
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Klossia loossi Nabih, 1938
Levine, N. D., 1977, Protozoology, v. 3, 173-182

(?)Klossia macrocoronata (Lueling, 1942) nov. comb.
Levine, N. D., 1977, Protozoology, v. 3, 173-182
Syn.: ? macrocoronata Lueling, 1942

Klossia perplexens Levine, Ivens and Kruidenier, 1955
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Klossia simplex Schneider, 1885
Levine, N. D., 1977, Protozoology, v. 3, 173-182
as syn. of Adelina simplex (Schneider, 1885) Hesse 1911

Klossia soror schneider, 1881
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Klossia variabilis Levine, Ivens and Kruidenier, 1955
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Klossia vitrina Moroff, 1911
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Klossiella
Landau, I., 1973, Mem. Mus. National Hist. Nat., Paris, n. s., s. A, Zool., v. 77, 1-62
Coccidiomorpha, life cycles, mechanisms assuring continuation of transmission, thesis

ssiella
aylor, J. L.; et al., 1979, Vet. Parasitol.,
. 5 (2-3), 137-144
Klossiella, literature review: geographical distribution, host range, life cycle, pathogenicity

ssiella spp.
lazebrook, J. S.; Campbell, R. S. F.; and
utchinson, G. W., 1977, Tropenmed. u. Parasi-
ol., v. 28 (4), 545-551
survey of feral rodents in six tropical habitats for infections of potential zoonotic importance: North Queensland, Australia

ssiella mabokensis Boulard et Landau, 1971,
.is.
andau, I., 1973, Mem. Mus. National Hist.
at., Paris, n. s., s. A, Zool., v. 77, 1-62
Coccidiomorpha, life cycles, mechanisms assuring continuation of transmission, thesis
Praomys jacksoni (nat. and exper.)
souris blanche (exper.)

llikerella Labbe, 1899
evine, N. D., 1976, Tr. Am. Micr. Soc., v. 95
4), Finley Mem. Issue, pt. 2, 695-702
as syn. of Lecudina Mingazzini, 1891

llikerella staurocephali (Mingazzini, 1891)
be, 1899
evine, N. D., 1976, Tr. Am. Micr. Soc., v. 95
4), Finley Mem. Issue, pt. 2, 695-702
as syn. of Lecudina staurocephali (Mingaz-
zini, 1891) n. comb.

likeria Mingazzini, 1891
evine, N. D., 1976, Tr. Am. Micr. Soc., v. 95
4), Finley Mem. Issue, pt. 2, 695-702
as syn. of Lecudina Mingazzini, 1891

likeria staurocephali Mingazzini, 1891
vine, N. D., 1976, Tr. Am. Micr. Soc., v. 95
, Finley Mem. Issue, pt. 2, 695-702
s syn. of Lecudina staurocephali (Mingaz-
zini, 1891) n. comb.

a
era, L., 1976, Med. Wet., v. 32 (10), 630-
1
Kudoa-infected Merluccius merluccius, muco-
olysaccharide changes in the muscles

Kudoa sp.
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102
presumably contaminants from heart muscle slit during smearing operation
Clupea harengus harengus: St. Andrews, New Brunswick
Macrozoarces americanus: St. Andrews, New Brunswick
Myoxocephalus octodecemspinosus: Kent Island, New Brunswick
Pholis gunnellus: Kent Island, New Brunswick
Fundulus heteroclitus: Woods Hole, Massachusetts
Pseudopleuronectes americanus: Woods Hole, Massachusetts
(blood of all)

Kudoa sp., illus.
Podlipaev, S. A., 1972, Parazitologiia, Leningrad, v. 6 (6), 506-508
Myxosporidia, 6 spp., lipid inclusions in spores and cysts

Kudoa [sp.], illus.
Voelker, F. A.; et al., 1978, Vet. Path., v. 15 (1), 40-48
Kudoa [sp.] in Chaetodon (skeletal muscle), ultrastructure, light and electron microscopy, differential diagnosis

Kudoa pericardialis n. sp., illus.
Nakajima, K.; and Egusa, S., 1978, Bull. Japan. Soc. Scient. Fish., v. 44 (2), 117-120
Seriola quinqueradiata (pericardial cavity): Ajiro Bay, Shizuoka prefecture

Kudoa quadratum (Thelohan, 1895), illus.
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
Syn.: Chloromyxum quadratum Thelohan, 1895
Trachurus mediterraneus ponticus
Syngnathus nigrolineatus
Blennius zvonimiri
Gobius melanostomus
(skeletal musculature of all): all from Black Sea

Kudoa sphyraeni n. sp., illus.
Narasimhamurti, C. C.; and Kalavati, C., 1979, Proc. Indian Acad. Sc., Sect. B, Animal Sc., v. 88 (4, pt. 1), 265-268
Sphyraena jello (muscles of gut wall): Offshore Fishing Station, Visakhapatnam, and Bheemunipatnam, Andhra Pradesh

Kudoa tetraspora n. sp., illus.
Narasimhamurti, C. C.; and Kalavati, C., 1979, Proc. Indian Acad. Sc., Sect. B, Animal Sc., v. 88 (1, pt. 1), 85-89
Mugil cephalus (brain tissue around optic lobes): India

Labyrinthomyxa sp., illus.
Li, M. F.; and Clyburne, S., 1979, J. Invert. Path., v. 33 (1), 108-110
Labyrinthomyxa sp., mortalities in Mytilus edulis: Prince Edward Island, Canada

Labyrinthomyxa marina (Mackin, Owen, and Collier, 1950) Mackin and Ray, 1966
Levine, N. D., 1978, J. Parasitol., v. 64 (3), 549
as syn. of Perkinsus marinus (Mackin, Owen, and Collier, 1950) comb. n.

Lainsonia n. g.
Landau, I., 1973, Mem. Mus. National Hist. Nat., Paris, n. s., s. A, Zool., v. 77, 1-62
Schellakiidae
tod: L. iguanae n. sp.

Lainsonia iguanae n. g., n. sp. (tod), illus.
Landau, I., 1973, Mem. Mus. National Hist. Nat., Paris, n. s., s. A, Zool., v. 77, 1-62
Coccidiomorpha, life cycles, mechanisms assuring continuation of transmission, thesis
Iguana iguana (sang): Exu, Pernambuco, Bresil
Aedes aegypti (exper.)

Lamblia
Moralev, L. N.; and Moskalev, V. G., 1973, Parazitologiia, Leningrad, v. 7 (4), 377-378
Lamblia, use of PAS reaction for identification in histological sections of host intestine

Lamblia canis
Padchenko, I. K., 1970, Parazitologiia, Leningrad, v. 4 (4), 394-396
Lamblia, infestation rate in humans of different age groups in rural and urban areas, high infestation rate in preschool rural children may be explained by contact with farm dogs which carry natural infections of L. canis and were proved experimentally susceptible to L. intestinalis: Ukraine

Lamblia intestinalis
Angyal, J., 1976, Rheumatol., Balneol., Allergol., v. 17 (1), 51-52
Lamblia intestinalis, human, chronic urticaria cured when parasitic infection treated, case report

Lamblia intestinalis
Dymowska, Z.; and Zembrzuski, K., 1977, Przegl. Epidemiol., v. 31 (1), 109-113
human intestinal parasites, comparison of statistics of incidence surveys in 1974 and 1975: Poland

Lamblia intestinalis
Padchenko, I. K., 1970, Parazitologiia, Leningrad, v. 4 (4), 394-396
Lamblia, infestation rate in humans of different age groups in rural and urban areas, high infestation rate in preschool rural children may be explained by contact with farm dogs which carry natural infections of L. canis and were proved experimentally susceptible to L. intestinalis: Ukraine

Lamblia intestinalis
Padelt, H., 1970, Taegl. Prax., v. 11 (1), 57-61
Lamblia intestinalis, humans, clinical aspects, diagnosis, therapy, review

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Wurbs, D.; and Classen, M., 1979, Deutsche Med. Wchnschr., v. 104 (33), 1156-1157
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Benazet, F.; et al., 1970, Scand. J. Infect. Dis., v. 2 (2), 139-143
intestinal and hepatic parasites, nitrohete ocyclic antiparasitics, laboratory studies of chemotherapeutic activity and toxicity i exper. animals

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Solov'ev, M. M.; and Chentsov, Iu. S., 1970, Parazitologiia, Leningrad, v. 4 (6), 510-514
Lamblia muris, cysts, ultrastructure

Lamblia muris, illus.
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Lamblia muris, Trichomonas muris, Octomitus muris, localization in white mice exposed t x-irradiation

Lambliasis. See Giardiasis.

Lambornella Keilin, 1921
Corliss, J. O.; and Coats, D. W., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 725-739
Tetrahymenidae
generic diagnosis; Syn.: Tetrahymena pro parte
ciliated protozoa found associated with culicine mosquitoes in nature, review (infectivity and host resistance, pathogenicity, mode of entry into host, facultative or obligate parasitism?, taxonomic and nomenclatural considerations, potential importan in biological control of mosquitoes)

Lambornella clarki n. sp., illus.
Corliss, J. O.; and Coats, D. W., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 725-739
Aedes sierrensis (body cavity): California

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redescription

Lankesterella [sp.]
Baker, J. R., 1974, J. Zool., London, v. 172 (2), 169-190
Corvus frugilegus
Passer domesticus
all from Britain

Lankesterella [sp.], similar to, illus.
da Costa, S. C. G.; Pereira, N. M.; and Martinez Bernaola, N., 1971, Atas Soc. Biol. Rio de Janeiro, v. 14 (3-4), 45-48
Leptodactylus ocellatus (blood): Brasil

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Landau, I., 1973, Mem. Mus. National Hist.
Nat., Paris, n. s., s. A, Zool., v. 77, 1-62
Coccidiomorpha, life cycles, mechanisms assuring continuation of transmission, thesis
Rana esculenta (sang, cellules reticulo-endotheliales des tissus): rivieres de la cote Est de Corse (Fiume Orbo, La Bravone, Stabaccio)

kesterella [sp.]
eirce, M. A.; and Mead, C. J., 1977, J. Nat.
ist., v. 11 (6), 597-600
Fringilla coelebs
Parus ater
Prunella modularis
Erithacus rubecula
Certhia familiaris
Passer montanus
all from near Tring, Hertfordshire

kesterella [sp.]
illiams, N. A.; and Bennett, G. F., 1978,
anad. J. Zool., v. 56 (4, pt. 1), 596-607
Quiscalus quiscula: Maryland, and/or New Jersey

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hiriac, E.; and Steopoe, I., 1977, Rev. Rou-
aine Biol., s. Biol. Animale, v. 22 (2), 139-
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Lacerta vivipara (capillaires et veinules des ovaires, corps jaune, capsules surrenales)

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ojtkova, L., 1976, Scripta Fac. Scient. Nat.
niv. Purkynianae Brun., Biol., v. 6 (5), 177-
09
description
Syn.: L. ranarum Lank
Rana esculenta (cervene krvinky, jatra, oko, ojedinele i slezina): CSSR

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ojtkova, L., 1976, Scripta Fac. Scient. Nat.
niv. Purkynianae Brun., Biol., v. 6 (5), 177-
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modified diagnosis

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ochitskii, P. Ia.; and Bryginskii, S. A.,
79, Vestnik Zool., Akad. Nauk Ukrainsk.
R, Inst. Zool. (6), 87
Lankesteria culicis as possible factor regulating mosquito populations
Anopheles maculipennis (opening of middle intestine)
Culex p. pipiens (opening of middle intestine)
Aedes excrucians (opening of middle intestine)
A. geniculatus
all from Ukraine

rania ranarum Grassi et Feletti, 1892
jtkova, L., 1976, Scripta Fac. Scient. Nat.
iv. Purkynianae Brun., Biol., v. 6 (5), 177-
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as syn. of Dactylosoma ranarum (Krusse, 1890) Labbe, 1894

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Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702
Lecudina, revision, checklist of species together with synonyms, hosts, body locations, geographic distribution, and key references

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synonymy

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Syn.: Lecudina arabellae [of] Schrevel, 1963

Lecudina arrhyncha Bogolepova, 1953
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702

Lecudina attenuata (Bogolepova, 1953) Theodorides, 1969
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702
Syn.: Cygnicollum attenuatum Bogolepova, 1953

Lecudina bhatiai n. nov.
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702
for: Lecudina eunicae Bhatia & Setna, 1938, preoccupied by L. eunicae (Lankester, 1866) n. comb.

Lecudina bogolepovae (Levine, 1971) n. comb.
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702
synonymy

Lecudina criodrilii (Sciacchitano, 1931) n. comb.
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702
Syn.: Doliocystis criodrilii Sciacchitano, 1931

Lecudina elongata (Mingazzini, 1891) Kamm, 1922
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702
synonymy

Lecudina eunicae (Lankester, 1866) n. comb.
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702
synonymy

Lecudina eunicae Bhatia & Setna, 1938
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renamed: L. bhatiai n. nov.

Lecudina euphrosynes Bogolepova, 1953
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Lecudina fluctus Iitsuka, 1931
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Syn.: Lecudina fluktus Iitsuka of Hoshide (1958) lapsus calami

Lecudina fluktus Iitsuka of Hoshide (1958) lapsus calami
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702
as syn. of Lecudina fluctus Iitsuka, 1931

Lecudina ganapatii Vivier, Ormieres & Tuzet, 1964
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Syn.: Lecudina pellucida (Koelliker) Mingazzini, 1891 of Ganapati (1946) non L. pellucida (von Koelliker, 1848) Mingazzini, 1891

Lecudina heterocephala (Mingazzini, 1891) Kamm, 1922
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702
synonymy

Lecudina laubieri Theodorides, 1969
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Syn.: Lecudina (Cygnicollum) laubieri Theodorides, 1969

Lecudina (Cygnicollum) laubieri Theodorides, 1969
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702
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Lecudina legeri (Brasil, 1909) Kamm, 1922
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702
synonymy

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Syn.: Lecudina lisidicae Bhatia & Setna, 1938 of Bogolepova (1953)

Lecudina mammilata H. Hoshide, 1944
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Lecudina nereicola (Bogolepova, 1953) n. comb.
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702
Syn.: Polyrhabdina nereicola Bogolepova, 1953

Lecudina pellucida (von Koelliker, 1848) Mingazzini, 1891
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. (4), Finley Mem. Issue, pt. 2, 695-702
synonymy

Lecudina platynereidis
Theodorides, J., 1978, Ann. Parasitol., v. 5 (4), 437-438
syzygy from Platynereis dumerilii at Villefranche-sur-Mer identified by Theodorides, 1969 a, as Lecudina platynereidis actually belongs to D. schreveli

Lecudina polydorae (Leger, 1893) Kamm, 1922
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synonymy

Lecudina postae n. sp., illus.
Theodorides, J.; and Desportes, I., 1978, Ann Parasitol., v. 53 (5), 443-446
Hyalinoecia tubicola (tube digestif): Cap Corse

Lecudina pyriformis Bogolepova, 1953
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 9 (4), Finley Mem. Issue, pt. 2, 695-702

Lecudina staurocephali (Mingazzini, 1891) n. comb.
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 9 (4), Finley Mem. Issue, pt. 2, 695-702
Syns.: Koellikeria staurocephali Mingazzini 1891; Koellikerella staurocephali (Mingazzini, 1891) Labbe, 1899

Leidyana gnyanagangai n. sp., illus.
Patil, C. C.; and Amoji, S. D., 1979, Current Sc., Bangalore, v. 48 (16), 736-737 [Letter]
Rhytinota impolita (midgut, hindgut): Gulbarga (Karnataka, India)

Leishmania
Abdalla, R. E.; and Sherif, H., 1978, Ann. Trop. Med. and Parasitol., v. 72 (4), 349-352
cutaneous leishmaniasis, human, description of first epidemic to be reported from the Sudan

Leishmania
Amato Neto, V.; da Silva, L. J.; and Camargo, M. E., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (2), 99-102
serodiagnostic tests for Chagas disease performed on sera of patients with visceral leishmaniasis gave positive results with several immunologic methods, indicates "group-reactions" within Trypanosomatidae

Leishmania
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leishmaniasis, epidemiology, review with emphasis on zoonotic cutaneous leishmaniasis of Ethiopia

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leishmaniasis, immunodiagnosis, review

Leishmania
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visceral leishmaniasis, human, review of previously reported autochthonous cases in Libya

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Dobrzhanskaia, R. S., 1979, Vestnik Dermat. i Venerol. (6), 56-59
leishmaniasis, human cutaneous, preparation of diagnostic leishmanial antigen based on deep freezing followed by thawing

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Evans, D. A., 1978, Methods Cultiv. Parasites in Vitro, 55-88
Kinetoplastida, cultivation, review

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Fuller, G. K.; et al., 1979, Ann. Trop. Med. and Parasitol., v. 73 (5), 417-431
kala-azar, humans, extensive epidemiologic survey using the leishmanin skin test: south-west Ethiopia

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Gardener, P. J., 1977, Trop. Dis. Bull., v. 74 (12), 1069-1088
Leishmania, early documentation of human leishmaniasis, taxonomic chronology of genus, synonymies of principal species, list of species not normally infective to humans, recent developments in leishmanial systematics

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identification of economically important parasites (use of anatomical, biochemical, and behavioral tests), brief review

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Grove, S. S., 1978, South African Med. J., v. 53 (18), 712-715
leishmaniasis, clinical and histological features of South West African form

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Gutteridge, W. E.; and Coombs, G. H., 1977, Biochemistry of parasitic protozoa, 172 pp., illus.
biochemistry of parasitic protozoa, textbook: methodology; catabolism and generation of energy; nucleic acid metabolism; protein metabolism; lipid metabolism; biochemical mechanism of drug action; isolation of parasitic protozoa from infected animals; culture of parasitic protozoa

ishmania
Hommel, M., 1978, Bull. Inst. Pasteur, Paris, v. 76 (1), 5-102
Leishmania spp., parasite biology, clinical aspects of leishmaniasis, extensive review and bibliography: taxonomy, morphology, transmission, development, disease in man and animals, experimental infections

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Inferrera, C.; et al., 1973, Riv. Anat. Patol. e Oncol., v. 38, 1972-1973, LVII-LXII
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Lumsden, W. H. R., 1974, Ciba Found. Symp., n.s. (20), 3-27
leishmaniasis and trypanosomiasis, causative organisms compared and contrasted, review

Leishmania
Lumsden, W. H. R., 1978, Handb. Exper. Immunol. (Weir), 3. ed., 42.1-42.47
demonstration of antibodies to Protozoa, extensive review

Leishmania
Luzzio, A. J.; McRoberts, M. J.; and Euliss, N. H., 1979, J. Infect. Dis., v. 140 (3), 370-377
Leishmania spp., immunized rabbits, infected hamsters (exper.), and humans, quantitative estimation of antibody titers by enzyme-linked immunosorbent assay, some comparisons with passive hemagglutination, complement fixation, and countercurrent immunoelectrophoresis

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Maekelt, G. A., 1972, Immun. Animal Parasites, 343-363
immune response to Leishmania, review

Leishmania
Martinez B., N., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (4), 179-180
Leishmania, experimental dog infections with human strains of espundia or uta types, dogs equally susceptible to both types

Leishmania
Mauel, J.; et al., 1974, Ciba Found. Symp., n.s. (25), 225-242
Leishmania, survival and death in macrophages, review

Leishmania
Mayrink, W.; et al., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (6), 676
human dermal leishmaniasis, field trial of vaccine, responses to Montenegro antigen after immunization with killed Leishmania promastigotes: Brazil

Leishmania
Mayrink, W.; et al., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (4), 385-387
American dermal leishmaniasis, humans, immunization, field trial using a vaccine containing killed promastigotes of 5 stocks of Leishmania, preliminary report of promising control method: Brazil

Leishmania
Mendes, E., 1979, Cellular Immunol., v. 42 (2), 424-427
transfer of delayed hypersensitivity to leishmanin (Montenegro reaction), remains to be established whether this alters clinical course of visceral leishmaniasis

Leishmania
Miles, M.; and Ward, R. D., 1978, Ann. Trop. Med. and Parasitol., v. 72 (4), 398-400
phlebotomine sandflies (vector of leishmaniasis), preliminary isoenzyme studies, possible application to identifying populations of medical importance

Leishmania, illus.
Moriearty, P. L.; et al., 1978, Rev. Inst. Med. Trop. S. Paulo, v. 20 (1), 15-21
borderline cutaneous leishmaniasis, clinical, immunological and histological differences from mucocutaneous leishmaniasis, patients from Bahia, Brazil

Leishmania
Newton, B. A., 1974, Ciba Found. Symp., n.s. (20), 285-307
trypanosomiasis, leishmaniasis, chemotherapy, review

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human mucocutaneous leishmaniasis, case report and autopsy findings of infection in Japanese patient who had never abroad: Osaka, Japan

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trypanosomiasis, leishmaniasis, drug resistance, review

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Ponirovskii, E. N., 1975, Parazitologiia, Leningrad, v. 9 (2), 139-141
Leishmania, comparison of 3 methods of identification of strains

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Price, S. M.; and Silvers, D. N., 1977, Arch. Dermat., Chicago, v. 113 (10), 1415-1416
cutaneous leishmaniasis resulting in ulcerous lesion on arm of college student who had recently returned from Peru, case report, diagnostic problems finally resolved by fluorescent antibody test: New York

Leishmania, illus.
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leishmaniasis, human cutaneous, histopathology in relationship to immunological mechanisms, logarithmic parasite index to provide framework of various disease forms and their inter-relationships

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Rondanelli, E. G.; et al., 1976, Recenti Prog. Med., v. 61 (2), 137-162
Leishmania donovani and L. tropica promastigote forms in vitro, basis for qualifying characters of ultrastructural organization of genus Leishmania and aspects of its reproduction and pathogenicity; promastigote and endomastigote phases discussed

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Rutherfoord, G. S.; and Uys, C. J., 1978, South African Med. J., v. 53 (18), 716-718
cutaneous leishmaniasis, 54-year-old woman, microscopical findings, ultrastructure of lesion presented to facilitate diagnosis in South West Africa

Leishmania
Saf'ianova, V. M., 1974, Parazitologiia, Leningrad, v. 8 (4), 336-347
leishmaniasis, characterization of nidi on basis of "transmissible factor" (species composition of sandfly vectors)

Leishmania
Sharma, M.; et al., 1979, Immune Reg. Transfer Factor, 563-569
human cutaneous Leishmania infection, transfer factor therapy, double blind clinical trial: Iran

Leishmania
Sharma, M. K., Anaraki, F.; and Ala, F., 1979, Clin. Immunol. and Immunopathol., v. 12 (2), 183-190
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Totescu, E.; and Susnea, S., 1978, Bacteriol., Virusol., Parazitol., Epidemiol., Bucuresti, v. 23 (4), 253-254
Leishmania, mother and son, case reports: Constanta, contracted in Libya

Leishmania
Trager, W., 1974, Ciba Found. Symp., n.s. (20), 225-254
trypanosomes, leishmanias, nutrition and biosynthetic capabilities, problems of in vitro cultivation and differentiation, review

Leishmania
Turk, J. L.; and Belehu, A., 1974, Ciba Found. Symp., n.s. (25), 101-122
infectious diseases including leishmaniasis and Chagas' disease, evidence for immunological basis of spectra of clinical manifestations, review

Leishmania
Vickerman, K., 1974, Ciba Found. Symp., n.s. (20), 171-198
trypanosomatid flagellates, ultrastructure, review with emphasis on changes during life cycles

Leishmania
Ward, R. D.; Lainson, R.; and Shaw, J. J., 1978, Ann. Trop. Med. and Parasitol., v. 72 (3), 269-276
techniques for membrane feeding and infecting of laboratory reared Lutzomyia vectors of leishmaniasis

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immunopathology due to cell-mediated (Type IV) reactions, review

Leishmania
Williams, P.; and Coelho, M. de V., 1978, Advances Parasitol., v. 16, 1-42
Leishmania, taxonomy and nomenclature, Phlebotominae as hosts for Trypanosomatidae, review

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Leishmania spp., characterization by isoenzyme electrophoresis, comparison of stocks from Kuwait with stocks from other parts of Old and New Worlds

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Leishmania spp., cellular levels and synthesis of polyamines during growth cycle, polyamines characteristics might serve as further criterion for strain identification and classification

shmania sp.
achrach, U.; et al., 1979, Exper. Parasitol., . 48 (3), 464-470
Leishmania spp., effect of ethidium, pentamidine and methylglyoxal-bis (guanylhydrazone) on growth and on polyamine, RNA, and DNA synthesis

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ettini, S.; Gradoni, L.; and Pozio, E., 1978, r. Roy. Soc. Trop. Med. and Hyg., v. 72 (4), 41-442 [Letter]
Leishmania, 3 strains isolated from Rattus rattus by inoculation of rat spleen into hamsters: area of Baccinello, Tuscany, Italy

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razil, R. P., 1978, Ann. Trop. Med. and Parasitol., v. 72 (3), 289-291
Leishmania spp., variation in electrophoretic mobility of the enzyme phosphoglucomutase in the parasite and its application to the differentiation of leishmanial strains

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ance, M. L.; et al., 1978, Ann. Trop. Med. d Parasitol., v. 72 (6), 533-542
Leishmania, identification of 68 strains from Aethiopian zoogeographical region on basis of biochemical and serological taxonomy (nuclear and kinetoplast DNA buoyant density, excreted factor serotypes, enzyme variant types), epidemiological implications

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âne, L. M.; da Silva, J. E.; and de Figueirlo, P. Z., 1974, Rev. Inst. Med. Trop. S. aulo, v. 16 (2), 68-69
porcupine, probably Coendou prehensilis prehensilis (liver, spleen): State of Piaui, Brazil

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det, J. P.; Derouin, F.; and Cornet, M., 1978, mpt. Rend. Acad. Sc., Paris, v. 286, s. D, . Nat. (3), 301-302
Phlebotomus duboscqi (estomac): Senegal

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Dias, M.; et al., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (6), 403-410
Leishmania sp., survey of wild and domestic animals as reservoir hosts, results suggest active role of dogs: Caratinga, Minas Gerais State, Brazil

Leishmania sp.
Hendricks, L. D.; Wood, D. E.; and Hajduk, M. E., 1978, Parasitology, v. 76 (3), 309-316
Leishmania spp., Trypanosoma spp., commercially available liquid media for rapid cultivation

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Killick-Kendrick, R.; Leaney, A. J.; and Ready, P. D., 1977, J. Med. Entom., v. 13 (4-5), 429-440
establishment, maintenance, and productivity of colony of Lutzomyia longipalpis from Brazil, results suggest that L. longipalpis may be suitable sand fly for life cycle and transmission studies of Leishmania spp.

Leishmania [sp.], L. braziliensis type
Lainson, R.; et al., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (2), 239-242
cutaneous leishmaniasis, observations on epidemiology in north Para State, Brazil
Lutzomyia umbratilis (pylorus, ileum, cardia, oesophagus, midgut)
L. whitmani (pylorus, ileum, cardia, oesophagus, midgut)
men (arms, hands)
hamsters (exper.)
Dasypus novemcinctus (spleen)
all from area near Monte Dourado

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Lainson, R.; et al., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (2), 239-242
Dasypus novemcinctus (heart-blood, spleen, liver): forest near Moju, north Para State, Brazil
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hamster (skin) (exper.)

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as syn. of Leishmania mexicana

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Trypanosoma cruzi culture forms, antigenic make-up, comparison with salivarian and some other stercorarian trypanosomes and Leishmania using immunoprecipitation in gels and immunoelectrophoresis, identification of component specific to T. cruzi

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Leishmania donovani, hamsters, superior efficacies of liposome-encapsulated meglumine antimoniate and sodium stibogluconate, efficacy of treatment influenced by lipid composition and charge of liposomes, morphologic evidence that liposomes travel to intracellular site of parasite, encapsulation and reduction of dose should minimize toxic reactions to antimonials

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Andrade, Z. A.; and Iabuki, K., 1972, Rev. Inst. Med. Trop. S. Paulo, v. 14 (1), 51-54
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Varanus sp.
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Leishmania spp., comparison of rate of parasite uptake by mouse peritoneal macrophages

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Ardehali, S. M.; Khoubyar, K.; and Rezai, H. R., 1979, Acta Trop., v. 36 (1), 15-21
Leishmania-macrophage interaction in vitro, effect of cytochalasin B, concluded that infection was by phagocytosis rather than activ penetration, cells from outbred mouse strain susceptible to L. tropica phagocytosed this species less efficiently than L. enriettii or L. donovani

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26 trypanosomatid species, cultivation in new chemically-defined medium RE III

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Leishmania enriettii in vitro grows well in guinea pig macrophages but fails to grow in mouse macrophages; medium containing excretory factors from L. enriettii culture (conditioned medium) promotes infection in mouse macrophages but similar medium from L. tropica does not; immune precipitation of medium by anti-Leishmania serum cancels infection-promoting effect; L. tropica infection of guinea pig macrophages enhanced by homologous medium

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Leishmania spp., Trypanosoma spp., commercially available liquid media for rapid cultivation

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Leishmania enriettii, destruction of intracellular organisms in macrophages activated by cocultivation with stimulated macrophages

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Leishmania enriettii
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parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

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Scorza, J. V.; et al., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (3), 293-298
Leishmania garnhami n. sp., epidemiology, pathology, behaviour in exper. hosts and in culture, associated with Lutzomyia townsendi, light and electron microscopy
human (cutaneous lesions): urban, suburban and rural areas in the Venezuelan Andes region, mainly from 800 to 1,800 m
Syrian hamsters (exper.)

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Leishmania herreri sp. n., illus.
Zeledon, R.; Ponce, C.; and Murillo, J., 1979, J. Parasitol., v. 65 (2), 275-279
Choloepus hoffmanni (skin of dorsum and ear, blood, liver, spleen)
Bradypus griseus (blood, spleen)
Lutzomyia ylephiletor
L. shannoni
L. trapidoi
all from Costa Rica

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Lainson, R.; and Shaw, J. J., 1977, Acta Amazonica, v. 7 (1), 51-57
Coendou prehensilis
Coendou nov. sp. (liver, spleen, skin)
all from Ponta de Pedras, Marajo Island, Para State, north Brazil

Leishmania hertigi deanei, illus.
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Leishmania hertigi hertigi, L. h. deanei, ultrastructure of promastigotes, amastigotes and virus-like particles observed within promastigotes; laboratory mammals were poor hosts with infection detectable only by culture, laboratory-bred Lutzomyia longipalpis developed poor infections

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Leishmania hertigi, taxonomic differentiation of strains from Panama and Brazil using morphological, biochemical and serological characteristics

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(micro-ELISA) technique for serodiagnosis using Leishmania infantum as soluble antige

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Leishmania, identification of 68 strains fro
Aethiopian zoogeographical region on basis of biochemical and serological taxonomy (nuclear and kinetoplast DNA buoyant density
excreted factor serotypes, enzyme variant types), epidemiological implications

Leishmania major
Dedet, J. P.; et al., 1979, Ann. Trop. Med. and Parasitol., v. 73 (5), 433-437
Leishmania major, search for reservoirs for human infection
Tatera gambiana (bone marrow)
Mastomys erythroleucus (spleen, liver)
all from Senegal, West Africa

Leishmania major Yakimoff & Schokhor, 1914 emend. Bray, Ashford & Bray, 1973
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Phlebotomus duboscqi, present and feeding on man, potential vector of Leishmania major in The Gambia

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Leishmania tropica major and L. mexicana, cross-immunity in mice, first reported observation of less virulent Leishmania species being able to confer immunity against a more virulent species

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Leishmania mexicana, mice, chronic infection, alterations of immune response, results suggest role for suppressor cells in pathogenesis of diffuse cutaneous leishmaniasis

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shmania mexicana
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each, D. H.; Holz, G. G., jr.; and Anekwe, G. , 1979, J. Parasitol., v. 65 (2), 203-216
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sera from humans infected with visceral or tegumentary leishmaniasis studied by electrophoresis using heterologous and homologous antigens; homologous antigens were more specific and detected higher titers, false positive reactions occurred only with Leishmania donovani

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Figueiredo, Y. P.; et al., 1976, Rev. Inst. Med. Trop. S. Paulo, v. 18 (5), 306-314
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Trypanosoma cruzi, Leishmania spp., factor in cell-free extracts that induces lysis of mammalian red cells and Vero cells, postulated that this lytic factor is involved in penetration and damage produced by T. cruzi in vertebrate cells

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Perez, H.; Arredondo, B.; and Gonzalez, M., 1978, Infect. and Immun., v. 22 (2), 301-307
Leishmania mexicana, 2 human strains (one from typical case of American cutaneous leishmaniasis and one from case of diffuse cutaneous leishmaniasis) in 2 strains of inbred mice, course of lesions, delayed hypersensitivity response, agglutinating antibodies, in vitro responses to leishmanial antigens and to mitogens, results show impaired immune response in BALB/c mice

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Leishmania mexicana, L. tropica major, cross immunity in mice, evidence of shared antigenic determinants which are involved in cell-mediated immune responses

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Leishmania mexicana, variations in response of 5 strains of mice (course of infection, delayed type hypersensitivity response, humoral antibody production), crossing experiments between resistant and susceptible strains suggest that resistance is inherited as dominant character

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mobility of the enzyme phosphoglucomutase i
the parasite and its application to the differentiation of leishmanial strains

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rapid method for isolation of purified amastigotes using cellulose column

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non-specific binding of 5 types of particle
(including Leishmania) to rat peritoneal cells, effect of various physical and chemical factors, lymphocytes as well as macrophages were able to bind Leishmania

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Lutzomyia flaviscutellata (mid- and fore-gut, 'hind-triangle'): Vega de Oropouche and Aripo-Waller Field, Trinidad, West Indies

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Leishmania mexicana mexicana, Mastomys natalensis readily infected and could serve as useful laboratory host for immunological and chemotherapeutic studies

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enzymes of ornithine-arginine metabolism,
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which may serve as biochemical markers in
classification of Trypanosomatidae

Leptomonas seymouri
Camargo, E. P.; Itow, S.; and Alfieri, S. C.
[1979], J. Parasitol., v. 64 (6), 1978, 1120
1121
Trypanosomatidae, 18 spp. of 6 genera, pro
lytic activities in cell extracts

Leptomonas seymouri
Fish, W. R.; Holz, G. G., jr.; and Beach, D. H., 1978, J. Parasitol., v. 64 (3), 546-547
26 trypanosomatid species, cultivation in r
chemically-defined medium RE III

Leptomonas seymouri
Galinari, S.; and Camargo, E. P., 1978, Exper
Parasitol., v. 46 (2), 277-282
trypanosomatid protozoa, 16 spp., survey fo
acetylornithinase and ornithine acetyltrans
ferase, metabolic and nutritional implications

Leptomonas seymouri
Goncalves de Lima, V. M. Q.; Roitman, I.; and
Kilgour, V., 1979, J. Protozool., v. 26 (4),
648-652
trypanosomatids, 7 species distinguished by
electrophoretic mobilities of some isoenzym

Leptomonas seymouri
Yoshida, N.; and Camargo, E. P., 1978, J. Bacteriol., v. 136 (3), 1184-1186
trypanosomatids, excretion of urea or ammonia or both, varies according to genus, may be of taxonomic use

theca hepseti Thelochan, 1895
aleva, A. A., 1966, Respublik. Mezhvedomstv.
rn., Akad. Nauk Ukrain. SSR, s. Biol. Moria,
38
nvasion extensity and intensity in relation
o host age
therina hepsetus (gall bladder): Black
ea (region of Karadag)

theca krogiusi
ovalov, S. M.; Shevliakov, A. G.; and
sin, V. K., 1970, Parazitologiia, Leningd, v. 4 (6), 547-556
arasite fauna of various groups of young
ncorhynchus nerka, comparative analysis
eveals 3 ecological groups: Lake Azaach'e, Kamchatka river basin

theca krogiusi
hovenko, E. T., 1972, Parazitologiia, Lengrad, v. 6 (4), 369-375
alvelinus alpinus inhabiting different ecoogical niches, possible use of differences
n parasite fauna between groups as biological tags: Lake Azabach'e, Kamchatka

cytozoon (or Akiba) [sp.]
er, J. R., 1974, J. Zool., London, v. 172
, 169-190
Corvus corone
C. frugilegus
C. monedula
Fringilla coelebs
Garrulus glandarius
Muscicapa striata
Parus caeruleus
Sturnus vulgaris
Turdus ericetorum
T. merula
Strix aluco
Gallinula chloropus
Columba palumbus
Lagopus scoticus
all from Britain

cytozoon [sp.]
nett, G. F.; Cameron, M.; and White, E.,
75, Canad. J. Zool., v. 53 (10), 1432-1442
ematozoa of passeriforms, prevalence,
ffect of climate, application of insecticide, and large-scale environmental alteration
Cyanocitta cristata
Coccyzus erythropthalmus
Carpodacus purpureus
Hesperiphona vespertina
Junco hyemalis
Melospiza georgiana
M. melodia
Passerculus sandwichensis
Pheucticus ludovicianus
Pooecetes gramineus
Spinus tristis
Spizella passerina
Zonotrichia albicollis
Agelaius phoeniceus
Molothrus ater
Quiscalus quiscula
Parus atricapillus
P. hudsonicus
Dendroica coronata
D. fusca
D. magnolia
D. palmarum
D. pensylvanica
D. petechia
D. striata
D. tigrina
Geothlypis trichas
Mniotilta varia

Leucocytozoon [sp.]-- Continued.
Bennett, G. F.; Cameron, M.; and White, E.,
1975, Canad. J. Zool., V. 53 (10), 1432-1442.--
Continued.
Oporornis philadelphia
Seiurus aurocapillus
S. novaboracensis
Setophaga ruticilla
Vermivora peregrina
V. ruficapilla
Wilsonia canadensis
W. pusilla
warblers, unknown
Colaptes auratus
Catharus guttatus
C. ustulatus
Turdus migratorius
Empidonax trailii
E. sp.
Nuttallornis borealis
Vireo olivaceous
sparrows, unidentified
Piranga olivacea
Catharus fuscescens
Hylocichla mustelina
Empidonax minimus
Melospiza lincolnii
all from New Brunswick

Leucocytozoon sp.
Bennett, G. F.; White, E. M.; and Williams, N.
A., 1977, J. Wildlife Dis., v. 13 (3), 251-257
Ploceus aurantius
P. nigerrimus
P. nigricollis
P. weynsi
Quelea erythrops
(blood of all): all from Uganda

Leucocytozoon sp. Gluschenko, 1963
Kairullaev, K. K., 1979, Izvest. Akad. Nauk
Kazakhsk. SSR, s. Biol. (1), 19-21
Phylloscopus trochilus
Sylvia borin
S. curruca
(blood of all): all from central course of
Ural river, Priural region, Ural oblast

Leucocytozoon [sp.]
Kocan, A. A.; Snelling, J.; and Greiner, E.
C., 1977, J. Wildlife Dis., v. 13 (3), 304-306
Strix varia
Bubo virginianus
Otus asio
Buteo jamaicensis
B. regalis
Falco sparverius
Buteo lineatus
Accipiter cooperii
Buteo lagopus
all from Oklahoma

Leucocytozoon n. sp., illus.
Samaddar, J.; Biswas, D. N.; and Ghose, A. N.,
1978, Indian Vet. J., v. 55 (7), 572-576
leucocytozoonosis, W[hite] L[eg] H[orn]
birds, quinine bisulphate and resotren failed
to ensure absolute recovery, though general
condition of treated birds improved

Leucocytozoon [sp.]
Williams, N. A., 1978, J. Parasitol., v. 64 (3), 556-558
Bombycilla cedrorum
Junco hyemalis
Melospiza melodia
Spinus pinus
Dendroica coronata
Oporornis tolmei
Vermivora celata
Colaptes auratus
Ixoreus naevius
Turdus migratorius
all from 16 km west of Courtenay, British Columbia

Leucocytozoon [spp.]
Williams, N. A.; and Bennett, G. F., 1978, Canad. J. Zool., v. 56 (4, pt. 1), 596-607
Anas rubripes: Maryland
Oxyura jamaicensis: "
Casmerodius albus: Maryland or New Jersey
Egretta thula: Maryland or New Jersey
Nycticorax nycticorax: New Jersey
Cathartes aura: Maryland or New Jersey
Corvus brachyrhynchos: Maryland
Cyanocitta cristata: Maryland, and/or New Jersey
Cardinalis cardinalis: Maryland, and/or New Jersey
Junco hyemalis: Maryland, and/or New Jersey
Passerella iliaca: " " " " "
Melospiza melodia: Maryland or New Jersey
Pipilo erythrophthalmus: Maryland, and/or New Jersey
Spizella passerina: Maryland or New Jersey
S. pusilla: Maryland or New Jersey
Zonotrichia albicollis: Maryland, and/or New Jersey
Agelaius phoeniceus: Maryland, and/or New Jersey
Euphagus carolinus: Maryland or New Jersey
Icterus spurius: Maryland
Molothrus ater: Maryland, and/or New Jersey
Quiscalus quiscula: " " " "
Dumetella carolinensis: Maryland or New Jersey
Parus bicolor: Maryland, and/or New Jersey
Dendroica magnolia: Maryland or New Jersey
D. pensylvanica: " " " "
Geothlypis formosa: Maryland or New Jersey
G. trichas: Maryland
Icteria virens: Maryland, and/or New Jersey
Setophaga ruticilla: Maryland
Colaptes auratus: Maryland or New Jersey
Passer domesticus: Maryland, and/or New Jersey
Sturnus vulgaris: Maryland, and/or New Jersey
Catharus guttatus: Maryland, and/or New Jersey
C. minimus: Maryland, and/or New Jersey
C. ustulatus: " " " "
Hylocichla mustelina: Maryland, and/or New Jersey
Sialia sialis: Maryland or New Jersey
Turdus migratorius: Maryland, and/or New Jersey
Vireo olivaceus: Maryland, and/or New Jersey
Toxostoma rufum: Maryland, and/or New Jersey

Leucocytozoon anatis Tartakovsky, 1913
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Leucocytozoon apiaster Zeiniev, 1975
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Leucocytozoon ardea
Bennett, G. F.; White, E. M.; and Williams, N. A., 1977, J. Wildlife Dis., v. 13 (3), 251-257
Butorides striatus (blood): Uganda

Leucocytozoon audieri Laveran and Nattan-Larrier 1911
Greiner, E. C.; and Kocan, A. A., 1977, Canad. J. Zool., v. 55 (5), 761-770
as syn. of Leucocytozoon toddi Sambon 1908

Leucocytozoon bacelari Tendeiro 1947
Greiner, E. C.; and Kocan, A. A., 1977, Canad. J. Zool., v. 55 (5), 761-770
as syn. of Leucocytozoon toddi Sambon 1908

Leucocytozoon beaurepairei Travassos Santos Dias 1954
Greiner, E. C.; and Kocan, A. A., 1977, Canad. J. Zool., v. 55 (5), 761-770
as syn. of Leucocytozoon toddi Sambon 1908

Leucocytozoon brimonti
Bennett, G. F.; White, E. M.; and Williams, N. A., 1977, J. Wildlife Dis., v. 13 (3), 251-257
Pycnonotus virens (blood): Uganda

Leucocytozoon brimonti
Wink, M.; and Bennett, G. F., 1976, J. Wildlife Dis., v. 12 (4), 587-590
Pycnonotus barbatus: Ghana

Leucocytozoon bubonis Fantham, 1926
Kairullaev, K. K., 1979, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (1), 19-21
Accipiter nisus (blood): central course of Ural river, Priural region, Ural oblast

Leucocytozoon caulleryi
Fujisaki, K.; et al., 1979, Japan. J. Vet. Sc., Tokyo, v. 41 (4), 385-390
Leucocytozoon caulleryi, chickens, auto-cold hemagglutinin in plasma

Leucocytozoon caulleryi, illus.
Kitaoka, S., 1978, Japan Agric. Research Quart., v. 12 (3), 157-160
Leucocytozoon caulleryi, chickens, immunoserological diagnosis, application in surveys and screening tests of prophylactic drugs, review

Leucocytozoon (= Akiba) caulleryi
Manuel, M. F.; Morales, E. G.; and Trovela, E., [1977], Philippine J. Vet. Med., v. 15 (1-2), 1976, 87-95
Leucocytozoon caulleryi, White Leghorn cockerels, superior prophylactic value of sulfamonomethoxine + pyrimethamine administered in feed under field conditions, no detrimental effects on growth rate or blood picture

Leucocytozoon (Akiba) caulleryi
Manuel, M. F.; Morales, E.; and Trovela, E., 1977, Philippine J. Vet. Med., v. 16 (1-2), 20-30
Leucocytozoon caulleryi, chickens under natural conditions, clopidol, halofuginone and furazolidone given in feed, clopidol 100% effective, no detrimental effects by any drug on host growth or red and white blood cell count

Leucocytozoon caulleryi
Manuel, M. F.; and Trovela, E., 1977, Philippine J. Vet. Med., v. 16 (1-2), 31-39
Leucocytozoon caulleryi, chickens under field conditions, efficacy of halofuginone and furazolidone alone and in combination, given with feed, furazolidone at high dosage showed some adverse host growth effects, neither drug showed adverse effect on blood picture

Leucocytozoon caulleryi (Akiba caulleryi)
Toriumi, T.; et al., 1978, Scient. Rep. Fac. Agric. Okayama Univ. (52), 49-53
Leucocytozoon caulleryi, chickens (exper.), sulfamonomethoxine and halofuginone in feed prevented infection

Leucocytozoon circaeti Sergent and Fabiani 1922
Greiner, E. C.; and Kocan, A. A., 1977, Canad. J. Zool., v. 55 (5), 761-770
as syn. of Leucocytozoon toddi Sambon 1908

Leucocytozoon dubreuili
Bennett, G. F.; Cameron, M.; and White, E., 1975, Canad. J. Zool., v. 53 (10), 1432-1442
hematozoa of passeriforms, prevalence, effect of climate, application of insecticide, and large-scale environmental alteration
Bombycilla cedrorum
Dumetella carolinensis
all from New Brunswick

Leucocytozoon dubreuili Mathis et Leger, 1910
Kairullaev, K. K., 1979, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (1), 19-21
Turdus philemelos
Phoenicurus phoenicurus
(blood of all): all from central course of Ural river, Priural region, Ural oblast

Leucocytozoon dubreuili
Peirce, M. A.; and Mead, C. J., 1977, J. Nat. Hist., v. 11 (6), 597-600
Turdus merula: near Tring, Hertfordshire

Leucocytozoon dubreuili, illus.
Wong, S. T. C.; and Desser, S. S., 1978, J. Protozool., v. 25 (3, pt. 2), 302-314
Leucocytozoon dubreuili, development of secondary schizonts in renal tubule cells of Turdus migratorius and profound parasite-induced changes in these cells, electron microscopy

Leucocytozoon franchini Franca 1927
Greiner, E. C.; and Kocan, A. A., 1977, Canad. J. Zool., v. 55 (5), 761-770
as syn. of Leucocytozoon toddi Sambon 1908

Leucocytozoon fringillinarum Woodcock, 1910
Kairullaev, K. K., 1979, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (1), 19-21
Fringilla coelebs
F. montifringilla
Coccothraustes coccothraustes
(blood of all): all from central course of Ural river, Priural region, Ural oblast

Leucocytozoon fringillinarum
Peirce, M. A.; and Mead, C. J., 1977, J. Nat. Hist., v. 11 (6), 597-600
Fringilla coelebs: near Tring, Hertfordshire

Leucocytozoon fringillinarum
Wink, M.; and Bennett, G. F., 1976, J. Wildlife Dis., v. 12 (4), 587-590
Lagonosticta rufopicta
Ploceus nigerrimus
all from Ghana

Leucocytozoon hirundinis Sergent and Sergent, 1905
Williams, N. A.; and Bennett, G. F., 1978, Canad. J. Zool., v. 56 (4, pt. 1), 596-607
Progne subis: Maryland or New Jersey

Leucocytozoon majoris Laveran, 1902
Kairullaev, K. K., 1979, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (1), 19-21
Parus major (blood): central course of Ural river, Priural region, Ural oblast

Leucocytozoon majoris
Peirce, M. A.; and Mead, C. J., 1977, J. Nat. Hist., v. 11 (6), 597-600
Parus major
P. caeruleus
all from near Tring, Hertfordshire

Leucocytozoon marchouxi Mathis & Leger
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
Syn.: Leucocytozoon turtur orientalis Yakunin, 1972

Leucocytozoon marchouxi Mathis and Leger, 1910
Williams, N. A.; and Bennett, G. F., 1978, Canad. J. Zool., v. 56 (4, pt. 1), 596-607
Zenaidura macroura: Maryland

Leucocytozoon martyi Commes 1918
Greiner, E. C.; and Kocan, A. A., 1977, Canad. J. Zool., v. 55 (5), 761-770
as syn. of Leucocytozoon toddi Sambon 1908

Leucocytozoon mathisi Franca 1912
Greiner, E. C.; and Kocan, A. A., 1977, Canad. J. Zool., v. 55 (5), 761-770
as syn. of Leucocytozoon toddi Sambon 1908

Leucocytozoon musajevi Zeiniev, 1975
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Leucocytozoon neavei
Wink, M.; and Bennett, G. F., 1976, J. Wildlife Dis., v. 12 (4), 587-590
Francolinus ahantensis: Ghana

Leucocytozoon simondi
Maley, G. J. M.; and Desser, S. S., 1977, Canad. J. Zool., v. 55 (2), 355-358
Leucocytozoon simondi, Pekin ducklings, quantification of anemia, gametocytemia, and osmotic fragility of erythrocytes

Leucocytozoon simondi
Tarshis, I. B., 1976, J. Med. Entom., v. 13 (3), 337-341
Cnephia ornithophilia, laboratory rearing and feeding, transmission of Leucocytozoon simondi to Anas platyrhynchos by bite and by intraperitoneal inoculation of triturated infected flies

Leucocytozoon simondi
Williams, N. A.; Calverley, B. K.; and Mahrt, J. L., 1977, J. Wildlife Dis., v. 13 (3), 226-229
Anas platyrhynchos
Anas acuta
(blood of all): all from Central Alberta and Mackenzie Delta, Northwest Territories

Leucocytozoon smithi
Alverson, D. R.; and Noblet, R., 1977, J. Med. Entom., v. 14 (1), 132-133
Leucocytozoon smithi, turkeys, spring relapse occurred in infected turkeys confined away from attack by simuliid vectors

Leucocytozoon smithi
Dick, J. W., 1978, Avian Dis., v. 22 (1), 82-85
Leucocytozoon smithi, turkey, persistence of gametocytes in peripheral blood for more than a year after a single short natural exposure: Marlboro and Sumter Counties, South Carolina

Leucocytozoon smithi
Gore, T. C.; and Noblet, G. P., 1978, Poultry Science, v. 57 (3), 603-607
Leucocytozoon smithi, turkeys, close correlation between turkey deep body temperature and L. smithi cyclic behavior under conditions of natural, reversed, and continuous light

Leucocytozoon smithi (Laveran and Lucet 1905)
Greiner, E. C.; and Forrester, D. J., 1979, J. Parasitol., v. 65 (2), 324-326
Leucocytozoon smithi in Simulium slossonae (nat. and exper.): Florida

Leucocytozoon smithi
Henry, C. W.; and Dick, J. W., 1978, Avian Dis., v. 22 (3), 542-546
Leucocytozoon smithi, isolation of gametocytes from whole peripheral turkey blood, Ficoll density-gradient system

Leucocytozoon smithi, illus.
Huchzermeyer, F. W.; and Sutherland, B., 1978, Avian Path., v. 7 (4), 645-649
turkeys (blood): near Onderstepoort, South Africa

Leucocytozoon smithi
Noblet, R.; Arnold, D. C.; and Snoddy, E. L., 1976, J. Econom. Entom., v. 69 (4), 481-483
Leucocytozoon smithi, distribution of Simulium vectors: South Carolina

Leucocytozoon tawaki, illus.
Allison, F. R.; Desser, S. S.; and Whitten, L. K., 1978, N. Zealand J. Zool., v. 5 (2), 371-374
Leucocytozoon tawaki, life cycle: schizogonic stages in Eudyptes pachyrhynchus described and parasitemia quantified; sporogonic stages in Austrosimulium ungulatum described; observations on transmission to penguin chicks: Jackson Head, south Westland
Eudyptes minor (blood) (exper.)

Leucocytozoon tawaki, illus.
Desser, S. S.; and Allison, F. B., 1979, J. Parasitol., v. 65 (5), 737-744
Leucocytozoon tawaki in Austrosimilium ungulatum, ultrastructure of sporogonic development, nature and significance of crystalloid inclusions

Leucocytozoon toddi Sambon 1908, illus.
Greiner, E. C.; and Kocan, A. A., 1977, Canad. J. Zool., v. 55 (5), 761-770
synonymy, redescription, variation
Accipiter badius: India
A. cooperi: United States
A. striatus: United States
A. tachiro: Tanzania
A. virgatus: Malaysia
Buteo brachypterus: Malagasy Republic (Madagascar)
B. jamaicensis: United States
B. lagopus: United States
B. lineatus: United States
B. regalis: United States
Falco peregrinus: Malaysia

Leucocytozoon toddi Sambon
Greiner, E. C.; and Mundy, P. J., 1979, J. Parasitol., v. 65 (1), 147-153
hematozoa from southern African vultures, prevalence, host age class, seasonal variation
Torgos tracheliotus
Gyps africanus
all from Rhodesia [and/or] South Africa

Leucocytozoon toddi Sambon, 1907
Williams, N. A.; and Bennett, G. F., 1978, Canad. J. Zool., v. 56 (4, pt. 1), 596-607
Buteo jamaicensis: Maryland

Leucocytozoon turtur orientalis Yakunin, 1972
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
as syn. of Leucocytozoon marchouxi Mathis & Leger

Leucocytozoon vulgaris Zeiniev, 1975
Peirce, M. A.; and Bennett, G. F., 1979, J. Protozool., v. 26 (3), 357-359
nomen nudum

Leucocytozoon ziemanni (Laveran), illus.
Kocan, A. A.; and Kocan, K. M., [1979], J. Parasitol., v. 64 (6), 1978, 1057-1059
Leucocytozoon ziemanni, elongate gametocytes, fine structure
Bubo virginianus: Oklahoma

Leucocytozoon ziemanni (Laveran, 1903)
Williams, N. A.; and Bennett, G. F., 1978, Canad. J. Zool., v. 56 (4, pt. 1), 596-607
Otus asio: Maryland

Lithocystis parguerae n. sp., illus.
Jones, I., 1971, Caribbean J. Sc., v. 11 (1-2), 47-50
[lapsus p. 47 as L. parquerae]
Siphonosoma cumanense (intestine): reefs in vicinity of La Parguera, Puerto Rico

Lithocystis parquerae [lapsus p. 47 for L. parguerae n. sp.]
Jones, I., 1971, Caribbean J. Sc., v. 11 (1-2), 47-50

Lomiella bergeri nova sp., illus.
de Puytorac, P.; and Rakotoarivelo, R., 1965, Arch. Zool. Exper. et Gen., v. 105 (2), 229-240
Pheretima rodericensis (tube digestif): Madagascar

Longa subgen. n.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31
subgen. of Cepedea
type sp.: Cepedea (Longa) longa (Bezzenberger, 1904) Metcalf, 1923

Longa
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
subgen. of Cepedea, key

Longa Amaro, 1966
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
subgen. of Cepedea
key to species

Lubetiella micheli de Puytorac, 1959, illus.
de Puytorac, P.; and Rakotoarivelo, R., 1965, Arch. Zool. Exper. et Gen., v. 105 (2), 229-240
description
Dichogaster inermis (intestin): Madagascar;
ile Maurice

Lucetina putorii (Railliet and Lucet, 1891) Henry and Leblois, 1926
Tadros, W.; and Laarman, J. J., 1978, Proc. K. Nederl. Akad. Wetensch., s. C, Biol. and Med. Sc., v. 81 (4), 466-468
as syn. of Sarcocystis putorii (Railliet and Lucet, 1891) comb. nov.

Madsenia Kahl, 1934
Iankovskii, A. V., 1973, Parazitologiia, Leningrad, v. 7 (3), 214-219
Hymenostomatida, Tetrahymenina, Entodiscidae fam. n.

Malamoeba locustae (Taylor and King), illus.
Hanrahan, S. A., 1973, Proc. Electron Microsc. Soc. South. Africa, v. 3, 19-20
Malamoeba locustae in laboratory bred Locusta migratoria (lumen of malpighian tubules and midgut), structure, effect on malpighian tubules

Malameba (=Malpighamoeba) locustae (King et Taylor 1936), illus.
Papillon, M.; and Cassier, P., 1978, Acrida, v. 7 (2), 101-114
Malameba locustae in Schistocerca gregaria (malpighian tubules), morphological and physiological disturbances due to M. locustae

Malaria
Ajao, O. G., 1978, J. Trop. Med. and Hyg., v. 81 (8), 153-155
malaria, overt attacks in humans as cause of post-operative fever, depressed acquired immunity resulting from stress of surgery, recommends routine administration of chloroquine prior to surgical procedures: Nigeria

Malaria
Alisangco-Gamez, L. L., 1979, Southeast Asian J. Trop. Med. and Pub. Health, v. 10 (2), 184-190
side effects of blood transfusions, humans, mentions malaria and other parasites: Philippines

Malaria
Amato Neto, V., 1973, Rev. Paul. Med., v. 81 (2), 93-104
human protozoan infections, drugs in current use, dosage recommendations, review

Malaria
Amato Neto, V.; et al., 1974, Rev. Soc. Brasil. Med. Trop., v. 8 (2), 83-86
human malarias, preliminary investigation of merits of testing blood donors for evidence of infection using the indirect immunofluorescence test

Malaria
Ambroise-Thomas, P., 1978, Israel J. Med. Sc., v. 14 (6), 690-691
malaria, diagnosis and seroepidemiologic study by immunofluorescence, indirect hemagglutination, and immunoenzymology, brief symposium presentation

Malaria
Andrianjafy, M.; Thuillier, M.; and Gandrille, M. C., 1976, Rev. Franc. Transfus. et Immuno-Hematol., v. 19 (4), 603-604
malaria in young woman given blood transfusions, donor of blood found to have high malarial antibody titers, successful treatment with nivaquine, case report: France

Malaria
Armon, P. J., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (3), 284-288
maternal deaths in the Kilimanjaro region, survey, includes information on amoebic colitis and malaria: Tanzania

Malaria
Arnold, K., 1978, Southeast Asian J. Trop. Med. and Pub. Health, v. 9 (2), 177-185
human parasitic diseases, trends in development of chemotherapy, review

Malaria
Azubuike, J. C.; Izuora, G. I.; and Obi, G. O., 1977, Trop. and Geogr. Med., v. 29 (4), 365-368
malaria and hookworm cited as causes of anemia in non-sickling children: Enugu region, Nigeria

Malaria
Bailey, D. L.; et al., 1979, Am. J. Trop. Med. and Hyg., v. 28 (5), 902-908
technology for sterilizing and packaging male Anopheles albimanus for field release in endemic malaria areas

Malaria
Barratt, T. M., 1979, Arch. Dis. Childhood, v. 54 (11), 825-830
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Service, M. W., 1977, Linn. Soc. Symposium Ser. (5), 151-164
human malaria, review of current control measures, problems in vector control

Malaria
Service, M. W.; Joshi, G. P.; and Pradhan, G. D., 1978, Ann. Trop. Med. and Parasitol., v. 72 (4), 377-386
survey of Anopheles gambiae and A. arabiensis (vectors of human malaria) in the Kisumu area after insecticidal spraying with fenitrothion: Kenya

Malaria
Service, M. W.; Martin, S. J. S.; and Invest, J. F., 1977, Cahiers O.R.S.T.O.M., s. Entom. Med. et Parasitol., v. 15 (3), 263-264
Anopheles moucheti as malaria vector: Lambarene, Gabon

Malaria
Shookhoff, H. B.; et al., 1979, Bull. N. York Acad. Med., v. 55 (4), 385-400
malaria, human, review of all cases reported in New York City between 1972 and 1977: geographical source of imported cases, age, sex, race, nationality, and occupational status of host; need for physicians to inform travelers of malaria risk and chemoprophylactic benefit

Malaria
Spencer, H. C.; et al., 1979, Am. J. Trop. Med. and Hyg., v. 28 (6), 927-932
malaria, human sera, micro enzyme-linked immunosorbent assay used with in vitro-cultured Plasmodium falciparum as antigen

Malaria
Spencer, H. C.; Collins, W. E.; and Skinner, J. C., 1979, Am. J. Trop. Med. and Hyg., v. 28 (6), 933-936
malaria, human sera, enzyme-linked immunosorbent assay using cultured Plasmodium falciparum as antigen compared with indirect fluorescent antibody test

Malaria
Spira, D. T., 1978, Israel J. Med. Sc., v. 14 (6), 707-711
immunodiagnosis of malaria, prospects for the future, symposium presentation

Malaria
Stella, V.; et al., 1978, J. Pharm. Sc., v. 67 (10), 1375-1377
α-(dibutylaminomethyl)-6,8-dichloro-2-(3', 4'-dichlorophenyl)-4-quinolinemethanol (an antimalarial), enhancement of bioavailability by formulation with oleic acid in soft gelatin capsule

Malaria
Stuerchler, D., 1978, Schweiz. Med. Wchnschr., v. 108 (46), 1810-1811
malaria, humans, epidemiology, risk of infection in travellers returning from Africa, importance of prophylaxis: Switzerland

Malaria
Stuiver, P. C., 1972, Tijdschr. Gastro-enterol., v. 15 (4), 238-247
tropical splenomegaly in humans thought to be lymphoproliferative disorder secondary to abnormal immunological reaction to malaria infection, prolonged antimalarial therapy treatment of choice

Malaria
Taylor, B., 1975, J. Med. Entom., v. 11 (6), 677-687
malaria vectors of the Anopheles punctulatus complex, historical background, distribution, effects of eradication programs, review: British Solomon Islands Protectorate

Malaria
de-The, G.; et al., 1978, Nature, London (5673), v. 274, 756-761
epidemiological evidence for causal relationship between Epstein-Barr virus and Burkitt's lymphoma from Ugandan prospective study, includes brief mention of possible association with malaria

Malaria
Thong, Y. H.; et al., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (4), 388-390
mefloquine (new antimalarial compound), effect on mitogen-induced human and mouse lymphocyte proliferative responses, effect on antibody responses and delayed-type hypersensitivity responses to sheep red blood cells in treated mice

Malaria
Thong, Y. H.; and Ferrante, A., 1978, Am. J. Trop. Med. and Hyg., v. 27 (2, pt. 1), 354-356
quinine, inhibition of mitogen-induced human lymphocyte proliferative responses, this immunosuppressive property may be undesirable side effect in treatment of malaria

Malaria
Toongsuwan, S.; Sovonnakote, T.; and Ariyamitr, P., 1979, Siriraj Hosp. Gaz., v. 31 (3), 386-390
malaria, pregnant woman living in malaria-free area who was exposed to infection while working in a laboratory that bred mosquitoes, case report, recommendations for diagnostic awareness

Malaria
Trigg, M. E., 1979, South. Med. J., v. 72 (5), 593-599
immune functions of the spleen, includes information on malaria with special reference to Plasmodium inui in rhesus monkeys

Malaria
Vaughan, J. P.; et al., 1977, Trop. and Geogr. Med., v. 29 (4), 369-373
hookworm and iron deficiency anemia in adults and children, therapy with tetrachlorethylene and oral iron, poor response to therapy by children suggests that malaria was causative factor: Tanzania

Malaria
Venkataraman, S.; Ahuja, G. K.; and Virmani, V., 1979, J. Ass. Physicians India, v. 27 (5), 421-428
cysticercosis, echinococcosis, malaria, differential diagnosis of neurological manifestations in humans

Malaria
Verhave, J. P., 1978, Israel J. Med. Sc., v. 14 (5), 599-600
malaria, introductory remarks on immunization with sporozoites, symposium presentation

Malaria
Verhave, J. P.; Meuwissen, J. H. E. T.; and Golenser, J., 1978, Israel J. Med. Sc., v. 14 (5), 611-616
arguments for the involvement of cell-mediated immunity in antisporozoite protection, symposium presentation

Malaria
Vittoz, P., 1977, Rev. Franc. Transfus. et Immuno-Hematol., v. 20 (1), 225-234
malaria, toxoplasmosis, screening of blood donors including those with possible parasitic infections

Malaria
Voller, A., 1976, Immunol. Parasit. Infect., 107-119
malaria, serodiagnosis, review

Malaria
Voller, A., 1978, New Trends and Develop. Vaccines, 299-309
schistosomiasis, trypanosomiasis, malaria, potential and progress towards vaccines, review

Malaria
Voorhoeve, A. M.; Muller, A. S.; and W'Oigo, H., 1979, Trop. and Geogr. Med., v. 31 (4), 607-627
agents, including malaria, that affect health of mothers and children in rural Kenya

Malaria
Wallach, D. F. H., 1979, Cell Biol. Internat. Rep., v. 3 (5), 395-408
malaria, membrane pathobiology, review

Malaria
Wise, D. L.; Gresser, J. D.; and McCormick, G. J., 1979, J. Pharm. and Pharmacol., v. 31 (4), 201-204
dual antimalarial system, sustained release of ^{3}H-labelled WR-7557 and ^{14}C-labelled WR-158122 in biodegradable carrier, rhesus monkeys, mice

Malaria
World Health Organization, 1975, WHO Chron., v. 29 (12), 474-481
human malaria, report of worldwide situation as of 1974

Malaria
World Health Organization. Division of Malaria and Other Parasitic Diseases, 1975, Manual on practical entomology in malaria. Part I. Vector bionomics and organization of anti-malaria activities, 160 pp., illus.
bionomics of mosquito populations and organization of anti-malaria activities

Malaria
World Health Organization. Division of Malaria and Other Parasitic Diseases, 1975, Manual on practical entomology in malaria. Part II. Methods and techniques, 191 pp., illus.
malaria, methods and techniques for collecting and studying vector mosquitoes

Malaria
Yekutiel, P., 1979, Israel J. Med. Sc., v. 15 (12), 976-982
infective diseases in Israel, changing patterns over 30 years, includes malaria, leishmaniasis, tick and flea borne infections

Marteilia
Desportes, I.; and Ginsburger-Vogel, T., 1977, Compt. Rend. Acad. Sc., Paris, v. 285, s. D, Sc. Nat., (11), 1111-1114
Marteilia, development, taxonomic implications, affinities with Myxosporida, Actinomyxida, and Paramyxida, "C'est dans ce sous-embranchement [Cnidospora] qui nous semble devoir etre maintenu et completement redefini que s'inserent en toute logique les Marteiliidae. . . . nous proposons pour l'instant de creer le nouvel et quatrieme ordre des Marteiliidae."

Marteilia sp., illus.
Desportes, I.; and Ginsburger-Vogel, T., 1977, Compt. Rend. Acad. Sc., Paris, v. 285, s. D, Sc. Nat., (11), 1111-1114
presumed sporogenesis

Marteiliidae
Desportes, I.; and Ginsburger-Vogel, T., 1977, Compt. Rend. Acad. Sc., Paris, v. 285, s. D, Sc. Nat., (11), 1111-1114
Marteilia, development, taxonomic implications, affinities with Myxosporida, Actinomyxida, and Paramyxida, "C'est dans ce sous-embranchement [Cnidospora] qui nous semble devoir etre maintenu et completement redefini que s'inserent en toute logique les Marteiliidae. . . . nous proposons pour l'instant de creer le nouvel et quatrieme ordre des Marteiliidae."

Mattesia sp.
Miln, A. J., 1978, N. Zealand Entom., v. 6 (4), 392-399
Mattesia sp. and Nosema takapauensis in Costelytra zealandica, incidence among larvae, seasonal distribution, growth and development of diseased larvae, mortality, transmission by soil: New Zealand

Mattesia geminata sp. n., illus.
Jouvenaz, D. P.; and Anthony, D. W., 1979, J. Protozool., v. 26 (3), 354-356
Mattesia geminata sp. n.-infected Solenopsis geminata, pathologic manifestations, biological control implications
Solenopsis geminata (hypodermis, apparently in oenocytes): Gainesville, Alachua Co., Florida
Solenopsis sp.: Brazil

Mattesia trogodermae Canning
Shapas, T. J.; Burkholder, W. E.; and Boush, G. M., 1977, J. Econom. Entom., v. 70 (4), 469-474
Mattesia trogodermae in biological control of Trogoderma glabrum, dissemination of protozoan pathogen by pheromone-baited spore-transfer sites

Maupasella Cepede
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key

Maupasella dichogasteri, nova sp., illus.
de Puytorac, P.; and Rakotoarivelo, R., 1965, Arch. Zool. Exper. et Gen., v. 105 (2), 229-240
Dichogaster inermis (partie anterieure et moyenne du tube digestif): ile Maurice

Maupasella dilatata, nova sp., illus.
de Puytorac, P.; and Rakotoarivelo, R., 1965, Arch. Zool. Exper. et Gen., v. 105 (2), 229-240
Dichogaster inermis (tube digestif): ile Maurice

Maupasella vacuolata Lom, 1959, illus.
de Puytorac, P.; and Rakotoarivelo, R., 1965, Arch. Zool. Exper. et Gen., v. 105 (2), 229-240
description
Pheretima hawayana (partie anterieure du tube digestif): Madagascar

Megatrypanum, subgenus
Godfrey, D. G., 1977, Protozoology, v. 3, 33-49
identification of morphologically similar trypanosomes of mammals

Merocystis sp. Paichuk, 1953
Levine, N. D., 1977, Protozoology, v. 3, 173-182
as syn. of Adelina sp. (Paichuk, 1953) nov. comb.

Mesastasia gen. n.
Palienko, L. P.; and Monchenko, V. I., 1979, Vestnik Zool., Akad. Nauk Ukrainsk. SSR, Inst. Zool. (6), 44-49
Euglenoididae
tod: M. mirabilis sp. n.

Mesastasia mirabilis sp. n. (tod), illus.
Palienko, L. P.; and Monchenko, V. I., 1979, Vestnik Zool., Akad. Nauk Ukrainsk. SSR, Inst. Zool. (6), 44-49
Mesastasia mirabilis sp. n., life cycle
Thermocyclops crassus
T. dybowskii
Mesocyclops leuckarti
(intestine of all): all from ponds in boundaries of Kiev, Ukrainian SSR

Mesnilella Cepede
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key

Metamera Duke, 1910 (type genus of fam.)
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
Metameridae fam. n.
diagnosis

Metameridae fam. n.
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
Apicomplexa
type genus: Metamera Duke, 1910

Metaradiophrya sp., illus.
Williams, G. W., 1942, J. Morphol., v. 70 (3), 545-589
earthworm (anterior midgut): wooded section, near Philadelphia, Pennsylvania

Metaradiophrya chlorotica n. sp., illus.
Williams, G. W., 1942, J. Morphol., v. 70 (3), 545-589
Helodrilus chlorotica (anterior section of mid-gut): banks of Tygart river, Elkins, West Virginia

Metaradiophrya falcifera Stein, illus.
Williams, G. W., 1942, J. Morphol., v. 70 (3), 545-589
description
Helodrilus chlorotica
H. calignosus (?)
(anterior section of mid-gut of all): all from banks of Tygart river, Elkins, West Virginia

Metaradiophrya lumbrici (Dujardin), illus.
Williams, G. W., 1942, J. Morphol., v. 70 (3), 545-589
Metaradiophrya lumbrici, detailed description, movement and attachment behavior, cytology of division; description of other Metaradiophrya spp. and comparison with M. lumbrici
Eisenia foetidus: botanical gardens of the University of Pennsylvania

Metaradiophrya wenrichi n. sp., illus.
Williams, G. W., 1942, J. Morphol., v. 70 (3), 545-589
earthworm (anterior midgut): cow manure heap, Institute, West Virginia

Microsporidia, illus.
Doroshenko, M. A.; and Pinchuk, L. E., 1978, Biol. Moria, Vladivostok (3), 88
Microsporidia, mucous glands in olfactory epithelium of some marine fishes are distinguishable morphologically and histochemically from epithelial cells affected by parasites

Microsporida
Palmieri, J. R.; and Sullivan, J. T., 1977, J. Invert. Path., v. 30 (2), 276
Microsporida, staining technique for location of spores in host tissues, used to locate Nosema sp. in Lymnaea rubiginosa snails and Tracheophilus sp. rediae

Microsporida, illus.
Shadduck, J. A.; Kelsoe, G.; and Helmke, R. J., 1979, J. Parasitol., v. 65 (1), 185-188
microsporida spontaneously contaminating cell culture prepared from baboon placental cells, ultrastructurally identical to Encephalitozoon cuniculi and different from Nosema connori

Microsporidia
Stunkard, H. W., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 725-739
Sporozoa, review: early taxonomic concepts, developmental cycles and taxonomic revisions, host-parasite relations, microsporidian infections of fishes

Microsporidia
Voronin, V. N.; and Issi, I. V., 1974, Parazitologiia, Leningrad, v. 8 (3), 272-273
Microsporidia, brief summary of some laboratory and field techniques

[Microsporidia sp.] mikrosporidiiami
Alikhanov, Sh. G., 1972, Parazitologiia, Leningrad, v. 6 (4), 381-384
Aedes vexans
Culex pipiens pipiens
C. theileri
all from Azerbaidzhan

Microsporidia [sp.], illus.
Berrebi, P.; and Bouix, G., 1978, Ann. Parasitol., v. 53 (1), 1-20
Microsporidia [sp.] probably new, cysts and xenomas (structure, localization, histology), epidemiology (geographic distribution, and and sex of host, seasonal distribution)
Atherina boyeri (paroi du tube digestif, cavite generale): lagunes saumatres du sud de Montpellier jusqu'a l'etang de Berre

Microsporidia [sp.]
Gaevskaia, A. V.; and Umnova, B. A., 1977, Biol. Moria, Vladivostok (4), 40-48
Limanda ferruginea
Glyptocephalus cynoglossus
all from near Nova Scotia, Northwest Atlantic

Microsporidia [sp.]
Gaevskaia, A. V.; and Umnova, B. A., 1977, Biol. Moria, Vladivostok (4), 40-48
Glyptocephalus cynoglossus (muscles, body): near Nova Scotia, Northwest Atlantic

Microsporidia gen. sp., illus.
Grabda, J., 1977, Acta Ichthyol. et Piscat., v. 7 (2), 15-34
degree of parasite infestation of Theragra chalcogramma (muscles), commercial value: imported from USSR

Microsporidia gen. sp.
Grozdilova, T. A., 1974, Parazitologiia, Leningrad, v. 8 (4), 293-298
Oncorhynchus gorbuscha: Umba [and/or] Keret rivers

Microsporidia sp. or spp., illus.
Hembree, S. C., 1979, Mosquito News, v. 39 (3), 575-582
Culex sp. (fat body): Thailand

Microsporidia [sp.], illus.
Jafri, R. H.; Ahmad, M.; and Idrees, K., 1976, Pakistan J. Zool., v. 8 (2), 234-236
Microcerotermes championi (fat body, lumen of proventriculus, body cavity): near Punjab University, New Campus, Lahore

Microsporida [sp.], illus.
Novilla, M. N.; and Kwapien, R. P., 1978, Avian Dis., v. 22 (1), 198-204
Microsporida [sp.] in Agapornis roseicollis (liver, kidney, small intestine), case history, pathology: purchased from commercial aviary

Microsporidia [sp.], illus.
Schuetz, A. W.; Selman, K.; and Samson, D., 1978, J. Exper. Zool., v. 204 (1), 81-94
Microsporidia [sp.]-infected Rana pipiens, class of unique abnormally enlarged and discolored follicles and oocytes identified in ovaries, various parameters of composition, function, and structure of these large oocytes: Vermont

Microsporidian, illus.
Hembree, S. C., 1979, Mosquito News, v. 39 (3), 575-582
Anopheles vagus (fat body): Thailand

Microsporidian, illus.
Hembree, S. C., 1979, Mosquito News, v. 39 (3), 575-582
Culex quinquefasciatus (fat body) (nat. and exper.): Thailand

Microsporidian species No. 1, illus.
Hembree, S. C., 1979, Mosquito News, v. 39 (3), 575-582
Aedes aegypti (fat body) (nat. and exper.): Thailand

Microsporidian species No. 2, illus.
Hembree, S. C., 1979, Mosquito News, v. 39 (3), 575-582
Aedes aegypti (midgut epithelium): Thailand

Microsporidian species No. 3, illus.
Hembree, S. C., 1979, Mosquito News, v. 39 (3), 575-582
Aedes aegypti: Thailand

Microsporidian
Paperna, I.; Sabnai, I.; and Castel, M., 1978, Ann. Parasitol., v. 53 (2), 123-130
microsporidian infection (tentatively Pleistophora) reported in Liza ramada in fibroblasts of metacercarial capsule of Heterophyes heterophyes, infection of metacercarial cyst resulted in hypertrophy of cyst wall and degeneration and eventual death of encapsulated metacercaria: Bardawil Lagoon, Mediterranean coast of Sinai Peninsula

Microsporidium aplysiae sp. n., illus.
Krauhs, J. M.; Long, J. L.; and Baur, P. S., jr., 1979, J. Protozool., v. 26 (1), 43-46
Aplysia californica (nucleus and cytoplasm of abdominal ganglion neurons): intertidal zone of Pacific Ocean, from Bodega Bay (California) to Baja Peninsula

Microsporidium giraudi (Leger and Hesse) (= Stempellia muelleri)
Bulnheim, H. P., 1978, Helgoland. Wissensch. Meeresuntersuchg., v. 31 (1-2), 1-33
Gammarus duebeni duebeni: Scotland
G. duebeni celticus: United Kingdom

Miescheria muris Blanchard, 1885
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
as syn. of Sarcocystis muris (Blanchard, 1885) Labbe, 1899

Minchinia sp., illus.
Hillman, R. E., 1978, J. Invert. Path., v. 31 (2), 265-266
Teredo navalis
T. bartschi
T. furcifera
(gills of all): all from Barnegat Bay, New Jersey

Minchinia sp. Marchand, 1975
Marchand, J.; and Sprague, V., 1979, J. Protozool., v. 26 (2), 179-185
as syn. of Minchinia cadomensis sp. n.

Minchinia armoricana nov. sp.
van Banning, P., 1977, J. Invert. Path., v. 30 (2), 199-206
pathogenic activity
Ostrea edulis (connective tissue): oyster culture of Yerseke, Netherlands, but originated in oyster areas of Brittany, France

Minchinia cadomensis sp. n., illus.
Marchand, J.; and Sprague, V., 1979, J. Protozool., v. 26 (2), 179-185
Syn.: Minchinia sp. Marchand, 1975
Rhithropanopeus harrisii tridentatus (cavite generale, sinus sanguins): Ouistreham (Canal de Caen a la Mer), Calvados, France

Minchinia chitonis Lankester
Baxter, J. M.; and Jones, A. M., 1978, Marine Biol., v. 46 (4), 305-313
Minchinia chitonis-infected mollusc, Lepidochitona cinereus, host growth and population structure, infection caused enhanced growth and deviation from normal growth curve
Lepidochitona cinereus: Easthaven, Scotland

Minchinia costalis
Couch, J. A.; and Rosenfield, A., 1968, Proc. 1967 National Shellfish Ass., v. 58, 51-59
Minchinia nelsoni and M. costalis in Crassostrea virginica introduced from non-infected area, incidence, annual and seasonal prevalence, mortality, sporulation, effect on host reproductive capacity, comparison with Delaware and lower Chesapeake Bays, practical implications for manipulation of introduced oysters: Chincoteague Bay, at Franklin City, Virginia

[Minchinia nelsoni] MSX
Andrews, J. D., 1964, Proc. 1962 National Shellfish Ass., v. 53, 65-84
[Minchinia nelsoni] MSX, cause of oyster mortality and poor seed quality, monitoring program, MSX activity in low- and high-salinity areas: James River public seed beds, Virginia

Minchinia nelsoni
  Andrews, J. D., 1967, Proc. 1966 National Shellfish. Ass., v. 57, 38-49
    Minchinia nelsoni (MSX), interaction with fungus pathogen Dermocystidium marinum, oysters, MSX prevents D. marinum from becoming epizootic

Minchinia nelsoni
  Andrews, J. D., 1968, Proc. 1967 National Shellfish Ass., v. 58, 23-26
    Minchinia nelsoni in Crassostrea virginica (susceptible imports, native oysters and progeny), seasonal patterns of morbidity and mortality, survival of early generations in MSX-prevalent areas suggests that acquired resistance is involved; hypotheses on origin of infection and life cycle of pathogen: Virginia waters

Minchinia nelsoni
  Couch, J. A.; and Rosenfield, A., 1968, Proc. 1967 National Shellfish Ass., v. 58, 51-59
    Minchinia nelsoni and M. costalis in Crassostrea virginica introduced from non-infected area, incidence, annual and seasonal prevalence, mortality, sporulation, effect on host reproductive capacity, comparison with Delaware and lower Chesapeake Bays, practical implications for manipulation of introduced oysters: Chincoteague Bay, at Franklin City, Virginia

Minchinia nelsoni
  Krantz, G. E.; et al., 1972, Proc. 1971 National Shellfish. Ass., v. 62, 83-85
    Crassostrea virginica: Wellfleet Harbor, Massachusetts

Minchinia nelsoni Haskin, Stauber and Mackin, illus.
  Myhre, J. L., 1969, Proc. 1968 National Shellfish Ass., v. 59, 52-59
    Minchinia nelsoni, nucleic acid and nucleic protein patterns in plasmodial stages

Mitraspora caspialosae Dogiel, 1939
  Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
    as syn. of Sphaerospora caspialosae (Dogiel, 1939)

Mitraspora caudata (Parisi)
  Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
    as syn. of Sphaerospora caudata Parisi, 1910

Molluskocystis gen. n.
  Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
    Gregarinidae
    tod: M. pterotracheae (Stuart, 1871) comb. n.

Molluskocystis pterotracheae (Stuart, 1871) comb. n. (tod)
  Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
    Syns.: Zygocystis pterotracheae Stuart, 1871; Gregarina pterotracheae (Stuart, 1971) Labbe (1899)
    Pterotrachea sp. (coelom): Odessa, U.S.S.R.

Monocercomonas Grassi, 1879
  Krishnamurthy, R.; and Sultana, T., 1976, Marathwada Univ. J. Sc. (Nat. Sc.), v. 15 (8), 132-141
    key to species from insects
    includes: Monocercomonas corydiae Bhaskar Rao 1970; M. osmaniae Bhaskar Rao, 1970; M. orthopterorum Parisi, 1910; M. digranulae n. sp.; M. phyllophagae Travis & Becker, 1931; M. trichopterae Mackinnon, 1910; M. laccotrephis Bhaskar Rao, 1970; M. axostylis Kirby, 1931; M. mackinnoni Kowlczyk, 1938; M. termitis Bernstein, 1928; M. passali Tanabe, 1926; M. leucophaeae Bhaskar Rao, 1970; M. indica n. sp.

Monocercomonas digranulaae [lapsus p. 133 for M. digranulae n. sp.]
  Krishnamurthy, R.; and Sultana, T., 1976, Marathwada Univ. J. Sc. (Nat. Sc.), v. 15 (8), 132-141

Monocercomonas digranulae n. sp.
  Krishnamurthy, R.; and Sultana, T., 1976, Marathwada Univ. J. Sc. (Nat. Sc.), v. 15 (8), 132-141
    key
    [lapsus p. 133 as M. digranulaae]
    Gryllotalpa africana (middle and hind gut): Aurangabad, Maharashtra, India

Monocercomonas (Monocercomonas) garnhami Krishnamurthy, 1968, illus.
  Madre, V. E.; and Krishnamurthy, R., 1976, Marathwada Univ. J. Sc. (Nat. Sc.), v. 15 (8), 143-147
    description
    Vipera russeli (rectum): Aurangabad, Maharashtra, India

Monocercomonas indica n. sp., illus.
  Krishnamurthy, R.; and Sultana, T., 1976, Marathwada Univ. J. Sc. (Nat. Sc.), v. 15 (8), 132-141
    key
    Pycnoscelus surinamensis (midgut): Aurangabad, Maharashtra, India

Monocercomonoides krishnamurthii n. sp., illus.
  Sultana, T., 1976, Marathwada Univ. J. Sc. (Nat. Sc.), v. 15 (8), 149-152
    Pycnoscelus surinamensis (hind gut): Aurangabad, Maharashtra, India

Monocercomonoides viperae n. sp., illus.
  Madre, V. E.; and Krishnamurthy, R., 1976, Marathwada Univ. J. Sc. (Nat. Sc.), v. 15 (8), 143-147
    Vipera russeli (rectum): Aurangabad, Maharashtra, India

Monocystis sp.
  Farmer, J. N.; et al., 1978, Vet. Rec., v. 102 (4), 78-80
    Vulpes vulpes (feces): North Wales

Monocystis abegbei sp. n., illus.
  Segun, A. O., 1978, J. Protozool., v. 25 (2), 157-162
    Libyodrilus violaceus (seminal vesicles, perivisceral coelom): Ibadan, Western Nigeria

Monocystis agilis Stein, 1848, illus.
  Barajas de Lopez, E., [1967], Rev. Soc. Mexicana Hist. Nat., v. 27, 1966, 29-36
    Monocystis agilis, morphology, life cycle
    Allolobophora longa (vesiculas seminales): bosque de Chapultepec, ciudad de Mexico, D. F.

Monocystis libyodrilii sp. n., illus.
Segun, A. O., 1978, J. Protozool., v. 25 (2), 157-162
Libyodrilus violaceus (perivisceral coelom): Ibadan, Western Nigeria

Monocystis pellucida (von Koelliker, 1848) Lankester, 1863
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702
as syn. of Lecudina pellucida (von Koelliker, 1848) Mingazzini, 1891

Monocystis pellucida (von Koelliker, 1848) Leger, 1893
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702
as syn. of Lecudina pellucida (von Koelliker, 1848) Mingazzini, 1891

Monocystis pontodrili n. sp.
Subba Rao, B. V. S. S. R.; Kalavati, C.; and Narasimhamurti, C. C., 1976, Proc. 1. Internat. Colloquium Invert. Path. and 9. Ann. Meet. Soc. Invert. Path. (Kingston, Canada, Aug. 29-Sept. 3), 390
Pontodrilus bermudensis (body musculature, seminal vesicles, coelomic fluid)

Monodontophrya Vejdovsky
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key

Mononema Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
Peranemidae, key
key to species

Mononema hanoiense Mich. et Wita
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Mononema harpacticdens Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Mononema ovale Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Mononema ovorum Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Mononema perpendiculare Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Mononema perpendiculare, illus.
Michajlow, W., 1977, Zhurnal Obsh. Biol., v. 38 (5), 657-675

Mononema reptans Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Mononema reptans, illus.
Michajlow, W., 1977, Zhurnal Obsh. Biol., v. 38 (5), 657-675

Mononema rostratum Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Mononema semilunare Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Mononema svennicum Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Mononema trepidans Mich.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key

Mrazekia bacilliformis (Leger et Hesse, 1922)
Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135
as syn. of Toxoglugea bacilliformis (Leger et Hesse, 1922) comb. nov.

Mrazekia brevicauda Leger et Hesse, 1916, illus.
Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135

Mrazekia limnodrili Jirovec, 1936, illus.
Janiszewska, J.; Kassner, J.; and Modrzejewska, M., 1978, Zool. Polon., v. 27 (1), 135-143
Mrazekia limnodrili, fine structure of surface of infected phagocytes of Limnodrilus hoffmeisteri: Odra River bank, Wroclaw

Mrazekia tetraspora (Leger et Hesse, 1922)
Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135
as syn. of Toxoglugea tetraspora (Leger et Hesse, 1922) comb. nov.

Mrazekiella Kijenskij
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key

MSX. See Minchinia nelsoni.

Myxidi[um], illus.
Koerting, W., 1977, Fisch u. Umwelt (4), 37-48
fish parasites, histopathological changes

Myxidium acinum n. sp., illus.
Hine, P. M., 1975, J. Roy. Soc. N. Zealand, v. 5 (2), 153-161
Anguilla australis
A. dieffenbachii
(gills of both): all from New Zealand

Myxidium acinum, illus.
Hine, P. M., 1979, N. Zealand J. Marine and Freshwater Research, v. 13 (2), 215-223
Myxidium zealandicum in Anguilla spp., factors affecting size and shape of spores, comparison with M. acinum
Anguilla dieffenbachii (gills): Makara Stream, Wellington, New Zealand

Myxidium gasterostei Noble
Lester, R. J. G., 1974, Syesis, v. 7, 195-200
Gasterosteus aculeatus (intestine, gall bladder): near Vancouver, British Columbia

Myxidium giardi
Jackson, P. B. N., 1978, J. South African Vet. Ass., v. 49 (1), 57-59
parasites of Anguilla mossambica, importance of disease control in aquaculture
Anguilla mossambica (gills): South Africa

Myxidium lieberkuehni Buetschli, 1882
Arthur, J. R.; Margolis, L.; and Arai, H. P., 1976, J. Fish. Research Bd. Canada, v. 33 (11), 2489-2499
Esox lucius (urinary bladder): Aishihik Lake and Stevens Lake, Yukon Territory

Myxidium lieberkuehni Butschli
Iziumova, N. A.; Mashtakov, A. V.; and Kashkovskii, V. V., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36) 64-68
[Esox lucius] (urinary bladder): Kamsk reservoir

Myxidium lieberkuehni Buetschli
Mashtakov, A. V.; et al., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36), 68-71
[Esox lucius] (urinary bladder): Votkinsk reservoir

Myxidium macrocapsularis Reuss
Iziumova, N. A.; Mashtakov, A. V.; and Timoshechkina, L. G., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36), 71-76
[Abramis brama]: Upper Kama

Myxidium macrocapsulare Auerbach, 1910, illus.
Mitchell, L. G., 1978, J. Protozool., v. 25 (1), 100-105
diagnosis
Notropis dorsalis
Catostomus commersonii
Ictalurus punctatus
Lepomis macrochirus
(gall bladder of all): all from Iowa

Myxidium minteri
Olson, R. E., 1978, Calif. Fish and Game, v. 64 (2), 117-120
Oncorhynchus kisutch (gall bladder): Pacific Ocean off Newport, Oregon

Myxidium ophiocephali Achmerov
Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
parasites of fishes: China

Myxidium polymorphus Nie et Lee
Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
parasites of fishes: China

Myxidium rhodei Leger
Iziumova, N. A.; Mashtakov, A. V.; and Timoshechkina, L. G., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36), 71-76
[Rutilus rutilus]: Upper Kama

Myxidium rostowstschikowi Schulman, 1962
Ashurova, M., 1973, Parazitologiia, Leningrad, v. 7 (2), 164-168
Schizothorax intermedius: Sarez Lake, central Pamir

Myxidium salvelini
Makhovenko, E. T., 1972, Parazitologiia, Leningrad, v. 6 (4), 369-375
Salvelinus alpinus inhabiting different ecological niches, possible use of differences in parasite fauna between groups as biological tags: Lake Azabach'e, Kamchatka

Myxidium serum n. sp., illus.
Hine, P. M., 1975, J. Roy. Soc. N. Zealand, v. 5 (2), 153-161
Anguilla dieffenbachii (alimentary musculature, sub-mucosa; mesenteric connective tissue): Burlings Stream, Wairarapa and Rough'ns Stream, Golden Downs State Forest, near Nelson, New Zealand

Myxidium zealandicum n. sp., illus.
Hine, P. M., 1975, J. Roy. Soc. N. Zealand, v. 5 (2), 153-161
Anguilla australis
A. dieffenbachii
(gills of both): all from New Zealand

Myxidium zealandicum Hine, 1975, illus.
Hine, P. M., 1979, N. Zealand J. Marine and Freshwater Research, v. 13 (2), 215-223
Myxidium zealandicum in Anguilla spp., factors affecting size and shape of spores, comparison with M. acinum
Anguilla dieffenbachii (gills)
A. australis (gills, gill arch)
all from Makara Stream, Wellington, New Zealand

Myxidium zealandicum, illus.
Hulbert, W. C.; et al., 1977, Canad. J. Zool., v. 55 (2), 438-447
Myxidium zealandicum, fine structure of sporogony and polar capsule development, trophozoite and cyst envelope structures

Myxidium zealandicum Hine, 1975, illus.
Komourdjian, M. P.; et al., 1977, Canad. J. Zool., v. 55 (1), 52-59
Myxidium zealandicum, description, histopathology, spore morphology
Anguilla rostrata (gills, kidney): St. Lawrence River near Quebec City and Cornwall

Myxobilatus sp.
Lester, R. J. G., 1974, Syesis, v. 7, 195-200
Gasterosteus aculeatus (kidney tubules): near Vancouver, British Columbia

Myxobilatus gasterostei (Parisi)
Lester, R. J. G., 1974, Syesis, v. 7, 195-200
Gasterosteus aculeatus (kidney tubules, urinary bladder): near Vancouver, British Columbia

Myxobilatus mictosporus
Riley, D. M., 1978, Tr. Am. Fish. Soc., v. 107 (1), 207-212
Micropterus salmoides: Florida

Myxobilatus ohioensis (Herrick, 1941) Davis, 1944
Cone, D. K.; and Anderson, R. C., 1977, Canad. J. Zool., v. 55 (9), 1410-1423
parasites of Lepomis gibbosus, prevalence and intensity in relation to host age and sex
Lepomis gibbosus (lumen of ureters, urinary bladder): Ryan Lake, Algonquin Park, Ontario

Myxobilatus platessae (Basikalowa, 1932), illus.
  Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
    synonymy
    Platichthys flesus luscus (urinary bladder): Black Sea

Myxobilatus yukonensis Arthur and Margolis, 1975
  Arthur, J. R.; Margolis, L.; and Arai, H. P., 1976, J. Fish. Research Bd. Canada, v. 33 (11), 2489-2499
    Cottus cognatus (kidney tubules, urinary bladder): Aishihik Lake, Yukon Territory

Myxobolus sp.
  Arthur, J. R.; Margolis, L.; and Arai, H. P., 1976, J. Fish. Research Bd. Canada, v. 33 (11), 2489-2499
    Cottus cognatus (stomach): Aishihik Lake, Yukon Territory

Myxobolus sp., illus.
  Desser, S. S.; and Paterson, W. B., 1978, J. Protozool., v. 25 (3, pt. 2), 314-326
    Myxobolus sp. in Notropis cornutus, spores and various stages of sporogenesis, ultrastructural and cytochemical observations

Myxobolus sp., illus.
  Ewers, W. H., 1973, Science N. Guinea, v. 1 (2), 16-20
    Litoria thesaurensis (testes): Lae, north coast of Papua New Guinea

Myxobolus sp.
  Markiw, M. E.; and Wolf, K., 1978, J. Fish. Research Bd. Canada, v. 35 (6), 828-832
    Myxosoma cerebralis, rabbits immunized with antigens extracted from mature spores or prespore stages, antisera and globulins used in fluorescent antibody techniques, direct fluorescent antibody test showed higher specificity than indirect FAT in cross reactions with other species of myxosporidians

Myxobolus sp.
  Mashtakov, A. V.; et al., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36), 68-71
    [Perca fluviatilis] (gills): Votkinsk reservoir

Myxobolus spp.
  Mitchell, L. G., 1978, J. Protozool., v. 25 (1), 100-105
    Campostoma anomalum (intestinal wall, gall bladder)
    Notropis cornutus (musculature, gall bladder, gill mucus)
    Notropis dorsalis (gall bladder, liver)
    Pimephales notatus (gills, mesenteries surrounding gall bladder)
    Semotilus atromaculatus (gall bladder, gills)
    Catostomus commersonii (gills)
    Carpiodes carpio (gills)
    Moxostoma macrolepidotum (gills)
    Pomoxis nigromaculatus (intestinal wall)
    all from Iowa

Myxobolus spp.
  Paperna, I.; and Baudin Laurencin, F., 1979, Aquaculture, v. 16 (2), 173-175
    Dicentrarchus labrax (gall bladder): marine cultures in France

Myxobolus sp.
  Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
    parasites of fishes: China

Myxobolus sp., illus.
  Wyatt, E. J., 1979, J. Protozool., v. 26 (1), 47-51
    description
    Cottus aleuticus (small intestine): Benton County, Oregon

Myxobolus albovi
  Podlipaev, S. A., 1974, Parazitologiia, Leningrad, v. 8 (6), 535-542
    Myxobolidae spp., polysaccharides in spores, nature of iodinophilous vacuole

Myxobolus argenteus
  Markiw, M. E.; and Wolf, K., 1978, J. Fish. Research Bd. Canada, v. 35 (6), 828-832
    Myxosoma cerebralis, rabbits immunized with antigens extracted from mature spores or prespore stages, antisera and globulins used in fluorescent antibody techniques, direct fluorescent antibody test showed higher specificity than indirect FAT in cross reactions with other species of myxosporidians

Myxobolus aristichthydis Nie et Yin
  Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
    parasites of fishes: China

Myxobolus bramae Reuss
  Iziumova, N. A.; Mashtakov, A. V.; and Timoshechkina, L. G., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36), 71-76
    [Abramis brama]
    [Rutilus rutilus]
    all from Upper Kama

Myxobolus bramae
  Linnik, V. Ia.; and Zen'kovich, E. M., 1970, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 8, 109-114
    [Abramis brama] (gills): Neman river basin

Myxobolus bramae
  Otvodova, G. D., 1975, Izvest. Gosudarstv. Nauchno-Issled. Inst. Ozer. i Rech. Ryb. Koziaistva, Leningrad, v. 93, 112-116
    [Abramis brama] (gills): Pskov-Chudskoe lake

Myxobolus bramae
  Podlipaev, S. A., 1974, Parazitologiia, Leningrad, v. 8 (6), 535-542
    Myxobolidae spp., polysaccharides in spores, nature of iodinophilous vacuole

Myxobolus cheni Schulman
  Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
    parasites of fishes: China

Myxobolus chimbuensis n. sp., illus.
  Ewers, W. H., 1973, Science N. Guinea, v. 1 (2), 16-20
    Litoria darlingtoni (testes): Mul, Chimbu District, Papua New Guinea

Myxobolus cybinae sp. n., illus.
  Mitenev, V. K., 1971, Parazitologiia, Leningrad, v. 5 (6), 556-558
    [Phoxinus] (cerebrum): Ponoy River near Ponoy Village, Kola Peninsula

Myxobolus cycloides Gurley, 1893, part.
Mitenev, V. K., 1971, Parazitologiia, Leningrad, v. 5 (6), 556-558
as syn. of M. lotae sp. n.

Myxobolus cyprinicola, illus.
Podlipaev, S. A., 1972, Parazitologiia, Leningrad, v. 6 (6), 506-508
Myxosporidia, 6 spp., lipid inclusions in spores and cysts

Myxobolus cyprinicola Reuss
Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
parasites of fishes: China

Myxobolus dechtiari Cone and Anderson, 1977
Cone, D. K.; and Anderson, R. C., 1977, Canad. J. Zool., v. 55 (9), 1410-1423
parasites of Lepomis gibbosus, prevalence and intensity in relation to host age and sex
Lepomis gibbosus (distal end of gill lamellae): Ryan Lake, Algonquin Park, Ontario

Myxobolus dispar
Linnik, V. Ia.; and Zen'kovich, E. M., 1970, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 8, 109-114
[Rutilus rutilus]: Neman river basin

Myxobolus dispar, illus.
Podlipaev, S. A., 1972, Parazitologiia, Leningrad, v. 6 (6), 506-508
Myxosporidia, 6 spp., lipid inclusions in spores and cysts

Myxobolus dispar
Podlipaev, S. A., 1974, Parazitologiia, Leningrad, v. 8 (6), 535-542
Myxobolidae spp., polysaccharides in spores, nature of iodinophilous vacuole

Myxobolus dispar Thelohan
Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
parasites of fishes: China

Myxobolus disparoides Schulman, 1962
Ashurova, M., 1973, Parazitologiia, Leningrad, v. 7 (2), 164-168
Schizothorax intermedius: Sarez Lake, central Pamir

Myxobolus drjagini
Wu, B.; et al., 1979, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 25 (1), 50-57
Myxobolus drjagini in Hypophthalmichthys molitrix, seasonal fluctuations with regard to occurrence and development of parasites, recommended prophylactic measures and treatment: hatchery ponds, Hangzhou region of Jiang Province

Myxobolus drjagini Achmerov, 1954, illus.
Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
Myxobolus drjagini in Hypophthalmichthys molitrix (lymphoid fluid in cavity of cranium, central nervous system, sense organs), symptoms, pathology, prevention and treatment: Hangchow region, Chekiang Province

Myxobolus elegans Kaschkowsky
Iziumova, N. A.; Mashtakov, A. V.; and Timoshechkina, L. G., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36), 71-76
[Rutilus rutilus]: Upper Kama; Chusovaia river

Myxobolus ellipsoides Thelohan
Wu, P. H.; et al., 1975, Tung Wu Hsuch Pao (Acta Zool. Sinica), v. 21 (2), 190-198
parasites of fishes: China

Myxobolus exiguus Thelohan
Armas, G., 1979, J. Fish Dis., v. 2 (6), 545-547
Mugil cephalus (gills, heart, liver, gall bladder, spleen, intestine, muscle tissue, kidney): Rio Moche coastal lagoon, northern Peru

Myxobolus exiguus Thelohan
Iziumova, N. A.; Mashtakov, A. V.; and Timoshechkina, L. G., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36), 71-76
[Abramis brama]: Chusovaia river

Myxobolus exiguus
Podlipaev, S. A., 1974, Parazitologiia, Leningrad, v. 8 (6), 535-542
Myxobolidae spp., polysaccharides in spores, nature of iodinophilous vacuole

Myxobolus exiguus
Siau, Y., 1977, Compt. Rend. Acad. Sc., Paris, v. 285, s. D, Sc. Nat., (6), 681-683
Myxobolus exiguus, early stages of development in vitro

Myxobolus exiguus Thelohan, 1895, illus.
Siau, Y., 1979, Compt. Rend. Acad. Sc., Paris, v. 288, s. D, Sc. Nat. (4), 403-404
Myxobolus exiguus, Ceratomyxa herouardi, synaptonemal complexes, electron microscopic observations, implications for life cycle and classification of myxosporidians

Myxobolus insidiosus
Markiw, M. E.; and Wolf, K., 1978, J. Fish. Research Bd. Canada, v. 35 (6), 828-832
Myxosoma cerebralis, rabbits immunized with antigens extracted from mature spores or prespore stages, antisera and globulins used in fluorescent antibody techniques, direct fluorescent antibody test showed higher specificity than indirect FAT in cross reactions with other species of myxosporidians

Myxobolus insidiosus
Wyatt, E. J., 1978, J. Parasitol., v. 64 (1), 169-170
Oncorhynchus kisutch (muscle tissue): Kalama, Lewis and Spleelyai River Hatcheries, Washington State

Myxobolus insidiosus Wyatt and Pratt, 1963
Wyatt, E. J., 1978, J. Fish Dis., v. 1 (3), 233-240
Myxobolus insidiosus in Oncorhynchus tshawytscha, epizootiology, factors affecting prevalence of infection in naturally contaminated waters, no infection could be induced in susceptible fish in disease free water supply: Oregon

Myxobolus insidiosus clarki sub. sp. n., illus.
Wyatt, E. J., 1979, J. Protozool., v. 26 (1), 47-51
Salmo clarki (somatic musculature): Linn and Lane counties, Oregon

Myxobolus kisutchi, illus.
Wyatt, E. J., 1978, J. Parasitol., v. 64 (1), 169-170
Oncorhynchus tshawytscha (mid-brain, spinal cord): McKenzie River, western Oregon

Myxobolus koi Kudo
Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
parasites of fishes: China

Myxobolus kozloffi sp. n., illus.
Wyatt, E. J., 1979, J. Protozool., v. 26 (1), 47-51
Catostomus luxatus (kidneys): Klamath County, Oregon

Myxobolus krokhini
Konovalov, S. M.; Shevliakov, A. G.; and Krasin, V. K., 1970, Parazitologiia, Leningrad, v. 4 (6), 547-556
parasite fauna of various groups of young Oncorhynchus nerka, comparative analysis reveals 3 ecological groups: Lake Azabach'e, Kamchatka river basin

Myxobolus krokhini
Makhovenko, E. T., 1972, Parazitologiia, Leningrad, v. 6 (4), 369-375
Salvelinus alpinus inhabiting different ecological niches, possible use of differences in parasite fauna between groups as biological tags: Lake Azabach'e, Kamchatka

Myxobolus lotae sp. n., illus.
Mitenev, V. K., 1971, Parazitologiia, Leningrad, v. 5 (6), 556-558
Syns.: M. mülleri Bütschli, 1882, part.; M. cycloides Gurley, 1893, part.
Lota lota (gill lobes): Ponoy River near Ponoy Village, Kola Peninsula

Myxobolus magnaspherus Cone and Anderson, 1977
Cone, D. K.; and Anderson, R. C., 1977, Canad. J. Zool., v. 55 (9), 1410-1423
parasites of Lepomis gibbosus, prevalence and intensity in relation to host age and sex
Lepomis gibbosus (parietal peritoneum of kidney): Ryan Lake, Algonquin Park, Ontario

Myxobolus minutus Nemeczek
Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
parasites of fishes: China

Myxobolus muelleri Buetshli, 1882
Andrews, C., 1979, J. Fish. Biol., v. 15 (2), 195-209
parasite fauna of Perca fluviatilis, host specificity, comparison with different localities in British Isles, factors affecting composition
Perca fluviatilis (buccal cavity): Llyn Tegid, Wales

Myxobolus muelleri Buetschli
Iziumova, N. A.; Mashtakov, A. V.; and Timoshechkina, L. G., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36), 71-76
[Abramis brama]: Upper Kama

Myxobolus mülleri Bütschli, 1882, part.
Mitenev, V. K., 1971, Parazitologiia, Leningrad, v. 5 (6), 556-558
as syn. of M. lotae sp. n.

Myxobolus muelleri, illus.
Podlipaev, S. A., 1972, Parazitologiia, Leningrad, v. 6 (6), 506-508
Myxosporidia, 6 spp., lipid inclusions in spores and cysts

Myxobolus muelleri, illus.
Podlipaev, S. A., 1974, Parazitologiia, Leningrad, v. 8 (6), 535-542
Myxobolidae spp., polysaccharides in spores, nature of iodinophilous vacuole

Myxobolus musculi Keysselitz, 1904
Ashurova, M., 1973, Parazitologiia, Leningrad, v. 7 (2), 164-168
Schizothorax intermedius: Sarez Lake, central Pamir

Myxobolus musculi Keysselitz
Mashtakov, A. V.; et al., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36), 68-71
[Pelecus cultratus] (muscle): Votkinsk reservoir

Myxobolus nemeczeki
Podlipaev, S. A., 1974, Parazitologiia, Leningrad, v. 8 (6), 535-542
Myxobolidae spp., polysaccharides in spores, nature of iodinophilous vacuole

Myxobolus neurobius Schuberg and Schroeder, 1905
Arthur, J. R.; Margolis, L.; and Arai, H. P., 1976, J. Fish. Research Bd. Canada, v. 33 (11), 2489-2499
Thymallus arcticus (brain, spinal cord): Aishihik Lake, Yukon Territory
Salvelinus namaycush (brain, spinal cord): Aishihik Lake, Yukon Territory
Prosopium cylindraceum (brain, spinal cord): Aishihik Lake, Yukon Territory
Coregonus clupeaformis (brain, spinal cord): Aishihik Lake and Stevens Lake, Yukon Territory

Myxobolus neurobius
Konovalov, S. M.; Shevliakov, A. G.; and Krasin, V. K., 1970, Parazitologiia, Leningrad, v. 4 (6), 547-556
parasite fauna of various groups of young Oncorhynchus nerka, comparative analysis reveals 3 ecological groups: Lake Azabach'e, Kamchatka river basin

Myxobolus neurobius
Makhovenko, E. T., 1972, Parazitologiia, Leningrad, v. 6 (4), 369-375
Salvelinus alpinus inhabiting different ecological niches, possible use of differences in parasite fauna between groups as biological tags: Lake Azabach'e, Kamchatka

Myxobolus neurobius
O'Grodnick, J. J., 1979, Tr. Am. Fish. Soc., v. 108 (2), 187-190
Salvelinus fontinalis (central nervous system): Tylersville hatchery

Myxobolus notemigoni
Riley, D. M., 1978, Tr. Am. Fish. Soc., v. 107 (1), 207-212
Notemigonus crysoleucas: Florida

Myxobolus obpyriformis Schulman, 1962
Ashurova, M., 1973, Parazitologiia, Leningrad, v. 7 (2), 164-168
Schizothorax intermedius: Sarez Lake, central Pamir

Myxobolus osburni Herrick, 1936
  Cone, D. K.; and Anderson, R. C., 1977, Canad. J. Zool., v. 55 (9), 1410-1423
    parasites of Lepomis gibbosus, prevalence and intensity in relation to host age and sex
    Lepomis gibbosus (pancreatic tissue dorsal to pyloric caeca, anterior to gall bladder): Ryan Lake, Algonquin Park, Ontario

Myxobolus oviformis Thelohan
  Iziumova, N. A.; Mashtakov, A. V.; and Timoshechkina, L. G., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36), 71-76
    [Abramis brama]: Upper Kama; Chusovaia river

Myxobolus parvus Schulman
  Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
    parasites of fishes: China

Myxobolus pavlovskii (Achmerov, 1954), illus.
  Molnar, K., 1979, Acta Vet., Budapest, v. 27 (3), 207-216
    Myxobolus pavlovskii in cultured fish, incidence, infection experiments, epizootiology, histopathology
    Hypophthalmichthys mollitrix (nat. and exper.) (stratified epithelium between gill lamellae)
    Aristichthys nobilis (nat. and exper.) (stratified epithelium between gill lamellae)
    all from Hungary

Myxobolus pavlovskii Achmerov
  Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
    parasites of fishes: China

Myxobolus pfeifferi
  Iskov, M. P., 1979, Gidrobiol. Zhurnal, v. 15 (4), 68-72
    parasites causing intensive epizootic disease of food fishes: Kremenchug reservoir

Myxobolus pfeifferi
  Kasim, M. H.; Rahemo, Z. L.; and Warsi, A. A., 1977, Mesopotamia J. Agric., v. 12 (2), 133-141, Arabic p. 15
    parasites of Acanthobrama marmid and Cyprinion macrostomus, intensity of infection, seasonal variations, sex of host
    Acanthobrama marmid (gill arches): river Tigris, Mosul (Iraq)

Myxobolus pseudodispar Gorbunova
  Iziumova, N. A.; Mashtakov, A. V.; and Timoshechkina, L. G., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36), 71-76
    [Rutilus rutilus]: Upper Kama

Myxobolus pseudodispar
  Linnik, V. Ia.; and Zen'kovich, E. M., 1970, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 8, 109-114
    [Blicca bjoerkna]: Neman river basin

Myxobolus rotundus Nemeczek
  Iziumova, N. A.; Mashtakov, A. V.; and Timoshechkina, L. G., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36), 71-76
    [Abramis brama]: Chusovaia river

Myxobolus sandrae Reuss
  Iziumova, N. A.; Mashtakov, A. V.; and Kashkovskii, V. V., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36) 64-68
    [Lucioperca lucioperca] (gills): Kamsk reservoir

Myxobolus schizopygopsis Dzhalilov et Aschurova, 1971
  Ashurova, M., 1973, Parazitologiia, Leningrad, v. 7 (2), 164-168
    Schizopygopsis stoliczkai: Sarez Lake, central Pamir

Myxobolus suturalis Schulman, 1962
  Ashurova, M., 1973, Parazitologiia, Leningrad, v. 7 (2), 164-168
    Schizothorax intermedius: Sarez Lake, central Pamir

Myxobolus symmetricus Nie et Lee
  Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
    parasites of fishes: China

Myxobolus uvuliferis Cone and Anderson, 1977
  Cone, D. K.; and Anderson, R. C., 1977, Canad. J. Zool., v. 55 (9), 1410-1423
    parasites of Lepomis gibbosus, prevalence and intensity in relation to host age and sex
    Lepomis gibbosus (striated muscle, mainly at base of unpaired fins): Ryan Lake, Algonquin Park, Ontario

Myxobolus wasjugani sp. n., illus.
  Bocharova, T. A.; and Donets, Z. C., 1974, Parazitologiia, Leningrad, v. 8 (1), 74-76
    Carassius carassius (muscles): Tukhemitor lake (middle course of Vasiugan river)

Myxoproteus Doflein 1898 emend, Davis 1917
  Sankurathri, C. S., 1977, Canad. J. Zool., v. 55 (7), 1147-1150
    redefinition; differentiation from Conispora gen. nov. and Sinuolinea

Myxoproteus abyssus Yoshino and Moser 1974
  Sankurathri, C. S., 1977, Canad. J. Zool., v. 55 (7), 1147-1150
    as syn. of Conispora abyssa (Yoshino and Moser 1974) n. comb.

Myxoproteus myoxocephali Fantham et al. 1940
  Sankurathri, C. S., 1977, Canad. J. Zool., v. 55 (7), 1147-1150
    does not belong to Myxoproteus, taxonomic status needs to be reassessed

Myxosoma sp.
  Arthur, J. R.; Margolis, L.; and Arai, H. P., 1976, J. Fish. Research Bd. Canada, v. 33 (11), 2489-2499
    Catostomus catostomus (gills): Aishihik Lake, Yukon Territory

Myxosoma sp.
  Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
    parasites of fishes: China

Myxosoma cartilaginis
Markiw, M. E.; and Wolf, K., 1978, J. Fish. Research Bd. Canada, v. 35 (6), 828-832
Myxosoma cerebralis, rabbits immunized with antigens extracted from mature spores or prespore stages, antisera and globulins used in fluorescent antibody techniques, direct fluorescent antibody test showed higher specificity than indirect FAT in cross reactions with other species of myxosporidians

Myxosoma cephalis
Markiw, M. E.; and Wolf, K., 1978, J. Fish. Research Bd. Canada, v. 35 (6), 828-832
Myxosoma cerebralis, rabbits immunized with antigens extracted from mature spores or prespore stages, antisera and globulins used in fluorescent antibody techniques, direct fluorescent antibody test showed higher specificity than indirect FAT in cross reactions with other species of myxosporidians

Myxosoma cerebralis, illus.
Griffin, B. R.; and Davis, E. M., 1978, J. Fish. Research Bd. Canada, v. 35 (9), 1186-1190
Myxosoma cerebralis-infected Salmo gairdneri, detection of circulating antibodies with indirect fluorescent antibody test

Myxosoma cerebralis, illus.
Hoffman, G. L., 1971, Parazitologiia, Leningrad, v. 5 (2), 182-185
Myxosoma cerebralis, causative agent of whirling disease of trout and salmon, distribution, research, and control measures in United States, review

Myxosoma cerebralis, illus.
Markiw, M. E.; and Wolf, K., 1978, J. Fish. Research Bd. Canada, v. 35 (6), 828-832
Myxosoma cerebralis, rabbits immunized with antigens extracted from mature spores or prespore stages, antisera and globulins used in fluorescent antibody techniques, direct fluorescent antibody test showed higher specificity than indirect FAT in cross reactions with other species of myxosporidians

Myxosoma cerebralis
O'Grodnick, J. J., 1979, Tr. Am. Fish. Soc., v. 108 (2), 187-190
Myxosoma cerebralis, susceptibility of fry and fingerling salmonids in contaminated water supply at Cedar Run, Clinton County, Pennsylvania; Salvelinus namaycush completely refractory to infection
Oncorhynchus nerka
Salvelinus fontinalis
Oncorhynchus tshawytscha
Salmo trutta
Oncorhynchus kisutch
Salmo gairdneri
(all exper.)

Myxosoma cerebralis
Taylor, R. L.; and Lott, M., 1978, J. Protozool., v. 25 (1), 105-106
Myxosoma cerebralis, transmission of whirling disease to Salmo gairdneri by feces of birds (Anas platyrhynchos, Nycticorax nycticorax) fed infected trout, first report that spores are still viable after passage through birds, necessity of maturation of spores in mud

Myxosoma cyprini
Markiw, M. E.; and Wolf, K., 1978, J. Fish. Research Bd. Canada, v. 35 (6), 828-832
Myxosoma cerebralis, rabbits immunized with antigens extracted from mature spores or prespore stages, antisera and globulins used in fluorescent antibody techniques, direct fluorescent antibody test showed higher specificity than indirect FAT in cross reactions with other species of myxosporidians

Myxosoma funduli Kudo, illus.
Current, W. L.; Janovy, J., jr.; and Knight, S. A., 1979, J. Protozool., v. 26 (4), 574-583
Myxosoma funduli, plasmodium wall, sporogenesis, ultrastructure
Fundulus kansae (gills): South Platte River, Keith County, Nebraska

Myxosoma lairdi n. sp., illus.
Narasimhamurti, C. C.; and Kalavati, C., 1979, Proc. Indian Acad. Sc., Sect. B, Animal Sc., v. 88 (4, pt. 1), 269-272
Liza macrolepis (gut muscles): Bheemunipatnam, Andhra Pradesh

Myxosoma lieni Nie et Lee
Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
parasites of fishes: China

Myxosoma notropis Fantham, Porter and Richardson, 1939, illus.
Desser, S. S.; and Paterson, W. B., 1978, Canad. J. Zool., v. 56 (2), 311-314
Myxosoma notropis, morphology and dimensions of spores, light and scanning electron microscopy, histochemistry
Notropis cornutus (liver): Lake Sasajewan, Algonquin Park, Ontario

Myxosoma squamalis
Olson, R. E., 1978, Calif. Fish and Game, v. 64 (2), 117-120
Oncorhynchus kisutch (scales): Pacific Ocean off Newport, Oregon

Myxosoma varia (Achmerov) Nie et Lee
Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
parasites of fishes: China

Myxosporidia
Donets, Z. S.; and Shul'man, S. S., 1973, Parazitologiia, Leningrad, v. 7 (2), 191-193
Myxosporidia, glycerine-gelatin slides prepared without preliminary fixation followed by use of phase-contrast microscopy recommended as best method to use for identification

Myxosporidia
Grasse, P. P.; and Lavette, A., 1978, Compt. Rend. Acad. Sc., Paris, v. 286, s. D, Sc. Nat. (10), 757-759
Sphaeromyxa sabrazesi, 'germinal' cells are complex of three cellular types, observations support independence of Myxosporidia from Protozoa and Metazoa

Myxosporidi[a], illus.
Koerting, W., 1977, Fisch u. Umwelt (4), 37-48
fish parasites, histopathological changes

[Myxosporidia sp.] miksosporidii
Linnik, V. Ia.; and Zen'kovich, E. M., 1970, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 8, 109-114
[Rutilus rutilus]
[Blicca bjoerkna]
[Leuciscus idus]
all from Neman river basin

Myxosporida [sp.], unidentified trophozoites
Mitchell, L. G., 1978, J. Protozool., v. 25 (1), 100-105
Lepisosteus platostomus
Notropis dorsalis
Semotilus atromaculatus
Carpiodes carpio
Moxostoma macrolepidotum
Lepomis macrochirus
Micropterus salmoides
(gall bladder of all): all from Iowa

Myxosporidia gen. sp.
Otvodova, G. D., 1975, Izvest. Gosudarstv. Nauchno-Issled. Inst. Ozer. i Rech. Ryb. Koziaistva, Leningrad, v. 93, 112-116
[Abramis brama] (gills): Pskov-Chudskoe lake

Myxosporidia gen. sp. (=Henneguya sargi Swarz., 1912)
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
Diplodus annularis (subcutaneous body tissue, fins): Black Sea

Naegleria
Botto, C.; and Rodriguez, N., 1975, Rev. Uruguaya Patol. Clin. y Microbiol., v. 13 (1), 3-16
human amoebic meningoencephalitis, etiology, epidemiology, pathology, diagnosis, therapy, review

Naegleria, illus.
Boyle, A. L.; et al., 1979, J. Clin. Path., v. 32 (3), 306-307 [Letter]
Naegleria, child, rapid detection of trophozoites in spinal fluid stained with bacterial stains

Naegleria-Hartmannella free-living amoebae, illus.
Carosi, G., 1974, Ann. Soc. Belge Med. Trop., v. 54 (4-5), 265-277
Entamoeba moshkovskii and free-living amoeba of Hartmannella-Naegleria group, ultrastructural comparisons

Naegleria-Hartmannella groups
Das, S. R., 1975, New Approaches Ident. Microorgan., 451-458
Naegleria-Hartmannella groups, agar plate method for isolation and cultivation of amoebae from infected humans and animals, soil, sewage sludge, fresh water, and other substrates

Naegleria
Dive, D.; et al., 1978, Ann. Microbiol., v. 129 B (2), 225-244
"Limax" amoebae in swimming pools, water purification systems, associated bacterial flora, temperature, pathogenicity to mice: North of France

Naegleria
Dutta, G. P., 1979, J. Scient. and Indust. Research, India, v. 38 (4), 199-218
pathogenic and nonpathogenic aerobic free-living amoebae, characterization of cytoplasmic inclusions, cytochemistry and ultrastructure, review

Naegleria
Feldman, R. E., 1979, Acta Bioquim. Clin. Latinoam., v. 13 (3), 309-311
pathogenic free-living amoebae, brief review

Naegleria
Gottlieb, B.; and Reyes, H., 1977, Rev. Med. Chile, v. 105 (7), 467-471
Naegleria, Acanthamoeba, Hartmannella, free-living amoeba now thought to be agents responsible for human meningoencephalitis, diagnosis, pathology, therapy, review

Naegleria
Hoffman, E. O.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (1, pt. 1), 29-38
primary amebic meningoencephalitis, human (brain, pancreas), clinical findings, postmortem studies, electron microscopy, immunohistologic studies, evidence slightly more indicative of Acanthamoeba than Naegleria infection: Louisiana

Naegleria, illus.
Janitschke, K.; et al., 1978, Zentralbl. Bakteriol., 1. Abt. Orig., Reihe B, v. 166 (2-3), 244-249
Naegleria, viability of pathogenic strain in water media (public water supply, swimming-pool, inland lake) at various temperatures

Naegleria
Johnson, C. M., 1977, Rev. Med. Panama, v. 2 (3), 141-144
human meningoencephalitis previously diagnosed by Herrera (Arch. Med. Panamenos, v. 1, 1952) as caused by Endamoeba histolytica is correctly diagnosed as resulting from infection with a species of Naegleria, opinion based on history, clinical course, morphology of amoeba and pathology: Panama

Naegleria
Rivera, F.; et al., 1979, Tr. Am. Micr. Soc., v. 98 (3), 465-469
protozoa polluting tap water, concentration and identification in culture: Federal District, Mexico City

Naegleria
Willaert, E., 1974, Ann. Soc. Belge Med. Trop., v. 54 (4-5), 429-440
Naegleria, Acanthamoeba spp. causing disease in man and other vertebrates, selected bibliography and tabular survey of cases

Naegleria
Wyburn-Mason, R., 1979, Med. Hypotheses, v. 5 (11), 1237-1249
Naegleria, possible cause of rheumatoid disease and many human cancers through chronic antigenic stimulation by the Naegleria, review of new medical concept

Naegleria sp.
De Jonckheere, J. F., 1979, Applied and Environment. Microbiol., v. 38 (4), 590-593
Naegleria and Acanthamoeba, incidence in aquaria, may be source of human infection: Belgium

Naegleria spp. Alexeieff 1912, emend. Calkins 1913
Derr-Harf, C.; et al., 1978, Ann. Parasitol., v. 53 (5), 467-477
isolation of 164 strains of free-living amoebae from public water supplies and swimming pools, some strains of Acanthamoeba polyphaga, A. castellanii, A. lenticulata, and A. comandoni were pathogenic for mice, none of the Naegleria isolated were pathogenic for mice, levels of bromine and chlorine used were insufficient to eliminate these amoebae, potential danger to users and suggested control measures: Strasbourg

Naegleria sp.
Scholten, T., 1979, Canad. Med. Ass. J., v. 120 (3), 264, 267-268 [Letter]
soil amoebae potentially pathogenic to man, existence in Canada, results of survey from various areas of Ontario

Naegleria aerobia
Das, S. R., 1974, Ann. Soc. Belge Med. Trop., v. 54 (4-5), 327-332
Naegleria aerobia flagellate stage, pathogenicity, bearing on epidemiology of exogenous amoebiasis

Naegleria aerobia (Singh & Das, 1970), illus.
Maitra, S. C.; et al., 1978, Current Sc., Bangalore, v. 47 (24), 939-941
Naegleria aerobia in infected mouse brain and in vitro, fate of special cytoplasmic inclusions (black bodies), nature and role of these bodies discussed

Naegleria fowleri, illus.
Brown, T., 1978, J. Med. Microbiol., v. 11 (3), 249-259
Naegleria fowleri, cytopathogenicity in mouse embryo-cell cultures, no evidence that cell-free cytotoxic factors play a part, damage seemed to occur only as result of direct contact with active amebae and appeared to be associated with phagocytic activity of trophozoites

Naegleria fowleri, illus.
Brown, T., 1979, J. Med. Microbiol., v. 12 (3), 355-362
Naegleria fowleri, cytopathogenicity in mouse embryo-cell cultures, inhibition by amoeba-specific antiserum and by cytochalasin B, concluded that cytopathogenicity was due to physical rather than biochemical or cytotoxic mechanisms and was associated with phagocytic activity of trophozoites

Naegleria fowleri, illus.
Brown, T., 1979, J. Med. Microbiol., v. 12 (3), 363-371
Naegleria fowleri, cytopathogenicity in mouse embryo-cell cultures, observations by immunofluorescence microscopy and electron microscopy, concluded that trophozoites destroy cultured cells by phagocytosis-like mechanism without aid of amoeba-associated cytotoxic or cytolytic agents

Naegleria fowleri
Carter, S. A., 1978, Internat. J. Environment. Studies, v. 12 (3), 199-205
Naegleria fowleri, human amoebic meningoencephalitis, water pollution as major source of infection, epidemiology, prevention and control, review

Naegleria fowleri
Cursons, R. T. M.; et al., 1979, J. Parasitol., v. 65 (1), 189-191
Acanthamoeba, Naegleria, semidefined media for cultivation of pathogenic and nonpathogenic free-living amebae

Naegleria fowleri
Cursons, R. T. M.; Brown, T. J.; and Keys, E. A., 1978, J. Parasitol., v. 64 (4), 744-745
Naegleria, Acanthamoeba, pathogenic vs. nonpathogenic strains, differences in level of production of phospholipase A may explain differences in invasiveness and virulence, likely that secretion of enzyme constitutes initial steps whereby host tissue is prepared for endocytosis by these amebae

Naegleria fowleri, illus.
Cursons, R. T.; Brown, T. J.; and Keys, E. A., 1978, N. Zealand J. Med. Lab. Tech., v. 32 (1), 11-14
primary amoebic meningo-encephalitis, humans, differential diagnosis and identification of aetiologic agents

Naegleria fowleri
Darby, C. P.; et al., 1979, Am. J. Dis. Child., v. 133 (10), 1025-1027
primary amebic meningoencephalitis, human, 3 fatal case reports, Naegleria fowleri isolated and identified premortem in 1: had been swimming in freshwater lakes in South Carolina

Naegleria fowleri
De Jonckheere, J., 1979, Path. Biol., v. 27 (8), 453-458
Naegleria fowleri, virulence for mice of isolates from environment, effect of axenic cultivation, brain passage, and passages in Vero cell cultures, mouse strain and age differences

Naegleria fowleri, illus.
Ferrante, A.; and Thong, Y. H., 1979, Internat. J. Parasitol., v. 9 (6), 599-601
Naegleri fowleri, antibody-induced capping and endocytosis of surface antigens, may allow amoeba to resist action of host's immune system

Naegleria fowleri
Haggerty, R. M.; and John, D. T., 1978, Infect. and Immun., v. 20 (1), 73-77
Naegleria fowleri in mice, infecting dose and age, sex, and strain of host are important variables that markedly affect innate resistance to infection

Naegleria fowleri, illus.
Hermanne, J.; et al., 1974, Rev. Prat., Paris, v. 24 (15), 1241-1242, 1245-1246, 1249-1253
Naegleria fowleri, human meningoencephalitis, general clinical review, 3 case reports: Belgium

Naegleria fowleri
Holbrook, T. W., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (4), 460-462
Naegleria fowleri, chick embryos are susceptible to infection and may represent useful host for experimental studies

Naegleria fowleri
Holbrook, T. W.; and Parker, B. W., 1979, Am. J. Trop. Med. and Hyg., v. 28 (6), 984-987
Naegleria fowleri incubated on chick embryos, effects of embryo age and temperature on maintenance, infectivity maintained after 25 serial passages

Naegleria fowleri
Kadlec, V.; Cerva, L.; and Skvarova, J., 1978, Science (4360), v. 201, 1025
reservoir of pathogenic Naegleria fowleri located in cracked wall of indoor swimming pool where repeated outbreaks of primary amoebic meningoencephalitis were observed between 1962 and 1965: northern Bohemia

Naegleria fowleri
Lawande, R. V.; et al., 1979, Am. J. Clin. Path., v. 71 (2), 201-203
Naegleria fowleri, children, soil amebas recovered from nasal passages during dusty harmattan period, cause of fatal meningoencephalitis in infant (cerebrospinal fluid, nose): Zaria

Naegleria fowleri, illus.
Lawande, R. V.; et al., 1979, Am. J. Clin. Path., v. 71 (5), 591-594
Naegleria fowleri, 8-month-old infant, fatal case of primary amebic meningoencephalitis: Zaria, Nigeria

Naegleria fowleri, illus.
Lawande, R. V.; et al., 1979, J. Trop. Med. and Hyg., v. 82 (4), 84-88
Naegleria fowleri, children, primary amoebic meningoencephalitis, possible inhalation of airborne cysts: Northern Nigeria

Naegleria fowleri, illus.
Lawande, R. V.; Ogunkanmi, A. E.; and Egler, L. J., 1979, Ann. Trop. Med. and Parasitol., v. 73 (1), 51-56
free-living pathogenic and non-pathogenic amoebae, prevalence survey in soils, Naegleria fowleri pathogenic in young Swiss white mice: Zaria, Nigeria

Naegleria fowleri
Lee, K. K.; et al., 1979, Antimicrob. Agents and Chemotherapy, v. 16 (2), 217-220
Naegleria fowleri, in vitro susceptibility to selected antimicrobial agents singly and in combination

Naegleria fowleri, illus.
Martin, J. J., 1974, Ann. Soc. Belge Med. Trop., v. 54 (4-5), 305-319
Naegleria fowleri, man, primary amoebic meningo-encephalitis, neuropathology of 3 fatal cases: Antwerp

Naegleria fowleri, illus.
Martinez, A. J.; and Janitschke, K., 1979, Immun. u. Infekt., v. 7 (2), 57-64
Naegleria fowleri, Acanthamoeba, human, histopathology, protozoology and taxonomy, epidemiology, treatment, animal models, review

Naegleria fowleri
Nerad, T. A.; and Daggett, P. M., 1979, J. Protozool., v. 26 (4), 613-615
Naegleria fowleri, N. gruberi, isoenzyme electrophoresis as effective method for separation of pathogenic and nonpathogenic Naegleria strains

Naegleria fowleri
Pringle, H. L.; Bradley, S. G.; and Harris, L. S., 1979, Antimicrob. Agents and Chemotherapy, v. 16 (5), 674-679
Naegleria fowleri, susceptibility to Δ^9-tetrahydrocannabinol and other cannabinoids

Naegleria fowleri, illus.
Proca-Cioban, M., 1978, Bacteriol., Virusol., Parazitol., Epidemiol., Bucuresti, v. 23 (3), 129-140
Naegleria fowleri, Acanthamoeba culbertsoni, human primary amoebic meningoencephalitis, broad review

Naegleria fowleri, illus.
Schuster, F. L.; and Dunnebacke, T. H., 1974, Ann. Soc. Belge Med. Trop., v. 54 (4-5), 359-370
Naegleria amoebae contain virus-like particles and an unassociated infectious agent, possible relationship to pathogenicity, review

Naegleria fowleri, illus.
Stevens, A. R.; Gallup, E. D.; and Willaert, E., 1978, J. Invert. Path., v. 31 (1), 63-76
Naegleria spp., evaluation of membrane-bound black bodies, found to be characteristic of trophozoites and unrelated to encystment

Naegleria fowleri
Thong, Y. H.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (2, pt. 1), 238-240
Naegleria fowleri, mice immunized with live parasites by intraperitoneal injection were found to be more resistant to subsequent intranasal challenge

Naegleria fowleri
Thong, Y. H.; et al., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (6), 650-652
Naegleria fowleri, mice immunized with live organisms acquire resistance to challenge, protective immunity can be transferred by immune serum but not by immune cells, mechanism of this immunity unknown

Naegleria fowleri
Thong, Y. H.; Ferrante, A.; and Shepherd, C., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (2), 207-209
Naegleria fowleri, differentiation from N. gruberi by comparison of phagocytic behavior towards baker's yeast

Naegleria fowleri
Thong, Y. H.; Rowan-Kelly, B.; and Ferrante, A., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (3), 336-337
Naegleria fowleri-infected mice, amphotericin B potentiated by tetracycline shows promise in treating meningoencephalitis

Naegleria fowleri, illus.
Vandepitte, J.; et al., 1974, Ann. Soc. Belge Med. Trop., v. 54 (4-5), 297-303
Naegleria fowleri, 14-year-old boy, fatal primary amoebic meningo-encephalitis after swimming in stream polluted by warm effluents of zinc and lead factory: Canal of Beverio, Balen-Wezel

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Weik, R. R.; and John, D. T., 1978, J. Parasitol., v. 64 (4), 746-747
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Naegleria fowleri, cell and mitochondria respiration

Naegleria fowleri
Willaert, E.; et al., 1974, Ann. Soc. Belge Med. Trop., v. 54 (4-5), 333-342
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Entamoeba moshkovskii and free-living amoeba of Hartmannella-Naegleria group, ultrastructural comparisons

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Cerva, L., 1971, Folia Parasitol., v. 18 (2), 171-176
Naegleria gruberi, Vitek strain, white mice, albino guinea pigs, 7 day-old chick embryos (exper. in all), pathogenicity studies

Naegleria gruberi
Cursons, R. T. M.; et al., 1979, J. Parasitol., v. 65 (1), 189-191
Acanthamoeba, Naegleria, semidefined media for cultivation of pathogenic and nonpathogenic free-living amebae

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  De Jonckheere, J. F., 1979, Applied and Environment. Microbiol., v. 38 (4), 590-593
    Naegleria and Acanthamoeba, incidence in aquaria, may be source of human infection: Belgium

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  Hermanne, J.; Jadin, J. B.; and Martin, J. J., 1972, Semaine Hop. Paris, v. 48 (21), 1539-1550 (Ann. Pediat., Paris, v. 19 (5), 425-436)
    Naegleria gruberi, human, fatal meningoencephalitis after swimming in public pools, amoeba discovered in spinal fluid, amphotericin B studied as possible therapy: Belgium

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  Jadin, J. B.; et al., 1971, Ann. Soc. Belges Med. Trop. Parasitol., v. 51 (2), 255-266
    Naegleria gruberi, human primary amoebic meningoencephalitis, case reports, swimming pools were source of infections: Western Europe

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  Malhotra, K. K.; et al., 1978, J. Trop. Med. and Hyg., v. 81 (6), 113-115
    Naegleria gruberi resulting in fatal human primary amoebic meningoencephalitis, report of atypical case documenting diagnostic and therapeutic problems: Jammu

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  Pernin, P.; and Riany, A., 1978, Ann. Parasitol., v. 53 (4), 333-344
    survey of swimming pools for presence of free-living amebae, potential danger for swimmers: Lyon

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  Schuster, F. L.; and Dunnebacke, T. H., 1974, Ann. Soc. Belge Med. Trop., v. 54 (4-5), 359-370
    Naegleria amoebae contain virus-like particles and an unassociated infectious agent, possible relationship to pathogenicity, review

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    Naegleria and Acanthamoeba, incidence in aquaria, may be source of human infection: Belgium

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  Godfrey, D. G., 1977, Protozoology, v. 3, 33-49
    identification of morphologically similar trypanosomes of mammals

Naupliicola sp., illus.
  Michajlow, W., 1970, Acta Parasitol. Polon., v. 18 (42-50), 473-482
    Acanthocyclops vernalis (nauplius eye)

Naupliicola sp., illus.
  Michajlow, W., 1977, Zhurnal Obsh. Biol., v. 38 (5), 657-675
    Macrocyclops sp. (eye)

Naupliicola spp.
  Michajlow, W., 1978, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 26 (4), 269-272
    Mesocyclops leuckarti: water body of Botanical Gardens, Brisbane and Melbourne, Australia

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  Michajlow, W., 1970, Acta Parasitol. Polon., v. 18 (42-50), 473-482
    Acanthocyclops vernalis
    Eucyclops serrulatus

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    Macrocyclops albidus

Naupliicola cystifingens Michajlow, 1968, illus.
  Michajlow, W., 1970, Acta Parasitol. Polon., v. 18 (42-50), 473-482
    Eucyclops macruroides

Naupliicola fennicus Michajlow, 1966
  Michajlow, W., 1970, Acta Parasitol. Polon., v. 18 (42-50), 473-482
    Acanthocyclops vernalis

Naupliicola kaunasiensis Michajlow, 1970
  Michajlow, W., 1970, Acta Parasitol. Polon., v. 18 (42-50), 473-482
    Eucyclops serrulatus

Naupliicola kenyensis sp. n., illus.
  Michajlow, W.; and Wita, I., 1976, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 24 (5), 283-288
    Microcyclops varicans (body cavity): bays of river Tsavo, Tsavo National Park, Kenya

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  Michajlow, W., 1970, Acta Parasitol. Polon., v. 18 (42-50), 473-482
    Acanthocyclops vernalis
    Cyclops vicinus

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  Michajlow, W., 1977, Zhurnal Obsh. Biol., v. 38 (5), 657-675
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Naupliicola necans Michajlow, 1965
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    Macrocyclops albidus

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  Michajlow, W., 1970, Acta Parasitol. Polon., v. 18 (42-50), 473-482
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    Eucyclops macruroides

Naupliicola vilniensis Michajlow, 1970
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    Eucyclops serrulatus

Nematocystis bunmii sp. n., illus.
  Segun, A. O., 1978, J. Protozool., v. 25 (2), 157-162
    Heliodrilus lagosensis (seminal vesicles): Oshogbo and Ile-Ife (Western), and Lagos, Nigeria

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  Newman, M. W., 1971, Proc. 1970 National Shellfish. Ass., v. 61, 59-63
    Crassostrea virginica (gill, mantle): New Haven Harbor, Connecticut

Nipponica subgen. n.
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34
    subgen. of Opalina
    type sp.: Opalina (Nipponica) japonica Sugiyama, 1920

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  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
    subgen. of Opalina, key

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    review of microsporidians reported from mammals

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  Pletcher, J. M., 1978, J. Am. Vet. Med. Ass., v. 172 (3), 216, 219 [Letter]
    ". . .continued use of 'nosema' as a synonym for 'encephalitozoon' is not in keeping with current knowledge"; morphologic differences and differing host specificities separate the two genera

Nosema
  Widder, H. U.; and Bremer, R., 1978, Monatsh. Vet.-Med., v. 33 (5), 184-187
    Nosema, bees, veterinary hygiene: Neubrandenburg

Nosema sp., illus.
  Grobov, O. F.; Karakuiumchian, M. K.; and Orlova-Sokol'skaia, I. A., 1975, Parazitologiia, Leningrad, v. 9 (2), 142-146
    [white rats] (brain): Gosudarstv Control Institute im. Tarasevich
    [Mus musculus] (peritoneal fluid) (exper.)

Nosema sp., illus.
  Jafri, R. H.; Asif, M.; and Aslamkhan, M., 1976, Pakistan J. Zool., v. 8 (2), 232-234
    Anopheles stephensi (lumen of stomach, epithelium of posterior part of stomach): Lahore, Pakistan

Nosema spp., morphologically similar to N. necatrix
  Knell, J. D.; and Zam, S. G., 1978, J. Invert. Path., v. 31 (3), 280-288
    Nosema spp., double immunodiffusion techniques used to investigate taxonomic relationships between 6 different microsporidian isolates
    ". . . Nosema necatrix isolates 942, 954, and 960 and Nosema plodiae . . . should be considered members of the genus Vairimorpha."

Nosema sp.
  Miln, A. J., 1978, N. Zealand Entom., v. 6 (4), 392-399
    Costelytra zealandica: New Zealand

Nosema sp., illus.
  Narayanan, K.; and Jayaraj, S., 1979, Current Sc., Bangalore, v. 48 (6), 276 [Letter]
    Spodoptera litura (nat. and exper.): India

Nosema sp.
  Narayanan, K.; and Jayaraj, S., 1979, Current Sc., Bangalore, v. 48 (18), 825 [Letter]
    mixed infections of Nosema sp. and virus in Pericallia ricini and Spodoptera litura: India

Nosema sp.
  Palmieri, J. R.; et al., 1978, Southeast Asian J. Trop. Med. and Pub. Health, v. 9 (2), 256-259
    Lymnaea rubiginosa
    Fasciola gigantica
    Echinostoma audyi
    Tracheophilus sp.
    (all exper.)

Nosema sp.
  Palmieri, J. R.; and Sullivan, J. T., 1977, J. Invert. Path., v. 30 (2), 276
    Microsporida, staining technique for location of spores in host tissues, used to locate Nosema sp. in Lymnaea rubiginosa snails and Tracheophilus sp. rediae

Nosema agrotidis Lipa et Issi
  Nilova, G. N.; and Strel'nikova, L. V., 1974, Parazitologiia, Leningrad, v. 8 (5), 463-468
    Plistophora schubergi, Nosema agrotidis, effect of ultraviolet radiation on viability of spores

Nosema algerae Vavra and Undeen
  Anthony, D. W.; Lotzkar, M. D.; and Avery, S. W., 1978, Mosquito News, v. 38 (1), 116-121
    Nosema algerae, infection rates, reproductive capacity, and longevity of Anopheles albimanus exposed at each larval instar, results indicate that introduction of N. algerae into natural A. albimanus populations of mixed larval instars could significantly reduce number and fertility of mosquito eggs

Nosema algerae
  Gajanana, A.; et al., 1979, Indian J. Med. Research, v. 70, 417-423
    partial suppression of Plasmodium gallinaceum and P. vivax in Aedes aegypti and Anopheles stephensi doubly infected with Nosema algerae and Plasmodium, epidemiological significance

Nosema algerae
  Kelly, J. F.; and Knell, J. D., 1979, J. Invert. Path., v. 33 (2), 252
    Nosema algerae, Amblyospora sp., modification of Ludox gradient spore purification technique

Nosema algerae
  Knell, J. D.; and Zam, S. G., 1978, J. Invert. Path., v. 31 (3), 280-288
    Nosema spp., double immunodiffusion techniques used to investigate taxonomic relationships between 6 different microsporidian isolates

Nosema algerae
  Maddox, J. V.; et al., 1977, Pakistan J. Zool., v. 9 (1), 19-22
    Nosema algerae, differences in susceptibility of Pakistan mosquitoes, potential as biological control agent
    Anopheles stephensi
    A. pulcherrimus
    A. culicifacies
    A. annularis
    A. subpictus
    Culex pipiens fatigans
    (all exper.)

Nosema algerae
Teetor-Barsch, G. E.; and Kramer, J. P., 1979, J. Invert. Path., v. 33 (3), 300-306
Octosporea muscaedomesticae, Nosema algerae, N. whitei, effect of lyophilization on infectivity of spores

Nosema antheraeae sp. n., illus.
Simchuk, P. A.; Lysenko, M. A.; and Chetkarova, E. M., 1979, Zool. Zhurnal, v. 58 (4), 477-482
Antheraea pernyi (fat body, cells of muscle tissue surrounding intestine and hemocytes): Ukraine (Kiev, Kivertsy Volynskoi oblast)
Hyphantria cunea (exper.)
Agrotis segetum (exper.)
Mamestra brassicae (exper.)
Galleria mellonella (exper.)
Plodia interpunctella (exper.)
Ephestia kuchniella (exper.)
Loxostege sticticalis (exper.)

Nosema apis Zander, 1909
Grobov, O. F.; and Ziuman, B. V., 1972, Parazitologiia, Leningrad, v. 6 (2), 176-179
Nosema apis, in vitro cultivation of infected honey-bee midgut tissue

Nosema apis Zander, illus.
Jacobs, F. J.; Kellner, N.; and Van der Vorst, E., 1978, Vlaams Diergeneesk. Tijdschr., v. 47 (3), 242-251
Nosema apis in Apis mellifera, development, diagnosis, and control, review

Nosema apis Zander
Kellner, N.; and Jacobs, F. J., 1978, Vlaams Diergeneesk. Tijdschr., v. 47 (3), 252-259
Nosema apis in Apis mellifera, enough spores reach ventriculus 10 to 15 minutes after inoculation to start maximal infection

Nosema apis Zander, illus.
Khsirsagar, K. K.; Chauhan, R. M.; and Singh, Y. K., 1977, Indian Bee J., v. 36 (1-4), 1974, 23-24
Apis cerana indica (gut): Pithoragarh, Kumaon hills, Uttar Pradesh, India

Nosema apis
Lehnert, T.; and Shimanuki, H., 1979, Apidologie, v. 10 (1), 17-22
Nosema apis, package bees, population change and spore levels in older vs. newly emerged bees, much of disease eliminated with death of older bees, effect of feeding fumagillin at time packages are installed

Nosema apis
Moffett, J. O.; and Lawson, F. A., 1975, J. Econom. Entom., v. 68 (5), 627-629
Nosema apis-infected honey bees, oxygen consumption

Nosema apis
Muresan, E.; et al., 1978, Apicultura Romania, v. 53 (4), 13-15, 16
Nosema apis, Fumidil B and NOSAN against Paramecium caudatum as substitute test agent (having sensitivity similar to N. apis)

Nosema apis
Muresan, E.; Duca, C.; and Papay, Z., 1975, 25. Internat. Apicult. Cong. (Grenoble, Sept. 8-14), 384-385
Nosema apis, healthy and infected Apis mellifica carpatica, histochemical indices of mid-gut

Nosema apis
Peroutka, M., 1975, 25. Internat. Apicult. Cong. (Grenoble, Sept. 8-14), 386-387
Nosema apis, drones, influence of host diet on development

Nosema apis Z.
Peroutka, M.; and Cihar, R., 1978, Apidologie, v. 9 (4), 291-304
Nosema apis in drones (exper.), effect of pollen, pharyngeal gland secretions, and pteridines (stimulation, inhibition, or no effect) on parasite reproduction

Nosema apis Zander
Rinderer, T. E.; and Elliott, K. D., 1977, J. Econom. Entom., v. 70 (4), 431-433
Nosema apis in Apis mellifera (exper.), influence of proteinaceous diet (pollen or pollen substitute) on worker bee longevity associated with infection, potential of protein feeding as component in standardized test of honey bee response to Nosema infection

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Rinderer, T. E.; and Sylvester, H. A., 1978, Ann. Entom. Soc. Am., v. 71 (3), 372-374
genetic characteristics of Apis mellifera, response to Nosema apis, longevity, and hoarding behavior

Nosema apis
Rousseau, M., 1975, 25. Internat. Apicult. Cong. (Grenoble, Sept. 8-14), 394-398
Nosema apis, bees, prophylactic control by sanitation and other management tactics, drugs used only on supplementary basis

Nosema apis
Singh, Y., 1977, Indian Bee J., v. 36 (1-4), 1974, 16
Nosema apis, honey bees, formaldehyde (some improvement), thymol (good results): apiary at Jeolikote, India

Nosema apis Zander, illus.
Steche, W.; and Held, T., 1978, Allg. Deutsche Imkerztg., v. 12 (11), 322-328
Nosema apis, structure of early developmental stages, scanning electron microscopy

Nosema apis
Van der Vorst, E.; and Jacobs, F. J., 1978, Vlaams Diergeneesk. Tijdschr., v. 47 (3), 260-266
Nosema apis spores, differential diagnosis in pollen by lugol staining

Nosema bombycis, illus.
Abe, Y., 1978, J. Sericult. Sc. Japan, v. 47 (3), 221-225
Nosema bombycis, alteration of staining properties of discharged sporoplasm in silkworm alimentary canal

Nosema bombycis Naegeli, illus.
Abe, Y., 1978, J. Sericult. Sc. Japan, v. 47 (4), 279-284
Nosema bombycis, propagative reproduction in silkworm larvae

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Jafri, R. H.; Khan, K. M. S.; and Iqbal, R., 1978, Pakistan J. Zool., v. 10 (2), 294-297
Nosema bombycis in Bombyx mori, development, histopathology, impact on sericulture: Pakistan and Azad Kashmir

Nosema bombycis
Knell, J. D.; and Zam, S. G., 1978, J. Invert. Path., v. 31 (3), 280-288
Nosema spp., double immunodiffusion techniques used to investigate taxonomic relationships between 6 different microsporidian isolates

Nosema carpocapsae Paillot
Simchuk, P. A.; and Sikura, A. I., 1978, Entom. Obozr., v. 57 (3), 495-499
Nosema carpocapsae, importance in regulating populations of Laspeyresia pomonella: Moldavia; Ukraine; Belorussia; RSFSR

Nosema cheisini Weiser, 1963, illus.
Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135

Nosema chironomi Lutz et Splendore, 1908, illus.
Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135

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Lipa, J. J.; Pruszynski, S.; and Bartkowski, J., 1975, Acta Parasitol. Polon., v. 23 (41-51), 453-461
role of pathogens and parasites in survival of Coccinellidae spp. during winter
Coccinella septempunctata
Adalia bipunctata
Coccinella quinquepunctata
Exochomus quadripustulatus
all from Poland

Nosema connori, illus.
Shadduck, J. A.; Kelsoe, G.; and Helmke, R. J., 1979, J. Parasitol., v. 65 (1), 185-188
microsporida spontaneously contaminating cell culture prepared from baboon placental cells, ultrastructurally identical to Encephalitozoon cuniculi and different from Nosema connori

Nosema costelytrae Hall, Oliver and Given
Miln, A. J., 1978, N. Zealand Entom., v. 6 (4), 392-399
Costelytra zealandica: New Zealand

Nosema (syn. Encephalitozoon) cuniculi
Nordstoga, K.; et al., 1978, Acta Vet. Scand., v. 19 (1), 150-152
Nosema cuniculi, Alopex lagopus, clinical signs of nosematosis in litter after artificial insemination and intrauterine injection of Nosema spores, possibility of transmission from male during mating

Nosema diplostomi sp. n., illus.
Shigina, N. G.; and Grobov, O. F., 1972, Parazitologiia, Leningrad, v. 6 (5), 469-475
hyperparasite of metacercariae found in eyes of fish
Diplostomum (parenchyma): salmon farm "Skhodnia", Moskovsk oblast

Nosema echinostomi Brumpt, 1922, illus.
Voronin, V. N., 1974, Parazitologiia, Leningrad, v. 8 (4), 359-364
Echinostomatidae: lake Dolgoe

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Canning, E. U.; Higby, G. C.; and Nicholas, J. P., 1979, Parasitology, v. 79 (3), 381-392
Nosema eurytremae, pathogenicity to Fasciola hepatica in Lymnaea truncatula

Nosema eurytremae, illus.
Higby, G. C.; et al., 1979, Parasitology, v. 78 (2), 155-170
Nosema eurytremae derived from trematode larvae, propagation in abnormal (insect) hosts and in tissue culture
Locusta migratoria
Periplaneta americana
Pieris brassicae
Spodoptera exempta
S. frugiperda
(all exper.)

Nosema eurytremae
Pilley, B. M.; Canning, E. U.; and Hammond, J. C., 1978, J. Invert. Path., v. 32 (3), 355-358
Nosema eurytremae in Pieris brassicae (exper.), microinjection procedure for large-scale production of minimally contaminated spores

Nosema fumiferanae
Wilson, G. G., 1977, Canad. J. Zool., v. 55 (1), 249-250
Nosema fumiferanae, feeding additional microsporidian spores to naturally infected Choristoneura fumiferana enhances adverse effects

Nosema fumiferanae
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Nosema fumiferanae, Pleistophora schubergi, incidence of infection in spruce budworm (Choristoneura fumiferana) 1 and 2 years after application of microsporidian spores to trees: Ontario

Nosema fumiferanae
Wilson, G. G., 1978, Canad. J. Zool., v. 56 (4), 578-580
feeding of Pleistophora schubergi to Choristoneura fumiferana naturally infected with Nosema fumiferanae results in greater adverse effects on budworms than either parasite alone, may play useful role in biological control

Nosema fumiferanae
Wilson, G. G., 1978, Proc. Entom. Soc. Ontario, v. 108, 1977, 144-145
Nosema fumiferanae in Choristoneura fumiferana, increase of incidence rates over six-year period: Uxbridge Forest of Southern Ontario

Nosema fumiferanae
Wilson, G. G.; and Kaupp, W. J., 1976, Insect Path. Research Inst., Inform. Rep. (IP-X-15), 14 pp.
Nosema fumiferanae, Pleistophora schubergi, control of Choristoneura fumiferana by spraying spruce and balsam fir trees with spores: Ontario

Nosema heliothidis, illus.
Brooks, W. M.; Cranford, J. D.; and Pearce, L. W., 1978, J. Invert. Path., v. 31 (2), 239-245
Nosema heliothidis, benomyl, at concentrations tested, not effective in eliminating infection in Heliothis zea in laboratory colonies, but definitely deleterious to microsporidium, potential should be further evaluated

Nosema heliothidis (Lutz and Splendor)
Jordan, J. A.; and Noblet, R., 1978, Tech. Bull. (1069) South Carolina Agric. Exper. Station, 5 pp.
Nosema heliothidis and Vairimorpha necatrix, susceptibility of selected insect pests, spore dose levels necessary to obtain LD_{50} in selected host age levels
Heliothis zea
H. virescens
Trichoplusia ni
Galleria mellonella
(all exper.)

Nosema heliothidis
Thompson, A. C.; and Sikorowski, P. P., 1979, Comp. Biochem. and Physiol., v. 63A (3), 325-328
Nosema heliothidis, effects on fatty and amino acids in infected Heliothis zea larvae and pupae

Nosema hermanniae n. sp., illus.
Purrini, K.; and Baeumler, W., 1977, Zool. Anz., Jena, v. 199 (1-2), 107-112
Hermannia gibba: Munchen (Ebersberger Forst; Grafrath)

Nosema hermanniae
Purrini, K.; Bukva, V.; and Baeumler, W., 1979, Pedobiologia, v. 19 (5), 329-339
Hermannia gibba (Nephrozyten in Caeca, Fettzellen): Nadelwaldbestanden in Suddeutschland

Nosema kingi
Armstrong, E., 1977, Ztschr. Parasitenk., v. 53 (3), 311-315
Nosema kingi in Drosophila willistoni, transmission by either parent to offspring during copulation, more offspring became infected when copulation was delayed 2 weeks after infection of parent

Nosema locustae Canning
Ewen, A. B.; and Mukerji, M. K., 1979, Canad. Entom., v. 111 (8), 973-974
Nosema locustae, susceptibility of 5 grasshopper species to field applications, degree of infestation, pathogen overwintering: near Biggar, Saskatchewan
Melanoplus sanguinipes
M. packardii
M. infantalis
Camnula pellucida
Aeropedellus clavatus

Nosema locustae Canning
Henry, J. E.; Oma, E. A.; and Onsager, J. A., 1978, J. Econom. Entom., v. 71 (4), 629-632
Nosema locustae spores for control of grasshoppers, 2 methods of application compared, application of spores on wheat bran was significantly better in reducing grasshopper densities than application in ultra low volume aqueous spray

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Menapage, D. M.; Sackett, R. R.; and Wilson, W. T., 1978, J. Econom. Entom., v. 71 (2), 304-306
Nosema locustae, adult Apis mellifera not susceptible, safe to use Nosema locustae in biological control of pest insects

Nosema locustae Canning
Mussgnug, G. L.; and Henry, J. L., 1979, Acrida, v. 8 (2), 77-81
Nosema locustae, compatibility with malathion for term control of Melanoplus sanguinipes

Nosema lunatum Kellen et al., 1967
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora californica (Kellen and Lipa, 1960) comb. n.

Nosema lymantriae (Weiser), illus.
Saftoiu, A.; and Caloianu-Iordachel, M., 1978, Trav. Mus. Hist. Nat. "Gr. Antipa", Bucuresti, v. 19, 83-87
Nosema lymantriae in Lymantria dispar, ultrastructural data on intracellular development, possible method of penetration into host cell nucleus, potential of parasite in control of forest pests

Nosema micrococcus (Leger et Hesse, 1921) Weiser, 1961, illus.
Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135

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as syn. of Vairimorpha necatrix (Kramer, 1965) [n. comb.]

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Orthocladius sp.: environs de Montpellier

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Krinsky, W. L.; and Hayes, S. F., 1978, J. Protozool., v. 25 (2), 177-186
Nosema parkeri, fine structure of sporogonic stages from Ornithodoros parkeri (exper.)

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Gorske, S. F.; and Maddox, J. V., 1978, J. Invert. Path., v. 32 (3), 235-243
Nosema pilicornis sp. n. in Schizocerella pilicornis, transovarial transmission, high mortality in infected larvae; affects performance of S. pilicornis as biological control agent
Schizocerella pilicornis (nat. and exper.): Champaign, Collinsville, and areas around Chicago, Illinois; Stanford, Florida
Heliothis zea (exper.)
Spodoptera frugiperda (exper.)
Diacrisia virginica (exper.)
Autographa precationis (exper.)

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Rhysotritia ardua (freien Nephrozyten): Nadelwaldbestanden in Suddeutschland

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Cercaria rhionica VIII (parenchymal cells): reservoirs in valley of Rioni river, Western Caucasus

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Mattesia sp. and Nosema takapauensis in Costelytra zealandica, incidence among larvae, seasonal distribution, growth and development of diseased larvae, mortality, transmission by soil: New Zealand

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brief description
Macrotermes estherae: Waltair, India

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Syn.: Thelohania veliae *of* Weiser, 1961

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Nosema whitei
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Nosema whitei-infected Tribolium castaneum, relationship between body weight gains and food consumption

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Teetor-Barsch, G. E.; and Kramer, J. P., 1979, J. Invert. Path., v. 33 (3), 300-306
Octosporea muscaedomesticae, Nosema algerae, N. whitei, effect of lyophilization on infectivity of spores

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Nosema xiphidiocercariae sp. n., illus.
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Nosema zavreli Weiser, 1946, illus.
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    Simocephalus vetulus (cellules intestinales)

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    avian Plasmodium spp. of 4 subgenera, list of type material in Garnham Collection of Wellcome Museum of Medical Science; neotypes designated

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  Krylov, M. V., 1971, Parazitologiia, Leningrad, v. 5 (3), 201-208
    Piroplasmida, distribution in different host groups and zoogeographic regions, speculations on phylogeny

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  Krylov, M. V.; and Krylova, N. P., 1972, Parazitologiia, Leningrad, v. 6 (6), 493-505
    Piroplasmida, analysis of host specificity

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  Hadani, A.; Kaufman, W.; and Barnett, S. F., 1978, J. Parasitol., v. 64 (3), 501-503
    Nuttallia danii, development within Hyalomma anatolicum excavatum salivary glands held in organ culture in vitro

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  Iakunin, M. P.; and Krivkova, A. M., 1971, Parazitologiia, Leningrad, v. 5 (5), 462-465
    Emberiza bruniceps (erythrocytes)

Nuttallia frugilegica sp. n., illus.
  Iakunin, M. P.; and Krivkova, A. M., 1971, Parazitologiia, Leningrad, v. 5 (5), 462-465
    Corvus frugilegus (erythrocytes)

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  Iakunin, M. P.; and Krivkova, A. M., 1971, Parazitologiia, Leningrad, v. 5 (5), 462-465
    Galerida cristata (erythrocytes): Southern Kazakhstan

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  Iakunin, M. P.; and Krivkova, A. M., 1971, Parazitologiia, Leningrad, v. 5 (5), 462-465
    Upupa epops (erythrocytes): Muiunkumy desert

Nuttallia mujunkumica sp. n., illus.
  Iakunin, M. P.; and Krivkova, A. M., 1971, Parazitologiia, Leningrad, v. 5 (5), 462-465
    Passer indicus (erythrocytes): Muiunkumy desert

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  Krylov, M. V.; Kostenko, L. A.; and Snigirevskaia, E. S., 1973, Parazitologiia, Leningrad, v. 7 (6), 481-484
    Nuttallia musculi, trophozoites, merozoites, fine structure; bacteria-like bodies often found in cytoplasm

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    H. hypomelas
    all from southern Turkmenistan

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  Iakunin, M. P.; and Krivkova, A. M., 1971, Parazitologiia, Leningrad, v. 5 (5), 462-465
    Hirundo rustica (erythrocytes)

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  Rosin, G.; Landau, I.; and Hugot, J. P., 1978, Ann. Parasitol., v. 53 (5), 447-459
    principal taxonomic characters

Nycteria erardi n. sp., illus.
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    Nycteris arge (foie, sang): Makokou (Gabon)

Nycteria gabonensis n. sp., illus.
  Rosin, G.; Landau, I.; and Hugot, J. P., 1978, Ann. Parasitol., v. 53 (5), 447-459
    Rhinolophus sylvestris (foie, sang): Belinga (Gabon)

Nycteria houini n. sp., illus.
  Rosin, G.; Landau, I.; and Hugot, J. P., 1978, Ann. Parasitol., v. 53 (5), 447-459
    Nycteris nana (foie, sang): Bengoue (Gabon)

Nycteria krampitzi n. sp., illus.
  Rosin, G.; Landau, I.; and Hugot, J. P., 1978, Ann. Parasitol., v. 53 (5), 447-459
    Syn.: Hepatocystis sp. Landau, 1973 pro parte
    Rhinolophus sp. (foie, sang): Meya (Congo Brazzaville)

Nycteria medusiformis Garnham et Heisch, 1953, illus.
  Rosin, G.; Landau, I.; and Hugot, J. P., 1978, Ann. Parasitol., v. 53 (5), 447-459
    description
    Nycteris grandis (foie, sang): Makokou (Gabon)

Nyctotheroides Grasse, 1928
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includes subgenera: Spiroperistomatus subgen. n.; Sigmaperistomatus subgen. n.; Aduncuperistomatus subgen. n.; Nyctotheroides; Rectiperistomatus subgen. n.

Nyctotheroides
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139
subgen. of Nyctotheroides
tod of subgen.: N. (N.) cordiformis (Ehrenberg, 1838) Grasse, 1928

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Nyctotheroides Grasse, 1928
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Nyctotheroides, synopsis of Brazilian species, subgenus Nyctotheroides, review of recent work

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Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (2), 91-94
Nyctotheroides, synopsis of Brazilian species, subgenus Nyctotheroides, review of recent work

Nyctotheroides sp.
Amaro, A.; Sena, S.; and dos Santos, E., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 123
Aparasphenodon brunoi
Hyla cuspidata
all from Recreio dos Bandeirantes, Lagoinha das Taxas, na cidade do Rio de Janeiro, Estado da Guanabara, Brasil

Nyctotheroides africanus (Castellani, 1905) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides amarali (Carini, 1933) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Rectiperistomatus) amarali (Carini, 1933) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides (Rectiperistomatus) amarali (Carini, 1933) Amaro & Sena, 1967, illus.
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 105-107
Syn.: Nyctotherus amarali Carini, 1933

Nyctotheroides amphisbaenae (Carini, 1939) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Rectiperistomatus) amphisbaenae (Carini, 1939) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides (Rectiperistomatus) amphisbaenae (Carini, 1939) Amaro & Sena, 1967, illus.
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 105-107
Syn.: Nyctotherus amphisbaenae Carini, 1939

Nyctotheroides ampullariarum (Cordero, 1928) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides bertarelli (Carini, 1939) comb. n.
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Nyctotheroides (Nyctotheroides) bertarelli (Carini, 1939) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides breviceps (Uttangi, 1957) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides bufonis (Uttangi, 1957) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Nyctotheroides) bufonis (Uttangi, 1957) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides cacopusi (Uttangi, 1951) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Rectiperistomatus) cacopusi (Uttangi, 1951) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides ceratophris (Carini, 1940) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Nyctotheroides) ceratophris (Carini, 1940) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides cinctus (Carini, 1939) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides cochlearis (Uttangi, 1948) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides coralli (Carini, 1933) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Rectiperistomatus) coralli (Carini, 1933) Amaro & Sena, 1967 (tod of subg.)
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides (Rectiperistomatus) coralli (Carini, 1933) Amaro & Sena, 1967, illus.
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 105-107
Syn.: Nyctotherus coralli Carini, 1933

Nyctotheroides (Nyctotheroides) cordiformis (Ehrenberg, 1838) Grasse, 1928 (tod of subgen.)
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides gaudichaudi [? i. e. crossodactyli] (Carini, 1945) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Sigmaperistomatus) crossodactyli (Carini, 1945) comb. n. (tod of subgen.)
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides cunhai (Pinto, 1926) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides curtipes (Uttangi, 1957) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Aduncuperistomatus) curtipes (Uttangi, 1957) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides elegans (Carini, 1939) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Nyctotheroides) elegans (Carini, 1939) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides faba (Jakoby & Schaudinn, 1899) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides faberi (Carini, 1939) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides fragilis (Carini, 1939) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Nyctotheroides) fragilis (Carini, 1939) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides fulvus (Carini, 1939) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Nyctotheroides) fulvus (Carini, 1939) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides gibber (Carini, 1939) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Nyctotheroides) gibber (Carini, 1939) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides gibbosus (Boisson, 1959) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Aduncuperistomatus) gibbosus (Boisson, 1959) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides giganteus (Krause, 1906) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides heteronucleatus (Carini, 1939) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides hylae (Stein, 1867) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Nyctotheroides) hylae (Stein, 1867) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides hylae gigantea (Kifer, 1953) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Nyctotheroides) hylae gigantea (Kifer, 1953) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides hylae hylae (Surowiak, 1937) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Nyctotheroides) hylae hylae (Surowiak, 1937) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides hylae pelobatidis (Kifer, 1953) comb. n.
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Nyctotheroides (Nyctotheroides) hylae pelobatidis (Kifer, 1953) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides incertus (Carini, 1939) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Nyctotheroides) incertus (Carini, 1939) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides jaegeri (Carini, 1933) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Rectiperistomatus) jaegeri (Carini, 1933) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides (Rectiperistomatus) jaegeri (Carini, 1933) Amaro & Sena, 1967, illus.
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 105-107
Syn.: Nyctotherus jaegeri Carini, 1933

Nyctotheroides kalii (Uttangi, 1951) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides kempi (Ghosh, 1921) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides leidyi sp. n., illus.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 153-155
Crossodactylus gaudichaudii (intestino grosso): Silvestre, Rio de Janeiro, Estado da Guanabara, Brasil

Nyctotheroides (Nyctotheroides) leidyi Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides limnocharis (Uttangi, 1957) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Nyctotheroides) limnocharis (Uttangi, 1957) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides loricatus (Carini, 1939) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Nyctotheroides) loricatus (Carini, 1939) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides macropharyngeus (Bezzenberger, 1904) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Spiroperistomatus) macropharyngeus (Bezzenberger, 1904) Amaro & Sena, 1967 (tod of subgen.)
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides magnus (Bezzenberger, 1904) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Aduncuperistomatus) magnus (Bezzenberger, 1904) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides magnus malabarica (Mello, 1932) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Aduncuperistomatus) magnus malabarica (Mello, 1932) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides mazzai (Jorg, 1930) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides mogyanus (Carini, 1939) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Nyctotheroides) mogyanus (Carini, 1939) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides multisporiferus (Walker, 1909) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Aduncuperistomatus) nankingensis (Nie, 1932) comb. n.
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides neivai (Otamendi, 1945) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Rectiperistomatus) neivai (Otamendi, 1945) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides ondinae (Carini, 1939) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides ophidiae (Fantham & Porter, 1950) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Aduncuperistomatus) ophidiae (Fantham & Porter, 1950) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides oswaldoi (Carini, 1939) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides paludicolae (Carini, 1940) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Rectiperistomatus) paludicolae (Carini, 1940) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides (Rectiperistomatus) paludicolae (Carini, 1940) Amaro & Sena, 1967, illus.
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 105-107
Syn.: Nyctotherus paludicolae Carini, 1940

Nyctotheroides panesthiae (Yamasaki, 1939) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides pangasia (Tripathi, 1954) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides papillatus (Dobell, 1910) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides parvus (Walker, 1909) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides paulistanus (Carini, 1939) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Rectiperistomatus) paulistanus (Carini, 1939) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides (Rectiperistomatus) paulistanus (Carini, 1939) Amaro & Sena, 1967, illus.
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 105-107
Syn.: Nyctotherus paulistanus Carini, 1939

Nycotheroides puytoraci Essawy, 1978, illus.
Sadek, I. A., 1979, Exper. Parasitol., v. 48 (2), 239-244
Nyctotheroides puytoraci, stimulation of encystment in Bufo regularis injected with urine of patients with rheumatoid arthritis, may be new biological test organism for detecting abnormalities in tryptophan metabolism of humans

Nyctotheroides pyriformis (Nie, 1932) comb. n.
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides reniformis (Bhatia & Gulati, 1927) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides rhinocrici (Mello, 1954) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides ruber (Carini, 1939) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Nyctotheroides) ruber (Carini, 1939) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides spirostomatus sp. n., illus.
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (5), 175-176
Bufo crucifer (intestino grosso): Boca do Rio, Salvador, Estado da Bahia, Brasil

Nyctotheroides spirostomatus Amaro & Sena, 1968, illus.
Amaro, A.; and Helmer, J. L., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (1-2), 27-28
description
Leptodactylus ocellatus (intestino grosso (regiao cecal)): Bairro de Muruipe, Vitoria, Espirito Santo, Brasil

Nyctotheroides spirostomatus Amaro & Sena, 1968, illus.
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (5), 191-192
Leptodactylus ocellatus (intestino grosso): Manguinhos, Rio de Janeiro, Estado da Guanabara, Brasil

Nyctotheroides (Sigmaperistomatus) spirostomatus Amaro & Sena, 1968, illus.
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (1), 21-25

Nyctotheroides systoma (Uttangi, 1957) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Nyctotheroides) systoma (Uttangi, 1957) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides tejerai (Pinto, 1926) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Aduncuperistomatus) tejerai (Pinto, 1926) Amaro & Sena, 1967 (tod of subgen.)
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides tieteanus (Carini, 1939) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Rectiperistomatus) tieteanus (Carini, 1939) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides (Rectiperistomatus) tieteanus (Carini, 1939) Amaro & Sena, 1967, illus.
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 105-107
Syn.: Nyctotherus tieteanus Carini, 1939

Nyctotheroides tipulae (Grasse, 1928) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides trachysauri (Johnston, 1932) comb. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Sigmaperistomatus) trachysauri (Johnston, 1932) Amaro & Sena, 1967
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides travassosi (Cunha & Pinto, 1927) comb. n.
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides untanha (Carini, 1940) comb. n.
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Nyctotheroides) untanha (Carini, 1940) Amaro & Sena, 1967
  Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides vesiculatus (Boisson, 1957) comb. n.
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Nyctotheroides) vesiculatus (Boisson, 1957) Amaro & Sena, 1967
  Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides viannai (Pinto, 1926) comb. n.
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides (Rectiperistomatus) viannai (Pinto, 1926) Amaro & Sena, 1967
  Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139

Nyctotheroides (Rectiperistomatus) viannai (Pinto, 1926) Amaro & Sena, 1967, illus.
  Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 105-107
    Syn.: Nyctotherus viannai Pinto, 1926

Nyctotheroides vorax (Carini, 1939) comb. n.
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides vulgaris (Carini, 1939) comb. n.
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotheroides woodi (Amrein, 1952) comb. n.
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 129-131

Nyctotherus Leidy, 1849
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 149-151
    Nyctotherus, provisional list of species

Nyctotherus Leidy, 1849
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120
    includes subgenera: Recurviperistomatus subg. n.; Curviperistomatus subg. n.; Nyctotherus

Nyctotherus
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120
    subgen. of Nyctotherus
    tod of subgen.: N. (N.) velox Leidy, 1849

Nyctotherus Leidy, 1849
  Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (1), 1-6
    Nyctotherus, synopsis of Brazilian species, review of recent work

Nyctotherus
  Uttangi, J. C., 1958, J. Univ. Bombay, n. s., v. 26 (5), Sect. B (43), 50-64
    diagnostic characters
    key to Indian species

Nyctotherus amarali Carini, 1933
  Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 105-107
    as syn. of Nyctotheroides (Rectiperistomatus) amarali (Carini, 1933) Amaro & Sena, 1967

Nyctotherus amphisbaenae Carini, 1939
  Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 105-107
    as syn. of Nyctotheroides (Rectiperistomatus) amphisbaenae (Carini, 1939) Amaro & Sena, 1967

Nyctotherus (Curviperistomatus) barberoi Schouten, 1934
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus (Nyctotherus) befasyi Tuzet & Manier, 1958
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus (Recurviperistomatus) beltrani Hegner, 1940
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus (Curviperistomatus) boipevae Carini, 1933
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus breviceps n. sp., illus.
  Uttangi, J. C., 1958, J. Univ. Bombay, n. s., v. 26 (5), Sect. B (43), 50-64
    key
    Rana breviceps (intestine): Dharwar, Bombay Karnatak

Nyctotherus bufonis n. sp., illus.
  Uttangi, J. C., 1958, J. Univ. Bombay, n. s., v. 26 (5), Sect. B (43), 50-64
    key
    Bufo melanostictus (rectum): Dharwar, Bombay Karnatak

Nyctotherus (Nyctotherus) buissoni Pinto, 1926
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus (Recurviperistomatus) cheni Wichterman, 1934
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus (Nyctotherus) congoi Tuzet, Manier & Jolivet, 1956
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus coralli Carini, 1933
  Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 105-107
    as syn. of Nyctotheroides (Rectiperistomatus) coralli (Carini, 1933) Amaro & Sena, 1967

Nyctotherus cordiformis (Ehrenberg, 1831)
  Frandsen, F., 1974, Acta Parasitol. Polon., v. 22 (1-11), 49-66
    Rana esculenta
    R. arvalis
    Bufo bufo
    R. temporaria
    (rectum of all): all from Denmark

Nyctotherus cordiformis Stein
  Uttangi, J. C., 1958, J. Univ. Bombay, n. s., v. 26 (5), Sect. B (43), 50-64
    key
    Rana curtipes
    R. breviceps
    all from Dharwar, Bombay Karnatak

Nyctotherus cordiformis
  Vinnikova, N. V.; and Golikova, M. N., 1978, Tsitologiia, v. 20 (7), 731-735
    Nyctotherus cordiformis, developed polytene chromosomes in macronuclear anlage, ultrastructure

Nyctotherus cordiformis Stein, 1859, illus.
  Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
    description
    Bombina bombina
    B. variegata
    Bufo bufo
    Pelobates fuscus
    Rana esculenta
    R. temporaria
    R. arvalis
    R. ridibunda
    all from CSSR

Nyctotherus cordiformis var. hylae Stein, 1859
  Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
    as syn. of N. hylae Surowiak, 1937

Nyctotherus curtipes n. sp., illus.
  Uttangi, J. C., 1958, J. Univ. Bombay, n. s., v. 26 (5), Sect. B (43), 50-64
    key
    Rana curtipes (rectum): Castle-rock, Dharwar, Bombay Karnatak

Nyctotherus (Nyctotherus) diplopodae Karandikar & Rodgi, 1956
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus (Curviperistomatus) gamarrai Schouten, 1937
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus (Curviperistomatus) gongylorrhus Karandikar & Rodgi, 1956
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus (Nyctotherus) hormeticae Carini, 1937
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus (Nyctotherus) hoyai Tuzet & Theorides, 1956
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus hylae Rosenberg, 1937
  Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
    as syn. of N. hylae Surowiak, 1937

Nyctotherus hylae Surowiak, 1937, illus.
  Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
    synonymy, description
    Hyla arborea
    Pelobates fuscus
    (strevo of all): all from CSSR

Nyctotherus (Curviperistomatus) inflatus Tuzet & Manier, 1958
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus (Curviperistomatus) jaboti Carini, 1938, illus.
  Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (1), 1-6

Nyctotherus jaegeri Carini, 1933
  Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 105-107
    as syn. of Nyctotheroides (Rectiperistomatus) jaegeri (Carini, 1933) Amaro & Sena, 1967

Nyctotherus limnocharis n. sp., illus.
  Uttangi, J. C., 1958, J. Univ. Bombay, n. s., v. 26 (5), Sect. B (43), 50-64
    key
    Rana limnocharis (rectal contents): Dharwar, Bombay Karnatak

Nyctotherus macropharyngeus Bezzenberger
  Uttangi, J. C., 1958, J. Univ. Bombay, n. s., v. 26 (5), Sect. B (43), 50-64
    key
    Rana curtipes
    R. verrucosa
    R. breviceps
    all from Dharwar, Bombay Karnatak

Nyctotherus (Nyctotherus) madagascari Tuzet & Manier, 1954
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus magnus Bezzenberger
  Uttangi, J. C., 1958, J. Univ. Bombay, n. s., v. 26 (5), Sect. B (43), 50-64
    key
    Rana limnocharis
    R. cyanophlyctis
    R. curtipes
    R. breviceps
    all from Dharwar, Bombay Karnatak

Nyctotherus (Nyctotherus) mandrakae Tuzet & Manier, 1954
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus (Nyctotherus) mardonii Tuzet, Manier & Vogeli-Zuber, 1952
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus (Curviperistomatus) neocurtillae Carini, 1937
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus (Curviperistomatus) ochoterenai
Schouten, 1937 (tod of subgen.)
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol.
  Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus (Curviperistomatus) osmodermae
Zeliff, 1933
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol.
  Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus (Nyctotherus) ovalis Leidy, 1849
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol.
  Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus ovalis
  Hayunga, E. G., 1977, Tr. Am. Micr. Soc., v.
  96 (1), 153-155
    parasitic protozoa, mixture of polyvinyl
    alcohol and Bouin's solution found to be
    satisfactory fixative and adhesive for
    smears, smears may be stored dry prior to
    staining with little apparent damage to
    protozoa

Nyctotherus paludicolae Carini, 1940
  Amaro, A.; and Sena, S., 1968, Atas Soc. Biol.
  Rio de Janeiro, v. 12 (3), 105-107
    as syn. of Nyctotheroides (Rectiperistomatus)
    paludicolae (Carini, 1940) Amaro & Sena,
    1967

Nyctotherus paulistanus Carini, 1939
  Amaro, A.; and Sena, S., 1968, Atas Soc. Biol.
  Rio de Janeiro, v. 12 (3), 105-107
    as syn. of Nyctotheroides (Rectiperistomatus)
    paulistanus (Carini, 1939) Amaro & Sena,
    1967

Nyctotherus (Recurviperistomatus) pintoi Carini,
1933, illus.
  Amaro, A.; and Sena, S., 1968, Atas Soc. Biol.
  Rio de Janeiro, v. 12 (1), 1-6

Nyctotherus (Nyctotherus) silvestrianus Kirby,
1932
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol.
  Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus (Recurviperistomatus) sokoloffi
Schouten, 1940
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol.
  Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus systoma n. sp., illus.
  Uttangi, J. C., 1958, J. Univ. Bombay, n. s.,
  v. 26 (5), Sect. B (43), 50-64
    key
    Uperodon systoma (rectum): Dharwar, Bombay
    Karnatak

Nyctotherus (Recurviperistomatus) teleacus
Geiman & Wichterman, 1937
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol.
  Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus (Nyctotherus) termitis Dobell, 1910
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol.
  Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus (Nyctotherus) thyropygus Karandikar
& Rodgi, 1956
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol.
  Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus tieteanus Carini, 1939
  Amaro, A.; and Sena, S., 1968, Atas Soc. Biol.
  Rio de Janeiro, v. 12 (3), 105-107
    as syn. of Nyctotheroides (Rectiperistomatus)
    tieteanus (Carini, 1939) Amaro & Sena, 1967

Nyctotherus (Recurviperistomatus) uianchoi
Kidder, 1937 (tod of subg.)
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol.
  Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus (Nyctotherus) velox Leidy, 1849
(tod of subgen.)
  Amaro, A.; and Sena, S., 1967, Atas Soc. Biol.
  Rio de Janeiro, v. 11 (3), 119-120

Nyctotherus viannai Pinto, 1926
  Amaro, A.; and Sena, S., 1968, Atas Soc. Biol.
  Rio de Janeiro, v. 12 (3), 105-107
    as syn. of Nyctotheroides (Rectiperistomatus)
    viannai (Pinto, 1926) Amaro & Sena, 1967

Octomitus dujardini Dobell 1909 p. p.
  Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
    as syn. of Spironucleus elegans Lavier (1936)

Octomitus dujardini Dobell (1909)
  Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
    as syn. of Hexamita intestinalis Dujardin, 1841

Octomitus intestinalis Jirovec (1953)
  Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
    as syn. of Hexamita intestinalis Dujardin, 1841

Octomitus muris, illus.
  Tumka, A. F., 1972, Parazitologiia, Leningrad, v. 6 (3), 222-228
    Lamblia muris, Trichomonas muris, Octomitus muris, localization in white mice exposed to x-irradiation

Octomitus neglecta Lavier, 1936, illus.
  Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
    description
    Syn.: Syndiomita neglecta Lavier (1936)
    Bombina bombina
    B. variegata
    Bufo bufo
    B. viridis
    B. calamita
    Pelobates fuscus
    Hyla arborea
    Rana esculenta
    R. temporaria
    R. dalmatina
    all from CSSR

Octosporea chironomi Weiser, 1943, illus.
  Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135

Octosporea effeminans Bulnheim & Vavra, 1968, illus.
  Bulnheim, H. P., 1978, Helgoland. Wissensch. Meeresuntersuchg., v. 31 (1-2), 1-33
    Octosporea effeminans and Thelohania herediteria in Gammarus duebeni duebeni, feminizing influence exerted on host's offspring by parasites, role of salinity and temperature on sex determination by parasites
    Gammarus duebeni duebeni: Federal Republic of Germany; Finland; Sweden; Denmark; France
    G. duebeni celticus: France

Octosporea muscaedomesticae
  Teetor-Barsch, G. E.; and Kramer, J. P., 1979, J. Invert. Path., v. 33 (3), 300-306
    Octosporea muscaedomesticae, Nosema algerae, N. whitei, effect of lyophilization on infectivity of spores

Octosporea procellioi n. sp.
  Kalavati, C.; and Narasimhamurti, C. C., 1976, Proc. 1. Internat. Colloquium Invert. Path. and 9. Ann. Meet. Soc. Invert. Path. (Kingston, Canada, Aug. 29-Sept. 3), 391
    Porcellio laevus (cells of haemocoelic fluid): Visakhapatnam (Andhra Pradesh, India)

Opalina Purkinje & Valentin, 1835
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34
    Opalinidae
    includes: Opalina; Nipponica subgen. n.; Angusta subgen. n.

Opalina
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34
    subgen. of Opalina
    type sp.: Opalina (O.) ranarum (Ehrenberg, 1831) Purkinje & Valentin, 1835

Opalina Purkinje & Valentin, 1835
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
    Opalininae
    key to subgenera, key

Opalina
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
    subgen. of Opalina, key

Opalina Purkinje & Valentin, 1835
  Amaro, A., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 149-152
    Opalina, subgenus Angusta, synopsis of Brazilian species

Opalina Purkinje & Valentin, 1835
  Earl, P. R., 1979, Tr. Am. Micr. Soc., v. 98 (4), 549-557
    Opalininae

Opalina antilliensis Metcalf, 1914
  Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (5-6), 307-314
    as syn. of Zelleriella (Z.) antilliensis (Metcalf, 1914) Metcalf, 1923

Opalina (Opalina) asiatica Metcalf, 1923
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina brasiliensis Pinto, 1918
  Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (5-6), 307-314
    as syn. of Zelleriella (Z.) brasiliensis (Pinto, 1918) Metcalf, 1923

Opalina (Opalina) bufoxena Metcalf, 1923
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Nipponica) camerunensis Metcalf, 1923
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Nipponica) cancrivora Boisson, 1957
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Opalina) cincta Corlin, 1913
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Nipponica) coracoidea Bezzenberger, 1904
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Nipponica) coracoidea lahorensis Bhatia & Gulati, 1927
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Nipponica) draytonii Metcalf, 1923
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Angusta) elongata Carini, 1937
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
    key

Opalina (Angusta) faber Carini, 1937
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
    key

Opalina (Opalina) gigantea Metcalf, 1923
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Nipponica) japonica Sugiyama, 1920 (tod of subgen.), illus.
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Nipponica) japonica dharwarensis Uttangi, 1953
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Nipponica) japonica javensis Metcalf, 1940
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Angusta) nebulosa Carini, 1937
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
    key

Opalina (Angusta) obtrigona Stein, 1864 (tod of subgen.), illus.
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Nipponica) panamensis Metcalf, 1923
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina paulista Pinto, 1926
  Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (5-6), 237-243
    as syn. of Zelleriella paulista (Pinto, 1926) comb. n.

Opalina (Opalina) raddei Metcalf, 1923
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Angusta) radiana Carini, 1937
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
    key

Opalina (Opalina) ranarum (Ehrenberg, 1831) Purkinje & Valentin, 1835 (tod of subg.), illus.
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina ranarum, illus.
  El Mofty, M. M., 1977, Proc. 1. Conf. Biol. Aspects Saudi Arabia, 246-256
    Opalina ranarum in Rana ribidunda, use as biological test animals for detecting carcinogenic tryptophan metabolites

Opalina ranarum (Ehrenberg, 1831)
  Frandsen, F., 1974, Acta Parasitol. Polon., v. 22 (1-11), 49-66
    Rana arvalis
    R. dalmatina
    Bufo bufo
    Rana temporaria
    Bufo viridis
    B. calamita
    (rectum of all): all from Denmark

Opalina ranarum Purkinje et Valentin, 1835, illus.
  Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
    description
    Bombina bombina
    B. variegata
    Bufo bufo
    B. viridis
    B. calamita
    Pelobates fuscus
    Hyla arborea
    Rana esculenta
    R. temporaria
    R. dalmatina
    R. ridibunda
    all from CSSR

Opalina (Opalina) ranarum arvalis Metcalf, 1923
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Opalina) ranarum cinctoidea Metcalf, 1923
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Opalina) ranarum lata Metcalf, 1923
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Opalina) ranarum latouchi Liu, 1958
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Opalina) ranarum orbiculata Metcalf, 1940
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Opalina) ranarum parvipalmatae Metcalf, 1923
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Opalina) ranarum smithi Metcalf, 1923
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalina (Opalina) ranarum truncata Metcalf, 1923
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 33-34

Opalinata Sandon, 1976
Earl, P. R., 1979, Tr. Am. Micr. Soc., v. 98 (4), 549-557
opalinids, taxonomy and problems of identification, factors of zoogeographical distribution (Metcalf's land bridges, continental drift and anuran dispersal)
includes: Opalinatea

Opalinatea Corliss & Balamuth, 1963
Earl, P. R., 1979, Tr. Am. Micr. Soc., v. 98 (4), 549-557
Opalinata
includes: Opalinida

Opalinida Poche, 1913
Earl, P. R., 1979, Tr. Am. Micr. Soc., v. 98 (4), 549-557
Opalinatea
includes: Opalinidae

Opalinidae Claus, 1874
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
key to subfamilies

[Opalinidae] opalinideos
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 125-127
[Opalinidae], paratypes of species in Metcalf collection deposited in American Museum of Natural History: United States

[Opalinidae] opalinideos
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (2), 47-49
list of recent species

Opalinidae Claus, 1874
Earl, P. R., 1979, Tr. Am. Micr. Soc., v. 98 (4), 549-557
Opalinida
includes: Protoopalininae; Hegneriellina [sic]; Opalininae

Opalininae Metcalf, 1920
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
Opalinidae
key to genera, key

Opalininae Claus, 1874
Earl, P. R., 1979, Tr. Am. Micr. Soc., v. 98 (4), 549-557
Opalinidae
includes: Opalina

Opalinopsidae
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key

Opalinopsis Foettinger
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key

Ophioidina Mingazzini, 1891
Levine, N. D., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 695-702
as syn. of Lecudina Mingazzini, 1891

Orcheobius Schuberg and Kunze, 1906
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Orcheobius cruzi Carini and Pinto, 1930
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Orcheobius herpobdellae Schuberg and Kunze, 1906
Levine, N. D., 1977, Protozoology, v. 3, 173-182

Orchitophrya Cepede
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key

Orientalis subgen. n.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 23-24
subgen. of Protoopalina
type sp.: Protoopalina (Orientalis) montana Metcalf, 1923

Orientalis
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
subgen. of Protoopalina, key

Ovalis subgen. n.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 23-24
subgen. of Protoopalina
type sp.: Protoopalina (Ovalis) ovoidea Metcalf, 1923

Ovalis
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
subgen. of Protoopalina, key

Ovoplasma orientale Marzinowsky & Bogrow, 1904
Gardener, P. J., 1977, Trop. Dis. Bull., v. 74 (12), 1069-1088
as syn. of Leishmania tropica (Wright, 1903) Luehe, 1906

Pachyporospora
Theodorides, J., 1979, Compt. Rend. Acad. Sc., Paris, v. 289, s. D, Sc. Nat. (3), 363-365
intestinal eugregarines of Brachyura, distribution among hosts confirms validity of new classification system for hosts

Pachyporospora retorta Ormieres pacifica ssp. nov., illus.
Theodorides, J., 1977, Canad. J. Zool., v. 55 (1), 237-241
Cancer productus (intestin): Nanaimo, Colombie-Britannique; ile San Juan, etat de Washington, U.S.A.

Paradinemula Monch.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
Peranemidae, key
key to species, includes: Paradinemula polonica Monch.; P. moraviensis Mich.; P. ocelli Mich.

Paradinemula ocelli, illus.
Michajlow, W., 1977, Zhurnal Obsh. Biol., v. 38 (5), 657-675

Paradistigma gen. n.
Wita, I., 1974, Acta Parasitol. Polon., v. 22 (35-44), 365-391
Euglenidae
[no type designated]

Paradistigma sp.
Michajlow, W., 1978, Bull. Acad. Polon. Sc., Cl. V, s. Sc. Biol., v. 26 (1), 51-54
Pseudoboeckella silvestri: reservoir on King George Island, Southern Shetland Islands, Antarctica

Paradistigma ergasili sp. n., illus.
Wita, I., 1974, Acta Parasitol. Polon., v. 22 (35-44), 365-391
Paradistigma n. spp., life cycles, flagellate locomotion, attraction to specific copepod host, speculation on evolution to parasitism
Ergasilus sieboldi (body cavity, limbs, and antennae of nauplii; egg bag): Strengiel Lake, Poland

Paradistigma hyperparasiticum sp. n., illus.
Wita, I., 1974, Acta Parasitol. Polon., v. 22 (35-44), 365-391
Paradistigma n. spp., life cycles, flagellate locomotion, attraction to specific copepod host, speculation on evolution to parasitism
Ergasilus sieboldi (body cavity, limbs, and antennae of nauplii): Swiecajty Lake, Poland

Paradistigma triangulatum sp. n., illus.
Wita, I., 1974, Acta Parasitol. Polon., v. 22 (35-44), 365-391
Paradistigma n. spp., life cycles, flagellate locomotion, attraction to specific copepod host, speculation on evolution to parasitism
Ergasilus sieboldi (body cavity of nauplii; eggs): Mamry Lake, Poland

Paradistigmoides g. n.
Michajlow, W.; and Wita, I., 1976, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 24 (5), 283-288
Euglenidae
mt: P. kenyensis sp. n.

Paradistigmoides bangalorensis sp. n., illus.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (8), 527-529
Mesocyclops leuckarti: Botanical Gardens Lal-Bagh, Bangalore, India (Karnataka)

Paradistigmoides kenyensis g. n., sp. n. (mt), illus.
Michajlow, W.; and Wita, I., 1976, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 24 (5), 283-288
Mesocyclops crassus
Microcyclops varicans
(body cavity of all): all from bays of river Tsavo, Tsavo National Park, Kenya

Paradistigmoides kenyensis Mich. et Wita, illus.
Michajlow, W., 1977, Zhurnal Obsh. Biol., v. 38 (5), 657-675
Microcyclops varicans

Parahaemoproteus (or Haemoproteus) [sp.]
Baker, J. R., 1974, J. Zool., London, v. 172 (2), 169-190
Fringilla coelebs
Garrulus glandarius
Muscicapa striata
Passer domesticus
Sturnus vulgaris
Turdus ericetorum
T. merula
Athene noctua
Columba palumbus
Lagopus scoticus
all from Britain

Parahistomonas wenrichi
Fine, P. E. M., 1979, J. Parasitol., v. 65 (2), 320-321
Heterakis gallinarum, egg output of Parahistomonas wenrichi-infected and uninfected strains, difference not statistically significant

Paramarteilia gen. nov.
Ginsburger-Vogel, T.; and Desportes, I., 1979, J. Protozool., v. 26 (3), 390-403
Marteiliidae
mt: P. orchestiae sp. n.

Paramarteilia orchestiae sp. n. (mt), illus.
Ginsburger-Vogel, T.; and Desportes, I., 1979, J. Protozool., v. 26 (3), 390-403
Paramarteilia orchestiae gen. n., sp. n., ultrastructure of sporulation
Orchestia gammarellus (tous les tissus a l'exception l'epithelium digestif et du systeme nerveux): Estuaire de la Penze (Nord-Finistere, France)

Paramoeba sp., illus.
Sawyer, T. K., 1969, Proc. 1968 National Shellfish Ass., v. 59, 60-64
Paramoeba sp. in Callinectes sapidus, rate of infection during and after peak periods of crab mortality: Chincoteague Bay, Virginia

Paranaplasma caudata
Rodriguez, O. N.; et al., 1975, Folia Vet., v. 19 (1-2), 249-255
Anaplasma marginale, Paranaplasma caudata, antigenic differences determined by passive hemagglutination and hemagglutination inhibition test

Parastasia coelomae, illus.
Michajlow, W., 1977, Zhurnal Obsh. Biol., v. 38 (5), 657-675

Parastasia cyclops, illus.
Michajlow, W., 1977, Zhurnal Obsh. Biol., v. 38 (5), 657-675

Parastasia kieviensis sp. n., illus.
Wita, I., 1978, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 26 (8), 555-560
Parastasia kieviensis sp. n., developmental cycle
Eucyclops sèrrulatus
E. macruroides
Acanthocyclops viridis
(intestine of all): all from fishery ponds in environs of Kiev (Sovki), Ukraine

Parastasia norvegica, illus.
Michajlow, W., 1977, Zhurnal Obsh. Biol., v. 38 (5), 657-675

Parastasia ucrainica sp. n., illus.
Wita, I., 1978, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 26 (7), 489-494
Acanthocyclops viridis
Eucyclops serrulatus
(gut of both): both from Sovki fishery ponds in environs of Kiev (Ukraine)

Parastasia ucrainica
Wita, I., 1978, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 26 (8), 555-560
Eucyclops serrulatus
Acanthocyclops viridis
all from fishery ponds in environs of Kiev (Sovki), Ukraine

Parastasiella naupliorum Mich.
Michajlow, W., 1978, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 26 (4), 269-272
Mesocyclops leuckarti: water body of Botanical Gardens, Brisbane and Melbourne, Australia

Parastasiella ovorum, illus.
Michajlow, W., 1977, Zhurnal Obsh. Biol., v. 38 (5), 657-675

Parastasiella tardiva sp. n., illus.
Michajlow, W., 1970, Acta Parasitol. Polon., v. 18 (42-50), 473-482
Acanthocyclops vernalis: Lithuanian SSR

Parastasiella vastans Michajlow, 1966
Michajlow, W., 1970, Acta Parasitol. Polon., v. 18 (42-50), 473-482
Eucyclops macruroides

Parastasiella velox Mich., illus.
Michajlow, W., 1977, Zhurnal Obsh. Biol., v. 38 (5), 657-675

Parathelohania Codreanu, 1966
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Thelohaniidae fam. n., key

Parathelohania, undescribed species, illus.
Hazard, E. I.; et al., 1979, J. Parasitol., v. 65 (1), 117-122
Amblyospora, meiotic configurations and synaptonemal complexes, Parathelohania, synaptonemal complexes; implications in life cycles
Anopheles bradleyi

Parathelohania africana Hazard and Anthony, 1974, illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Syn.: Thelohania legeri: Fantham et al. (in part), 1941

Parathelohania anomala (Sen, 1941) Hazard and Anthony, 1974
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
synonymy

Parathelohania anophelis (Kudo)
Hazard, E. I.; et al., 1979, J. Parasitol., v. 65 (1), 117-122
Amblyospora, meiotic configurations and synaptonemal complexes, Parathelohania, synaptonemal complexes; implications in life cycles
Anopheles quadrimaculatus

Parathelohania anophelis (Kudo, 1924) Hazard and Anthony, 1974, illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
synonymy, description

Parathelohania barra (Pillai, 1968) comb. n.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Syn.: Thelohania barra Pillai, 1968

Parathelohania chagrasensis sp. n., illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Aedeomyia squamipennis (adipose tissue): Chagras River, Canal Zone

Parathelohania illinoisensis (Kudo, 1921) Hazard and Anthony, 1974, illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
synonymy

Parathelohania indica (Kudo, 1929) Hazard and Anthony, 1974
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
synonymy

Parathelohania legeri: Simmers, 1974
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Parathelohania illinoisensis (Kudo, 1921) Hazard and Anthony, 1974

Parathelohania legeri (Hesse, 1904) Codreanu, 1966
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
synonymy

Parathelohania legeri (Hesse, 1904) Hazard et Anthony, 1974
Kilochitskii, P. Ia., 1977, Vestnik Zool., Akad. Nauk Ukrainsk. SSR, Inst. Zool. (4), 71-75
Anopheles maculipennis
A. hyrcanus
all from southern Ukraine

Parathelohania obesa (Kudo)
Hazard, E. I.; et al., 1979, J. Parasitol., v. 65 (1), 117-122
Amblyospora, meiotic configurations and synaptonemal complexes, Parathelohania, synaptonemal complexes; implications in life cycles
Anopheles crucians

Parathelohania obesa (Kudo, 1924) Hazard and Anthony, 1974, illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
synonymy, description

Parathelohania obscura (Kudo, 1929) Hazard and Anthony, 1974
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
synonymy

Parathelohania octolagenella Hazard and Anthony, 1974, illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
description

Parathelohania opacita: Simmers, 1974
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora opacita (Kudo, 1922) comb. n.

Parathelohania periculosa (Kellen and Wills, 1962) Hazard and Anthony, 1974
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
synonymy

Paratrichodina alburni (Vojtek, 1957) Lom, 1963, illus.
Lom, J.; and Haldar, D. P., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 527-541
trichodinids endoparasitic in fishes, survey of literature records together with some new observations
Syn.: Trichodina alburni Vojtek, 1957
Alburnus alburnus: Tisa river, Hungary

Paratrichodina degiustii n. sp., illus.
Lom, J.; and Haldar, D. P., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 527-541
trichodinids endoparasitic in fishes, survey of literature records together with some new observations
Notropis heterodon (urinary bladder and ureters): vicinity of Detroit, Michigan

Paratrichodina phoxini Lom, 1963, illus.
Lom, J.; and Haldar, D. P., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 527-541
trichodinids endoparasitic in fishes, survey of literature records together with some new observations
Phoxinus phoxinus: vicinity of Prague, Czechoslovakia

Paucinucleata subgen. n.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31
subgen. of Cepedea
type sp.: Cepedea (Paucinucleata) lanceolata (Bezzenberger, 1904) Metcalf, 1923

Paucinucleata
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
subgen. of Cepedea, key

Pectenita gen. n.
Iankovskii, A. V., 1973, Parazitologiia, Leningrad, v. 7 (3), 214-219
Hymenostomatida, Tetrahymenina, Entodiscidae fam. n.
tod: P. golikowi sp. n.

Pectenita golikowi gen. et sp. n. (tod), illus.
Iankovskii, A. V., 1973, Parazitologiia, Leningrad, v. 7 (3), 214-219
Mizuhopecten yessoensis (intestine): sublittoral lagoon of Busse, north Sakhalin; Veslo peninsula, north Kunashiri

Pegmatheca gen. n.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Thelohaniidae fam. n., key
tod: P. simulii sp. n.

Pegmatheca simulii sp. n. (tod), illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Simulium tuberosum (adipose tissue): Hatchet Creek on Waldo Road, near Gainesville, Florida, U.S.A.

Pentatrichomonas hominis
Engbaek, K., 1978, Ugeskr. Laeger, v. 140 (1), 14-17
Giardia lamblia and other intestinal parasites of children in day-nurseries and kindergartens, incidence, clinical symptoms, management: Denmark

Pentatrichomonas hominis
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Pentatrichomonas hominis (Davaine), illus.
Warton, A.; and Honigberg, B. M., 1979, J. Protozool., v. 26 (1), 56-62
Hypotrichomonas acosta, Trichomonas vaginalis, Pentatrichomonas hominis, Tritrichomonas foetus, scanning electron microscopy

Pentatrichomonas intestinalis, illus.
Harvey, J., 1973, Saguenay Med., v. 20 (3), 146-156
parasitic flagellates, life cycle, infection in man, clinical signs, therapy, brief review

Peranemidae Klebs (Hollande)
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (2), 101-106
key to genera, parasitic species
includes: Dinemina; Dinema; Paradinemula; Dinemula; Mononema

Perezia nelsoni (Sprague, 1950) comb. n.
Vivares, C. P.; and Sprague, V., 1979, J. Invert. Path., v. 33 (1), 40-52
Syn.: Ameson nelsoni (Sprague, 1950) Sprague, 1977

Perkinsea class n.
Levine, N. D., 1978, J. Parasitol., v. 64 (3), 549
includes: Perkinsida ord. n.

Perkinsida ord. n.
Levine, N. D., 1978, J. Parasitol., v. 64 (3), 549
Perkinsea class n.
includes: Perkinsidae fam. n.

Perkinsidae fam. n.
Levine, N. D., 1978, J. Parasitol., v. 64 (3), 549
Perkinsida ord. n.
includes: Perkinsus gen. n.

Perkinsus gen. n.
Levine, N. D., 1978, J. Parasitol., v. 64 (3), 549
Perkinsidae fam. n.
tod: P. marinus (Mackin, Owen, and Collier, 1950) comb. n.

Perkinsus marinus (Mackin, Owen, and Collier, 1950) comb. n. (tod)
Levine, N. D., 1978, J. Parasitol., v. 64 (3), 549
Syns.: Dermocystidium marinum Mackin, Owen, and Collier, 1950; Labyrinthomyxa marina (Mackin, Owen, and Collier, 1950) Mackin and Ray, 1966

Petalomonas parasitica sp. n., illus.
Michajlow, W., 1977, Bull. Acad. Polon. Sc., Cl. II, s. Sc. Biol., v. 25 (6), 391-394
Eucyclops serrulatus: water reservoir, Ranganathittu bird preserve, near Mysore, India

Phrynomantidis subgen. n.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31
subgen. of Cepedea
type sp.: Cepedea (Phrynomantidis) phrynomantidis Metcalf, 1923

Phrynomantidis
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
subgen. of Cepedea, key

Phytomonas davidi
Fish, W. R.; Holz, G. G., jr.; and Beach, D. H., 1978, J. Parasitol., v. 64 (3), 546-547
26 trypanosomatid species, cultivation in new chemically-defined medium RE III

Phytomonas davidi
Simpson, L., 1979, Proc. National Acad. Sc., v. 76 (4), 1585-1588
hemoflagellate protozoa, method for isolation of maxicircle component of kinetoplast DNA

Pileocephalus heerii Koelliker, illus.
Baudoin, J., 1965, Arch. Zool. Exper. et Gen., v. 105 (2), 245-258
description

Pileocephalus hovassei, n. sp., illus.
Baudoin, J., 1965, Arch. Zool. Exper. et Gen., v. 105 (2), 245-258
Limnophilus sp.: Auvergne (bras herbeux de l'Allier a Joze; etang de Saint-Jean-d'Heurs; lac d'Aydat)

Pileocephalus sinensis Schneider, illus.
Baudoin, J., 1965, Arch. Zool. Exper. et Gen., v. 105 (2), 245-258
description

Pilosporella gen. n.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Thelohaniidae fam. n., key
tod: P. fishi sp. n.

Pilosporella chapmani sp. n., illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Aedes triseriatus (adipose tissue, in head, and sixth abdominal segment): near Lake Charles, Louisiana, U.S.A.

Pilosporella fishi sp. n. (tod), illus.
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Wyeomyia vanduzeei (adipose tissue): near Vero Beach, Florida, U.S.A.

Pilosporella fishi Hazard and Oldacre, 1975
Frank, J. H.; and Curtis, G. A., 1977, Mosquito News, v. 37 (3), 487-489
Pilosporella fishi in Wyeomyia vanduzeei, greater mortality of female than male mosquitoes, incidence too low to affect size of mosquito population: near Vero Beach, Florida

Piroplasma bigeminum
Nikol'skii, S. N.; Nikiforenko, V. I.; and Pozov, S. A., 1977, Veterinariia, Moskva (4), 71-75
Piroplasma jakimovi, cattle, morphological and biological comparison with P. bigeminum, epizootiology (Ixodes ricinus as main vector; frequent association with leptospirosis), treatment: Siberia

Piroplasma canis
Ramadan, P.; and Bauer, M., 1978, Vet. Arhiv, Zagreb, v. 48 (5), 251-256
Piroplasma canis, dogs, clinical pathology, particular reference to haemogram

Piroplasma donovani Laveran & Mesnil, 1903
Gardener, P. J., 1977, Trop. Dis. Bull., v. 74 (12), 1069-1088
as syn. of Leishmania donovani (Laveran & Mesnil, 1903) Ross, 1903

Piroplasma jakimovi
Nikol'skii, S. N.; Nikiforenko, V. I.; and Pozov, S. A., 1977, Veterinariia, Moskva (4), 71-75
Piroplasma jakimovi, cattle, morphological and biological comparison with P. bigeminum, epizootiology (Ixodes ricinus as main vector; frequent association with leptospirosis), treatment: Siberia

Piroplasma ovis
Charyev, O. Ch., 1977, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (6), 70-71
piroplasmids, sheep, pure and mixed infections: Turkmenia

Piroplasmida
Krylov, M. V., 1971, Parazitologiia, Leningrad, v. 5 (3), 201-208
Piroplasmida, distribution in different host groups and zoogeographic regions, speculations on phylogeny

Piroplasmida
Krylov, M. V.; and Krylova, N. P., 1972, Parazitologiia, Leningrad, v. 6 (6), 493-505
Piroplasmida, analysis of host specificity

Piroplasmosis
Dubovyi, S. Z.; et al., 1977, Veterinariia, Moskva (3), 71-72
babesiasis, piroplasmosis, cattle, dimidine as effective chemoprophylaxis under pasture conditions with presence of vector, Boophilus calcaratus, comparison with azidine

Piroplasmosis
Mamaev, N. Kh.; Golin, P. I.; and Omarov, M. V., 1978, Veterinariia, Moskva (11), 73-74
Hypoderma, B[oophilus] calcaratus, cattle, chlorophos treatment, prophylaxis against piroplasmosis: Dagestan

Piroplasmosis
Meshkov, S.; Iotov, I.; and Sirachev, D., 1977, Vet. Sbirka, v. 75 (5), 32-33
piroplasmosis, berenil, cattle, control of Boophilus calcaratus with chlorophos

Piroplasmosis
Nickel, S., 1978, Monatsh. Vet.-Med., v. 33 (23), 915-917
parasitoses of farm animals, potential effects of soil and water improvement programs

Piroplasmosis
Rakhimov, T. Kh.; et al., 1977, Veterinariia, Moskva (10), 75-77
piroplasmosis, francaiellosis, cattle, dimidine and imidocarb tested in various doses, recommended for control

Piroplasmosis
Valiev, B., 1977, Veterinariia, Moskva (10), 21-22
piroplasmosis, cattle, control, elimination of tick vectors: Ishtykhansk region, Samarkandsk oblast

Piroplasmosis
Vulchovski, Ia.; et al., 1977, Vet. Sbirka, v. 75 (5), 26-29
tick control important in prophylaxis of piroplasmosis under range conditions: Bulgaria

Piroplasmosis
Zhiliaev, E. A., 1977, Veterinariia, Moskva (7), 57-59
piroplasmosis, reindeer, Ixodes persulcatus found in insignificant numbers and only in summer, need to consider other possible vectors: Khabarovsk Krai

Plasmodium
Ambroise-Thomas, P., 1976, Rev. Franc. Transfus. et Immuno-Hematol., v. 19 (2), 331-336
human malaria, detection of latent Plasmodium carriers, immunofluorescence test, prophylactic survey of potential blood donors

Plasmodium
Ayala, S. C., 1978, J. Protozool., v. 25 (1), 87-100
Plasmodium from reptiles, checklist, host index, annotated bibliography

Plasmodium
Bailenger, J.; et al., 1972, Therapeutique, v. 48 (2), 117-128
dangers of cortisone therapy in the presence of human parasitic pathology, clinical review

Plasmodium
Besson, P.; et al., 1976, Rev. Franc. Transfus. et Immuno-Hematol., v. 19 (2), 369-373
human malaria, case reports of infection from blood transfusions, recommendations that the immunofluorescence test be used along with other methods of screening potential donors

Plasmodium
Bouckaert, A.; and Wery, M., 1974, Ann. Soc. Belge Med. Trop., v. 54 (2), 103-119
Plasmodium, numerical taxonomy, 3 methods of cluster analysis used to build hierarchical classification of 33 members of genus

Plasmodium
Brown, K. N., 1974, Ciba Found. Symp., n.s. (25), 35-51
antigenic variation and immunity to malaria, review

Plasmodium
Brown, K. N., 1976, Immunol. Parasit. Infect., 268-295
malaria, immunity, review

Plasmodium
Brown, K. N., 1978, Colloque Immun. Parasit. Dis. (Thiverval-Grignon, Sept. 5-9, 1977), 59-70
Trypanosoma, Plasmodium, Babesia, antigenic variation (nature, consequences for protective immunity, possible implications for other protozoan infections), colloquium presentation

Plasmodium
Cohen, S., 1978, Israel J. Med. Sc., v. 14 (5), 537-541
malaria, mechanisms of acquired immunity to erythrocytic stage, symposium presentation

Plasmodium
Cohen, S., 1979, Proc. Roy. Soc. London, s. B., Biol. Sc. (1153), v. 203, 323-345
immunity to malaria with emphasis on vaccination, review lecture

Plasmodium
Cohen, S.; and Mitchell, G. H., 1978, Current Topics Microbiol. and Immunol., v. 80, 97-137
malaria, prospects for immunization, review

Plasmodium
Cox, F. E. G., 1978, Nature, London (5669), v. 274, 312
malaria, piroplasmosis, and endotoxin, brief review of recent work

Plasmodium
Dei Cas, E.; et al., 1979, Ann. Parasitol., v. 54 (5), 567-570
Plasmodium, diagnosis, advantages of Errecart's modified technique for making thick blood films

Plasmodium
Diggs, C. L., 1973, Immunol. Series, v. 2, 169-196
malaria, experimental approaches to study of acquired immunity, review

Plasmodium
Dymowska, Z., 1975, Przegl. Epidemiol., v. 29 (1), 65-68
malaria, human, diagnosis, brief review

Plasmodium
Faugere, B.; and Ranque, J., 1976, Rev. Franc. Transfus. et Immuno-Hematol., v. 19 (2), 349-355
human malaria, screening blood donors for possible latent Plasmodium carriers, immunofluorescence test, suggested prophylactic measures

lasmodium
Gabaldon, A.; and Ulloa, G., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (3), 311-312
Plasmodium, quick and easy method to determine sporozoite index in vector mosquitoes

Plasmodium
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
avian Plasmodium spp. of 4 subgenera, list of type material in Garnham Collection of Wellcome Museum of Medical Science; neotypes designated

Plasmodium
Godfrey, D. G., 1978, Nature, London (5664), v. 273, 600-604
identification of economically important parasites (use of anatomical, biochemical, and behavioral tests), brief review

Plasmodium
Gutteridge, W. E.; and Coombs, G. H., 1977, Biochemistry of parasitic protozoa, 172 pp., illus.
biochemistry of parasitic protozoa, textbook: methodology; catabolism and generation of energy; nucleic acid metabolism; protein metabolism; lipid metabolism; biochemical mechanism of drug action; isolation of parasitic protozoa from infected animals; culture of parasitic protozoa

Plasmodium
Jerusalem, C., 1978, Israel J. Med. Sc., v. 14 (6), 620-650
immunopathology of malaria, extensive review, symposium presentation

asmodium
Jiang, J. B., 1978, Exper. Parasitol., v. 46 (2), 339-352
Plasmodium, experimental animals for human malaria and research needs, review

asmodium
Lumsden, W. H. R., 1978, Handb. Exper. Immunol. (Weir), 3. ed., 42.1-42.47
demonstration of antibodies to Protozoa, extensive review

Plasmodium
McGregor, I. A., 1978, Israel J. Med. Sc., v. 14 (5), 523-536
malaria, epidemiological aspects, symposium presentation: failure of eradication programs; malaria in pregnancy; interaction between parasite and human erythrocyte

lasmodium
Nozais, J. P.; Truong Minh Ky, D.; and Doucet, J., 1979, Med. Trop., v. 39 (5), 549-553
malaria, newborn infants and young children living in stable hypoendemic area, evaluation of antimalarial antibody titers using Plasmodium berghei as antigen: Abidjan dispensary, Ivory Coast

Plasmodium
Nurse, G. T., 1979, Lancet, London (8149), v. 2, 938-940
humans with thalassaemia, role of iron in host tissue as protective mechanism against Plasmodium infections

Plasmodium
Playfair, J. H. L., 1978, Current Topics Microbiol. and Immunol., v. 80, 37-64
effective and ineffective immune responses to parasites, evidence from experimental models, review with emphasis on malaria and trypanosomiasis

Plasmodium
Tin, F., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (4), 552-557
Toxoplasma gondii, Plasmodium, young school children living in 2 areas of different phases of an antimalarial control program, survey for presence of malaria and toxoplasmosis antibodies using the indirect fluorescent antibody test: Burma

Plasmodium
Trigg, P. I., 1978, Methods Cultiv. Parasites in Vitro, 89-110
Plasmodium, cultivation, review

Plasmodium
Varnai, F.; and Ecker, A., 1977, Therap. Hungar., v. 25 (4), 131-133
malaria, humans travelling to endemic areas, drug prophylaxis, comparative study, least unwanted side effects and lowest morbidity rate recorded with pyrimethamine: Hungary

Plasmodium
Voller, A., 1976, Immunol. Parasit. Infect., 107-119
malaria, serodiagnosis, review

Plasmodium
Wallach, D. F. H., 1979, Cell Biol. Internat. Rep., v. 3 (5), 395-408
malaria, membrane pathobiology, review

Plasmodium
Watanabe, Y., 1975, Rinsho Fujinka Sanka (Clin. Gynec. and Obst.), v. 29 (8), 625-628
pathological changes in human placenta due to infection, includes Toxoplasma gondii, Plasmodium, and trypanosomiasis

Plasmodium
Wilson, R. J. M., 1978, Colloque Immun. Parasit. Dis. (Thiverval-Grignon, Sept. 5-9, 1977), 87-101
circulating antigens of parasites, source, nature, fate, and possible effects on immune response, colloquium presentation

Plasmodium
Wright, I. G., 1979, Gen. Pharmacol., v. 10 (4), 319-325
Babesia, Plasmodium, Trypanosoma, kallikrein-kinin system, mechanisms of activation (parasite enzymes, immune complexes), role in hypotensive shock syndrome of infected animals, review

Plasmodium [sp.] 'tropiduri'-like plasmodium
Ayala, S. C.; Moreno, E.; and Bolanos, R., 1978, J. Parasitol., v. 64 (2), 330-335
Anolis limifrons
A. humilis
A. lionotus
all from 'La Selva' near Puerto Viejo, Heredia Province, Costa Rica

Plasmodium [sp.]
Baker, J. R., 1974, J. Zool., London, v. 172 (2), 169-190
Corvus monedula
Turdus ericetorum
T. merula
all from Britain

Plasmodium [sp.]
Bennett, G. F.; Cameron, M.; and White, E., 1975, Canad. J. Zool., v. 53 (10), 1432-1442
hematozoa of passeriforms, prevalence, effect of climate, application of insecticide, and large-scale environmental alteration
Melospiza georgiana
M. lincolnii
M. melodia
Passerculus sandwichensis
Zonotrichia albicollis
Agelaius phoeniceus
Molothrus ater
Quiscalus quiscula
Dumetella carolinensis
Dendroica coronata
D. magnolia
D. pensylvanica
D. petechia
D. striata
Geothlypis trichas
Oporornis philadelphia
Setophaga ruticilla
Vermivora ruficapilla
Wilsonia pusilla
Catharus ustulatus
Turdus migratorius
Empidonax sp.
Tyrannus tyrannus
Vireo olivaceous
Colaptes auratus
Philohela minor
Catharus fuscescens
all from New Brunswick

Plasmodium spp.
Biswas, A. K., 1978, Water Supply & Management, v. 2 (4), 283-297
discussion on environmental implications of water development for developing countries: possibility of decreased human parasitism by improving potable water and by reducing human contacts with vectors of water-borne or water-based infections, also possibility of spread of parasitism through development of irrigation canals

Plasmodium sp.
Deane, L. M.; and Ferreira, J. A. (Neto), 1969, Rev. Inst. Med. Trop. S. Paulo, v. 11 (5), 299-305
Alouatta fusca clamitans: Estado do Rio Grande do Sul, Brasil

Plasmodium sp.
Guerrero, S.; and Ayala, S. C., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (5), 283-288
Thecadactylus rapicauda (sangre): selva amazonica, Huanuco, Peru

Plasmodium [sp.]
Kocan, A. A.; Snelling, J.; and Greiner, E. C., 1977, J. Wildlife Dis., v. 13 (3), 304-306
Falco sparverius: Oklahoma

Plasmodium (Novyella) sp.
Peirce, M. A.; and Mead, C. J., 1977, J. Nat. Hist., v. 11 (6), 597-600
Phylloscopus trochilis: near Tring, Hertfordshire

Plasmodium sp.
Powell, R. D., 1978, Am. J. Trop. Med. and Hyg., v. 27 (1, pt. 1), 1-5
Plasmodium spp., servicemen returning from Vietnam, relationships between chemoprophylaxis history and occurrence and timing of post-departure episodes of malaria: Iowa City VA Hospital

Plasmodium sp.
Stafford, E. E.; Galdikas-Brindamour, B.; and Beaudoin, R. L., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (1), 106-107 [Letter]
Pongo pygmaeus (blood): Southern Kalimantan (Indonesian Borneo)

Plasmodium species
Telford, S. R., jr., 1978, Internat. J. Parasitol., v. 8 (2), 133-138
Saurocytozoon tupinambi, study of initial infection in juvenile Tupinambis teguixin provides evidence that schizogonic cycle in circulating cells may occur, but identity of intralymphocytic asexual stages with this species cannot be established due to presence of concurrent infection by small Plasmodium species, if confirmed data would justify removing Saurocytozoon from Leucocytozoidae: Venezuela

Plasmodium sp. indet., probably P. chiricahuae
Telford, S. R., jr., 1978, J. Parasitol., v. 64 (3), 553-554
Sceloporus poinsetti
Urosaurus inornatus
all from Texas

Plasmodium sp.
Telford, S. R., jr., 1979, Ann. Parasitol., v. 54 (2), 129-144
synopsis; reported earlier as Plasmodium tropiduri
Anolis limifrons: central Panama

Plasmodium [spp.]
Williams, N. A.; and Bennett, G. F., 1978, Canad. J. Zool., v. 56 (4, pt. 1), 596-607
Anas acuta: Maryland
Casmerodius albus: Maryland and/or New Jersey
Egretta thula: Maryland or New Jersey
Nycticorax nycticorax: New Jersey
Vireo olivaceus: Maryland and/or New Jersey
Corvus brachyrhynchos: Maryland
Cyanocitta cristata: Maryland and/or New Jersey
Cardinalis cardinalis: Maryland and/or New Jersey
Junco hyemalis: Maryland or New Jersey
Melospiza lincolnii: Maryland or New Jersey
M. melodia: Maryland and/or New Jersey
Pipilo erythrophthalmus: Maryland and/or New Jersey
Spizella passerina: Maryland or New Jersey
S. pusilla: Maryland or New Jersey
Zonotrichia albicollis: Maryland or New Jersey
Hirundo rustica: Maryland or New Jersey
Riparia riparia: New Jersey
Agelaius phoeniceus: Maryland and/or New Jersey
Molothrus ater: Maryland or New Jersey
Quiscalus quiscula: Maryland and/or New Jersey
Sturnella magna: Maryland
Dumetella carolinensis: Maryland and/or New Jersey
Parus bicolor: Maryland or New Jersey
Geothlypis trichas: Maryland
Icteria virens: Maryland and/or New Jersey
Passer domesticus: " " " " "
Porzana carolinensis: Maryland
Calidris pusilla: "
Otus asio: Maryland
Sturnus vulgaris: Maryland and/or New Jersey
Piranga olivacea: Maryland
Catharus minimus: Maryland or New Jersey
C. ustulatus: Maryland and/or New Jersey
Hylocichla mustelina: Maryland and/or New Jersey
Sialia sialis: Maryland or New Jersey
Turdus migratorius: Maryland and/or New Jersey
Zenaidura macroura: Maryland

Plasmodium [sp.]
Winchell, E. J., 1978, J. Parasitol., v. 64 (3), 558-559
Dendroica magnolia: El Salvador

Plasmodium sp., probably P. knowlesi
Yap, L. F.; et al., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (3), 345-353
Presbytis obscura
P. melalophos
Macaca fascicularis
all from Jenderak Utara, Malaysia

Plasmodium (Giovannolaia) anasum Manwell and Kuntz, 1965
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
parahapantotype designated

Plasmodium aurulentum Telford, 1971
Telford, S. R., jr., 1978, Internat. J. Parasitol., v. 8 (5), 341-353
measurements
Thecadactylus rapicaudus (erythrocytes): Municipio Manrique and Araure, Venezuela

Plasmodium balli
Ayala, S. C.; Moreno, E.; and Bolanos, R., 1978, J. Parasitol., v. 64 (2), 330-335
Anolis limifrons
A. humilis
all from 'La Selva' near Puerto Viejo, Heredia Province, Costa Rica

Plasmodium balli, illus.
Guerrero, S.; and Ayala, S. C., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (5), 283-288
Anolis fuscoauratus (sangre): selva amazonica, Huanuco, Peru

Plasmodium basilisci
Ayala, S. C.; Moreno, E.; and Bolanos, R., 1978, J. Parasitol., v. 64 (2), 330-335
Basiliscus vittatus: 'La Selva' near Puerto Viejo, Heredia Province, Costa Rica

Plasmodium beebei new species, illus.
Telford, S. R., jr., 1978, Internat. J. Parasitol., v. 8 (5), 341-353
effects upon host cells
Gonatodes taniae (erythroblasts, proerythrocytes, erythrocytes, thrombocytes): Parque Nacional Henri Pittier (Rancho Grande), Estado Aragua, Venezuela

Plasmodium berghei
Agarwal, K.; Puri, S. K.; and Dutta, G. P., 1979, Indian J. Med. Research, v. 69, 577-582
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Plasmodium berghei, ability of hyperimmune serum to neutralize infective inoculum, isolation and characterization of protective antibody, antibody combined mainly with free parasites and not with infected RBC

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Plasmodium berghei-infected mice (exper.), vitamin E deficiency moderates severity of infection since premature, oxidant-induced hemolysis of infected erythrocytes prevents orderly parasite maturation, restoration of susceptibility to malaria by vitamin E supplementation, observations provide basis for selective advantage of G-6-PD deficiency in areas of endemic malaria

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Plasmodium spp. in laboratory animals, thioquinazoline analogues synthesized and tested under laboratory conditions showed substantial suppressive antimalarial and prophylactic activity when compared with reference compounds, analogues also retained potent antimalarial effects against strains resistant to common antimalarials; the most active compound, 2,4-diamino-6-[(α,α,α-trifluoro-m-tolyl)thio]quinazoline has been designated for preclinical toxicologic studies

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Plasmodium berghei, undiminished mefloquine accumulation by erythrocytes infected with chloroquine-resistant strain provides explanation for superiority of mefloquine in treating chloroquine-resistant malaria, but competition observed between chloroquine and mefloquine raises possibility that same process of accumulation serves both drugs

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Plasmodium berghei, evidence that erythrocyte surface components determine affinity with which chloroquine is accumulated and thereby determine whether or not the malaria parasite will be susceptible to the drug

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Plasmodium berghei, mice, anti-plasmodial activity of chloroquine does not appear to be associated with inhibition of erythrocytic glucose-6-phosphate dehydrogenase

Plasmodium berghei
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Plasmodium berghei, successful cultivation in vitro of infective exoerythrocytic schizonts in primary monolayer cultures of rat liver cells

Plasmodium berghei, illus.
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Plasmodium berghei, successful method for obtaining suspensions of hepatocytes parasitized with viable infective exoerythrocytic schizonts from donor rats originally infected with sporozoites

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Plasmodium berghei-infected mice, cellular changes: differential counts of bone marrow, 3-day cultures of bone marrow, [^{3}H]thymidine uptake by splenocytes and bone marrow cells, interaction of adherent and nonadherent splenic and bone marrow cells, symposium presentation

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Plasmodium, delayed-type hypersensitivity to sheep erythrocytes was depressed during fatal P. berghei and self-limiting P. yoelii infections in mice, immunological lesion found to be at level of DTH expression (i.e., inflammatory response) rather than at level of T cell sensitization

Plasmodium berghei
Freeman, R. R.; and Parish, C. R., 1978, Clin. and Exper. Immunol., v. 32 (1), 41-45
Plasmodium berghei, P. yoelii, mice, numbers of 'background' plaque-forming cells secreting IgM specific for either sheep or horse erythrocytes elevated in spleens during infection or in spleens of uninfected mice injected with non-infectious extracts of parasitized mouse red blood cells, results provide corroborating evidence for hypothesis that B-cell mitogen is associated with blood stage of malaria parasites, possible involvement in immunosuppression

Plasmodium berghei
Freeman, R. R.; and Parish, C. R., 1978, Immunology, v. 35 (3), 479-484
Plasmodium, spleen cell changes during fatal P. berghei vs. those during self-limiting P. yoelii infections in mice, protective immunity is associated with marked and sustained increases in numbers of Ig$^+$ and Thy-1·2$^+$ spleen cells and in fatal infections these proliferative responses are apparently suppressed

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Plasmodium berghei, comparison of infected mice subjected to electric shock stimulation and infected controls showed that mice subjected to stress were more resistant to infection than were controls

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human malarias, systematic screening of all potential blood donors recommended using the indirect immunofluorescence test and Plasmodium berghei antigen

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Plasmodium berghei, immunization of chloroquinized rats against sporozoites by bites of infected mosquitoes: influence of number of exposures to infected mosquitoes on antibody titers and protection; influence of exposure to different numbers of infective mosquitoes on antibody production and protection; specificity of antiplasmodial antibodies; influence of passive transfer of sera from rats immune to sporozoites or to erythrocytic forms on development of sporozoites, symposium presentation

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Plasmodium berghei-infected mouse erythrocytes, formation of myelin-like forms, light and scanning electron microscopy

Plasmodium berghei
Gorenflot, A.; et al., 1979, Ann. Pharm. Franc., v. 37 (7-8), 275-284
Plasmodium berghei, mice (exper.), cause-effect relationship between myelin-like form of erythrocytes and that of neutrophilic granulocytes or monocytes which have phagocytized pigment grains

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Gorenflot, A.; and Piette, M., 1977, Ann. Pharm. Franc., v. 35 (9-10), 337-342
Plasmodium berghei-infected mice, changes in red blood cell morphology after penetration and intracellular development of parasite

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Plasmodium berghei in inbred rats, macrophage-cytophilic antibody specific for malarial antigens, identification and characterization, demonstration of role in protection, acts synergistically with opsonizing antibody

Plasmodium berghei
Greenblatt, C. L.; and Jacobson, R. L., 1978, Israel J. Med. Sc., v. 14 (5), 596-598
malaria, speculation on use of adoptive immunity in vaccination, some experiments with Plasmodium berghei in hamsters, symposium presentation

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Plasmodium berghei
Hamburger, J.; and Zuckerman, A., 1978, Israel J. Med. Sc., v. 14 (5), 557-562
Plasmodium berghei, partial separation of putative species-specific immunobiologically active antigen from soluble extract, symposium presentation

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Plasmodium berghei, IgM and IgG antisporozoite antibodies in mice immunized with irradiation-attenuated sporozoites, detection by indirect fluorescent antibody test, correlation with protection, some cross-reaction with blood stage antigens but test should still prove useful

Plasmodium berghei
Heumann, A. M.; et al., 1979, Infect. and Immun., v. 24 (3), 829-836
Plasmodium berghei, high and low antibody responder lines of mice and their interline hybrids, antibody response induced by vaccination with irradiated parasitized erythrocytes, innate resistance and protective efficacy of vaccination, results indicate vaccination-induced immunity is essentially due to antibody response

Plasmodium berghei, illus.
Howard, R. J.; and Battye, F. L., 1979, Parasitology, v. 78 (3), 263-270
Plasmodium berghei, dye-labelling technique for sorting uninfected, singly infected, doubly infected, and more heavily infected red cells from mouse blood based on their DNA content

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Howard, R. J.; Battye, F. L.; and Mitchell, G. F., 1979, J. Histochem. and Cytochem., v. 27 (1), 803-813
Plasmodium berghei-infected red blood cells analyzed and sorted by flow fluorimetry with the DNA-binding dye 33258 Hoechst, possible applications in biochemical and immunochemical analyses and in clinical diagnosis

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Plasmodium berghei, uninfected cells separated from infected mouse blood were cleared more rapidly than normal mouse erythrocytes after injection into normal mice

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Plasmodium berghei, removal of leucocytes from red cells in infected mouse blood, purification of schizont-infected cells

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Jacobson, R. L.; Zuckerman, A.; and Greenblatt, C. L., 1978, Clin. and Exper. Immunol., v. 33 (1), 25-29
Plasmodium berghei, neonate rats from normal or immune mothers receiving spleen cells from normal or immune mothers and fostered to normal or immune mothers in various combinations, results suggest that immune response was suppressed in presence of passively transferred maternal antibody

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Plasmodium berghei, white rats (exper.), electron microscopic study of resulting nephrotic syndrome showed direct relationship with antibody formation

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3- and 5-aminoquinolines, potential antimalarials, synthesis, testing of some against Plasmodium berghei in mice, Leishmania donovani in hamsters, or P. cynomolgi in rhesus monkeys

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P[lasmodium] berghei, 646 1-aryl-2-(alkylamino)ethanol antimalarials, quantitative structure-activity relationship, mathematical analysis

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Plasmodium berghei, mice infected with clindamycin-resistant parasites, uptake of clindamycin and its metabolites by erythrocytes, impaired uptake is not mechanism of resistance

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Plasmodium berghei-infected rats, serum urea and electrolyte levels

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Plasmodium yoelii- and P. berghei-infected mice and vaccinated mice challenged with homologous parasites, changes in phagocytic and adherent cell numbers, development and suppression of population of late-adhering macrophages

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Liew, F. Y.; Dhaliwal, S. S.; and Teh, K. L., 1979, Immunology, v. 37 (1), 35-44
Plasmodium berghei, mice, effect of infection and of supernatant obtained from cultures of infected red cells on humoral (enhanced or suppressed) and cell-mediated (suppressed) immune responses to unrelated antigens

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polychlorinated biphenyl- and hexachlorobenzene-treated mice, impaired resistance to bacterial endotoxin and to Plasmodium berghei, data indicate that environmental chemicals impair host resistance and that the alteration may be related to presence of the chemicals in the lymphoreticular organs

Plasmodium berghei
Mack, S. R.; Samuels, S.; and Vanderberg, J. P., 1979, J. Parasitol., v. 65 (1), 130-136
Plasmodium berghei-infected and uninfected Anopheles stephensi, free amino acids in hemolymph

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Mack, S. R.; Samuels, S.; and Vanderberg, J. P., 1979, J. Parasitol., v. 65 (2), 217-221
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Plasmodium berghei, energy metabolism of sporozoites

Plasmodium berghei
Mack, S. R.; and Vanderberg, J. P., 1978, J. Parasitol., v. 64 (5), 918-923
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Mack, S. R.; Vanderberg, J. P.; and Nawrot, R., 1978, J. Parasitol., v. 64 (1), 166-168
Plasmodium berghei, DEAE-cellulose column procedure for separation of sporozoites from infected mosquitoes, technique permits preparation of cleaner suspensions of sporozoites than any previously described procedure

Plasmodium berghei
McQuistion, T. E., 1979, Am. J. Trop. Med. and Hyg., v. 28 (1), 12-14
Plasmodium berghei, mice acclimated to 22°C or 5°C before infection, some treated with clofibrate and some briefly exposed to -35°C after infection, parasitemia and plasma free fatty acid levels

Plasmodium berghei
Maier, W. A.; and Piekarski, G., 1979, Immun. u. Infekt., v. 7 (3), 75-82
malaria, human, diagnosis, indirect immunofluorescent test using Plasmodium berghei or P. falciparum as antigen

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Plasmodium berghei, mice, phagocytosis of parasitized erythrocytes by mouse peritoneal macrophages, concluded that both humoral and cellular factors were important

Plasmodium berghei
Meuwissen, J. H. E. T.; Golenser, J.; and Verhave, J. P., 1978, Israel J. Med. Sc., v. 14 (5), 601-605
Plasmodium berghei, rats under prophylactic treatment with various drug regimens, development of effective antisporozoite immunity by natural bites of infected mosquitoes, symposium presentation

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Michel, J. C.; Lagrange, P. H.; and Hurtrel, B., 1979, Parasite Immunol., v. 1 (4), 267-275
Plasmodium-infected mice, profound alteration of inductive phase of delayed-type hypersensitivity and antibody formation to sheep erythrocytes when sensitization with antigen was performed intravenously at critical time of disease but not after subcutaneous immunization, suggests major role for spleen in mechanism of immunodepression

Plasmodium berghei
Mitchell, G. F.; Handman, E.; and Howard, R. J., 1978, Austral. J. Exper. Biol. and Med. Sc., v. 56 (5), 553-559
Plasmodium berghei, Babesia rodhaini, mice, attempts to raise host-protective sera using variety of immunization manipulations (BCG injection, P. yoelii infection, others)

Plasmodium berghei
Momen, H., 1979, Ann. Trop. Med. and Parassitol., v. 73 (2), 109-115
Plasmodium, Babesia, and Anthemosoma spp. in mouse erythrocytes, identification of enzymes of parasite origin using starch-gel electrophoresis

Plasmodium berghei
Momen, H., 1979, Ann. Trop. Med. and Parasitol., v. 73 (2), 117-121
Plasmodium, Babesia and Anthemosoma spp., comparative study of glucose catabolism by infected mouse erythrocytes, glucose utilization and lactate production of parasites

Plasmodium berghei, illus.
Moser, G.; et al., 1978, J. Protozool., v. 25 (1), 119-124
Plasmodium berghei, P. knowlesi, P. cynomolgi, purification of sporozoites by passage through DEAE-cellulose column, retention of ability to produce infection, to induce protective immunity, and to react with known antisera

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Murphy, J. R., 1979, Infect. and Immun., v. 24 (3), 707-712
Plasmodium berghei, mice, analysis of mechanisms of immunity generated in response to immunization with formalin-killed blood-stage parasites

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Murphy, J. R.; and Lefford, M. J., 1978, Infect. and Immun., v. 22 (3), 798-803
Plasmodium berghei, mice, induction of protracted state of immunity with formalin-killed blood parasite vaccine in combination with Corynebacterium parvum and/or living BCG

lasmodium berghei
Murphy, J. R.; and Lefford, M. J., 1979, Am. J. Trop. Med. and Hyg., v. 28 (1), 4-11
Plasmodium berghei, mice, successful vaccination by using formolized blood parasites

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Nardin, E. H.; and Nussenzweig, R. S., 1978, Nature, London (5666), v. 274, 55-57
Plasmodium berghei-mouse and P. knowlesi-rhesus monkeys systems, detection of stage and species specific antisporozoite antibodies with circumsporozoite precipitation and indirect immmunofluorescence methods, preliminary application to P. falciparum in humans with similar results

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Nasr, M.; Nabih, I.; and Burckhalter, J. H., 1978, J. Med. Chem., v. 21 (3), 295-298
Plasmodium gallinaceum, Plasmodium berghei, pyrimido[5,4-c]quinolines and derivatives, laboratory trials, inactive as antimalarials

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Nozais, J. P.; Truong Minh Ky, D.; and Doucet, J., 1979, Med. Trop., v. 39 (5), 549-553
malaria, newborn infants and young children living in stable hypoendemic area, evaluation of antimalarial antibody titers using Plasmodium berghei as antigen: Abidjan dispensary, Ivory Coast

asmodium berghei
Orjih, A. U.; and Nussenzweig, R. S., 1979, Clin. and Exper. Immunol., v. 38 (1), 1-8
Plasmodium berghei, mice, suppression of antibody response to sporozoite stage by acute blood infection

smodium berghei
Pacheco, N. D.; et al., 1979, J. Parasitol., v. 65 (3), 414-417
Plasmodium berghei, rapid large-scale isolations of sporozoites from infected mosquitoes, modification of discontinuous gradient technique

asmodium berghei
Palmer, T. T., 1978, J. Parasitol., v. 64 (3), 493-496
Plasmodium berghei, rats, effect of primary patent infection during pregnancy upon course of infection and humoral antibody response in offspring, passive transfer of protective antibody through milk, in utero sensitization by soluble malaria antigens may also exert protective effect

Plasmodium berghei
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Plasmodium berghei 'NS lines' form distinct taxon within P. yoelii complex and should be referred to as P. yoelii subspecies on basis of isoenzyme, DNA, and cross-immunity relationships with other rodent Plasmodium spp.

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Phillips, R. S.; Brown, K. N.; and Hills, L. A., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (1), 92-94
Plasmodium berghei, reduced protective activity of immune spleen cells from completely cured rats if cells are transferred to rats which have been infected for 5-8 days before cell transfer

Plasmodium berghei
Pirson, P.; et al., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (3), 347 [Letter]
Plasmodium berghei, mice (exper.), liposome-entrapped primaquine used as therapy against pre-erythrocytic stage infection, less toxic than free primaquine

Plasmodium berghei
Playfair, J. H. L.; et al., 1979, Nature, London (5740), v. 282, 731-734
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Plasmodium berghei
Playfair, J. H. L.; and De Souza, J. B., 1979, Parasite Immunol., v. 1 (3), 197-208
Plasmodium yoelii- or P. berghei-vaccinated mice, immunofluorescent antibody response with particular reference to antibody class and subclass, correlation with protection, passive transfer experiments, effect of macrophage stimulation and inhibition on antibody and on protection

Plasmodium berghei, illus.
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Plasmodium berghei, mice, immunization, possible role of plasmodial antigens exposed on surface of infected reticulocytes in induction of protective immunity, observations on entry of parasites into red blood cells, symposium presentation

Plasmodium berghei
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Plasmodium berghei, mice, formation of two types of immune complex (one with and one lacking plasmodial antigens) and their deposition in renal glomeruli, immune complexes lacking parasite antigen may be involved in secondary autoimmune (anti-smooth muscle) process, possible induction mechanism of autoantibodies, symposium presentation

Plasmodium berghei
Quinn, T. C.; and Wyler, D. J., 1979, J. Immunol., v. 123 (5), 2245-2249
Plasmodium berghei, rats, mechanisms of action of hyperimmune serum in mediating protective immunity

Plasmodium berghei
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Plasmodium berghei
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Plasmodium berghei in chloroquine resistant white mice, results of treatment with combinations of proguanil and dapsone

Plasmodium berghei
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Plasmodium berghei-infected Mesocricetus auratus, cerebral lesions, transmission and scanning electron microscopy, hypothesized that pathogenesis relates to immune complex formation

Plasmodium berghei
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Plasmodium berghei, correlation of in vitro erythrophagocytosis with dynamics of early-onset anemia and reticulocytosis in mice

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malaria and Giemsa type blood stains, use of lower ratio azure B to methylene blue offers superior staining effects

Plasmodium berghei, illus.
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Schistosoma japonicum, schistosomal pigment, purification and histochemical characteristics, comparison with malarial (Plasmodium berghei) pigment

Plasmodium berghei
Sauvager, F.; and Fauconnier, B., 1978, Biomedicine Express, v. 29 (6), 184-187
Plasmodium berghei, mice, protective effect of endogenous interferon in mouse malaria demonstrated by increase in death rate and in % parasitized erythrocytes in infected mice treated with anti-interferon globulins

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Plasmodium berghei, 1,9-Phenazine-bis(dialkylaminocarboxamides) synthesized and screened as potential antimalarials, no significant activity

Plasmodium berghei
Schmidt, L. H.; et al., 1978, Antimicrob. Agents and Chemotherapy, v. 14 (3), 292-314
Plasmodium falciparum in Aotus trivirgatus griseimembra and human volunteers, P. cynomolgi in Macaca mulatta, antimalarial activities of various 9-phenanthrenemethanols with special attention to WR-122,455 and WR-171,669, includes some [apparently unpublished] results of other workers for P. berghei and these same compounds

Plasmodium berghei
Schmidt, L. H.; et al., 1978, Antimicrob. Agents and Chemotherapy, v. 14 (3), 420-435
Plasmodium falciparum and P. vivax in Aotus trivirgatus griseimembra, antimalarial activities of various 4-pyridinemethanols with special attention to WR-172,435 and WR-180,409, includes some [apparently unpublished] results of other workers for P. berghei and these same compounds

Plasmodium berghei, illus.
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Plasmodium berghei, illus.
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Plasmodium berghei-infected erythrocytes, phagocytosis by peritoneal macrophages from normal, infected, or immune mice, light and scanning electron microscopy

Plasmodium berghei
Shamblee, D. A.; and Gillespie, J. S., jr., 1979, J. Med. Chem., v. 22 (1), 86-89
Plasmodium berghei, mice, activity of trichloronaphthalene amino alcohols

Plasmodium berghei
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Plasmodium berghei, mice, xanthine oxidase activity in liver

Plasmodium berghei
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Plasmodium berghei, mice, effect of starvation, infection, and interactions between the two on lipid peroxide and protein levels of liver and spleen

Plasmodium berghei
Shear, H. L.; Nussenzweig, R. S.; and Bianco, C., 1979, J. Exper. Med., v. 149 (6), 1288-1298
Plasmodium berghei-infected mice, phagocytosis of erythrocytes by spleen macrophages appears to be mediated by Ig on surface; other indications of spleen macrophage activation; phagocytosis is inhibited later in infection by serum factors possibly immune complexes; high levels of anti-Forssman antibodies

Plasmodium berghei
Shetty, R. V.; and Blanton, C. D., jr., 1978, J. Med. Chem., v. 21 (9), 995-998
2-substituted primaquine analogues synthesized and evaluated in laboratory animals against Plasmodium berghei, P. cynomolgi, Trypanosoma rhodesiense, Leishmania donovani and Schistosoma mansoni; significant activity was observed against P. berghei and L. donovani

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Plasmodium berghei, 17-X and NYU-2 strains, mice, immunosuppression using triamcinolone acetonide

lasmodium berghei
Smrkovski, L. L.; and Strickland, G. T., 1978, J. Immunol., v. 121 (4), 1257-1261
Plasmodium berghei, mice, single or multiple immunizations with BCG and/or irradiated sporozoites (varying degrees of protection), immunization with irradiated sporozoites before BCG (suppression of protective immunity against sporozoite challenge)

lasmodium berghei
Song, M.; and Di Luzio, N. R., 1979, Lysosomes Applied Biol. and Therap., v. 6, 533-547
yeast glucan and immunotherapy of infectious diseases, review including section on Plasmodium berghei

lasmodium berghei, illus.
Sterling, C. R., 1978, J. Parasitol., v. 64 (4), 747-749
Plasmodium berghei, rapid method for concentrating and recovering spleen-derived schizont-infected erythrocytes

asmodium berghei
Strambachova-McBride, J.; and Micklem, H. S., 1979, Parasite Immunol., v. 1 (2), 141-157
Plasmodium berghei- and P. yoelii yoelii-infected mice, immunosuppression, secondary response to bovine serum albumin

asmodium berghei
Strambachova-McBride, J.; and Micklem, H. S., 1979, Immunology, v. 36 (3), 607-614
Plasmodium berghei, P. y. yoelii, mice, primary antibody response to immunogenic alum-adsorbed bovine serum albumin (BSA) was depressed and induction of immunological memory by soluble polymerized BSA was suppressed in both infections but neither interfered with induction of low-zone tolerance by monomeric BSA

asmodium berghei, illus.
Suzuki, M.; et al., 1979, Bull. World Health Organ., v. 57 (1), 129-132
Plasmodium berghei-parasitized rat erythrocytes, isolation by carrier-free electrophoresis

asmodium berghei
Tanabe, K.; Takada, S.; and Suzuki, M., 1978, J. Parasitol., v. 64 (2), 367-368
Plasmodium berghei-infected mice, sensitization to sheep red blood cells by subcutaneous vs. intravenous routes, difference in cell-mediated immune reaction in regional lymph nodes and spleen, results show difference between iv and sc sensitized mice in immunodepression by malaria

asmodium berghei, illus.
Terzakis, J. A.; et al., 1979, J. Protozool., v. 26 (3), 385-389
Plasmodium berghei, exoerythrocytic stages in rat liver, possible example of phagocytosis by Kupffer cells, electron microscopy

asmodium berghei
Verhave, J. P.; et al., 1978, J. Immunol., v. 121 (3), 1031-1033
Plasmodium berghei, transfer of protective immunity with lymphoid cells from mice immune to malaria sporozoites

Plasmodium berghei
Waki, S., 1976, Kiseichugaku Zasshi (Japan. J. Parasitol.), v. 25 (6), 441-446
Plasmodium berghei, mice, protective immunity induced by repeated infections followed by radical chemotherapy with sulfamonomethoxine

Plasmodium berghei
Warhurst, D. C.; and Thomas, S. C., 1978, Ann. Trop. Med. and Parasitol., v. 72 (3), 203-211
Plasmodium berghei, effect of some metabolic inhibitors upon chloroquine-induced pigment clumping

Plasmodium berghei
Wedderburn, N.; et al., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (6), 610-614
Plasmodium yoelii infection in mice followed by either one or repeated infections with P. berghei, induction of chronic malaria, course of parasitemia, fluctuations in reticulocyte levels, antibody titres, immunoglobulin deposition in tissues, changes in spleen and liver

Plasmodium berghei
Weiss, M. L., 1978, Israel J. Med. Sc., v. 14 (5), 554-556
Plasmodium berghei, infectivity and immunogenicity, symposium presentation

Plasmodium berghei
Weiss, M. L., 1978, Israel J. Med. Sc., v. 14 (6), 655-658
Plasmodium berghei-infected mice, immune complexes in lungs, symposium presentation

Plasmodium berghei
Weiss, M. L., 1978, Israel J. Med. Sc., v. 14 (6), 682-686
Plasmodium berghei in splenectomized and/or spleen-transposed gerbils and rats, course of infection, histological findings in liver and spleen, specific arming of macrophages

Plasmodium berghei
Weissberger, H.; and Spira, D. T., 1978, Israel J. Med. Sc., v. 14 (5), 590-595
Plasmodium berghei, lymphoblast transformation in rats convalescent for one week to one year from infections of varying intensities, symposium presentation

Plasmodium berghei
Wellde, B. T.; and Diggs, C. L., 1978, Exper. Parasitol., v. 44 (2), 197-201
Plasmodium berghei, mice, antiserum treatment of infections resulted in population of parasites with altered antiserum susceptibility and virulence

Plasmodium berghei
Werbel, L. M.; et al., 1978, J. Med. Chem., v. 21 (4), 337-339
Plasmodium berghei, 6-[(aryloxy- and arylthio-)methyl]-2,4-pteridinediamines and -pteridinediamine 8-oxides, synthesis for antimalarial evaluation, none showed significant activity in trials with mice

Plasmodium berghei
Worth, D. F.; et al., 1978, J. Med. Chem., v. 21 (4), 331-337
Plasmodium gallinaceum, P. berghei, 6-[[(aryl and aralkyl)amino]methyl]-2,4-pteridinediamines and -pteridinediamine 8-oxides, synthesized for antimalarial evaluation, laboratory trials with experimental animals

Plasmodium berghei
Wyler, D. J.; Oppenheim, J. J.; and Koontz, L. C., 1979, Infect. and Immun., v. 24 (1), 151-159
Plasmodium berghei, P. yoelii, mice, effects of infection on ability of adherent mononuclear cells to elaborate soluble mediators that regulate lymphocyte activation in vitro

Plasmodium berghei
van Zon, A.; Eling, W.; and Jerusalem, C., 1978, Israel J. Med. Sc., v. 14 (6), 659-672
Plasmodium berghei, histo- and immunopathology in 6 different mouse strains, symposium presentation

Plasmodium berghei berghei
Bafort, J., 1970, Ann. Soc. Belges Med. Trop. Parasitol., v. 50 (2), 247-262
Plasmodium berghei berghei, differences between various isolates, assumed to be pleomorphic species, characteristics compared with P.b. killicki and P.b. yoelii

Plasmodium berghei berghei, illus.
Bafort, J. M., 1971, Ann. Soc. Belges Med. Trop. Parasitol., v. 51 (1), 5-203
Plasmodium v. vinckei; P. b. berghei, biology of rodent malaria, monograph covering life cycle in vertebrate and invertebrate hosts, light and electron microscopic study of morphology

Plasmodium berghei berghei
Boonpucknavig, V.; Boonpucknavig, S.; and Bhamarapravati, N., 1979, Arch. Path. and Lab. Med., v. 103 (11), 567-572
Plasmodium berghei berghei-infected mice treated with chloroquine phosphate, focal glomerulonephritis in hyperimmune state, clinical, immunopathologic, and histopathologic findings

Plasmodium berghei berghei
Endardjo, S.; et al., 1978, J. Trop. Med. and Hyg., v. 81 (2-3), 25-31
Plasmodium berghei berghei, Swiss albino mice infected intraperitoneally, capacity of cyclophosphamide to suppress immune response and increase pathology attributed to suppressive effects of drug upon protein synthesis, cell division and activity of reticuloendothelial cells

Plasmodium berghei berghei
Gillet, J.; and Herman, F., 1971, Ann. Soc. Belges Med. Trop. Parasitol., v. 51 (6), 655-677
Plasmodium berghei berghei, erythrocytic forms inoculated into mouse embryos, development, reproduction, mice at birth had either no evidence of infection or had overwhelming parasitemia with extended period of parasite development

Plasmodium berghei berghei
Gillet, J.; and Herman, F., 1976, Ann. Soc. Belge Med. Trop., v. 56 (3), 143-150
Trypanosoma equiperdum, absence of transplacental passage in mice, comparison of similar results with congenital malaria in mice

Plasmodium berghei berghei, illus.
Howells, R. E.; and Bafort, J. M., 1970, Ann. Soc. Belges Med. Trop. Parasitol., v. 50 (5), 587-594
Plasmodium berghei berghei, pre-erythrocytic schizonts in liver of Thamnomys surdaster, presence of various respiratory enzymes demonstrated, light microscope histochemical techniques

Plasmodium b. berghei
Jadin, J.; Timperman, G.; and De Ruysser, F., 1975, Ann. Soc. Belge Med. Trop., v. 55 (6), 603-608
Plasmodium b. berghei, pyrimethamine-resistant strain preserved at very low temperature for 11 years, maintained virulence but lost drug resistance, gametocytogenesis increased, cyclical transmission was successful, parasites crossed blood-brain barrier indicating this strain could serve as laboratory model for P. falciparum cerebral malaria

Plasmodium berghei berghei, illus.
van der Kaay, H. J.; and Boorsma, L., 1977, Acta Leidensia, v. 45, 13-19
Plasmodium berghei berghei, selection of a fully susceptible strain of Anopheles atroparvus, not possible to obtain a completely refractive strain, effect of reciprocal matings between susceptible and refractive strains and of backcrosses of F1 offspring with these strains on susceptibility

Plasmodium berghei berghei
Moyou-Somo, R.; et al., 1978, Compt. Rend. Acad. Sc., Paris, v. 286, s. D, Sc. Nat. (12), 993-996
Plasmodium berghei berghei, mice infected and maintained in hot ambient temperature undergo chronic infection whereas controls at laboratory temperature develop acute and lethal infection, the hot environmental temperature does not seem to affect the parasites' pathogenicity but to stimulate host immune defenses

Plasmodium berghei berghei, illus.
Moyou-Somo, R.; et al., 1978, Compt. Rend. Acad. Sc., Paris, v. 286, s. D, Sc. Nat. (18), 1315-1317
Plasmodium berghei berghei in mice maintained at high temperature (35°C), certain parasites (less than 30%) show atypical morphology (gigantism), amount of DNA is higher than in parasites from mice grown at 20-22°C, no evidence of relationship between increase in DNA and morphological modification

Plasmodium berghei berghei
Parbtani, A.; and Cameron, J. S., 1979, Kidney Internat., v. 16 (1), 53-63
Plasmodium spp., exper. infections in mice, nephritis dependent upon deposition of immune complexes in the kidney accompanies acute infections

smodium berghei berghei
an Ros, G.; et al., 1972, Ann. Soc. Belge Med.
rop., v. 52 (6), 515-530
Plasmodium berghei berghei, inbred mouse strain partially resistant to infection, surveillance of hemoglobins and glucose-6-phosphate-dehydrogenase changes from 12th day of gestation to 3rd day after birth, no correlations with Plasmodium resistance

smodium berghei berghei
incke, I. H., 1970, Ann. Soc. Belges Med.
rop. Parasitol., v. 50 (3), 339-358
Plasmodium berghei berghei, mice, action of pyrimethamine and sulphormethoxine on pre-erythrocytic and sporogonous cycles

smodium berghei berghei
dderburn, N., 1974, Ciba Found. Symp., n.s.
25), 123-135, 147-159
Plasmodium berghei subspp.-infected mice, immunodepression

smodium berghei killicki
ufort, J., 1970, Ann. Soc. Belges Med. Trop.
arasitol., v. 50 (2), 247-262
Plasmodium berghei berghei, differences between various isolates, assumed to be pleomorphic species, characteristics compared with P.b. killicki and P.b. yoelii

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n, 1968
andau, I., 1973, Mem. Mus. National Hist.
at., Paris, n. s., s. A, Zool., v. 77, 1-62
Coccidiomorpha, life cycles, mechanisms assuring continuation of transmission, thesis
Thamnomys rutilans: Congo-Brazzaville

odium berghei yoelii
fort, J., 1970, Ann. Soc. Belges Med. Trop.
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Plasmodium berghei berghei, differences between various isolates, assumed to be pleomorphic species, characteristics compared with P.b. killicki and P.b. yoelii

smodium berghei yoelii
ogren, C. H.; and Weidanz, W. P., 1978, Pro-
es Biol. Fluids, v. 26, 233-237
Plasmodium berghei yoelii, mice, antibodies and antigens studied by quantitative immunoelectrophoresis

odium berghei yoelii
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22 (6), 694-699
Plasmodium spp. in mice and rhesus monkeys, synthesis and antimalarial activity of some 4-substituted 8-amino-6-methoxyquinolines, 4-ethylprimaquine was approximately as active and was less toxic than primaquine

smodium berghei yoeli, illus.
yman, B. C.; and MacInnis, A. J., 1979, J.
arasitol., v. 65 (3), 421-425
Plasmodium spp., rapid detection in blood smears by fluorescence microscopy with 4'6 diamidino-2-phenylindole; can also be used to stain Toxoplasma and microfilariae

smodium berghei yoelii
ayawardena, A. N.; et al., 1978, Immunology,
. 34 (1), 157-165
Plasmodium berghei yoelii (P. yoelii), mice, passive transfer of immunity with serum and cells

Plasmodium berghei yoelii Landau et Killick-Kendrick, 1966
Landau, I., 1973, Mem. Mus. National Hist. Nat., Paris, n. s., s. A, Zool., v. 77, 1-62
Coccidiomorpha, life cycles, mechanisms assuring continuation of transmission, thesis
Thamnomys rutilans: Republique Centrafricaine
T. surdaster (exper.)
T. sp. proches de surdaster (exper.)
rats blancs (exper.)
souris blanches (exper.)
Steatomys (exper.)

Plasmodium berghei yoeli, illus.
Miller, F. W.; and Ilan, J., 1978, Parasitology, v. 77 (3), 345-365
Plasmodium berghei yoeli, isolation of ultrastructurally intact viable parasites free from detectable host ribosome contamination, isolation of ribosomes in high yield from these parasites, ribosomal RNA analysis

Plasmodium berghei yoelii
Parbtani, A.; and Cameron, J. S., 1979, Kidney Internat., v. 16 (1), 53-63
Plasmodium spp., exper. infections in mice, nephritis dependent upon deposition of immune complexes in the kidney accompanies acute infections

Plasmodium berghei yoelii
Rane, D. S.; and Kinnamon, K. E., 1979, Am. J. Trop. Med. and Hyg., v. 28 (6), 937-947
sporozoite-induced Plasmodium berghei in mice, development of high volume tissue schizonticidal drug screen based upon mortality of infected mice

Plasmodium berghei yoelii
Rosenberg, Y. J., 1978, Nature, London (5667), v. 274, 170-172
Plasmodium berghei yoelii, mice, plaque-forming cell assays used to reveal pattern of both total and antigen-specific splenic B lymphocyte activation and to define anti-erythrocytic autoimmune response, both responses shown to be T-cell dependent

Plasmodium berghei yoelii
Wedderburn, N., 1974, Ciba Found. Symp., n.s. (25), 123-135, 147-159
Plasmodium berghei subspp.-infected mice, immunodepression

Plasmodium berghei yoelii
Weinbaum, F. I.; et al., 1978, J. Immunol., v. 121 (2), 629-636
Plasmodium berghei yoelii (substrain 17X nonlethal) in BALC/c mice, kinetics of various specific and nonspecific cellular and humoral responses during course of infection

Plasmodium brasilianum, illus.
Ayala, S. C., 1978, Vet. Med. and Small Animal Clin., v. 73 (2), 217-218
Plasmodium brasilianum in pet Cebus capucinus (blood), case report, chloroquine, good results, public health implications: Cali Colombia, acquired from Catatumbo River region along Colombia-Venezuela border, South America

Plasmodium brasilianum
Deane, L. M.; et al., 1969, Rev. Inst. Med. Trop. S. Paulo, v. 11 (2), 71-86
malaria parasites of Brazilian monkeys, summary of survey done from 1964-1968 (hosts, vectors, environmental characteristics of areas covered, discussion on zoonotic potential)

Plasmodium brasilianum
Deane, L. M.; et al., 1971, Rev. Inst. Med. Trop. S. Paulo, v. 13 (3), 167-170
Alouatta belzebul: Estado do Para, Brasil

Plasmodium brasilianum
Deane, L. M.; et al., 1971, Rev. Inst. Med. Trop. S. Paulo, v. 13 (5), 311-319
simian malaria, Anopheles cruzi as vector in State of Sao Paulo and probably also in eastern and southern areas, A. neivai suspected as vector in Manaus area: Brazil

Plasmodium brasilianum, illus.
Deane, L. M.; d'Andretta, C., jr.; and Kameyama, I., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (2), 144-148
Alouatta sp., probably A. caraya (sangue): Parque Nacional do Xingu, Estado de Mato Grosso

Plasmodium brasilianum
Deane, L. M.; and Ferreira, J. A. (Neto), 1969, Rev. Inst. Med. Trop. S. Paulo, v. 11 (5), 299-305
Plasmodium brasilianum, P. simium, Anopheles cruzi as possible vector
Alouatta fusca clamitans: Estado do Rio Grande do Sul, Brasil

Plasmodium brasilianum
Deane, L. M.; and Ferreira, J. A. (Neto), 1973, Rev. Inst. Med. Trop. S. Paulo, v. 15 (3), 107-111
Alouatta caraya (sangue): Municipio de Monte do Carmo, vale do Rio Tocantins, Estado de Goias

Plasmodium brasilianum
Ferreira, J. A. (Neto); and Deane, L. M., 1973, Rev. Inst. Med. Trop. S. Paulo, v. 15 (3), 112-115
Callicebus moloch cupreus
Cebus apella macrocephalus
Saimiri sciureus
Cacajao rubicundus rubicundus
Alouatta seniculus seniculus
Lagothrix lagotricha poeppigii
(sangue cardiaco of all): all from Municipio de Cruzeiro do Sul, Estado do Acre, Brasil

Plasmodium brasilianum
Ferreira, J. A. (Neto); Deane, L. M.; and de Almeida, F. B., 1972, Rev. Inst. Med. Trop. S. Paulo, v. 14 (4), 231-234
Callicebus moloch cupreus (sangue): Seringal Ouro, Sena Madureira

Plasmodium brasilianum Gonder & Berenberg-Gossler, 1908
Ferreira, J. A. (Neto); Deane, L. M.; Carneiro, E. W. B., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (3), 169-174
Alouatta belzebul belzebul (sangue): Santa Luzia and Imperatriz, Estado do Maranhao, Brasil

Plasmodium (Haemamoeba) cathemerium Hartman, 192
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
neohapantotype designated

Plasmodium cathemerium Hartman, 1927
Kairullaev, K. K., 1979, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (1), 19-21
Emberiza citrinella
Fringilla coelebs
(blood of all): all from central course of Ural river, Priural region, Ural oblast

Plasmodium cathemerium
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Plasmodium chabaudi
Buengener, W., 1979, Tropenmed. u. Parasitol., v. 30 (2), 198-205
Plasmodium spp. in mice, parasitization of mature vs. immature erythrocytes

Plasmodium chabaudi
Carter, R., 1978, Parasitology, v. 76 (3), 241-267
Plasmodium berghei, P. yoelii, P. vinckei, P. chabaudi, and their subspecies, electrophoretic variation of enzymes glucose phosphate isomerase, 6-phosphogluconate dehydrogenase, lactate dehydrogenase, and glutamate dehydrogenase, detailed description of technique, genetic and taxonomic implications, key for identification of murine plasmodia by enzyme type

Plasmodium (Vinckeia) chabaudi
Chance, M. L.; et al., 1978, Ann. Trop. Med. an Parasitol., v. 72 (1), 13-22
Plasmodium (Vinckeia) spp., value of DNA studies employing buoyant density determinations and measurements of nucleotide sequence homology in systematics and identification

Plasmodium chabaudi, illus.
David, P. H.; et al., 1978, Proc. National Acad. Sc., v. 75 (10), 5081-5084
Plasmodium chabaudi, method of merozoite isolation: release of merozoites from schizonts bound to immobilized concanavalin A

Plasmodium chabaudi
Eugui, E. M.; and Allison, A. C., 1979, Parasitology, v. 79 (2), 267-275
Plasmodium, 3 murine spp., density-gradient centrifugation in metrizamide for separating uninfected erythrocytes from erythrocytes containing parasites in different developmental stages

Plasmodium chabaudi
Fabia, F.; et al., 1979, Ann. Parasitol., v. 54 (1), 1-10
Plasmodium chabaudi-infected mice, increase in sialic acids, neutral hexoses, and fucose on surface of red blood cells, coincides with reticulocytosis

asmodium chabaudi
Leke, R. G. F.; and Viens, P., 1978, IRCS J.
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Plasmodium chabaudi-infected mice (T-cell deprived, sham-thymectomized and normal), course of infection, comparison with P. yoelii; T-cell deprived mice with P. chabaudi (unlike P. yoelii) gave reactions similar to those of normal and sham-deprived mice indicating that this malaria model is not thymus dependent

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Plasmodium chabaudi, thymectomized mice more susceptible to infection than controls; adoptive transfer of immunity with enriched populations of spleen T and B lymphocytes

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chloroquine) and drug sensitive strains, re-
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Trypanosoma lewisi, Babesia rodhaini, Plasmo-
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rypanosoma lewisi, rats, presence of anti-
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ion with cold-activated hemagglutinin and
mmunoconglutinin, temporal relationships
ith anemia and parasitemia

Plasmodium chabaudi adami
Carter, R., 1978, Parasitology, v. 76 (3), 241-267
Plasmodium berghei, P. yoelii, P. vinckei, P. chabaudi, and their subspecies, electrophoretic variation of enzymes glucose phosphate isomerase, 6-phosphogluconate dehydrogenase, lactate dehydrogenase, and glutamate dehydrogenase, detailed description of technique, genetic and taxonomic implications, key for identification of murine plasmodia by enzyme type

Plasmodium chabaudi chabaudi
Carter, R., 1978, Parasitology, v. 76 (3), 241-267
Plasmodium berghei, P. yoelii, P. vinckei, P. chabaudi, and their subspecies, electrophoretic variation of enzymes glucose phosphate isomerase, 6-phosphogluconate dehydrogenase, lactate dehydrogenase, and glutamate dehydrogenase, detailed description of technique, genetic and taxonomic implications, key for identification of murine plasmodia by enzyme type

Plasmodium (Giovannolaia) circumflexum Kikuth, 1931
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P[lasmodium] coatneyi
Areekul, S., 1979, Southeast Asian J. Trop. Med. and Pub. Health, v. 10 (3), 438-443
malaria, humans and monkeys (exper.), bleeding diathesis

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Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
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Plasmodium cyclopsi n. sp., illus.
Landau, I.; and Chabaud, A. G., 1978, Ann. Parasitol., v. 53 (3), 247-253
Hipposideros cyclops: Makokou (Gabon)

Plasmodium cynomolgi Mayer, illus.
Aikawa, M.; et al., 1979, J. Protozool., v. 26 (2), 273-279
Plasmodium spp., sporozoites before and after incubation in immune serum, freeze-fracture study, antibody-induced changes of pellicular membrane

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Plasmodium knowlesi infection in Macaca mulatta terminated by curative therapy and followed by P. cynomolgi infection greatly increases gametocyte infectivity of latter species

Plasmodium cynomolgi
Carroll, F. I.; et al., 1979, J. Med. Chem., v. 22 (6), 694-699
Plasmodium spp. in mice and rhesus monkeys, synthesis and antimalarial activity of some 4-substituted 8-amino-6-methoxyquinolines, 4-ethylprimaquine was approximately as active and was less toxic than primaquine

Plasmodium cynomolgi
Carroll, F. I.; Berrang, B.; and Linn, C. P., 1978, J. Med. Chem., v. 21 (4), 326-330
Plasmodium berghei, P. cynomolgi, experimental animals, resolution of antimalarial agents via complex formation with α-(2,4,5,7-tetranitro-9-fluorenylideneaminooxy)propionic acid, significant differences in toxicity

Plasmodium cynomolgi
Khan, M. Sami; and LaMontagne, M. P., 1979, J. Med. Chem., v. 22 (8), 1005-1008
3- and 5-aminoquinolines, potential antimalarials, synthesis, testing of some against Plasmodium berghei in mice, Leishmania donovani in hamsters, or P. cynomolgi in rhesus monkeys

Plasmodium cynomolgi
Moser, G.; et al., 1978, J. Protozool., v. 25 (1), 119-124
Plasmodium berghei, P. knowlesi, P. cynomolgi, purification of sporozoites by passage through DEAE-cellulose column, retention of ability to produce infection, to induce protective immunity, and to react with known antisera

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Schmidt, L. H., 1979, Antimicrob. Agents and Chemotherapy, v. 16 (4), 475-485
Plasmodium spp. in monkeys, floxacrine, lacking in radical curative activity, significant prophylactic activity but with requirement for daily dosage, untoward host reaction

Plasmodium cynomolgi
Schmidt, L. H.; et al., 1978, Antimicrob. Agents and Chemotherapy, v. 13 (6), 1011-1030
Plasmodium falciparum in Aotus trivirgatus griseimembra, pilot appraisals of activities of 12 4-quinolinemethanols, further appraisal of mefloquine with P. vivax in Aotus trivirgatus and P. cynomolgi in Macaca mulatta

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Schmidt, L. H.; et al., 1978, Antimicrob. Agents and Chemotherapy, v. 14 (3), 292-314
Plasmodium falciparum in Aotus trivirgatus griseimembra and human volunteers, P. cynomolgi in Macaca mulatta, antimalarial activities of various 9-phenanthrenemethanols with special attention to WR-122,455 and WR-171,669, includes some [apparently unpublished] results of other workers for P. berghei and these same compounds

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Plasmodium cynomolgi, monkeys infected with Ro or Ro/PM strains, appraisals of various activities of CI-679

Plasmodium cynomolgi
Scovill, J. P.; et al., 1979, J. Med. Chem., v. 22 (10), 1164-1167
Plasmodium cynomolgi in rhesus monkeys, Leishmania donovani in golden hamster, derivatives of 4-amino-2-methoxyacridine

Plasmodium cynomolgi
Shetty, R. V.; and Blanton, C. D., jr., 1978, J. Med. Chem., v. 21 (9), 995-998
2-substituted primaquine analogues synthesized and evaluated in laboratory animals against Plasmodium berghei, P. cynomolgi, Trypanosoma rhodesiense, Leishmania donovani and Schistosoma mansoni; significant activit was observed against P. berghei and L. donovani

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Plasmodium cynomolgi-rhesus monkey antimalarial screening of 5-phenylthio and 5-anilino derivatives of primaquine in an attempt to diminish toxicity of primaquine

Plasmodium cynomolgi bastianellii
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

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Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
hapantotype designated

Plasmodium dissanaikei de Jong
Laird, M., 1978, J. Protozool., v. 25 (1), 21-24
transferred from subgenus Novyella to Giovannolaia

Plasmodium (Giovannolaia) durae Herman, 1941
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neohapantotype designated

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Laird, M., 1978, J. Protozool., v. 25 (1), 21-24
Pavo cristatus (blood): Jos, Benue Plateau, Nigeria

Plasmodium (Huffia) elongatum Huff, 1930
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
neohapantotype designated

Plasmodium elongatum
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Plasmodium relictum, P. elongatum in Spheniscus demersus, diagnostic methods evaluated, chloroquine phosphate and primaquine phosphate therapy: Baltimore Zoo

Plasmodium falciparum
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Plasmodium spp., differential diagnosis, review

Plasmodium falciparum
Al Tawil, N., 1977, J. Trop. Med. and Hyg., v. 80 (12), 230-237
Plasmodium falciparum, strain resistant to chloroquine therapy discovered in Vientiane, Lao People's Democratic Republic

Plasmodium falciparum
Al Tawil, N., 1978, Southeast Asian J. Trop. Med. and Pub. Health, v. 9 (3), 409-413
Plasmodium falciparum, semi-immune humans, clearance of asexual parasitaemia with single dose sulfadoxine-pyrimethamine, comparison with standard dose of chloroquine over 3 days: Laos

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malaria, humans and monkeys (exper.), bleeding diathesis

lasmodium falciparum
Astaf'eva, N. V.; et al., 1979, Sovet. Med. (5), 93-95
malaria, humans, complication of pregnancy, case reports

[lasmodium] falciparum
Atangana, S.; et al., 1979, Med. Trop., v. 39 (5), 537-543
onchocerciasis, malaria, humans, epidemiological and vector survey; no evidence of schistosomiasis but potential vectors are present; little evidence of Toxoplasma gondii: lac de retenue de Bamendjin, Cameroun

lasmodium falciparum
Baruzzi, R. G.; et al., 1971, Ann. Soc. Belges Med. Trop. Parasitol., v. 51 (2), 205-213
high incidence of splenomegaly in Indians of Alto Xingu region, serological investigation for various parasitic diseases indicated only malaria as a possible cause: Brazil

lasmodium] falciparum
Baruzzi, R. G.; et al., 1976, Rev. Inst. Med. Trop. S. Paulo, v. 18 (5), 322-348
P[lasmodium] spp., association with tropical splenomegaly syndrome in Indians from Alto Xingu region, Central Brazil

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Plasmodium falciparum, man, renal disease associated with acute infection, extensive review

asmodium falciparum
Boreham, P. F. L.; et al., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (4), 418-423
incidence of multiple feeding by Anopheles gambiae; strong correlation between ahaptoglobinaemia and malaria infections in villagers: Barmawa, Garki District, Kano State, Nigeria

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malarias, humans, clinical trials using sulphones and sulphonamides with a pyrimidine derivative

asmodium falciparum
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human malarias, clinical findings, brief review

P[lasmodium] falciparum
Bray, R. S., 1978, J. Parasitol., v. 64 (3), 410
absence of circumsporozoite antibodies (to P[lasmodium] falciparum) in areas of hyperendemic malaria in The Gambia, appears that antibodies to sporozoites of human malaria parasites are not generated in nature

Plasmodium falciparum
Bray, R. S.; and Anderson, M. J., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (4), 427-431
Plasmodium falciparum, prevalence and density in pregnant women (by age/parity), recently pregnant women, and infants, malarial antibody levels in cord blood, seasonal variations: The Gambia

Plasmodium falciparum
Brohult, J.; et al., 1979, Ann. Trop. Med. and Parasitol., v. 73 (4), 327-331
malaria, humans, analysis of chemoprophylactic habits and reasons for breakdowns in therapy, small mining town: Yekepa, Liberia

Plasmodium falciparum
Brohult, J.; et al., 1979, Ann. Trop. Med. and Parasitol., v. 73 (5), 401-405
Plasmodium falciparum, chloroquine use by non-immunes when in endemic areas, analysis of drug concentrations in human serum during short and long term malaria prophylaxis, recommendations for standard vs. double dosage, clinical implications

Plasmodium falciparum
Buck, A. A.; Anderson, R. I.; and MacRae, A. A., 1978, Tropenmed. u. Parasitol., v. 29 (3), 253-268
polyparasitism, humans, epidemiology, assessment of combined effects of multiple infections on an individual's state of health, comparative analysis of field data obtained from several tropical villages

Plasmodium falciparum
Cabrera, B. D.; and Arambulo, P. V. III, 1977, Acta Trop., v. 34 (3), 265-279
Plasmodium spp., epidemiologic review: Philippines

Plasmodium falciparum
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Plasmodium gallinaceum-parasitized chicken erythrocytes used in a practical hemagglutination test for IgM antibodies in human malarias, useful complementary tool for seroepidemiologic studies

Plasmodium falciparum
Chan, K. L.; Goh, K. T.; and Koh, T. S., 1976, Singapore Med. J., v. 17 (3), 124-132
malaria, human, large post-war outbreak in highly urbanized area, epidemiology, vector survey, outbreak traced to at least one imported case from neighboring country where malaria is endemic: Whampoa-Kallang area of Singapore

P[lasmodium] falciparum
Charoenlarp, P.; Vanijanonta, S.; and Chatpanyaporn, P., 1979, Southeast Asian J. Trop. Med. and Pub. Health, v. 10 (1), 127-131
falciparum malaria, humans, prednisolone administered with quinine sulfate did not increase red cell survival

Plasmodium falciparum
Chin, W., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (3), 334-335
malaria, method for large-volume cultivation of parasites based on principle of Trager-Jensen culture method

[Plasmodium] falciparum
Chongsuphajaisiddhi, T.; et al., 1979, Southeast Asian J. Trop. Med. and Pub. Health, v. 10 (1), 132-137
falciparum malaria, children, chloroquine resistance, efficacy of quinine and fansidar, clinical study: Thailand

Plasmodium falciparum
Collins, W. E.; et al., 1978, J. Parasitol., v. 64 (3), 497-500
Plasmodium falciparum, Burma (Thau.) strain, passage from man to Aotus trivirgatus monkey and from monkey to monkey by means of blood parasites or sporozoites, some characteristics of infection in monkey and in Anopheles freeborni

Plasmodium falciparum
Collins, W. E.; et al., 1979, J. Parasitol., v. 65 (4), 605-608
Plasmodium vivax, Plasmodium falciparum, Aotus trivirgatus, effect of prior malarial experience with one species on subsequent malarial infection with another species in terms of parasitemia and mosquito infectivity

Plasmodium falciparum
Collins, W. E.; et al., 1979, J. Parasitol., v. 65 (5), 763-767
Plasmodium falciparum, West African I strain, parasitemia in Aotus trivirgatus subspp., mosquito infectivity and transmission studies
Anopheles freeborni
A. maculatus
A. culicifacies
A. balabacensis balabacensis
A. albimanus
Aotus trivirgatus trivirgatus
A. trivirgatus griseimembra
(all exper.)

Plasmodium falciparum
Collins, W. E.; et al., 1979, Mosquito News, v. 39 (3), 466-472
Plasmodium falciparum, P. vivax (2 strains), Anopheles freeborni (exper.), susceptibility of natural and selected pupal color phenotypes to infection

Plasmodium falciparum
Cornille Broegger, R.; et al., 1978, Bull. World Health Organ., v. 56 (4), 579-600
Plasmodium falciparum, P. malariae, changing patterns in humoral immune response before, during, and after application of control measures: Nigeria

Plasmodium falciparum
Cornille-Brøgger, R.; et al., 1979, Ann. Trop. Med. and Parasitol., v. 73 (2), 173-183
malaria in normal subjects and those with sickle cell trait, determination of plasma immunoglobulins and antimalarial antibodies, findings suggest that during infancy early phagocytosis of parasitized cells led to enhanced processing of antigen and hence earlier immune response to sickle cell trait

Plasmodium falciparum
Coulaud, J. P.; et al., 1979, Ann. Med. Int., Paris, v. 130 (10), 693-696
imported malaria, increased incidence, inadequate prophylaxis, frequent re-infestation in black Africans living in France and returning periodically to endemic areas, special risks for pregnant women: Paris

Plasmodium falciparum, illus.
Cuartas, F.; et al., 1972, South. Med. J., v. 65 (6), 523, 546
Plasmodium falciparum, soldiers, diagnosis by bone marrow aspiration: Texas, recently returned from Vietnam

Plasmodium falciparum
Das, P. K.; Reuben, R.; and Batra, C. P., 1979, Indian J. Med. Research, v. 69, 403-411
Plasmodium vivax, P. falciparum in Anopheles stephensi (nat. and exper.), only likely vector in Salem, Tamil Nadu

Plasmodium falciparum
Degremont, A., 1977, Schweiz. Med. Wchnschr., v. 107 (18), 641-642
human malarias and amoebiasis, brief review of current diagnostic methods

Plasmodium falciparum, illus.
Dei Cas, E.; et al., 1979, Ann. Parasitol., v. 54 (5), 567-570
Plasmodium, diagnosis, advantages of Errecart's modified technique for making thick blood films

Plasmodium falciparum
Dias, L. C. S.; and Baruzzi, R. G., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (2), 210 [Letter]
malaria, children, prevalence, comparison with findings prior to start of control program: Xingu Indian Reservation, Mato Grosso State, Brazil

Plasmodium falciparum
Donaldson, P.; et al., 1978, Stain Tech., v. 53 (4), 225-227
Plasmodium spp., diagnosis using borax methylene blue, spectroscopic and staining data

Plasmodium falciparum
Draper, C. C.; et al., 1973, Rev. Soc. Brasil. Med. Trop., v. 7 (4), 231-235
Plasmodium falciparum, P. vivax, statistics of survey using thick blood films and indirect fluorescent antibody test, results showed low prevalence of infection and that P. vivax was the predominate parasite: Amazonas

Plasmodium falciparum
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malaria, epidemiological model applied to transmission, includes both human and entomological factors

Plasmodium falciparum
Dutta, H. M.; and Dutt, A. K., 1978, Social Sc. and Med., v. 12 (2D), 69-84
malaria ecology, a global perspective, extensive review

P[lasmodium] falciparum
Eales, L., 1974, S. African Med. J., v. 48 (32), 1386-1389
acute falciparum malaria, human, complications and treatment, review

Plasmodium falciparum
Eke, R. A., 1979, Am. J. Trop. Med. and Hyg., v. 28 (6), 1074-1075
Plasmodium falciparum, possible chloroquine-resistant strain, recrudescence of infection in 42-year-old hospital worker after chloroquine therapy, radical cure with sulfadiazine and pyrimethamine: Nigeria

Plasmodium falciparum
Ellis, C. J.; et al., 1979, Brit. Med. J. (6160), v. 1, Feb. 10, 385-388
Plasmodium spp., humans emigrating or returning from endemic areas, striking rise in incidence, need for use of prophylactic drugs by travelers to endemic areas and for increased vigilance in diagnosing and treating malaria: East Birmingham Hospital and St. Thomas's Hospital, London

Plasmodium falciparum
Elslager, E. F.; et al., 1978, J. Med. Chem., v. 21 (10), 1059-1070
Plasmodium spp. in laboratory animals, thioquinazoline analogues synthesized and tested under laboratory conditions showed substantial suppressive antimalarial and prophylactic activity when compared with reference compounds, analogues also retained potent antimalarial effects against strains resistant to common antimalarials; the most active compound, 2,4-diamino-6-[(α,α,α-trifluoro-m-tolyl)thio]quinazoline has been designated for preclinical toxicologic studies

Plasmodium falciparum
Elslager, E. F.; et al., 1979, J. Med. Chem., v. 22 (10), 1247-1257
Plasmodium spp., antimalarial activity of 2,4-diamino-6-(2-naphthylsulfonyl)quinazoline and related 2,4-diamino-6-[(phenyl and naphthyl)sulfinyl and sulfonyl]quinazolines

Plasmodium falciparum
Facer, C. A.; Bray, R. S.; and Brown, J., 1979, Clin. and Exper. Immunol., v. 35 (1), 119-127
Plasmodium falciparum, Gambian children, direct Coombs antiglobulin reactions, incidence and class specificity, results indicate that sensitization of non-parasitized erythrocytes contributes to pathogenesis of anemia

P[lasmodium] falciparum
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[Letter]
P[lasmodium] falciparum, no significant difference in frequency of ABO antigens between children with malaria and controls, confirms previous investigations: The Gambia

Plasmodium falciparum
Fascio, M. de N. C.; Pinheiro, A. D.; and de Paula e Souza, A., 1976, Rev. Goiana Med., v. 22 (1-2), 109-110
Plasmodium falciparum, patient in coma and renal failure treated with exchange transfusions, 2% of patient's red blood cells remained parasitized after therapy, 60% parasitized prior to transfusions: Cristalandia, Brasil

P[lasmodium] falciparum
Faulk, W. P.; et al., 1978, Protides Biol. Fluids, v. 26, 427-430
several conditions of abnormal pregnancy including 3 patients with P[lasmodium] falciparum malaria, deposition of complement components within placenta

Plasmodium falciparum
Fein, I. A.; Rackow, E. C.; and Shapiro, L., 1978, Am. Rev. Resp. Dis., v. 118 (2), 425-429
Plasmodium falciparum, woman, acute pulmonary edema as a complication of parasite infection, case findings suggest that the edema was the result of altered capillary membrane permeability

Plasmodium falciparum
Fernex, M., 1977, Schweiz. Med. Wchnschr., v. 107 (18), 643-644
human malarias and amoebiasis, brief review of current therapeutic and prophylactic measures

Plasmodium falciparum
Fernex, M., 1979, Therap. Umschau, v. 36 (3), 205-210
tropical protozoan diseases, humans travelling to endemic areas, preventive measures, review

Plasmodium falciparum
Ferraroni, J. J.; and Hayes, J., 1977, Acta Amazonica, v. 7 (3), 401-406
Plasmodium falciparum, outbreak in 2 indian tribes, Anopheles darlingi only vector mosquito present, tribal differences in infection based on proximity to vector population: Roraima, Brazil

Plasmodium falciparum
Ferraroni, J. J.; and Hayes, J., 1979, Am. J. Trop. Med. and Hyg., v. 28 (5), 909-911
Plasmodium falciparum outbreak among indigenous indian tribe, 3 cases resistant to chloroquine responded favorably to fansidar therapy: Uauaris, Territory of Roraima, Brazil

Plasmodium falciparum
Ferraroni, J. J.; Waki, S.; and Suzuki, M., 1977, Acta Amazonica, v. 7 (1), 147-148
Plasmodium falciparum, humans, chloroquine resistance, confirmed in vitro: Manaus, Amazonas

[Plasmodium] falciparum
Finau, S. A., 1979, N. Zealand Med. J. (649), v. 90, 465-466
[Plasmodium] falciparum, woman who had previously travelled in Papua New Guinea, case report: Tonga

Plasmodium falciparum
Fitch, C. D.; Chevli, R.; and Gonzalez, Y., 1974, Antimicrob. Agents and Chemotherapy, v. 6 (6), 757-762
Plasmodium falciparum, Aotus trivirgatus erythrocytes infected with chloroquine-susceptible vs. chloroquine-resistant strain, effect of substrate (glucose) on chloroquine and amodiaquin accumulation

Plasmodium falciparum, illus.
Fleischer, N. K. F., 1979, Med. Welt., v. 30 (44), 1625-1630
parasitic tropical diseases, humans, central nervous system involvement, clinical review

Plasmodium falciparum
Fleming, A. F.; et al., 1979, Ann. Trop. Med. and Parasitol., v. 73 (2), 161-172
Plasmodium spp., human, prevalence of abnormal hemoglobins, relationships between sickle cell trait, malaria and survival of host: Garki District, Kano State, Nigeria

Plasmodium falciparum
Fogh, S.; Jepsen, S.; and Effersøe, P., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (2), 228-229
Plasmodium falciparum, chloroquine resistant strain in non-immune male, treatment with fansidar terminated infection: Danish tourist to Kenya

Plasmodium falciparum
Friedman, M. J., 1978, Proc. National Acad. Sc., v. 75 (4), 1994-1997
Plasmodium falciparum, development in cells with sickle cell hemoglobin, results suggest that mechanism of sickle cell resistance in vivo may be due solely to intraerythrocytic conditions

Plasmodium falciparum, illus.
Friedman, M. J., 1979, J. Protozool., v. 26 (2), 195-199
Plasmodium falciparum, ultrastructural damage to parasite in sickled erythrocytes

Plasmodium falciparum
Friedman, M. J., 1979, Nature, London (5719), v. 280, 245-247
Plasmodium falciparum, α- and β-thalassaemia trait red cells from adults, fetal red cells, and glucose-6-phosphate dehydrogenase deficiency red cells are refractory to parasite development because of oxidant sensitivity

Plasmodium falciparum
Friedman, M. J.; et al., 1979, Am. J. Trop. Med. and Hyg., v. 28 (5), 777-780
Plasmodium falciparum, in vitro cultures, host cell competence of abnormal hemoglobin-containing erythrocytes, evolutionary significance of results

Plasmodium falciparum
Friedman, M. J.; et al., 1979, Exper. Parasitol., v. 47 (1), 73-80
Plasmodium falciparum, intraerythrocytic conditions in infected cells, changes which would alter sickling behavior of infected AS cells, potassium levels in sickled AS cells, effect of decreased potassium on parasite survival

P[lasmodium] falciparum
Galbraith, R. M.; et al., 1978, Protides Biol. Fluids, v. 26, 229-232
P[lasmodium] falciparum, 10 Gambians with pregnancies complicated by maternal malaria, marked increase in complement components in placentae

Plasmodium falciparum
Gao, M. X.; et al., 1979, Acta Microbiol. Sinica, v. 19 (1), 88-90
Plasmodium falciparum, continuous in vitro culture following deep-freezing of infected human blood

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Plasmodium falciparum, human, transmission by platelet concentrate transfusion, case report

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malaria, human, treatment recommendations

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Plasmodium falciparum, Nigerian woman, severe malarial attack within hours of giving birth to normal twins, at age 2 months one infant developed non-febrile hemolytic anemia resulting from prenatal parasitic infection, other infant possibly protected by partial glucose-6-phosphate dehydrogenase deficiency: Switzerland

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Plasmodium falciparum, acute cerebral manifestations in man, complicated by blackwater fever and diffuse intravascular coagulation, case report, clinical management: Basel, Switzerland (had traveled to North Africa)

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Plasmodium falciparum
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[Plasmodium] falciparum, problems in clinical management of infected persons, importance of providing tourists to endemic areas with information on malaria: Switzerland

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nutritional and health status survey of children under five, includes malaria, Ascaris and hookworm among most frequently diagnosed diseases: Mentawai Island Sipora/Indonesia

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Anopheles minimus (malaria vector), responses to DDT residual spraying in cleared forested foothill area, appeared to reduce estimated vectorial capacities but effect was not maintained and malaria transmission was not interrupted: central Thailand

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Plasmodium b. berghei, pyrimethamine-resistant strain preserved at very low temperature for 11 years, maintained virulence but lost drug resistance, gametocytogenesis increased, cyclical transmission was successful, parasites crossed blood-brain barrier indicating this strain could serve as laboratory model for P. falciparum cerebral malaria

Plasmodium falciparum
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Plasmodium falciparum, gametocytogenesis in continuous cultures, crescent development with regard to timing of sequential stages, current culture methods cannot produce continuous supply of functional gametes for further studies

Plasmodium falciparum
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Plasmodium falciparum, establishment of new strains in continuous culture

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Plasmodium falciparum, recent advances in cultivation, symposium presentation

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Plasmodium falciparum, comparison of dihydrofolate reductase from parasite and from Aotus trivirgatus

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Plasmodium falciparum, chloroquine-resistance, 2 case reports of people having travelled in Africa

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Plasmodium falciparum
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Plasmodium falciparum, erythrocytic cycle in vitro, ultrastructure, comparison with in viv (Aotus trivirgatus) life cycle stages

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Plasmodium falciparum, protection given by haemoglobin S in heterozygotes for sickle cell gene, rebuttal to hypothesis of Pasvol, G.; Weatherall, D. J.; and Wilson, R. J. M., 1978, Nature, London (5672), v. 274, 701-703

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malaria, American civilians, detailed analysis of cases acquired during travel abroad, 1970-1976

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malaria, observations on imported human cases, diagnostic importance, clinical aspects: SSSR

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Plasmodium falciparum
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malaria, human, diagnosis, indirect immunofluorescent test using Plasmodium berghei or P. falciparum as antigen

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malaria, standardization of indirect fluorescent antibody test

Plasmodium falciparum
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Plasmodium spp., detection and measurement of species-specific malarial antibodies using standardized indirect fluorescent antibody test

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malaria, humans, role in tropical splenomegaly syndrome, current appraisal, review

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Plasmodium falciparum, low erythrocyte pyridoxal-kinase activity in serum of non-infected Black and White races in comparison to that of infected Black persons; possible relation to malarial infection, possibly requirement of parasite

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Plasmodium falciparum, infected Nigerian children, survey of blood group antigens, no significant difference from healthy children

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Plasmodium falciparum as cause of acute urticaria, 2 case reports: Ethiopia

Plasmodium falciparum, illus.
Matzen, P.; and Starklint, H., 1976, Ugeskr. Laeger, v. 138 (3), 157-158
Plasmodium falciparum, man, associated Salmonella bacteremia, death due to malarial cerebral edema, infected during conducted tourist trip to Gran Canaria and Gambia: Denmark

Plasmodium falciparum
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malaria, 4 yearly surveys in Surinam with indirect hemagglutination test, general comments on seroepidemiologic surveys and mathematical models for assessment of malaria transmission rates, symposium presentation

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human acute falciparum malaria, changes in serum protein patterns studied using polyacrylamide gel electrophoresis, other blood biochemical parameters

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Plasmodium falciparum, P. malariae, serological study comparing infants exposed to or protected from malaria: Nigeria

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Molineaux, L.; et al., 1979, Ann. Trop. Med. and Parasitol., v. 73 (4), 301-310
sickle cell disease subjects living in hyperendemic malarial area, numbers of malaria-infected persons, seroimmunologic test results, immunoglobulin levels, and age groups compared with subjects without sickle cell trait: Sudan savanna of Nigeria

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Plasmodium falciparum, epidemiological evaluation of mathematical model, baseline data tested against data collected over 3-year period during which propoxur and fenitrothion were used for mosquito control, relatively good agreement between actual observations and model: Garki District, Nigeria and Kisumu District, Kenya

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human malaria, contributions of author to the study of malaria and its eradication in Ecuador

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Plasmodium falciparum, woman, fatal infection accompanied by jaundice and renal failure, unresponsive to quinine therapy, review of clinical aspects: Cayenne (Guyane)

Plasmodium falciparum, illus.
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Plasmodium falciparum-infected erythrocytes from long-term culture, concentration and separation by gradient centrifugation

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Plasmodium spp., diagnosis, current therapeutic measures, review

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children with syndrome of enlarged parotids, localized forehead edema, heavy infestation with Ascaris lumbricoides, and unusual freedom from malaria, piperazine treatment of ascariasis resulted in attacks of malaria, suppression of malaria may be nutritional consequence of severe ascariasis and may represent ecological balance for optimum co-survival of host and two parasites: Anjouan, Comorro archipelago

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parasitic (malaria, Schistosoma) and non-parasitic infections, Somali nomads, adverse effect of iron repletion on course of certain infections

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malaria in Israel and Israeli-held territories, assessment of increasing incidence 1967-1971

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malaria, suggested approach to control, methodology applicable in different epidemiologic situations: the Americas

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Plasmodium falciparum, human, antibodies to sporozoites, occurrence in 4 age groups: Keneba, West Kiang district, The Gambia

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Plasmodium berghei-mouse and P. knowlesi-rhesus monkeys systems, detection of stage and species specific antisporozoite antibodies with circumsporozoite precipitation and indirect immmunofluorescence methods, preliminary application to P. falciparum in humans with similar results

Plasmodium falciparum
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Plasmodium falciparum, P. vivax, prevalence survey in hospital patients, discussion of changes in prevalence with introduction of chloroquine resistant strains of P. falciparum, treatment trials with various malarial drugs: Brazil

Plasmodium falciparum
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Plasmodium falciparum, African strain, production of chloroquine resistance in vitro

Plasmodium falciparum, illus.
Nielsen, R. L.; et al., 1979, Am. J. Med. Sc., v. 277 (3), 325-329
Plasmodium falciparum, woman, intense parasitemia, deepening coma, and acute renal failure, exchange transfusions as useful adjunct to conventional chemotherapy, numbers of infected circulating erythrocytes reduced: United States, had previously travelled in Ghana

Plasmodium falciparum
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children with Burkitt's lymphoma, serum immunoglobulin levels, specific antibody titers to Plasmodium falciparum and P. malariae

Plasmodium falciparum
Omer, A. H. S., 1978, Am. J. Trop. Med. and Hyg., v. 27 (5), 853-857
Plasmodium falciparum, human, clinical trial of response to chloroquine, little evidence of resistance: Gezira and Bor areas, Sudan

Plasmodium falciparum
Omer, A. H. S., 1978, Am. J. Trop. Med. and Hyg., v. 27 (5), 858-863
Plasmodium spp., human, prevalence by parasite species and by host age group, dramatic response to mass chemoprophylaxis with chloroquine: Gezira and Bor regions, Sudan

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body build and nutritional status of three ethnic groups inhabiting same locality, effects of infections including malaria, Schistosoma haematobium and hookworm: Northern Nigeria

P[lasmodium] falciparum
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males of 3 ethnic groups and 3 age groups inhabiting same locality, haematological status (including anemia), spleen and liver enlargement, immunoglobulin status, malaria parasite rates, other parasite infections, possible associations between these and other factors (including nutrition, sickle cell trait, altered immune response to malaria): Northern Nigeria

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Plasmodium falciparum, humans, role of major histocompatibility complex in antibody response under natural conditions: Tanzania

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Pasvol, G.; Weatherall, D. J.; and Wilson, R. J. M., 1978, Nature, London (5672), v. 274, 701-703
Plasmodium falciparum, hemoglobin S has detrimental effect on parasite proliferation, this involves both invasion into red cell and growth once inside and requires conditions of low oxygen tension, actual sickling of cells concerned is not necessary, provides explanation for protection of sickle cell heterozygotes against P. falciparum malaria and thus for high frequency of sickle-cell gene in parts of world where malaria is or has been endemic

Plasmodium falciparum
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Plasmodium falciparum
Phillips, R. S.; Wilson, R. J. M.; and Pasvol, G., 1978, J. Protozool., v. 25 (3, pt. 2), 394-398
Plasmodium falciparum, differentiation of gametocytes in microcultures of infected human blood, can develop both from ring-stage parasites introduced into culture and from merozoites released during subsequent schizogonies in vitro, incorporation of [^{3}H]-isoleucine by developing gametocytes

Plasmodium falciparum
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Plasmodium falciparum, woman, case report, probable resistance to chloroquine, successfully treated with quinine: South Africa (had recently returned from Mozambique)

Plasmodium falciparum
Ponnampalam, J. T., 1978, J. Trop. Med. and Hyg., v. 81 (10), 198-203
human malarias, comparative study of prophylaxis using chloroquine and a combination of sulfadoxine and pyrimethamine: residents of rubber estate in central Malaysia

Plasmodium falciparum
Powell, R. D., 1978, Am. J. Trop. Med. and Hyg., v. 27 (1, pt. 1), 1-5
Plasmodium spp., servicemen returning from Vietnam, relationships between chemoprophylaxis history and occurrence and timing of post-departure episodes of malaria: Iowa City VA Hospital

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Plasmodium falciparum, chloroquine-resistant strain in Aotus trivirgatus, 2 chlorinated lincomycin analogues cured blood-induced infections

Plasmodium falciparum
du Preez, O.; and Cockcroft, C. R., 1975, South African Med. J., v. 49 (34), 1407-1408
malaria prophylaxis trials, army personnel camping in endemic area, 3 drug combinations, no infections reported in trial groups while local population acting as control reported 250 falciparum cases: Caprivi Strip, South Africa

Plasmodium falciparum
Reese, R. T.; et al., 1978, Proc. National Acad. Sc., v. 75 (11), 5665-5668
Plasmodium falciparum, immunization of Aotus monkeys grouped according to karyotype, antigenic material obtained from parasites cultivated in vitro for over a year, protective immunity can be induced without use of complete Freund's adjuvant if sufficient antigen is used together with synthetic muramyl dipeptide

Plasmodium falciparum
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Plasmodium falciparum, inhibition of in vitro growth by immune serum and purified immunoglobulin from Aotus sp.

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Plasmodium spp., issue devoted to current status of human malarias with discussion on: life cycle, geographic distribution, immunopathology, diagnosis, clinical management, treatment and prophylaxis

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Plasmodium falciparum in continuous culture, effects of pyrimethamine and chloroquine on parasite growth and viability

Plasmodium falciparum
Rieckmann, K. H.; Mrema, J. E.; and Campbell, G. H., 1978, J. Parasitol., v. 64 (4), 750-752
Plasmodium falciparum parasites obtained from culture are capable of inducing pronounced immunity to malaria in Aotus trivirgatus griseimembra

Plasmodium falciparum
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Plasmodium falciparum, male drug addict infected by using common injection equipment, case report

Plasmodium falciparum
Robinson, C. R.; Wilson, D.; and Kaplan, E. P., 1979, Canad. Med. Ass. J., v. 120 (2), 130 [Letter]
Plasmodium falciparum, boy, chloroquine, case report, recently travelled to Liberia and Ivory Coast; importance of determining whether patient has travelled outside United States or Canada: Massachusetts

Plasmodium falciparum, illus.
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Plasmodium falciparum, 36-year-old male after travel to Senegal, case report, severe infection cured by exchange blood transfusion in conjunction with classical drug therapy: Argentina

Plasmodium falciparum
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Plasmodium falciparum, increased sickling propensity of infected red cell under conditions of total and partial deoxygenation in vitro, results lend support to concept that heterozygotes for Hb S in malarious region may have improved fitness for survival which in turn maintains balanced polymorphism for Hb S gene

P[lasmodium] falciparum
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P[lasmodium] vivax, P. falciparum, humans, presumptive treatment with 600 mg. chloroquine base, good response, no RII or RIII type chloroquine resistance seen: Dharmapuri and North Arcot districts, Tamil Nadu State

Plasmodium falciparum
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Plasmodium falciparum still sensitive to quinine, in vitro and in vivo (humans): Thailand

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Plasmodium falciparum
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Plasmodium falciparum, humans with chloroquine-resistant infection (exper.), acetylator phenotype does not influence therapeutic response to sulfalene or sulfalene combined with pyrimethamine

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    soluble parasite antigens, possible modes of interference with immune response, review

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    P[lasmodium] spp., humans, general clinical review

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    human malaria, report of worldwide situation as of 1974

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    Babesia, Plasmodium, Trypanosoma, kallikrein-kinin system, mechanisms of activation (parasite enzymes, immune complexes), role in hypotensive shock syndrome of infected animals, review

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    Plasmodium falciparum, response of sensitized and unsensitized human lymphocyte subpopulations to malaria antigen

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    Gyps africanus: Rhodesia

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    Anolis concolor (blood): San Andres Island, western Caribbean

Plasmodium floridense Thompson and Huff 1944
  Telford, S. R., jr., [1979], J. Parasitol., v. 64 (6), 1978, 1126-1127
    Anolis carolinensis
    Sceloporus undulatus
    all from Florida

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  Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
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Plasmodium fragile
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    Plasmodium fragile in Macaca mulatta as model system for study of malarial vaccines

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  Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
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  Carter, R.; Gwadz, R. W.; and McAuliffe, F. M., 1979, Exper. Parasitol., v. 47 (2), 185-193
    Plasmodium gallinaceum, chickens, comparative effectiveness of gametocyte- and gamete-containing preparations in inducing transmission-blocking immunity

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  Ceneviva, A. C.; and Camargo, M. E., 1979, Am. J. Trop. Med. and Hyg., v. 28 (4), 622-626
    Plasmodium gallinaceum-parasitized chicken erythrocytes used in a practical hemagglutination test for IgM antibodies in human malarias, useful complementary tool for seroepidemiologic studies

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    Plasmodium spp. in laboratory animals, thioquinazoline analogues synthesized and tested under laboratory conditions showed substantial suppressive antimalarial and prophylactic activity when compared with reference compounds, analogues also retained potent antimalarial effects against strains resistant to common antimalarials; the most active compound, 2,4-diamino-6-[(α,α,α-trifluoro-m-tolyl)thio]quinazoline has been designated for preclinical toxicologic studies

Plasmodium gallinaceum
  Elslager, E. F.; et al., 1979, J. Med. Chem., v. 22 (10), 1247-1257
    Plasmodium spp., antimalarial activity of 2,4-diamino-6-(2-naphthylsulfonyl)quinazoline and related 2,4-diamino-6-[(phenyl and naphthyl)sulfinyl and sulfonyl]quinazolines

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Aedes aegypti feeding on Plasmodium gallinaceum-infected chickens: take less blood and lay fewer eggs than those feeding on uninfected hosts in inverse correlation with degree of parasitemia, and ingest blood in amounts directly proportional to amount of time spent on hosts (whereas there is no relationship between host contact and blood meal size when feeding on uninfected hosts); infected chickens are less attractive to mosquitoes than uninfected chickens

Plasmodium gallinaceum
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partial suppression of Plasmodium gallinaceum and P. vivax in Aedes aegypti and Anopheles stephensi doubly infected with Nosema algerae and Plasmodium, epidemiological significance

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Plasmodium gallinaceum
Kazim, M.; Puri, S. K.; and Dutta, G. P., 1979, Chemotherapy, v. 25 (4), 222-226
Plasmodium gallinaceum, chicks (exper.), minocycline and doxycycline, blood schizontocidal activity compared with that of known antibiotics, both more effective than oxytetracycline and tetracycline in controlling acute infection

Plasmodium gallinaceum
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Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

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Plasmodium gallinaceum
Worth, D. F.; et al., 1978, J. Med. Chem., v. 21 (4), 331-337
Plasmodium gallinaceum, P. berghei, 6-[[(aryl and aralkyl)amino]methyl]-2,4-pteridinediamines and -pteridinediamine 8-oxides, synthesized for antimalarial evaluation, laboratory trials with experimental animals

Plasmodium (Giovannolaia) garnhami Guindy, Hoogstraal and Mohamed, 1965
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Gonatodes albogularis (exper.)

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"P. gonzalezi must be considered a nomen dubium"

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reported earlier as Plasmodium cnemidophori
Plica plica: vicinity of Georgetown, Guyana

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Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
hapantotype designated

Plasmodium hexamerium, illus.
Ayala, S. C.; et al., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (6), 411-416
Zenaida auriculata caucae (blood): Cauca River valley, Colombia

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Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
parahapantotype designated

Plasmodium (Huffia) huffi Muniz, Saores and Batista, 1951
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
neohapantotype designated

Plasmodium inui, illus.
Dei Cas, E.; et al., 1979, Ann. Parasitol., v. 54 (5), 567-570
Plasmodium, diagnosis, advantages of Errecart's modified technique for making thick blood films

Plasmodium inuei
Scalise, G.; et al., 1978, J. Med. Primatol., v. 7 (2), 114-118
Plasmodium inuei-infected Macaca mulatta had enhanced susceptibility to hepatitis B virus

Plasmodium inui
Trigg, M. E., 1979, South. Med. J., v. 72 (5), 593-599
immune functions of the spleen, includes information on malaria with special reference to Plasmodium inui in rhesus monkeys

Plasmodium (Novyella) juxtanucleare Versiani Gomes, 1941
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
neohapantotype designated

Plasmodium juxtanucleare, illus.
Krettli, A. U., 1972, Rev. Inst. Med. Trop. S. Paulo, v. 14 (4), 235-245
Plasmodium juxtanucleare, new foci, prevalence survey, pathology, recently isolated strain and 20 year old strain compared for forms and virulence
Gallus gallus domesticus (nat. and exper.) (spleen, liver, bone marrow, brain, blood): State of Minas Gerais, Brazil

Plasmodium knowlesi, illus.
Aikawa, M.; et al., 1978, J. Cell Biol., v. 77 (1), 72-82
Plasmodium knowlesi, invasion of erythrocytes by merozoites investigated by electron microscopy, findings include junction between erythrocytes and merozoites, movement of junction during invasion, and fate of surface coat on merozoites

Plasmodium knowlesi, illus.
Aikawa, M.; et al., 1979, J. Protozool., v. 26 (2), 273-279
Plasmodium spp., sporozoites before and after incubation in immune serum, freeze-fracture study, antibody-induced changes of pellicular membrane

P[lasmodium] knowlesi
Areekul, S., 1979, Southeast Asian J. Trop. Med. and Pub. Health, v. 10 (3), 438-443
malaria, humans and monkeys (exper.), bleeding diathesis

Plasmodium knowlesi
de Arruda-Mayr, M.; Cochrane, A. H.; and Nussenzweig, R. S., 1979, Am. J. Trop. Med. and Hyg., v. 28 (4), 627-633
Plasmodium knowlesi infection in Macaca mulatta terminated by curative therapy and followed by P. cynomolgi infection greatly increases gametocyte infectivity of latter species

Plasmodium knowlesi
Butcher, G. A.; Mitchell, G. H.; and Cohen, S., 1978, Immunology, v. 34 (1), 77-86
Plasmodium knowlesi, Macaca mulatta, antibody mediated mechanisms associated with sterilizing immunity induced by merozoite vaccination role of Freund's complete adjuvant

Plasmodium knowlesi
Carter, R.; Gwadz, R. W.; and Green, I., 1979, Exper. Parasitol., v. 47 (2), 194-208
Plasmodium gallinaceum, chickens, elaboration of antigamete antibodies during immunization and infection and their relationship to fertilization of malaria gametes in vitro and to transmission-blocking immunity in vivo, comparison with rhesus monkeys immunized with P. knowlesi gametes

Plasmodium knowlesi
Chen, Z. R.; et al., 1978, Acta Microbiol. Sinica, v. 18 (4), 332-335
Plasmodium knowlesi, cultivation by continuous inoculation technique, infectivity to monkeys

Plasmodium knowlesi
Collins, W. E.; Contacos, P. G.; and Chin, W., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (6), 662-663
Plasmodium knowlesi infections in Saimiri sciureus (exper.), may prove to be suitable host for vaccine studies

Plasmodium knowlesi
Deans, J. A.; Dennis, E. D.; and Cohen, S., 1978, Parasitology, v. 77 (3), 333-344
Plasmodium knowlesi, antigenic analysis of sequential erythrocytic stages using crossed immuno-electrophoresis

Plasmodium knowlesi
Dutta, G. P.; Singh, P. P.; and Banyal, H. S., 1978, Indian J. Med. Research, v. 68, 923-926
Plasmodium knowlesi in Macaca assamensis (exper.), possible alternate host for experimental studies

Plasmodium knowlesi
Gutteridge, W. E.; Dave, D.; and Richards, W. H. G., 1979, Biochim. et Biophys. Acta, v. 582 (3), 390-401
Kinetoplastida spp., Plasmodium spp., conversion of dihydroorotate to orotate, mechanism of reaction different in these 2 groups of protozoa, possible target of chemotherapeutic attack

Plasmodium knowlesi
Gwadz, R. W.; and Green, I., 1978, J. Exper. Med., v. 148 (5), 1311-1323
Plasmodium knowlesi, rhesus monkeys, vaccine effective against both sexual and asexual stages

Plasmodium knowlesi
Langhorne, J.; and Cohen, S., 1979, Parasitology, v. 78 (1), 67-76
Plasmodium knowlesi in Callithrix jacchus investigated as possible model for immunological studies, course of infection, differential susceptibility, resistance to challenge infection

Plasmodium knowlesi, illus.
McLaren, D. J.; et al., 1979, Parasitology, v. 79 (1), 125-139
Plasmodium knowlesi, interaction between malaria parasite and host erthrocyte, freeze fracture studies of internal cytoarchitecture of surface membranes

Plasmodium knowlesi, illus.
Miller, L.; et al., 1979, J. Exper. Med., v. 149 (1), 172-184
Plasmodium knowlesi, interaction between cytochalasin B-treated merozoites and erythrocytes, attachment and junction formation, results suggest that defect in invasion of Duffy-negative RBCs is at the step of junction formation

Plasmodium knowlesi
Molineaux, L.; et al., 1979, Ann. Trop. Med. and Parasitol., v. 73 (4), 301-310
sickle cell disease subjects living in hyperendemic malarial area, numbers of malaria-infected persons, seroimmunologic test results, immunoglobulin levels, and age groups compared with subjects without sickle cell trait: Sudan savanna of Nigeria

Plasmodium knowlesi
Moser, G.; et al., 1978, J. Protozool., v. 25 (1), 119-124
Plasmodium berghei, P. knowlesi, P. cynomolgi, purification of sporozoites by passage through DEAE-cellulose column, retention of ability to produce infection, to induce protective immunity, and to react with known antisera

Plasmodium knowlesi
Nardin, E. H.; and Nussenzweig, R. S., 1978, Nature, London (5666), v. 274, 55-57
Plasmodium berghei-mouse and P. knowlesi-rhesus monkeys systems, detection of stage and species specific antisporozoite antibodies with circumsporozoite precipitation and indirect immmunofluorescence methods, preliminary application to P. falciparum in humans with similar results

Plasmodium knowlesi
Onabanjo, A. O., 1978, Agents and Actions, v. 8 (1-2), 139-140
Plasmodium knowlesi in Macaca mulatta, plasma kinin concentration, physiopathological effects of kinin extracts from blood of infected monkeys when injected intradermally into guinea pigs

Plasmodium knowlesi
Schmidt-Ullrich, R.; and Wallach, D. F. H., 1978, Proc. National Acad. Sc., v. 75 (10), 4949-4953
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Plasmodium knowlesi
Shakespeare, P. G.; Trigg, P. I.; and Tappenden, L., 1979, Ann. Trop. Med. and Parasitol., v. 73 (4), 333-343
Plasmodium knowlesi, surface properties of normal rhesus monkey erythrocytes and of infected erythrocytes, externally disposed protein used as probes of surface changes, pigment-free preparation of membrane proteins obtained, possible application in preparing specific antigens

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Schmidt-Ullrich, R.; Wallach, D. F. H.; and Lightholder, J., 1979, J. Exper. Med., v. 150 (1), 86-99
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Plasmodium knowlesi
Shakespeare, P. G.; et al., 1979, Ann. Trop. Med. and Parasitol., v. 73 (5), 407-415
Plasmodium knowlesi, glucose metabolism, activities of the glycolytic and pentose phosphate pathways during the intraerythrocytic cycle, studies suggest that parasite carries its own complement of glycolytic enzymes

Plasmodium knowlesi, illus.
Spangler, W. L.; et al., 1978, Vet. Path., v. 15 (1), 83-91
Plasmodium knowlesi in Macaca mulatta (exper.), pathology

Plasmodium knowlesi
Storey, J.; et al., 1979, Ann. Trop. Med. and Parasitol., v. 73 (4), 311-315
malaria, immunoglobulins and antimalarial antibodies in haemoglobin AC individuals, little difference from rest of population except for higher IgG levels, suggests that haemoglobin C gene's geographical relationship to malaria may be coincidence: Sudan savanna of Nigeria

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Babesia, Plasmodium, Trypanosoma, kallikrein-kinin system, mechanisms of activation (parasite enzymes, immune complexes), role in hypotensive shock syndrome of infected animals, review

Plasmodium lainsoni new species, illus.
Telford, S. R., jr., 1978, Internat. J. Parasitol., v. 8 (5), 341-353
effects upon host cells
Phyllodactylus ventralis (erythroblasts, proerythrocytes, erythrocytes): Fundo Vega Honda, 25 km N Guanare, Municipio Guanare, Estado Portuguesa, Venezuela

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Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
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Plasmodium lophurae, illus.
Hollingdale, M. R.; and Kilejian, A., 1979, J. Protozool., v. 26 (4), 616-619
Plasmodium lophurae, exoerythrocytic merozoites grown in embryonic turkey brain cells, purification by ion exchange column

Plasmodium lophurae, illus.
Hyman, B. C.; and MacInnis, A. J., 1979, J. Parasitol., v. 65 (3), 421-425
Plasmodium spp., rapid detection in blood smears by fluorescence microscopy with 4'6 diamidino-2-phenylindole; can also be used to stain Toxoplasma and microfilariae

Plasmodium lophurae
Kilejian, A., 1978, Science (4359), v. 201, 922-924
Plasmodium lophurae, ducklings, successful immunization with purified and characterized histidine-rich protein as antigen, use of adjuvant is not required for this protective effect and immunity can be passively transferred with serum

Plasmodium lophurae
Platzer, E. G.; and Kassis, J. A., 1978, J. Protozool., v. 25 (4), 556-559
Plasmodium lophurae, pyridoxine kinase in trophozoites and in duckling erythrocytes, results suggest that vitamin B_6 metabolism of malaria parasites is distinct and separate from that of host erythrocytes

Plasmodium lophurae, illus.
Sherman, I. W.; and Jones, L. A., 1979, J. Protozool., v. 26 (3), 489-501
Plasmodium lophurae, membrane proteins of erythrocyte-free plasmodia and malaria-infected red cells

Plasmodium lophurae, illus.
Takahashi, Y.; and Sherman, I. W., 1978, Exper. Parasitol., v. 44 (2), 145-154
Plasmodium lophurae, differentiation of parasite membrane, parasitophorous vacuole membrane, and duck erythrocyte membrane with cationized ferritin staining as an electron microscope cytochemical method

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Yamada, K. A.; and Sherman, I. W., 1979, Exper. Parasitol., v. 48 (1), 61-74
Plasmodium lophurae, hemozoin (malarial pigment), composition and properties

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Telford, S. R., jr., 1979, J. Parasitol., v. 65 (3), 409-413
Egernia cunninghami
E. striolata
E. whitei (exper.)
all from Southern Queensland, Australia (presumably)

Plasmodium malariae
Abranches, P., 1975, Rev. Portug. Med. Mil., v. 23 (1-2), 64-66
Plasmodium spp., differential diagnosis, review

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Alvarez Chacon, R.; Rodriguez Encarnacion, L.; and Diaz Herrera, A., 1974, Salud Pub. Mexico, v. 15 (2), 213-216
Plasmodium malariae, hospitalized children, infected by blood transfusions, clinical aspects of 12 cases, diagnosis, therapy: Mexico, D.F., Mexico

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Baruzzi, R. G.; et al., 1976, Rev. Inst. Med. Trop. S. Paulo, v. 18 (5), 322-348
P[lasmodium] spp., association with tropical splenomegaly syndrome in Indians from Alto Xingu region, Central Brazil

Plasmodium malariae
Boreham, P. F. L.; et al., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (4), 418-423
incidence of multiple feeding by Anopheles gambiae; strong correlation between ahaptoglobinaemia and malaria infections in villagers: Barmawa, Garki District, Kano State, Nigeria

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(31), 1351-1352
malarias, humans, clinical trials using sulphones and sulphonamides with a pyrimidine derivative

Plasmodium malariae
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human malarias, clinical findings, brief review

Plasmodium malariae
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nephrotic syndrome in Malawian Blacks attributed to Plasmodium malariae and use of mercury-containing skin lightening cream

Plasmodium malariae
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polyparasitism, humans, epidemiology, assessment of combined effects of multiple infections on an individual's state of health, comparative analysis of field data obtained from several tropical villages

Plasmodium malariae
Cabrera, B. D.; and Arambulo, P. V. III, 1977, Acta Trop., v. 34 (3), 265-279
Plasmodium spp., epidemiologic review: Philippines

Plasmodium malariae
Cornille Broegger, R.; et al., 1978, Bull. World Health Organ., v. 56 (4), 579-600
Plasmodium falciparum, P. malariae, changing patterns in humoral immune response before, during, and after application of control measures: Nigeria

Plasmodium malariae
Cornille-Brøgger, R.; et al., 1979, Ann. Trop. Med. and Parasitol., v. 73 (2), 173-183
malaria in normal subjects and those with sickle cell trait, determination of plasma immunoglobulins and antimalarial antibodies, findings suggest that during infancy early phagocytosis of parasitized cells led to enhanced processing of antigen and hence earlier immune response to sickle cell trait

Plasmodium malariae
Coulaud, J. P.; et al., 1979, Ann. Med. Int., Paris, v. 130 (10), 693-696
imported malaria, increased incidence, inadequate prophylaxis, frequent re-infestation in black Africans living in France and returning periodically to endemic areas, special risks for pregnant women: Paris

Plasmodium malariae
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human malarias and amoebiasis, brief review of current diagnostic methods

Plasmodium malariae
Dias, L. C. S.; and Baruzzi, R. G., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (2), 210 [Letter]
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Ellis, C. J.; et al., 1979, Brit. Med. J. (6160), v. 1, Feb. 10, 385-388
Plasmodium spp., humans emigrating or returning from endemic areas, striking rise in incidence, need for use of prophylactic drugs by travelers to endemic areas and for increased vigilance in diagnosing and treating malaria: East Birmingham Hospital and St. Thomas's Hospital, London

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parasitic tropical diseases, humans, central nervous system involvement, clinical review

lasmodium malariae
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Plasmodium spp., human, prevalence of abnormal hemoglobins, relationships between sickle cell trait, malaria and survival of host: Garki District, Kano State, Nigeria

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malaria, human, treatment recommendations

lasmodium malariae
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Gimeno de Sande, A., 1975, Rev. San. e Hig. Pub., v. 49 (2), 109-140
human malarias, epidemiologic survey of diagnosed cases that have occurred in Spain since 1964 (all malarias thought to have been eradicated as of that date); all reported cases since that time were found to be imported by persons from endemic areas

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malaria, humans, current status in southern islands of Singapore

lasmodium malariae
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human malaria imported into Switzerland by travellers to tropical endemic areas, statistical review of reported cases, suggestions for diagnosis and clinical management

lasmodium malariae
Hendrickse, R. G.; and Adeniyi, A., 1979, Kidney Internat., v. 16 (1), 64-74
Plasmodium malariae, children, causing immune complex nephritis, presenting clinical and biochemical findings, renal pathology

Plasmodium malariae
Jopling, W. H., 1979, Lancet, London (8130), v. 1, 1340 [Letter]
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P[lasmodium] malariae
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malaria, human, situation 1970-1974 in Singapore

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Plasmodium malariae, infections in humans who had received transfusions of blood from infected donor: Sardinia

Plasmodium malariae
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malaria, human, diagnosis, indirect immunofluorescent test using Plasmodium berghei or P. falciparum as antigen

Plasmodium malariae
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malaria, standardization of indirect fluorescent antibody test

Plasmodium malariae
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Plasmodium malariae
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sickle cell disease subjects living in hyperendemic malarial area, numbers of malaria-infected persons, seroimmunologic test results, immunoglobulin levels, and age groups compared with subjects without sickle cell trait: Sudan savanna of Nigeria

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Plasmodium spp., diagnosis, current therapeutic measures, review

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Plasmodium malariae, 15-year-old girl, after blood transfusion, blood donor was asymptomatic carrier (from Nigeria) who satisfied currently recommended standards for blood donations: Tennessee

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children with Burkitt's lymphoma, serum immunoglobulin levels, specific antibody titers to Plasmodium falciparum and P. malariae

Plåsmodium malariae
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Omer, A. H. S., 1978, Am. J. Trop. Med. and Hyg., v. 27 (5), 858-863
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body build and nutritional status of three ethnic groups inhabiting same locality, effects of infections including malaria, Schistosoma haematobium and hookworm: Northern Nigeria

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males of 3 ethnic groups and 3 age groups inhabiting same locality, haematological status (including anemia), spleen and liver enlargement, immunoglobulin status, malaria parasite rates, other parasite infections, possible associations between these and other factors (including nutrition, sickle cell trait, altered immune response to malaria): Northern Nigeria

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malaria, humans, incidence of parasites in peripheral blood of out-patients: Siriraj Hospital

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malaria, human, treatment, prophylaxis, review

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asmodium malariae
Storey, J.; et al., 1979, Ann. Trop. Med. and Parasitol., v. 73 (4), 311-315
malaria, immunoglobulins and antimalarial antibodies in haemoglobin AC individuals, little difference from rest of population except for higher IgG levels, suggests that haemoglobin C gene's geographical relationship to malaria may be coincidence: Sudan savanna of Nigeria

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Plasmodium spp., humans, incidence survey, distribution by age and sex of host, multiple infections more common in children: rural areas of Zambia

asmodium malariae
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P[lasmodium] spp., humans, general clinical review

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Williamson, W. A.; and Gilles, H. M., 1978, Ann. Trop. Med. and Parasitol., v. 72 (4), 323-328
malariometric study of young children (seasonal distribution, age-specific parasite prevalence, splenic indices): Malumfashi, Northern Nigeria

Plasmodium marginatum sp. nov.
Telford, S. R., jr., 1979, Ann. Parasitol., v. 54 (2), 129-144
reported earlier as Plasmodium tropiduri and P. minasense
Anolis frenatus: Frijoles River, 5 km N of Gamboa, Canal Zone, Panama

Plasmodium (Haemamoeba) matutinum Huff, 1937
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
neohapantotype designated

Plasmodium minasense Carini and Rudolph, 1912
Telford, S. R., jr., 1979, Ann. Parasitol., v. 54 (4), 409-422
taxonomic revision; definition, range, hosts

Plasmodium minasense anolisi subsp. nov., illus.
Telford, S. R., jr., 1979, Ann. Parasitol., v. 54 (4), 409-422
diagnosis
Anolis limifrons: Frijoles River, 4.8 km N Gamboa, Canal Zone, Panama; San Blas Territory, Panama

Plasmodium minasense capitoi subsp. nov., illus.
Telford, S. R., jr., 1979, Ann. Parasitol., v. 54 (4), 409-422
diagnosis
Anolis capito: Frijoles River, 4.8 km N Gamboa, Canal Zone, Panama

Plasmodium minasense carinii Leger and Mouzels, 1917, illus.
Telford, S. R., jr., 1979, Ann. Parasitol., v. 54 (4), 409-422
diagnosis
Iguana iguana: Salvador, Bahia; presumably in the vicinity of Barranquilla, Colombia

Plasmodium minasense diminutivum Telford, 1973, new combination, illus.
Telford, S. R., jr., 1979, Ann. Parasitol., v. 54 (4), 409-422
diagnosis
Ameiva ameiva praesignis: Guayabalito Village, Rio Chagres, Colon Province, Panama

Plasmodium minasense minasense Carini and Rudolph, 1912, illus.
Telford, S. R., jr., 1979, Ann. Parasitol., v. 54 (4), 409-422
diagnosis
Mabuya agilis: River Paranahyba and its tributary Bagagem, Minas Gerais, Brasil

Plasmodium minasense plicae subsp. nov., illus.
Telford, S. R., jr., 1979, Ann. Parasitol., v. 54 (4), 409-422
diagnosis
Plica umbra
Plica plica
all from vicinity of Georgetown, Guyana

Plasmodium minasense tegui subsp. nov., illus.
Telford, S. R., jr., 1979, Ann. Parasitol., v. 54 (4), 409-422
diagnosis
Tupinambis teguixin: San Jorge, Municipio Piritu, Estado Portuguesa, Venezuela

Plasmodium (Novyella) nucleophilum Manwell, 1935
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
neohapantotype designated

Plasmodium (Novyella) octamerium Manwell, 1970
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
neohapantotype designated

Plasmodium ovale
Abranches, P., 1975, Rev. Portug. Med. Mil., v. 23 (1-2), 64-66
Plasmodium spp., differential diagnosis, review

P[lasmodium] ovale
Atangana, S.; et al., 1979, Med. Trop., v. 39 (5), 537-543
onchocerciasis, malaria, humans, epidemiological and vector survey; no evidence of schistosomiasis but potential vectors are present; little evidence of Toxoplasma gondii: lac de retenue de Bamendjin, Cameroun

Plasmodium ovale
Bounameaux, Y., 1977, Schweiz. Med. Wchnschr., v. 107 (18), 638-639
human malarias, clinical findings, brief review

Plasmodium ovale
Cabrera, B. D.; and Arambulo, P. V. III, 1977, Acta Trop., v. 34 (3), 265-279
Plasmodium spp., epidemiologic review: Philippines

Plasmodium ovale
Coulard, J. P.; et al., 1979, Ann. Med. Int., Paris, v. 130 (10), 693-696
imported malaria, increased incidence, inadequate prophylaxis, frequent re-infestation in black Africans living in France and returning periodically to endemic areas, special risks for pregnant women: Paris

Plasmodium ovale
Degremont, A., 1977, Schweiz. Med. Wchnschr., v. 107 (18), 641-642
human malarias and amoebiasis, brief review of current diagnostic methods

Plasmodium ovale
Dutta, H. M.; and Dutt, A. K., 1978, Social Sc. and Med., v. 12 (2D), 69-84
malaria ecology, a global perspective, extensive review

Plasmodium ovale
Ellis, C. J.; et al., 1979, Brit. Med. J. (6160), v. 1, Feb. 10, 385-388
Plasmodium spp., humans emigrating or returning from endemic areas, striking rise in incidence, need for use of prophylactic drugs by travelers to endemic areas and for increased vigilance in diagnosing and treating malaria: East Birmingham Hospital and St. Thomas's Hospital, London

Plasmodium ovale
Fernex, M., 1979, Therap. Umschau, v. 36 (3), 205-210
tropical protozoan diseases, humans travelling to endemic areas, preventive measures, review

Plasmodium ovale
Fleming, A. F.; et al., 1979, Ann. Trop. Med. and Parasitol., v. 73 (2), 161-172
Plasmodium spp., human, prevalence of abnormal hemoglobins, relationships between sickle cell trait, malaria and survival of host: Garki District, Kano State, Nigeria

Plasmodium ovale
Gear, J. H. S.; et al., 1978, South African Med. J., v. 53 (23), 921 [Letter]
malaria, human, treatment recommendations

Plasmodium ovale
Gimeno de Sande, A., 1975, Rev. San. e Hig. Pub., v. 49 (2), 109-140
human malarias, epidemiologic survey of diagnosed cases that have occurred in Spain since 1964 (all malarias thought to have been eradicated as of that date); all reported cases since that time were found to be imported by persons from endemic areas

Plasmodium ovale
Hatz, B.; et al., 1978, Schweiz. Med. Wchnschr., v. 108 (39), 1495-1499
human malaria imported into Switzerland by travellers to tropical endemic areas, statistical review of reported cases, suggestions for diagnosis and clinical management

Plasmodium ovale
Jopling, W. H., 1979, Lancet, London (8130), v. 1, 1340 [Letter]
malaria, travellers, chemoprophylaxis, recommendations

Plasmodium ovale
Le Bras, J.; and Payet, M., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (5), 552 [Letter]
Plasmodium spp., human, concentration of parasitized erythrocytes by centrifugation on Ficoll R solution, useful method when attempting to confirm doubtful diagnosis

Plasmodium ovale
Loban, K. M.; and Polozok, E. S., 1978, Sovet. Med. (5), 59-61
human malaria, errors in diagnosing cases imported into non-endemic areas

Plasmodium ovale
Lopez, C. E.; Ruebush, T. K. II; and Schultz, M. G., 1979, J. Infect. Dis., v. 139 (2), 255-260
malaria, American civilians, detailed analysis of cases acquired during travel abroad, 1970-1976

Plasmodium ovale
Maier, W. A.; and Piekarski, G., 1979, Immun. u. Infekt., v. 7 (3), 75-82
malaria, human, diagnosis, indirect immunofluorescent test using Plasmodium berghei or P. falciparum as antigen

Plasmodium ovale
Muehlemann, P., 1977, Riv. Med. Svizzera Ital., v. 42 (12), 495-498
Plasmodium spp., diagnosis, current therapeutic measures, review

Plasmodium ovale
Omer, A. H. S., 1978, Am. J. Trop. Med. and Hyg., v. 27 (5), 858-863
Plasmodium spp., human, prevalence by parasite species and by host age group, dramatic response to mass chemoprophylaxis with chloroquine: Gezira and Bor regions, Sudan

lasmodium ovale
Rev. Prat., Paris, v. 27 (37), 1977, 2341-2409
Plasmodium spp., issue devoted to current status of human malarias with discussion on: life cycle, geographic distribution, immunopathology, diagnosis, clinical management, treatment and prophylaxis

lasmodium ovale
Sheehy, T. W., 1979, Current Therapy (Conn), 37-39
malaria, human, treatment, prophylaxis, review

lasmodium ovale
Shin, J. W., 1974, Taehan Uihak Hyophoe Chi (J. Korean Med. Ass.), v. 17 (8), 569-573
Plasmodium spp., human, life cycle, therapeutic recommendations

lasmodium ovale
Smith, D. H.; Warren, K. S.; and Mahmoud, A. A. F., 1979, Am. J. Trop. Med. and Hyg., v. 28 (2), 220-229
Schistosoma mansoni, human, morbidity in relation to prevalence and intensity, host age and sex, importance of environmental factors such as ecology of transmission and presence of malaria: Nduru, Kisumu, Kenya

lasmodium ovale
Storey, J.; et al., 1979, Ann. Trop. Med. and Parasitol., v. 73 (4), 311-315
malaria, immunoglobulins and antimalarial antibodies in haemoglobin AC individuals, little difference from rest of population except for higher IgG levels, suggests that haemoglobin C gene's geographical relationship to malaria may be coincidence: Sudan savanna of Nigeria

lasmodium ovale
Thompson, R. G.; et al., 1979, Brit. Med. J. (6168), v. 1, 952 [Letter]
malaria, immigrants and travellers, incidence, recommendations for educating travellers and the general public: Wolverhampton

asmodium ovale
Trenholme, G. M.; and Carson, P. E., 1978, J. Am. Med. Ass., v. 240 (21), 2293-2295
malaria, therapy and prophylaxis recommendations, primaquine, quinine, chloroquine hydrochloride

asmodium ovale
Trzaska, B.; Czesnel, Z.; and Golba, J., 1974, Przegl. Epidemiol., v. 28 (3), 389-395
Plasmodium ovale, clinical aspects and diagnostic criteria of infections in 2 African natives living in Poland

asmodium ovale
Van Ros, G.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (4), 659-663
Plasmodium spp.-infected patients, hemoglobin A_2 levels not statistically different from healthy controls, malaria is thus unlikely to influence results of surveys for β-thalassemia in malaria-endemic areas

Plasmodium ovale
Vu Thi Phan, 1977, Sante Pub., v. 20 (2), 147-156
human malarias, current status, eradication programs: Republique Socialiste du Vietnam

Plasmodium ovale
Wenlock, R. W., 1978, East African Med. J., v. 55 (6), 268-276
Plasmodium spp., humans, incidence survey, distribution by age and sex of host, multiple infections more common in children: rural areas of Zambia

Plasmodium ovale
Wenlock, R. W., 1979, J. Trop. Med. and Hyg., v. 82 (5), 90-98
malaria, hookworm, Schistosoma haematobium, humans in rural environment, epidemiology in relation to malnutrition and host age, importance as public health problems: Zambia

Plasmodium ovale
Willcox, M.; et al., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (2), 209-211
malaria influence on haemoglobin A_2 levels is not a significant factor in assessing β-thalassaemia trait in a malarial endemic area: northern Liberia

Plasmodium ovale
Williamson, W. A.; and Gilles, H. M., 1978, Ann. Trop. Med. and Parasitol., v. 72 (4), 323-328
malariometric study of young children (seasonal distribution, age-specific parasite prevalence, splenic indices): Malumfashi, Northern Nigeria

P[lasmodium] ovale
Wolfensberger, H. R., 1979, Therap. Umschau, v. 36 (3), 250-253
P[lasmodium] spp., humans, general clinical review

Plasmodium ovale/vivax
Armstrong, J. C., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (4), 342-344
Plasmodium vivax/ovale, prevalence rates for Nilotic and Hamitic-Semitic populations residing together in small town show that the two ethnic groups are innately different in susceptibility to patent infection with vivax malaria: Ethiopia

Plasmodium (Haemamoeba) paddae Brumpt, 1935
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
neohapantotype designated

Plasmodium (Giovannolaia) pediocetii Schillinger, 1942 emend. (Stabler and Kitzmuller, 1973)
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
parahapantotype designated

Plasmodium (Giovannolaia) pedioecetii
Stabler, R. M., [1979], J. Parasitol., v. 64 (6), 1978, 1125-1126
Tympanuchus pallidicinctus (blood): Chaves County, New Mexico; Lipscomb County, Texas

Plasmodium pessoai sp. n., illus.
Ayala, S. C.; Moreno, E.; and Bolanos, R., 1978, J. Parasitol., v. 64 (2), 330-335
Spilotes pullatus (blood): 'La Selva' near Puerto Viejo, Heredia Province, Costa Rica
Lachesis muta (blood): Bananito Sur, Limon Province, about 100 km to southeast, Costa Rica

Plasmodium (Giovannolaia) pinotti Muniz and Soares, 1954
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
neohapantotype designated

Plasmodium pitheci, presumably
Rijksen, H. D., 1978, Mededel. Landbouwhogeschool, Wageningen, v. 78 (2), 420 pp.
Pongo pygmaeus abelii (blood): Ketambe area, Sumatra

Plasmodium polare, illus.
Ayala, S. C.; and Varela, C. E., 1975, Rev. Inst. Med. Trop. S. Paulo, v. 17 (4), 253-256
Plasmodium polare, immature Chordeiles minor (blood), first report from South America, potential for intercontinental spread of avian malarias: Cali, Colombia (on migration route from North to South America)

Plasmodium (Giovannolaia) polare Manwell, 1934
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
neohapantotype designated

Plasmodium polare Manwell, 1934
Kairullaev, K. K., 1979, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (1), 19-21
Phylloscopus trochilus (blood): central course of Ural river, Priural region, Ural oblast

Plasmodium praecox relictum
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Plasmodium (Haemamoeba) relictum Grassi and Feletti, 1891
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
neohapantotype designated

Plasmodium relictum Grassi et Feletti, 1890
Kairullaev, K. K., 1979, Izvest. Akad. Nauk Kazakhsk. SSR, s. Biol. (1), 19-21
Luscinia svecica
Parus major
P. coeruleus
(blood of all): all from central course of Ural river, Priural region, Ural oblast

Plasmodium relictum, illus.
Kingston, N.; et al., 1976, J. Wildlife Dis., v. 12 (4), 562-565
Plasmodium relictum in falcons, treated with chloroquine
Falco peregrinus
F. rusticolus
(blood of all): all from breeding facility near Fort Collins, Colorado

Plasmodium relictum
Stoskopf, M. K.; and Beier, J., 1979, J. Am. Vet. Med. Ass., v. 175 (9), 944-947
Plasmodium relictum, P. elongatum in Spheniscus demersus, diagnostic methods evaluated, chloroquine phosphate and primaquine phosphate therapy: Baltimore Zoo

Plasmodium (Haemamoeba) relictum lutzi Lucena, 1939
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
neohapantotype designated

Plasmodium rhadinurum Thompson et Huff, 1944, illus.
Telford, S. R., jr., 1979, Ann. Parasitol., v. 54 (4), 409-422
"recognized as a distinct species at present"

Plasmodium (Novyella) rouxi (Sergent, Sergent and Catanei, 1928)
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
neohapantotype designated

Plasmodium scorzai new species, illus.
Telford, S. R., jr., 1978, Internat. J. Parasitol., v. 8 (5), 341-353
effects upon host cells
Phyllodactylus ventralis (proerythrocytes, erythrocytes): Fundo Vega Honda, 25 km N Guanare, Municipio Guanare, Estado Portuguesa, Venezuela

Plasmodium simiovale
Collins, W. E.; and Contacos, P. G., 1979, J. Parasitol., v. 65 (4), 609-612
Plasmodium simiovale, infection and transmission studies with Macaca mulatta and Anopheles spp.
Anopheles freeborni
A. b. balabacensis
A. maculatus
A. stephensi
A. atroparvus
A. quadrimaculatus
(all exper.)

Plasmodium simium
Deane, L. M.; et al., 1969, Rev. Inst. Med. Trop. S. Paulo, v. 11 (2), 71-86
malaria parasites of Brazilian monkeys, summary of survey done from 1964-1968 (hosts, vectors, environmental characteristics of areas covered, discussion on zoonotic potential)

Plasmodium simium
Deane, L. M.; et al., 1971, Rev. Inst. Med. Trop. S. Paulo, v. 13 (5), 311-319
simian malaria, Anopheles cruzi as vector in State of Sao Paulo and probably also in eastern and southern areas, A. neivai suspected as vector in Manaus area: Brazil

Plasmodium simium
Deane, L. M.; and Ferreira, J. A. (Neto), 1969, Rev. Inst. Med. Trop. S. Paulo, v. 11 (5), 299-305
Plasmodium brasilianum, P. simium, Anopheles cruzi as possible vector
Alouatta fusca clamitans: Estado do Rio Grande do Sul, Brasil

Plasmodium (Haemamoeba) subpraecox Grassi and Feletti, 1892
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
neohapantotype designated

Plasmodium (Novyella) tenue (Laveran and Marullaz, 1914 emend. Manwell, 1970)
Garnham, P. C. C., 1979, J. Trop. Med. and Hyg., v. 82 (9-10), 178-182
neohapantotype designated

Plasmodium traguli sp. n., illus.
Garnham, P. C. C.; and Edeson, J. F. B., 1962, Riv. Malariol., v. 41 (1-3), 1-8
Tragulus javanicus (blood, liver): Malaya

asmodium tropiduri sensu Telford 1974, illus.
Guerrero, S.; and Ayala, S. C., 1977, Rev.
Inst. Med. Trop. S. Paulo, v. 19 (5), 283-288
Anolis fuscoauratus (sangre): selva amazonica, Huanuco, Peru

asmodium (Sauramoeba) tropiduri, illus.
Scorza, J. V., 1972, Rev. Inst. Med. Trop. S.
Paulo, v. 14 (2), 97-105
Plasmodium tropiduri, P. berghei, P. gallinaceum, intraerythrocytic stages, morphological and enzyme cytochemical observations on phagotrophy

asmodium tropiduri Aragao and Neiva, 1909
Telford, S. R., jr., 1979, Ann. Parasitol.,
v. 54 (2), 129-144
definition; taxonomic reconsideration of saurian Plasmodium populations previously identified as P. tropiduri

asmodium tropiduri aquaticum subsp. nov.
Telford, S. R., jr., 1979, Ann. Parasitol.,
v. 54 (2), 129-144
Anolis lionotus: Achiote, Colon Province, Panama
A. poecilopus: central Panama
Anolis lionotus [and/or] A. poecilopus: Costa Rica

asmodium tropiduri panamense subsp. nov.
Telford, S. R., jr., 1979, Ann. Parasitol.,
v. 54 (2), 129-144
Anolis biporcatus: El Aguacate, Panama Province, Panama

asmodium tropiduri tropiduri Aragao and Neiva,
09
Telford, S. R., jr., 1979, Ann. Parasitol.,
v. 54 (2), 129-144
diagnosis; description
Tropidurus: Brazil
T. hispidus: central Venezuela
T. torquatus: Guyana

asmodium vacuolatum
Telford, S. R., jr., 1979, Ann. Parasitol.,
v. 54 (2), 129-144
reported earlier as Plasmodium tropiduri
Plica umbra: Guyana

asmodium vaughani
Bennett, G. F.; White, E. M.; and Williams, N.
A., 1977, J. Wildlife Dis., v. 13 (3), 251-257
Ispidina picta
Merops superciliosus
Anthus leucophrys
Motacilla flava
Parisoma plumbeum
Ploceus weynsi
Quelea erythrops
(blood of all): all from Uganda

asmodium (Novyella) vaughani Novy and MacNeal,
04
Garnham, P. C. C., 1979, J. Trop. Med. and
Hyg., v. 82 (9-10), 178-182
neohapantotype designated

asmodium vaughani Novi and MacNeal, 1907
Kairullaev, K. K., 1979, Izvest. Akad. Nauk
Kazakhsk. SSR, s. Biol. (1), 19-21
Phoenicurus phoenicurus (blood): central course of Ural river, Priural region, Ural oblast

Plasmodium (Novyella) vaughani Novy and MacNeal
1904
Williams, N. A.; and Bennett, G. F., 1978,
J. Parasitol., v. 64 (1), 165-166
Plasmodium vaughani, Culiseta morsitans considered functional exper. vector and possible natural vector in Tantramar Marshes, New Brunswick, status of Mansonia perturbans as potential vector requires further clarification
Agelaius phoeniceus (nat. and exper.): Tantramar Marshes, New Brunswick
Culiseta morsitans (salivary glands, midgut) (exper.)
Mansonia perturbans (exper.)

Plasmodium vaughani Novy and MacNeal
Winchell, E. J., 1978, J. Parasitol., v. 64
(3), 558-559
Icteria virens
Turdus grayi
all from El Salvador

Plasmodium vaughani
Wink, M.; and Bennett, G. F., 1976, J. Wildlife Dis., v. 12 (4), 587-590
Cyanomitra olivacea: Ghana

Plasmodium (Novyella) vaughani merulae Corradetti and Scanga, 1973
Garnham, P. C. C., 1979, J. Trop. Med. and
Hyg., v. 82 (9-10), 178-182
hapantotype designated

Plasmodium vinckei
Carter, R., 1978, Parasitology, v. 76 (3), 241-267
Plasmodium berghei, P. yoelii, P. vinckei, P. chabaudi, and their subspecies, electrophoretic variation of enzymes glucose phosphate isomerase, 6-phosphogluconate dehydrogenase, lactate dehydrogenase, and glutamate dehydrogenase, detailed description of technique, genetic and taxonomic implications, key for identification of murine plasmodia by enzyme type

Plasmodium vinckei
Cox, F. E. G., 1978, Parasitology, v. 76 (1),
55-60
Plasmodium vinckei, Babesia microti, heterologous immunity, simultaneous elimination of the two species from blood of doubly infected mice

Plasmodium vinckei, illus.
Heidrich, H. G.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 151-159
Plasmodium vinckei, separation of infected and uninfected mouse erythrocytes and isolation of free parasites by free-flow electrophoresis

Plasmodium vinckei
Nickel, P.; et al., 1978, Arzneimittel-Forsch.,
v. 28 (4), 578-581
Plasmodium vinckei, synthesis of 2- and 4-amino derivatives of 6-(4-diethylamino-1-methylbutylamino)-5,8-dimethoxyquinoline, laboratory trials in mice for possible antimalarial activity

Plasmodium vinckei
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Plasmodium vinckei brucechwatti
Carter, R., 1978, Parasitology, v. 76 (3), 241-267
Plasmodium berghei, P. yoelii, P. vinckei, P. chabaudi, and their subspecies, electrophoretic variation of enzymes glucose phosphate isomerase, 6-phosphogluconate dehydrogenase, lactate dehydrogenase, and glutamate dehydrogenase, detailed description of technique, genetic and taxonomic implications, key for identification of murine plasmodia by enzyme type

Plasmodium vinckei chabaudi Landau, 1965
Landau, I., 1973, Mem. Mus. National Hist. Nat., Paris, n. s., s. A, Zool., v. 77, 1-62
Coccidiomorpha, life cycles, mechanisms assuring continuation of transmission, thesis
Thamnomys rutilans: Republique Centrafricaine
Thamnomys sp. proches de surdaster (exper.)

Plasmodium vinckei lentum
Carter, R., 1978, Parasitology, v. 76 (3), 241-267
Plasmodium berghei, P. yoelii, P. vinckei, P. chabaudi, and their subspecies, electrophoretic variation of enzymes glucose phosphate isomerase, 6-phosphogluconate dehydrogenase, lactate dehydrogenase, and glutamate dehydrogenase, detailed description of technique, genetic and taxonomic implications, key for identification of murine plasmodia by enzyme type

Plasmodium vinckei lentum Landau et coll., 1969
Landau, I., 1973, Mem. Mus. National Hist. Nat., Paris, n. s., s. A, Zool., v. 77, 1-62
Coccidiomorpha, life cycles, mechanisms assuring continuation of transmission, thesis
Thamnomys rutilans: Congo-Brazzaville

Plasmodium vinckei lentum
Michel, J. C.; Lagrange, P. H.; and Hurtrel, B., 1979, Parasite Immunol., v. 1 (4), 267-275
Plasmodium-infected mice, profound alteration of inductive phase of delayed-type hypersensitivity and antibody formation to sheep erythrocytes when sensitization with antigen was performed intravenously at critical time of disease but not after subcutaneous immunization, suggests major role for spleen in mechanism of immunodepression

Plasmodium vinckei petteri
Carter, R., 1978, Parasitology, v. 76 (3), 241-267
Plasmodium berghei, P. yoelii, P. vinckei, P. chabaudi, and their subspecies, electrophoretic variation of enzymes glucose phosphate isomerase, 6-phosphogluconate dehydrogenase, lactate dehydrogenase, and glutamate dehydrogenase, detailed description of technique, genetic and taxonomic implications, key for identification of murine plasmodia by enzyme type

Plasmodium vinckei petteri
Clark, I. A., 1978, Lancet, London (8080), v. 2 75-77
acute malaria and babesiosis, hypothesis that endotoxin (lipopolysaccharide) causes both th disease and the parasite death, experiments in mice

Plasmodium vinckei petteri
Clark, I. A., 1979, Infect. and Immun., v. 24 (2), 319-325
Babesia spp., Plasmodium vinckei petteri, mice, protection conferred by pretreatment with extract of Coxiella burnetii, possible involvement of interferon or tumor necrosis factor

Plasmodium vinckei petteri, illus.
Eugui, E. M.; and Allison, A. C., 1979, Parasitology, v. 79 (2), 267-275
Plasmodium, 3 murine spp., density-gradient centrifugation in metrizamide for separating uninfected erythrocytes from erythrocytes containing parasites in different developmental stages

Plasmodium vinckei vinckei Rodhain 1952, illus.
Bafort, J. M., 1971, Ann. Soc. Belges Med. Trop. Parasitol., v. 51 (1), 5-203
Plasmodium v. vinckei; P. b. berghei, biology of rodent malaria, monograph covering life cycle in vertebrate and invertebrate hosts, light and electron microscopic study of morphology

Plasmodium vinckei vinckei
Carter, R., 1978, Parasitology, v. 76 (3), 241-267
Plasmodium berghei, P. yoelii, P. vinckei, P. chabaudi, and their subspecies, electrophoretic variation of enzymes glucose phosphate isomerase, 6-phosphogluconate dehydrogenase, lactate dehydrogenase, and glutamate dehydrogenase, detailed description of technique, genetic and taxonomic implications, key for identification of murine plasmodia by enzyme type

Plasmodium (Vinckeia) vinckei vinckei
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Plasmodium (Vinckeia) spp., value of DNA studies employing buoyant density determinations and measurements of nucleotide sequence homology in systematics and identification

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Plasmodium vivax- and P. falciparum-infected Anopheles albimanus (exper.), susceptibility of natural pupal phenotypes to infection

Plasmodium vivax
Weinstok, H.; et al., 1979, Bull. Pan Am. Health Organ., v. 13 (3), 257-263
Plasmodium vivax, P. falciparum, human, survey, indirect immunofluorescence test, prevalence of antibodies: Costa Rica

Plasmodium vivax
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Plasmodium falciparum- and P. vivax-infected Thai adults, loss of circulating T lymphocyt with normal levels of B and 'null' lymphocyt

Plasmodium vivax
Wenlock, R. W., 1978, East African Med. J., v. 55 (6), 268-276
Plasmodium spp., humans, incidence survey, distribution by age and sex of host, multipl infections more common in children: rural areas of Zambia

Plasmodium vivax
Wenlock, R. W., 1979, J. Trop. Med. and Hyg., v. 82 (5), 90-98
malaria, hookworm, Schistosoma haematobium, humans in rural environment, epidemiology in relation to malnutrition and host age, importance as public health problems: Zambia

P[lasmodium] vivax
Wolfensberger, H. R., 1979, Therap. Umschau, v. 36 (3), 250-253
P[lasmodium] spp., humans, general clinical review

Plasmodium vivax hibernans Nikolaev, 1949
Bruce-Chwatt, L. J., 1977, Protozoology, v. 3, 143-148
Plasmodium vivax hibernans, history

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Plasmodium yoelii
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Plasmodium spp. in mice, parasitization of mature vs. immature erythrocytes

Plasmodium yoeli
Carter, R., 1978, Parasitology, v. 76 (3), 241-267
Plasmodium berghei, P. yoelii, P. vinckei, P. chabaudi, and their subspecies, electrophoretic variation of enzymes glucose phosphate isomerase, 6-phosphogluconate dehydrogenase, lactate dehydrogenase, and glutamate dehydrogenase, detailed description of technique, genetic and taxonomic implications, key for identification of murine plasmodia by enzyme type

Plasmodium (Vinckeia) yoelii subsp.
Chance, M. L.; et al., 1978, Ann. Trop. Med. an Parasitol., v. 72 (1), 13-22
Plasmodium (Vinckeia) spp., value of DNA studies employing buoyant density determinations and measurements of nucleotide sequence homology in systematics and identification

Plasmodium yoelii
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Plasmodium berghei and P. yoelii-vaccinated mice, manifestations of cell-mediated immunit

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Plasmodium, 3 murine spp., density-gradient centrifugation in metrizamide for separating uninfected erythrocytes from erythrocytes containing parasites in different developmental stages

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Freeman, R. R., 1978, Cellular Immunol., v. 41 (2), 373-379
Plasmodium, delayed-type hypersensitivity to sheep erythrocytes was depressed during fatal P. berghei and self-limiting P. yoelii infections in mice, immunological lesion found to be at level of DTH expression (i.e., inflammatory response) rather than at level of T cell sensitization

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Freeman, R. R.; and Parish, C. R., 1978, Clin. and Exper. Immunol., v. 32 (1), 41-45
Plasmodium berghei, P. yoelii, mice, numbers of 'background' plaque-forming cells secreting IgM specific for either sheep or horse erythrocytes elevated in spleens during infection or in spleens of uninfected mice injected with non-infectious extracts of parasitized mouse red blood cells, results provide corroborating evidence for hypothesis that B-cell mitogen is associated with blood stage of malaria parasites, possible involvement in immunosuppression

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reeman, R. R.; and Parish, C. R., 1978, Immuology, v. 35 (3), 479-484
Plasmodium, spleen cell changes during fatal P. berghei vs. those during self-limiting P. yoelii infections in mice, protective immunity is associated with marked and sustained increases in numbers of Ig^+ and Thy-1·2^+ spleen cells and in fatal infections these proliferative responses are apparently suppressed

smodium yoelii
nter, K. W., jr.; et al., 1979, J. Immunol., 123 (1), 133-137
Plasmodium yoelii, defective resistance in CBA/N mice, demonstrates that X-linked gene that affects B cell function influences malarial resistance in mice

modium yoelii
ayawardena, A. N.; et al., 1978, Immunology, 34 (1), 157-165
Plasmodium berghei yoelii (P. yoelii), mice, passive transfer of immunity with serum and cells

modium yoelii
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Plasmodium yoelii in intact and T cell-deprived mice carrying CBA/N X chromosome, course of infection, specific fluorescent antibody levels, anti-erythrocyte autoantibody responses; effect of CBA/N X chromosome on secondary responses

modium yoelii, illus.
ndau, I.; et al., 1979, Ann. Parasitol., 54 (2), 145-161
Plasmodium yoelii, gametocytes, morphology, development, infectivity

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Landau, I.; et al., 1979, Compt. Rend. Acad. Sc., Paris, v. 288, s. D, Sc. Nat. (5), 521-522
Plasmodium yoelii, gametocytes, morphological characters as indication of age, infectivity, and periodicity

Plasmodium yoelii
Leke, R. G. F.; and Viens, P., 1978, IRCS J. Med. Sc., v. 6 (3), 99
Plasmodium chabaudi-infected mice (T-cell deprived, sham-thymectomized and normal), course of infection, comparison with P. yoelii; T-cell deprived mice with P. chabaudi (unlike P. yoelii) gave reactions similar to those of normal and sham-deprived mice indicating that this malaria model is not thymus dependent

Plasmodium yoelii
Lelchuk, R.; et al., 1979, Parasite Immunol., v. 1 (1), 61-78
Plasmodium yoelii- and P. berghei-infected mice and vaccinated mice challenged with homologous parasites, changes in phagocytic and adherent cell numbers, development and suppression of population of late-adhering macrophages

Plasmodium yoelii
Mitchell, G. F.; Handman, E.; and Howard, R. J., 1978, Austral. J. Exper. Biol. and Med. Sc., v. 56 (5), 553-559
Plasmodium berghei, Babesia rodhaini, mice, attempts to raise host-protective sera using variety of immunization manipulations (BCG injection, P. yoelii infection, others)

Plasmodium yoelii
Murphy, J. R.; and Lefford, M. J., 1979, Infect. and Immun., v. 23 (2), 384-391
Plasmodium yoelii, mice, defense mechanism against infection is mediated by humoral factors in absence of demonstrable cell-mediated immunity

Plasmodium yoelii
Playfair, J. H. L.; et al., 1979, Nature, London (5740), v. 282, 731-734
Plasmodium yoelii- or P. berghei-vaccinated mice, cell-mediated immunity in liver

Plasmodium yoelii
Playfair, J. H. L.; and De Souza, J. B., 1979, Parasite Immunol., v. 1 (3), 197-208
Plasmodium yoelii- or P. berghei-vaccinated mice, immunofluorescent antibody response with particular reference to antibody class and subclass, correlation with protection, passive transfer experiments, effect of macrophage stimulation and inhibition on antibody and on protection

Plasmodium yoelii
Roberts, D. W.; and Weidanz, W. P., 1978, Infect. and Immun., v. 20 (3), 728-731
Plasmodium yoelii in immunologically competent mice and mice with defined immunological deficiencies, results indicate that splenomegaly, enhanced phagocytosis, and anemia are thymus-dependent responses to malaria infection

Plasmodium yoelii
Roberts, D. W.; and Weidanz, W. P., 1979, Am. J. Trop. Med. and Hyg., v. 28 (1), 1-3
Plasmodium yoelii, B-cell deficient mice drug-rescued from otherwise lethal infections resisted subsequent challenge despite lack of detectable antibody

Plasmodium yoelii
Strickland, G. T.; DeSilva, S.; and Sayles, P. C., 1979, Tropenmed. u. Parasitol., v. 30 (1), 35-42
Plasmodium yoelii infection in mice and P. falciparum and P. vivax infection in humans, changes in lymphocyte populations during acute infections thought to be related to development of malarial immunity and immunodepression

Plasmodium yoelii
Wedderburn, N.; et al., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (6), 610-614
Plasmodium yoelii infection in mice followed by either one or repeated infections with P. berghei, induction of chronic malaria, course of parasitemia, fluctuations in reticulocyte levels, antibody titres, immunoglobulin deposition in tissues, changes in spleen and liver

Plasmodium yoelii
Wyler, D. J.; Oppenheim, J. J.; and Koontz, L. C., 1979, Infect. and Immun., v. 24 (1), 151-159
Plasmodium berghei, P. yoelii, mice, effects of infection on ability of adherent mononuclear cells to elaborate soluble mediators that regulate lymphocyte activation in vitro

Plasmodium yoelii subspecies
Peters, W.; et al., 1978, Ann. Trop. Med. and Parasitol., v. 72 (1), 23-36
Plasmodium berghei 'NS lines' form distinct taxon within P. yoelii complex and should be referred to as P. yoelii subspecies on basis of isoenzyme, DNA, and cross-immunity relationships with other rodent Plasmodium spp.

Plasmodium yoelii killicki
Carter, R., 1978, Parasitology, v. 76 (3), 241-267
Plasmodium berghei, P. yoelii, P. vinckei, P. chabaudi, and their subspecies, electrophoretic variation of enzymes glucose phosphate isomerase, 6-phosphogluconate dehydrogenase, lactate dehydrogenase, and glutamate dehydrogenase, detailed description of technique, genetic and taxonomic implications, key for identification of murine plasmodia by enzyme type

Plasmodium yoelii nigeriensis
Carter, R., 1978, Parasitology, v. 76 (3), 241-267
Plasmodium berghei, P. yoelii, P. vinckei, P. chabaudi, and their subspecies, electrophoretic variation of enzymes glucose phosphate isomerase, 6-phosphogluconate dehydrogenase, lactate dehydrogenase, and glutamate dehydrogenase, detailed description of technique, genetic and taxonomic implications, key for identification of murine plasmodia by enzyme type

Plasmodium (Vinckeia) yoelii nigeriensis
Chance, M. L.; et al., 1978, Ann. Trop. Med. an Parasitol., v. 72 (1), 13-22
Plasmodium (Vinckeia) spp., value of DNA studies employing buoyant density determinations and measurements of nucleotide sequence homology in systematics and identification

Plasmodium yoelii nigeriensis
Dei Cas, E.; et al., 1979, Ann. Parasitol., v. 54 (5), 567-570
Plasmodium, diagnosis, advantages of Errecart's modified technique for making thick blood films

Plasmodium yoelii nigeriensis, illus.
Dubremetz, J. F.; et al., 1979, Compt. Rend. Acad. Sc., Paris, v. 288, s. D, Sc. Nat. (6), 623-626
Plasmodium yoelii nigeriensis sporozoites, structure of pellicle, freeze-fracture study

Plasmodium yoelii nigeriensis
Landau, I.; et al., 1979, Ann. Parasitol., v. 54 (2), 145-161
Plasmodium yoelii, gametocytes, morphology, development, infectivity

Plasmodium yoelii nigeriensis
Mendis, K. N.; and Targett, G. A. T., 1979, Nature, London (5695), v. 277, 389-391
Plasmodium yoelii nigeriensis, mice, successful immunization against sexual stages using formalin-fixed gametes, vaccinated mice were also strongly protected against asexual erythrocytic stages

Plasmodium yoelii nigeriensis
Momen, H., 1979, Ann. Trop. Med. and Parasitol., v. 73 (2), 109-115
Plasmodium, Babesia, and Anthemosoma spp. in mouse erythrocytes, identification of enzymes of parasite origin using starch-gel electrophoresis

Plasmodium yoelii nigeriensis
Momen, H., 1979, Ann. Trop. Med. and Parasitol., v. 73 (2), 117-121
Plasmodium, Babesia and Anthemosoma spp., comparative study of glucose catabolism by infected mouse erythrocytes, glucose utilization and lactate production of parasites

Plasmodium yoelii nigeriensis
Peters, W.; et al., 1978, Ann. Trop. Med. and Parasitol., v. 72 (1), 23-36
Plasmodium berghei 'NS lines' form distinct taxon within P. yoelii complex and should be referred to as P. yoelii subspecies on basis of isoenzyme, DNA, and cross-immunity relationships with other rodent Plasmodium spp.

Plasmodium yoelii nigeriensis
Toye, P. J.; Sinden, R. E.; and Canning, E. U., 1977, Ztschr. Parasitenk., v. 53 (2), 133-141
Plasmodium yoelii nigeriensis mature gametocytes incubated with various metabolic inhibitor antibiotics, effects on microgametogenesis studied, results show de novo synthesis of axonemes and proteins essential to microgametogenesis, some results suggest possibility that de novo RNA synthesis is also required

Plasmodium yoelii yoelii
Carter, R., 1978, Parasitology, v. 76 (3), 241-267
Plasmodium berghei, P. yoelii, P. vinckei, P. chabaudi, and their subspecies, electrophoretic variation of enzymes glucose phosphate isomerase, 6-phosphogluconate dehydrogenase, lactate dehydrogenase, and glutamate dehydrogenase, detailed description of technique, genetic and taxonomic implications, key for identification of murine plasmodia by enzyme type

Plasmodium (Vinckeia) yoelii yoelii
Chance, M. L.; et al., 1978, Ann. Trop. Med. and Parasitol., v. 72 (1), 13-22
Plasmodium (Vinckeia) spp., value of DNA studies employing buoyant density determinations and measurements of nucleotide sequence homology in systematics and identification

Plasmodium yoelii yoelii
Landau, I.; et al., 1979, Ann. Parasitol., v. 54 (2), 145-161
Plasmodium yoelii, gametocytes, morphology, development, infectivity

Plasmodium yoelii yoelii
McBride, J. S.; and Micklem, H. S., 1977, Immunology, v. 33 (2), 253-259
Plasmodium yoelii yoelii-infected mice, depressed primary response to bovine serum albumin

Plasmodium yoelii yoelii
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Plasmodium berghei 'NS lines' form distinct taxon within P. yoelii complex and should be referred to as P. yoelii subspecies on basis of isoenzyme, DNA, and cross-immunity relationships with other rodent Plasmodium spp.

Plasmodium yoelii yoelii
Strambachova-McBride, J.; and Micklem, H. S., 1979, Immunology, v. 36 (3), 607-614
Plasmodium berghei, P. y. yoelii, mice, primary antibody response to immunogenic alum-adsorbed bovine serum albumin (BSA) was depressed and induction of immunological memory by soluble polymerized BSA was suppressed in both infections but neither interfered with induction of low-zone tolerance by monomeric BSA

asmodium yoelii yoelii
Strambachova-McBride, J.; and Micklem, H. S., 1979, Parasite Immunol., v. 1 (2), 141-157
Plasmodium berghei- and P. yoelii yoelii-infected mice, immunosuppression, secondary response to bovine serum albumin

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. jerichaense (Kirk, 1949)
Gardener, P. J., 1977, Trop. Dis. Bull., v. 74 (12), 1069-1088
as syn. of Leishmania tropica (Wright, 1903) Luehe, 1906

lata subgen. n.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31
subgen. of Cepedea
type sp.: Cepedea (Plata) plata Metcalf, 1940

Plata
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
subgen. of Cepedea, key

Pleistophora. See also Plistophora.

Pleistophora sp.
Kaya, H. K., 1979, Proc. Hawaiian Entom. Soc. (1976), v. 13 (1), 91-94
Pleistophora sp., Vairimorpha necatrix, retention of infectivity after passage through gut of Zelus exsanguis
Dasychira basiflava
Estigmene acrea (exper.)

Pleistophora sp., illus.
Kelly, J. F., 1979, J. Invert. Path., v. 33 (3), 331-339
Thelohania duorara, Agmasoma penaei, and Pleistophora sp. in Penaeus duorarum, pathology, tissue specificity: southern Biscayne Bay

Pleistophora [sp.], tentatively, illus.
Paperna, I.; Sabnai, I.; and Castel, M., 1978, Ann. Parasitol., v. 53 (2), 123-130
microsporidian infection (tentatively Pleistophora) reported in Liza ramada in fibroblasts of metacercarial capsule of Heterophyes heterophyes, infection of metacercarial cyst resulted in hypertrophy of cyst wall and degeneration and eventual death of encapsulated metacercaria: Bardawil Lagoon, Mediterranean coast of Sinai Peninsula

Pleistophora blatellae n. sp.
Narasimhamurti, C. C.; and Kalavati, C., 1976, Proc. 1. Internat. Colloquium Invert. Path. and 9. Ann. Meet. Soc. Invert. Path. (Kingston, Canada, Aug. 29-Sept. 3), 377
Blatella germanica (epithelium of Malpighian tubules, lumen of tubules): Andhra University Campus at Waltair (India)

Pleistophora chironomi (Debaisieux, 1928) comb. nov., illus.
Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135
Syns.: Thelohania chironomi Debaisieux, 1928; Plistophora thienemanni (Debaisieux, 1928)

Pleistophora jiroveci Weiser, 1942, illus.
Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135

Pleistophora ovariae
Nagel, M. L.; and Summerfelt, R. C., 1977, Proc. Oklahoma Acad. Sc., v. 57, 61-63
Pleistophora ovariae, absence of infection in Carassius auratus despite prolonged natural exposure and artificial exposure by intraperitoneal and intramuscular injection of spores: Stoutland, Missouri

Pleistophora schubergi Zwolfer, 1927
Simchuk, P. A., 1978, Biol. Nauki, Min. Vyssh. i. Sredn. Spets. Obraz. SSR (178) (10), 51-54
Laspeyresia pomonella
L. funebrana
Carpocapsa pyrivora
(all exper.)

Pleistophora schubergi
Wilson, G. G., 1978, Bi-Month. Research Notes Canad. Forestry Serv., v. 34 (3), 16
Nosema fumiferanae, Pleistophora schubergi, incidence of infection in spruce budworm (Choristoneura fumiferana) 1 and 2 years after application of microsporidian spores to trees: Ontario

Pleistophora schubergi
Wilson, G. G., 1978, Canad. J. Zool., v. 56 (4), 578-580
feeding of Pleistophora schubergi to Choristoneura fumiferana naturally infected with Nosema fumiferanae results in greater adverse effects on budworms than either parasite alone, may play useful role in biological control

Pleistophora schubergi
Wilson, G. G.; and Kaupp, W. J., 1976, Insect Path. Research Inst., Inform. Rep. (IP-X-15), 14 pp.
Nosema fumiferanae, Pleistophora schubergi, control of Choristoneura fumiferana by spraying spruce and balsam fir trees with spores: Ontario

Pleistophora simulii (Lutz & Splendore), illus.
Loubes, C., 1979, J. Protozool., v. 26 (2), 200-208
Microsporida, synaptonemal complexes demonstrated in 6 genera but not in Nosema, implications for life cycles

Pleistophora simulii (Lutz et Splendore, 1904), illus.
Maurand, J.; and Loubes, C., 1978, Ztschr. Parasitenk., v. 56 (2), 131-146
microsporidians from simuliid larvae, ultrastructural studies
simuliid larvae: region languedocienne (Sud de la France)

Pleistophora spraguei (Debaisieux, 1931) nom. nov., illus.
Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135
Syn.: Plistophora chironomi Debaisieux, 1931

Pleistophora sulci Rasin 1936, 1949 (Sprague and Vavra, 1977) (Cocconema sulci)
Raikova, E. V.; Suppes, V. C.; and Hoffman, G. L., 1979, J. Parasitol., v. 65 (5), 804-810
Polyodon spathula (oocytes): Osage River, Missouri

Plistophora. See also Pleistophora.

Plistophora [sp.] or Thelohania [sp.], illus.
Herman, R. L.; and Putz, R. E., 1970, J. Wildlife Dis., v. 6 (3), 173
Ictalurus punctatus (ventricle of heart, intestinal sub-mucosa): Maryland

Plistophora sp. ?
Jackson, P. B. N., 1978, J. South African Vet. Ass., v. 49 (1), 57-59
parasites of Anguilla mossambica, importance of disease control in aquaculture
Anguilla mossambica (muscles, serosa): South Africa

Plistophora sp.
Miln, A. J., 1978, N. Zealand Entom., v. 6 (4), 392-399
Costelytra zealandica: New Zealand

Plistophora chironomi Debaisieux, 1931
Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135
as syn. of Pleistophora spraguei (Debaisieux, 1931) nom. nov.

Plistophora elegans
Iskov, M. P., 1979, Gidrobiol. Zhurnal, v. 15 (4), 68-72
parasites causing intensive epizootic disease of food fishes: Kremenchug reservoir

Plistophora schubergi Zwoelfer
Metspalu, L., 1976, Eesti NSV Teaduste Akad. Toimetised, Biol., v. 25 (1), 13-19
Plistophora schubergi, effect of microsporidiosis on development of diapause in noctuids, biological control
Barathra brassicae
Mamestra dissimilis
M. persicariae
M. pisi
(all exper.)

Plistophora schubergi Zwoelfer
Nilova, G. N.; and Strel'nikova, L. V., 1974, Parazitologiia, Leningrad, v. 8 (5), 463-468
Plistophora schubergi, Nosema agrotidis, effect of ultraviolet radiation on viability of spores

Plistophora schubergi aporiae
Issi, I. V., 1971, Parazitologiia, Leningrad, v. 5 (4), 297-301
Plistophora schubergi, subspecies are morphologically identical but characterized by a certain degree of host specificity and possibly by divergence of biochemical and physiological characters, implications for possible methods of speciation among the Microsporidia

Plistophora schubergi balbiani [?n. rank]
Issi, I. V., 1971, Parazitologiia, Leningrad, v. 5 (4), 297-301
Plistophora schubergi, subspecies are morphologically identical but characterized by a certain degree of host specificity and possibly by divergence of biochemical and physiological characters, implications for possible methods of speciation among the Microsporidia

Plistophora schubergi hyphantriae
Issi, I. V., 1971, Parazitologiia, Leningrad, v. 5 (4), 297-301
Plistophora schubergi, subspecies are morphologically identical but characterized by a certain degree of host specificity and possibly by divergence of biochemical and physiological characters, implications for possible methods of speciation among the Microsporidia

Plistophora schubergi noctuidae
Issi, I. V., 1971, Parazitologiia, Leningrad, v. 5 (4), 297-301
Plistophora schubergi, subspecies are morphologically identical but characterized by a certain degree of host specificity and possibly by divergence of biochemical and physiological characters, implications for possible methods of speciation among the Microsporidia

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Issi, I. V., 1971, Parazitologiia, Leningrad, v. 5 (4), 297-301
Plistophora schubergi, subspecies are morphologically identical but characterized by a certain degree of host specificity and possibly by divergence of biochemical and physiological characters, implications for possible methods of speciation among the Microsporidia

istophora schubergi schubergi
Issi, I. V., 1971, Parazitologiia, Leningrad, v. 5 (4), 297-301
Plistophora schubergi, subspecies are morphologically identical but characterized by a certain degree of host specificity and possibly by divergence of biochemical and physiological characters, implications for possible methods of speciation among the Microsporidia

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Levchenko, N. G.; Tokarev, G. G.; and Grinin, V. S., 1974, Parazitologiia, Leningrad, v. 8 (6), 543-547
Plistophora tabani sp. n., pathogenicity Tabanus sp. (fat body, hemolymph): USSR, Turgen river basin, Zailiiskii Alatau, Kazakhstan

istophora thienemanni (Debaisieux, 1928)
oste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135
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urinucleata subgen. n.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 23-24
subgen. of Protoopalina
type sp.: Protoopalina (Plurinucleata) axonucleata Metcalf, 1923

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Amaro, A., 1966, Atas Soc. Biol. Rio de aneiro, v. 10 (3), 49-52
subgen. of Protoopalina, key

umocystis
hatterjee, S. N.; Gottleib, L.; and Berne, . V., 1978, Surg., Gynec. and Obst., v. 147 (4), 583-587
fulminant pulmonary infections in renal transplant recipients, etiology including Pneumocystis, postoperative complications

eumocystis carinii
Amato Neto, V., 1973, Rev. Paul. Med., v. 81 (2), 93-104
human protozoan infections, drugs in current use, dosage recommendations, review

eumocystis carinii
Andersen, H. A.; Miller, W. E.; and Bernatz, . E., 1973, Surg. Clin. North Am., v. 53 (4), 85-793
Pneumocystis carinii and other lung lesions, human, diagnosis by lung biopsy (transbronchoscopic, percutaneous, and open)

eumocystis carinii
Ball, F.; et al., 1975, Roentgen-Blaetter, v. 28 (10), 443-454
Pneumocystis carinii interstitial pneumonia in children treated with immunosuppressive and cytostatic drugs for malignant systemic disease, radiologic diagnosis

Pneumocystis carinii
Barkin, R. M.; et al., 1975, Rocky Mountain Med. J., v. 72 (8), 340-344
medical screening of recently adopted Vietnamese orphans, scabies, Entamoeba histolytica, Giardia lamblia, and Pneumocystis carinii among medical problems encountered: Denver area, Colorado

Pneumocystis carinii, illus.
Bartlett, M. S.; Verbanac, P. A.; and Smith, J. W., 1979, J. Clin. Microbiol., v. 10 (6), 796-799
Pneumocystis carinii, successful culture with WI-38 human embryonic lung fibroblasts

Pneumocystis carinii
Benatar, S. R., 1977, South African Med. J., v. 52 (25), 1019-1024
pulmonary complications in immunocompromised patients, includes information on Pneumocystis carinii

Pneumocystis carinii
Blank, N.; and Castellino, R. A., 1975, Seminars Roentgenol., v. 10 (1), 63-72
pulmonary infections in patients with altered immunity, radiologic diagnosis and follow-up, includes information on Pneumocystis carinii and Toxoplasma gondii

Pneumocystis carinii, illus.
Cash, R. L.; Cutts, W. B.; and Quinn, R. F., 1976, Rocky Mountain Med. J., v. 73 (1), 31-32
Pneumocystis carinii pneumonia in immunocompromised man who was receiving continuously assisted ventilation via tracheostomy tube, diagnosis using tracheostomy tube adapter for fiberbronchoscopy

Pneumocystis carinii, illus.
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perro
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Protoopalina (Caudiculata) brevis Boisson, 1959, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 89-94

Protoopalina (Longicorpus) canevi Sukhanova, 1960, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Caudicula) caudata (Zeller, 1877) Metcalf, 1923 (tod of subgen.), illus.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 23-24

Protoopalina caudata (Zeller, 1877)
Frandsen, F., 1974, Acta Parasitol. Polon., v. 22 (1-11), 49-66
Rana esculenta
Bombina bombina
(rectum of all): all from Denmark

Protoopalina (Caudicula) caudata attenuata Metcalf, 1923
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Caudicula) caudata filiformis Liu, 1958, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Caudiculata) curcurbitacea Boisson, 1959, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 89-94

Protoopalina (Caudiculata) dakariensis Boisson, 1959, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 89-94

Protoopalina (Caudiculata) daloalensis Tuzet & Zuber-Vogeli, 1954, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 89-94

Protoopalina delphini n. sp. (Fig. 1 of Delphin et al., 1972)
Earl, P. R., 1979, Tr. Am. Micr. Soc., v. 98 (4), 549-557
Microhyla ornata: Burma

Protoopalina (Longicorpus) dharwarensis Uttangi, 1951, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Caudiculata) diplocarya Metcalf, 1923 (tod of subgen.), illus.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 23-24

Protoopalina (Caudiculata) drachi Tuzet & Knoepffler, 1968, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Caudiculata) duboscqui Lavier, 1936, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 89-94

Protoopalina (Filiformis) filiformis Metcalf, 1923 (tod of subgen.), illus.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 23-24

Protoopalina (Longicorpus) filiformis Metcalf, 1923 (tod of subgen.)
Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (5-6), 237-243

Protoopalina (Plurinucleata) formosae Metcalf, 1923
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Caudiculata) globulata Boisson, 1959, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 89-94

Protoopalina (Plurinucleata) hexasoma Yagiu, 1936, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Caudicula) indica Uttangi, 1951, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Caudiculata) intestinalis (Stein, 1856) Metcalf, 1923
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina intestinalis (Stein, 1856), illus.
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
description
Bombina bombina
B. variegata
Pelobates fuscus
Rana esculenta
R. temporaria
(kloaka of all): all from CSSR

Protoopalina (Acuminata) karnatakensis Uttangi, 1951, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Caudiculata) lamottei Tuzet & Knoepffler, 1968, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Caudiculata) larvata Uttangi, 1961, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 89-94

Protoopalina (Caudicula) lemuriae Uttangi, 1961, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Longicorpus) limnocharis Nie, 1932
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Caudiculata) macronucleata Tuzet & Knoepffler, 1968, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Longicorpus) malabarica Uttangi, 1961, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Longicorpus) metcalfi Uttangi, 1961, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Caudicula) mitotica (Metcalf, 1912) Metcalf, 1923
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Orientalis) montana Metcalf, 1923 (tod of subgen.), illus.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 23-24

Protoopalina (Caudicula) orientalis Metcalf, 1923
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Ovalis) ornata Uttangi, 1961, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Ovalis) ovalis Fantham, 1929
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Ovalis) ovoidea Metcalf, 1923 (tod of subgen.), illus.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 23-24

Protoopalina (Caudiculata) peguyi Tuzet & Knoepffler, 1968, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Caudiculata) petiti Tuzet & Knoepffler, 1968, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Acuminata) regularis Metcalf, 1923 (tod of subgen.), illus.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 23-24

Protoopalina (Caudiculata) rhopaloides Wach, 1953, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Protoopalina) scaphiopodos Metcalf, 1923 (tod of subgen.), illus.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 23-24

Protoopalina (Caudiculata) senegalensis Boisson, 1959, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 89-94

Protoopalina (Caudiculata) spinosa Fernandez-Galiano, 1965, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 89-94

Protoopalina (Caudiculata) stevensoni saigonensis Boisson, 1957, illus.
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 89-94

Protoopalina symphysodonis nov. spec., illus.
Foissner, W.; Schubert, G.; and Wilbert, N., 1979, Zool. Anz., Jena, v. 202 (1-2), 71-85
Protoopalina symphysodonis nov. spec., morphology, infraciliature, and silverline system
Symphysodon aequifasciata (Rectum)

Protoopalina (Longicorpus) tenuis (Raff, 1912) Metcalf, 1923
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalina (Caudicula) yunnanensis Metcalf, 1940
Amaro, A., 1970, Atas Soc. Biol. Rio de Janeiro, v. 13 (3-4), 121-127

Protoopalininae Metcalf, 1920
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
Opalinidae
key to genera, key

Protoopalininae Metcalf, 1920
Earl, P. R., 1979, Tr. Am. Micr. Soc., v. 98 (4), 549-557
Opalinidae
includes: Protoopalina; Bezzenbergia; Cepedea

Protozoa
Polianskii, Iu. I., 1977, Protozoology, v. 3, 17-23
aspects of species concept in asexually reproducing protozoa

Protozoa
Whittaker, R. H.; and Margulis, L., 1978, Bio Systems, v. 10 (1-2), 3-18
classification of living world into kingdoms (Monera, Protista, Fungi, Animalia, Plantae) with emphasis on protist classification

Protozoa [sp.], illus.
Carpenter, J. W.; et al., 1979, J. Am. Vet. Med. Ass., v. 175 (9), 948-951
Protozoa [sp.] in Grus canadensis, disseminated granulomas, histopathology, electron microscopy: Patuxent Wildlife Research Center

[Protozoa sp.], illus.
Durfort, M., 1978, Bol. R. Soc. Espan. Hist. Nat., Secc. Biol., v. 75, 1977, 71-86
Mytilicola intestinalis (ovulos, ovocitos)

Protozoans resembling Hexamita sp.
Kocan, A. A.; Hannon, L.; and Eve, J. H., 1979, Proc. Oklahoma Acad. Sc., v. 59, 20-22
Colinus virginianus (intestine): Oklahoma

Protozoa [sp.] cysts
Meyers, T. R., 1978, Proc. Helminth. Soc. Washington, v. 45 (1), 120-128
Pomatomus saltatrix (pericardium): Raritan Bay, New Jersey

Psorospermium sp., illus.
Voronin, V. N., 1975, Izvest. Gosudarstv. Nauchno-Issled. Inst. Ozer. i Rech. Ryb. Khoziaistva, Leningrad, v. 93, 125-126
Gammarus lacustris: lakes of Baikal District

Ptyssostoma Hentschel, 1924
Iankovskii, A. V., 1973, Parazitologiia, Leningrad, v. 7 (3), 214-219
Hymenostomatida, Tetrahymenina, Entodiscidae fam. n.

Pulchra subgen. n.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31
subgen. of Cepedea
type sp.: Cepedea (Pulchra) pulchra Metcalf, 1923

Pulchra
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
subgen. of Cepedea, key

Pyxinia firmus (Leger, 1892), illus.
Ormieres, R., 1977, Ztschr. Parasitenk., v. 53 (1), 13-22
Pyxinia firmus, ultrastructure of epimerite, possible functions in fixation and nutrition
Dermestes frischi (intestin)

Quadruspinospora atractomorpha [lapsus p. 45 for Q. atractomorphii n. sp.]
Haldar, D. P.; and Chakraborty, N., 1978, Indian J. Zool., v. 6 (1), 43-47

Quadruspinospora atractomorphii n. sp., illus.
Haldar, D. P.; and Chakraborty, N., 1978, Indian J. Zool., v. 6 (1), 43-47
[lapsus p. 43 as Q. atractomorpii; lapsus p. 45 as Q. atractomorpha]
Atractomorpha crenulata (hepatic caeca, midgut): Kaiyani, West Bengal

Quadruspinospora atractomorpii [lapsus p. 43 for Q. atractomorphii n. sp.]
Haldar, D. P.; and Chakraborty, N., 1978, Indian J. Zool., v. 6 (1), 43-47

Radiophrya lumbrici [i.e. ? lumbriculi] Cheissin to Anthonyella gen. nov. [comb. not made]
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519

Ramicephalus cecheni n. sp., illus.
Massot, N.; and Ormieres, R., 1979, Ann. Parasitol., v. 54 (3), 267-275
Cechenus pyrenaeus (intestin): Hautes-Pyrenees (Lac d'Oncet; Soum de Grum)

Ramicephalus hirsutus
Massot, N.; and Ormieres, R., 1979, Ann. Parasitol., v. 54 (3), 267-275
Chrysocarabus auronitens festivus
Carabus auratus

Ramicephalus nebriae n. sp., illus.
Massot, N.; and Ormieres, R., 1979, Ann. Parasitol., v. 54 (3), 267-275
Nebria lafresnayei: Lac d'Oncet (Hautes-Pyrenees)

Ramicephalus rostratus n. sp., illus.
Massot, N.; and Ormieres, R., 1979, Ann. Parasitol., v. 54 (3), 267-275
Nebria lafresnayei: Hautes-Pyrenees (Lac d'Oncet; Soum de Grum; Cirque de Troumouse)

Rasajeyna nannyla
Beesley, J. E., 1977, J. Invert. Path., v. 30 (2), 249-254
Rasajeyna nannyla in Tipula paludosa and T. vittata, incidence throughout 2-year sampling period, incidence as a function of temperature: 2 different sites (one damp and one dry) in Northumberland, England

Rasajeyna nannyla
Beesley, J. E., 1978, J. Invert. Path., v. 31 (2), 255-259
Rasajeyna nannyla, seasonal abundance of three life cycle stages in Tipula paludosa and T. vittata: Northumberland, United Kingdom

Rectiperistomatus subgen. n.
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139
subgen. of Nyctotheroides
tod of subgen.: N. (R.) coralli (Carini, 1933) Amaro & Sena, 1967

Recurviperistomatus subgen. n.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 119-120
subgen. of Nyctotherus
tod of subgen.: N. (R.) uianchoi Kidder, 1937

Retortomonas intestinalis, illus.
Harvey, J., 1973, Saguenay Med., v. 20 (3), 146-156
parasitic flagellates, life cycle, infection in man, clinical signs, therapy, brief review

Retortamonas viperae n. sp., illus.
Krishnamurthy, R.; and Madre, V. E., 1976, Marathwada Univ. J. Sc. (Nat. Sc.), v. 15 (8), 129-131
Vipera russeli (rectum): Aurangabad, Maharashtra, India

Retortomonas. See Retortamonas.

Rhabdospora thelohani Laguesse, 1895, illus.
Mayberry, L. F.; et al., 1979, J. Protozool., v. 26 (2), 168-178
Rhabdospora thelohani, chronological listing of published reports, new host and geographic records, ultrastructure, evidence in support of inclusion among Apicomplexa
Alburnoides bipunctatus ohridanus (gall bladder, intestine, kidney, stomach, swim bladder): Yugoslavia
Anguilla anguilla (gall bladder, intestine, spleen): Yugoslavia
Rutilus rubilio (kidney): Yugoslavia
Borostomias antarcticus (stomach): N. Atlantic; Antarctic
Carassius auratus (intestine): Yugoslavia
Cetonurus globiceps (urinary bladder): Indonesia
Chaenocephalus aceratus (urinary bladder): Antarctic
Chalinura leptolepis (urinary bladder): Farallon Is.
Chondrostoma nasus ohridanus (liver, swim bladder, spleen): Yugoslavia
Leuciscus cephalus albus (gall bladder, intestine, kidney, liver, swim bladder, spleen): Yugoslavia
Nezumia stelgiolepis (gall bladder): Vancouver Is.
Pachychlon pictum (intestine, liver, swim bladder): Yugoslavia
Anoplopoma fimbria (urinary bladder): California
Ventrifossa atherodon (gall bladder): Hawaiian Is.

Rhabdostyla scyphidiformis Vavra, 1961, illus.
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
description
Bombina variegata
Pelobates fuscus
Bufo viridis
Hyla arborea
Rana esculenta
R. temporaria
all from CSSR

Rhizionella sp.
Massot, N.; and Ormieres, R., 1979, Ann. Parasitol., v. 54 (3), 267-275
Pelor obesus

Rhizionella tenuis Baudoin, 1971
Massot, N.; and Ormieres, R., 1979, Ann. Parasitol., v. 54 (3), 267-275
Cechenus pyrenaeus

Rhizocaryum Caullery et Mesnil
Delphy, J., [1937], Bull. Mus. National Hist. Nat., Paris, 2. s., v. 8 (6), 1936, 516-519
key

Rhopalonia stella Leger, 1894
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
as syn. of Chilogregarina stella (Leger, 1894) comb. n.

Rhytidocystidae n. fam.
Levine, N. D., 1979, J. Protozool., v. 26 (2), 167-168
Agamococcidiorida ord. n.
includes: Rhytidocystis

Rhytidocystis Henneguy, 1907
Levine, N. D., 1979, J. Protozool., v. 26 (2), 167-168
Rhytidocystidae n. fam.
Syn.: Dehornia Porchet-Hennere, 1972

Rhytidocystis henneguyi de Beauchamp, 1912
  Levine, N. D., 1979, J. Protozool., v. 26 (2), 167-168
    diagnosis

Rhytidocystis opheliae Henneguy, 1907
  Levine, N. D., 1979, J. Protozool., v. 26 (2), 167-168
    diagnosis

Rhytidocystis sthenelais (Porchet-Hennere, 1972) comb. n.
  Levine, N. D., 1979, J. Protozool., v. 26 (2), 167-168
    diagnosis
    Syn.: Dehornia sthenelais Porchet-Hennere, 1972

Saccamoeba sp., illus.
Pernin, P.; and Riany, A., 1978, Ann. Parasitol., v. 53 (4), 333-344
survey of swimming pools for presence of free-living amebae, potential danger for swimmers: Lyon

Sarcocyst III
Lai, P. F., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (3), 417-419
Rattus annandeli
R. r. diardii
all from Malacca, Malaysia

Sarcocyst IV
Lai, P. F., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (3), 417-419
Rattus annandeli
R. jalorensis
R. r. diardii
all from Malacca, Malaysia

Sarcocyst V
Lai, P. F., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (3), 417-419
Rattus exulans: Malacca, Malaysia

Sarcocystis
Aryeetey, M. E.; and Piekarski, G., 1978, Ztschr. Parasitenk., v. 56 (3), 211-218
Sarcocystis antibodies in young humans and rats, gradually decreased after birth, probably passed to newborn through placenta

Sarcocystis
Beier, T. V.; Shibalova, T. A.; and Kostenko, L. A., 1978, [Cytology of coccidia], 185 pp., illus.

[Sarcocystis] sarkotsistami
Bogush, A. A., 1973, et. Nauka--Proizvod., Trudy, Minsk, v. 11, 175-178
[Sarcocystis], pork, physical and chemical changes (tenderness, color, odor, consistency, quantity of juice in cooking, peroxidase activity, peptone and polypeptides, amino-ammonium nitrogen, volatile fatty acids); infected pork after 48 hours at room temperature unfit for eating

[Sarcocystis] sarkotsistami
Bogush, A. A., 1974, Vet. Nauka--Proizvod., Trudy, Minsk, v. 12, 133-136
[Sarcocystis], swine, levels of free amino acids in infested meat

[Sarcocystis] sarkotsist
Bogush, A. A., 1977, Vet. Nauka--Proizvod., Trudy, Minsk, v. 15, 121-126
[Sarcocystis], toxic substances in pork, mutton and beef, tests with rabbits, cooking methods to render the substances harmless

[Sarcocystis] sarkotsistami
Bogush, A. A.; and Pyshko, I. I., 1976, Vet. Nauka--Proizvod., Trudy, Minsk, v. 14, 173-174
[Sarcocystis]-infected swine, glycogen content in muscles and liver

[Sarcocystis] sarkotsistami
Bogush, A. A.; and Urbanovich, N. A., 1977, Vet. Nauka--Proizvod., Trudy, Minsk, v. 15, 117-121
[Sarcocystis], increased prevalence of bacteria in meat and organs of infected swine and cattle, public health implications

Sarcocystis
Box, E. D.; and McGuinness, T. B., 1978, J. Parasitol., v. 64 (1), 161-162
Sarcocystis recovered in beef from retail outlets by using a digestion technique

Sarcocystis
Collins, G. H.; and Charleston, W. A. G., 1979, N. Zealand Vet. J., v. 27 (4), 80-84
feral Felis catus, survey on distribution, feeding, and breeding, probably important source of sporozoan infections for domesticated animals: New Zealand

Sarcocystis Lankester, 1882
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
definition; proposed classification and nomenclature
proposed as new type species: S. muris (Blanchard, 1885) Labbe, 1899

Sarcocystis
Heydorn, A. O., 1979, Berl. u. Munchen. Tierarztl. Wchnschr., v. 92 (11), 214-220
coccidia of cats, life cycle, epidemiology, review

Sarcocystis
Jørgensen, R. J.; and Otwelo, J. A., 1978, Dansk Vet.-Tidsskr., v. 61 (9), 417-423
Sarcocystis, cattle, 100% prevalence in small cadaver survey, suggested control measures for human consumption of raw beef

Sarcocystis
de Kruijf, J. M.; and Bibo, T. M., 1976, Tijdschr. Diergeneesk., v. 101 (19), 1093-1095
Sarcocystis, prevalence in sheep, predilection for masseter muscle: Netherlands

Sarcocystis, illus.
Markus, M. B., 1978, Advances Vet. Sc. and Comp. Med., v. 22, 159-193
Sarcocystis and sarcocystosis in domestic animals and man, extensive review (life cycle; host specificity; pathogenicity and pathology; immunology and serology)

Sarcocystis
Markus, M. B., 1979, J. Parasitol., v. 65 (5), 699
Sarcocystis, technique for obtaining intact tissue cysts from bovine heart muscle, not satisfactory for separation of sarcocysts from skeletal muscle

Sarcocystis
Markus, M. B., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (3), 346-347 [Letter]
Sarcocystis, "serological tests for human sarcocystis have little or no diagnostic value"

Sarcocystis
Mehlhorn, H.; et al., 1979, Ann. Biol., 4. s., v. 18 (3-4), 97-120
Sarcocystis, Theileria, life cycle, systematic position, pathology, review

Sarcocystis, illus.
Mehlhorn, H.; and Heydorn, A. O., 1978, Advances Parasitol., v. 16, 43-91
Sarcosporidia, life cycle and fine structure, review

Sarcocystis
  Munday, B. L., 1978, N. Zealand Vet. J., v. 26 (6), 166 [Letter]
    Sarcocystis, sarcocysts derived from cats take between 8 and 14 months to reach infectivity in sheep

Sarcocystis
  Prasad, B. N.; and Jorgensen, R. J., 1978, v. 17 (1-2), 197-202
    Sarcocystis, cattle, isolation and identification using various diagnostic methods, differentiation from Toxoplasma with dye-test: Denmark

Sarcocystis
  Rice, D. A.; and Calderon M., J. E., 1979, Trop. Animal Health and Prod., v. 11 (2), 85-86
    Sarcocystis, prevalence in slaughtered cattle (hearts): El Salvador

Sarcocystis
  Rommel, M., 1978, Ztschr. Parasitenk., v. 57 (3), 269-283
    cyst-forming coccidia, life cycle, taxonomy, comparative review

Sarcocystis
  Stalheim, O. H.; et al., 1976, Proc. 19. Ann. Meet., Am. Ass. Vet. Lab. Diagn., 317-328
    Sarcocystis from dogs, experimental infections in pregnant Holstein-Friesian and Jersey cows, clinical signs, serology, abortions and fetal deaths

Sarcocystis
  Theis, J. H.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (4), 837-839
    diaphragm samples from 297 humans at autopsy over 20-month period revealed 3 cases of Trichinella spiralis and no cases of Sarcocystis: California

Sarcocystis
  Thomas, V.; and Dissanaike, A. S., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (3), 303-306
    Sarcocystis, human, indirect fluorescent antibody test using Sarcocystis fusiformis as antigen, antibody prevalence varies among 4 ethnic groups (Orang Asli, Malays, Indians, Chinese), some sera also reacted positively to Toxoplasma: West Malaysia

Sarcocystis, illus.
  Viles, J. M.; and Powell, E. C., 1977, Proc. 14. Electron Micr. Colloq., 43-59
    Sarcocystis in Mus musculus (exper.) (abdominal musculature), pathology

Sarcocystis
  Zasukhin, D. N.; Vel'iaminov, K. S.; and D'iakonov, L. P., 1979, Veterinariia, Moskva (1), 49-55
    Sarcocystis spp., domestic animals, morphology, life cycles, pathology, prophylaxis, brief review

Sarcocystis sp.
  de Araujo, F. G., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (2), 105-111
    Toxoplasma gondii, survey of serum samples from blood donors, Sabin-Feldman dye test and indirect immunofluorescence tests compared; Sarcocystis sp. in local animals may have some bearing on test results: Belo Horizonte, Brazil

Sarcocystis [sp.]
  Ashford, R. W., 1978, Ann. Trop. Med. and Parasitol., v. 72 (1), 37-43
    Microtus agrestis
    Apodemus sylvaticus
    Mustela nivalis (feces)

Sarcocystis [sp.]
  Asteriades, G.; and Charalambides, S., 1978, Ellen. Kteniatrike, v. 21 (1), 25-28
    Sarcocystis [sp.], ram, stallion, localization in bulbocavernosus muscle, diagnosis: Greece

Sarcocystis [sp.]
  Ayala, S. C.; et al., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (6), 411-416
    Zenaida auriculata caucae: Cauca River valley, Colombia

Sarcocystis [spp.], illus.
  Beaver, P. C.; Gadgil, R. K.; and Morera, P., 1979, Am. J. Trop. Med. and Hyg., v. 28 (5), 819-844
    Sarcocystis in man, 5 new case reports, review of previous reports, classification into 7 morphological types
    humans (muscles, heart): Singapore; Bombay; Uganda; Costa Rica

Sarcocystis spp.
  Benko, L.; and Gaspar, P., 1977, Bull. Animal Health and Prod. Africa, v. 25 (3), 325
    Sarcocystis spp., cattle, incidence in relation to rainy vs. dry seasons: abattoir, Zambia

Sarcocystis spec., illus.
  Boch, J.; et al., 1979, Berl. u. Munchen. Tierarztl. Wchnschr., v. 92 (7), 137-141
    Sarcocystis spp., Toxoplasma, sheep, parasitological and serological studies: slaughterhouses in Bavaria

Sarcocystis spp.
  Boch, J.; Boehm, A.; and Weiland, G., 1979, Berl. u. Munchen. Tierarztl. Wchnschr., v. 92 (12), 240-243
    coccidia, dogs (feces), coprological and serological survey: South Germany

Sarcocystis spp.
  Bogush, A. A., 1976, [Parasites in muscles of swine and measures for their prophylaxis (trichinosis and sarcocystosis)], 94 pp., illus.
    trichinosis, sarcocystosis, swine muscles, general aspects, emphasis on diagnosis and prophylaxis, review: Belorussia

[Sarcocystis sp.] sarkotsistami
  Bogush, A. A.; and Pyshko, I. I., 1974, Vet. Nauka--Proizvod., Trudy, Minsk, v. 12, 130-132
    [Equus caballus]
    [Bos taurus]
    [Sus scrofa]
    [pig, wild]
    [Alces alces]
    [Felis catus]
    (muscles of all): all from Belorussia

Sarcocystis [sp.], illus.
  Box, E. D.; and Duszynski, D. W., 1978, J. Parasitol., v. 64 (4), 682-688
    Sarcocystis [sp.], experimental transmission from Molothrus ater and Cassidix mexicanus to Passer domesticus and Serinus canarius by sporocysts from Didelphis virginiana

Sarcocystis sp., resembling S. ovicanis
Chhabra, M. B.; and Mahajan, R. C., 1978, Vet. Rec., v. 103 (25), 562-563
Sarcocystis sp. from goats slaughtered at meat market, successful transmission to pups (feces) raised hygienically on meat-free diet, implications for continued transmission: India

Sarcocystis spp.
Collins, G. H.; and Charleston, W. A. G., 1979, N. Zealand Vet. J., v. 27 (7), 134-135
Sarcocystis spp., wild and feral animals, occurrence, prevalence; transmission from rats and rabbits to cats, and from goats to dogs
Cervus elaphus
Capra hircus
Sus scrofa
Oryctolagus cuniculi
Mus musculus
Rattus norvegicus
F[elis] catus (exper.)
C[anis] familiaris (exper.)
all from New Zealand

Sarcocystis spp.
Collins, G. H.; and Crawford, S. J. S., 1978, N. Zealand Vet. J., v. 26 (11), 288 [Letter]
Capra hircus (diaphragmatic muscle): Hawke's Bay area, New Zealand
dogs (faeces) (exper.)

Sarcocystis-like, illus.
Cornaglia, E., 1977, Ann. Fac. Med. Vet. Torino, v. 24, 288-290
Sarcocystis-like structure, bovine, mild meningoencephalitis, light electron microscopy

Sarcocystis sp.
Diaz, L.; Rioseco, H.; and Cubillos, V., 1977, Bol. Chileno Parasitol., v. 32 (3-4), 86-89
survey of endoparasites of wild cervids
Ovis musimon (corazon)
Pudu pudu (musculatura esqueletica y cardiaca)
all from sur de Chile

Sarcocystis sp.
Douglass, E. M.; and Hansen, B., 1979, Vet. Med. and Small Animal Clin., v. 74 (10), 1534-1535
Sarcocystis sp. in Thrautis bonariensi (striated musculature), fatal infection: Memphis Zoo and Aquarium

Sarcocystis spp.
Dubey, J. P., 1979, Am. J. Vet. Research, v. 40 (6), 867-868
Sarcocystis spp., incidence in slaughtered pigs (diaphragms, esophagi, heart, tongues), unsuccessful attempt to transmit to dogs and cats: Xenia, Ohio

Sarcocystis sp.
Dubey, J. P.; Fayer, R.; and Seesee, F. M., 1978, J. Am. Vet. Med. Ass., v. 173 (9), 1167-1170
Sarcocystis sporocysts, coyotes (feces), prevalence, infectivity for sheep and cattle, dogs fed tissues of inoculated and control sheep and cattle shed sporocysts, results indicated that coyotes are source of ovine and possibly bovine Sarcocystis in Montana

Sarcocystis [sp.]
Duszynski, D. W.; and Box, E. D., 1978, J. Parasitol., v. 64 (2), 326-329
Sarcocystis-infected muscles from Molothrus ater, Cassidix mexicanus, Quiscalus quiscula, and Anas acuta produced patent infections when fed to Didelphis virginiana but infected muscle from A. carolinensis and Spatula clypeata did not, sporocysts of cowbird and grackle origin designated as S. debonei, those of duck origin not given species designation, opossum likely to be important definitive host for Sarcocystis of icterid birds: Texas

Sarcocystis sp., illus.
Entzeroth, R.; Scholtyseck, E.; and Greuel, E., 1978, Naturwissenschaften, v. 65 (7), 395
"The application of different techniques led to the conclusion that, besides three ultramorphologically different sarcosporidian cysts, the roe-deer [Capreolus capreolus] muscles contain coccidia of the genera Toxoplasma and Hammondia."

Sarcocystis spec., illus.
Erber, M.; Boch, J.; and Barth, D., 1978, Berl. u. Munchen. Tierarztl. Wchnschr., v. 91 (24), 482-486
brief description
Capreolus capreolus (Muskulatur): Sudbayern

Sarcocystis [sp.]-like sporocysts
Erdman, L. F., 1978, Iowa State Univ. Vet., v. 40 (3), 112
Mephitis mephitis (heart, diaphragm, esophagus, and iliopsoas muscles): Green County, Iowa
Canis familiaris (exper.)

Sarcocystis [sp.]
Ernst, S.; Raby, J.; and Rebolledo, C., 1977, Bol. Chileno Parasitol., v. 32 (3-4), 81-83
Sarcocystis [sp.], incidence survey in sheep, relationships of sex, age and weight to infection rate: Province of Valdivia, Chile

Sarcocystis spp.
Glazebrook, J. S.; Campbell, R. S. F.; and Hutchinson, G. W., 1977, Tropenmed. u. Parasitol., v. 28 (4), 545-551
survey of feral rodents in six tropical habitats for infections of potential zoonotic importance: North Queensland, Australia

Sarcocystis [sp.], illus.
Goregliad, Kh. S.; Litvinov, V. F.; and Litvenkova, A. E., 1977, Vestsi Akad. Navuk BSSR, s. Biial. Navuk (3), 118-121
[Sus scrofa]
[Bos taurus]
[Alces alces]
[pig, wild]
[Bison bonasus]
[Ursus]
[Rattus rattus]
[Capreolus capreolus]
[Nyctereutes procyonoides]
[Vulpes vulpes]
[Martes martes]
[Canis lupus]
all from northern sections of Belorussia

Sarcocystis [sp.]
Gupta, S. L.; Gautam, O. P.; and Bhardwaj, R. M., 1979, Indian J. Animal Sc., v. 49 (11), 971
Sarcocystis [sp.], sheep (diaphragm, heart muscle), prevalence, peptic digestion technique: Hissar, India

Sarcocystis species cysts, Type I, illus.
Kan, S. P., 1979, Internat. J. Parasitol., v. 9 (5), 475-480
Sarcocystis spp. from rodents, ultrastructure of cyst wall
Rattus r. diardii (skeletal muscles)
R. exulans (subcutaneous tissues)
R. jalorensis (abdominal muscle)
Bandicota indica (skeletal muscle)
all from Malaysia

Sarcocystis species cysts, Type II, illus.
Kan, S. P., 1979, Internat. J. Parasitol., v. 9 (5), 475-480
Sarcocystis spp. from rodents, ultrastructure of cyst wall
Rattus r. diardii (abdominal muscles, subcutaneous tissues)
R. exulans (subcutaneous tissues)
R. annandalei (subcutaneous tissues)
Bandicota indica (skeletal muscle)
all from Malaysia

Sarcocystis species cysts, Type III, illus.
Kan, S. P., 1979, Internat. J. Parasitol., v. 9 (5), 475-480
Sarcocystis spp. from rodents, ultrastructure of cyst wall
Rattus r. diardii (subcutaneous tissues)
R. exulans (subcutaneous tissues)
all from Malaysia

Sarcocystis sp., illus.
Kan, S. P., and Dissanaike, A. S., 1977, Ztschr. Parasitenk., v. 52 (3), 219-227
Sarcocystis sp., ultrastructure, possible taxonomic relationships
Rattus rattus diardii (diaphragm, gastrocnemius and abdominal muscles): Klang, Selangor, Malaysia

Sarcocystis sp., illus.
Kan, S. P.; Prathap, K.; and Dissanaike, A. S., 1979, Am. J. Trop. Med. and Hyg., v. 28 (4), 634-642
Sarcocystis sp. from Macaca fascicularis (femoral muscle), ultrastructure of cyst wall and zoites, comparison with Sarcocystis spp. from other monkeys and from moonrat: Malaysia

Sarcocystis [sp.]-like
Keymer, I. F., 1978, Vet. Rec., v. 103 (25), 548-552
parasitic and other diseases of tortoises, necropsy survey
Testudo denticulata (skeletal muscles)

Sarcocystis [sp.]
Landsverk, T., 1978, Norsk Vet.-Tidsskr., v. 90 (4), 237
okse: Telemark, Norge

Sarcocystis-like protozoon [sp.], illus.
Landsverk, T.; Gamlem, H.; and Svenkerud, R., 1978, Vet. Path., v. 15 (2), 186-195
Sarcocystis-like protozoon in ewe (lymph nodes, myocardium, capillary endothelium), description (light and electron microscopy), distribution in host, clinical signs, pathology, case report: Gotland

Sarcocystis sp., illus.
Mehlhorn, H.; and Heydorn, A. O., 1978, Advances Parasitol., v. 16, 43-91
Sarcosporidia, life cycle and fine structure, review

Sarcocystis sp., illus.
Ogassawara, S.; et al., 1977, Rev. Microbiol., Sao Paulo, v. 8 (1), 21-23
Sarcocystis sp., incidence of sporocysts in dogs (feces): city of Sao Paulo

Sarcocystis sp., illus.
Ohbayashi, M.; Masegi, T.; and Kubota, K., 1972, Japan. J. Vet. Research, v. 20 (3), 50-56
Urotrichus talpoides (skeletal muscles): Yumoto, Japan

Sarcocystis spp.
Perrotin, C.; and Graber, M., 1977, Rev. Elevage et Med. Vet. Pays Trop., n.s., v. 30 (4), 377-382
Sarcocystis spp., domestic animals, review of life cycle and differential characters from Toxoplasma gondii and Hammondia hammondi

Sarcocystis [sp.], illus.
Prathap, K.; and Dissanaike, A. S., 1978, Southeast Asian J. Trop. Med. and Pub. Health, v. 9 (3), 452-453
man (soft tissue of neck): Malaysia

Sarcocystis [sp.]
Raju, N. R.; and Munro, R., 1978, Austral. Vet. J., v. 54 (12), 599 [Letter]
Sarcocystis, survey, incidence in pigs and cattle slaughtered at abattoir: Fiji

Sarcocystis sp.
Sinniah, B., 1979, Southeast Asian J. Trop. Med. and Pub. Health, v. 10 (1), 115-121
distribution and prevalence
Rattus annandalei
R. argentiventer
R. rattus diardii
R. exulans
R. tiomanicus
R. norvegicus
R. sabanus
Bandicota indica
all from Peninsular Malaysia

Sarcocystis [sp.], illus.
Srivastava, C. P.; Sinha, B. K.; and Sahai, B. N., 1977, Indian J. Animal Health, v. 16 (2), 105-106
Sarcocystis [sp.] in heart muscle of cattle and sheep, pathogenic effect, 3 cases

Sarcocystis sp., illus.
Tadros, W.; and Laarman, J. J., 1978, Proc. K. Nederl. Akad. Wetensch., s. C, Biol. and Med. Sc., v. 81 (4), 469-491
Sarcocystis spp., structure of muscle cyst walls, comparative study, light and electron microscopy, useful but not infallible tool for recognizing different species
Alabuia multicarinata (sub-dermal connective tissue and adjacent pectoral muscle): India (purchased from pet shop in The Netherlands)

Sarcocystis sp., illus.
Tadros, W.; and Laarman, J. J., 1978, Proc. K. Nederl. Akad. Wetensch., s. C, Biol. and Med. Sc., v. 81 (4), 469-491
Sarcocystis spp., structure of muscle cyst walls, comparative study, light and electron microscopy, useful but not infallible tool for recognizing different species
Cyanocorax chrysops (pectoral muscle)

Sarcocystis [sp.], illus.
Tadros, W.; and Laarman, J. J., 1979, Ztschr. Parasitenk., v. 58 (3), 195-200
Sarcocystis [sp.] in Mustela nivalis (abdominal, pectoral, and femoral muscles; subepithelium of small intestinal villi) and Strix aluco (droppings) (exper.), sarcocysts "strongly reminiscent of" and sporocysts "morphologically indistinguishable from" Sarcocystis sebeki, oocysts "morphologically identical to" S. putorii

Sarcocystis sp.
Todd, K. S., jr.; and Seaman, W. J., 1978, Vet. Med. and Small Animal Clin., v. 73 (6), 821-822
sheep (heart): Canton, Illinois

Sarcocystis sp.
Wade, S. E.; Haschek, W. M.; and Georgi, J. R., 1979, Cornell Vet., v. 69 (3), 198-205
Bison bison (heart muscle): New York State

Sarcocystis [sp.]
Wobeser, G., 1976, J. Wildlife Dis., v. 12 (4), 566-571
Rangifer tarandus caribou: northern Saskatchewan

Sarcocystis asinus n. sp., illus.
Gadaev, A., 1978, Uzbek. Biol. Zhurnal (1), 47-48
Equus asinus (muscles of heart, neck, esophagus, and diaphragm, peripheral blood): Kalininsk and Verkhnechirchiksk regions, Tashkent oblast

Sarcocystis bertrami Doflein, 1901
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
nomen dubium

Sarcocystis bigemina
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
nomen dubium

Sarcocystis blanchardi
Grundboeck, M., 1977, Biul. Vsesoiuz. Ordena Lenin. Inst. Eksper. Vet., v. 30, 14-15
protozoa, cattle, as cofactors in leucotic process

Sarcocystis booliati, illus.
Prathap, K.; and Dissanaike, A. S., 1978, Southeast Asian J. Trop. Med. and Pub. Health, v. 9 (3), 452-453

Sarcocystis bovicanis, illus.
Boch, J.; Laupheimer, K. E.; and Erber, M., 1978, Berl. u. Munchen. Tierarztl. Wchnschr., v. 91 (21), 426-431
brief description
Rinder (Muskulatur): Schlachthofen Suddeutschlands

Sarcocystis bovicanis
Dissanaike, A. S.; and Kan, S. P., 1978, Ztschr. Parasitenk., v. 55 (2), 127-138
as syn. of S. cruzi

Sarcocystis bovicanis
Fassi-Fehri, N.; et al., 1978, Ann. Recherches Vet., v. 9 (3), 409-417
Sarcocystis spp., cattle, sheep, frequency of infestation, host age and sex, localization in various muscles, pathology: Morocco

Sarcocystis bovicanis, illus.
Fayer, R., 1977, Proc. Helminth. Soc. Washington, v. 44 (2), 206-209
Sarcocystis bovicanis, calves (exper.), life cycle, first asexual generation

Sarcocystis bovicanis Heydorn et al., 1975
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
definition, intermediate and final hosts, synonymy

Sarcocystis bovicanis, illus.
Hinaidy, H. K.; et al., 1979, Wien. Tierarztl. Monatsschr., v. 66 (5), 181-184
Sarcocystis spp., cattle, monthly occurrence, localization, diagnosis
Rinder: Osterreich

Sarcocystis bovicanis, illus.
Jungmann, R.; et al., 1977, Monatsh. Vet.-Med., v. 32 (23), 885-889
Sarcocystis bovicanis, calves (exper.), clinical course of infection, pathomorphological findings

Sarcocystis bovicanis, illus.
Mehlhorn, H.; and Heydorn, A. O., 1978, Advances Parasitol., v. 16, 43-91
Sarcosporidia, life cycle and fine structure, review

Sarcocystis bovifelis, illus.
Boch, J.; Laupheimer, K. E.; and Erber, M., 1978, Berl. u. Munchen. Tierarztl. Wchnschr., v. 91 (21), 426-431
brief description
Rinder (Muskulatur): Schlachthofen Suddeutschlands

Sarcocystis bovifelis
Dissanaike, A. S.; and Kan, S. P., 1978, Ztschr. Parasitenk., v. 55 (2), 127-138
as syn. of S. hirsuta

Sarcocystis bovifelis and/or S. bovihominis
Fassi-Fehri, N.; et al., 1978, Ann. Recherches Vet., v. 9 (3), 409-417
Sarcocystis spp., cattle, sheep, frequency of infestation, host age and sex, localization in various muscles, pathology: Morocco

Sarcocystis bovifelis Heydorn et al., 1975
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
definition, intermediate and final hosts, synonymy

Sarcocystis bovifelis, illus.
Hinaidy, H. K.; et al., 1979, Wien. Tierarztl. Monatsschr., v. 66 (5), 181-184
Sarcocystis spp., cattle, monthly occurrence, localization, diagnosis
Rinder: Osterreich

Sarcocystis bovihominis, illus.
Boch, J.; Laupheimer, K. E.; and Erber, M., 1978, Berl. u. Munchen. Tierarztl. Wchnschr., v. 91 (21), 426-431
brief description
Rinder (Muskulatur): Schlachthofen Suddeutschlands

Sarcocystis bovihominis and/or S. bovifelis
Fassi-Fehri, N.; et al., 1978, Ann. Recherches Vet., v. 9 (3), 409-417
Sarcocystis spp., cattle, sheep, frequency of infestation, host age and sex, localization in various muscles, pathology: Morocco

Sarcocystis bovihominis Heydorn et al., 1975
  Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
    definition, intermediate and final hosts, synonymy

Sarcocystis bovihominis, illus.
  Hinaidy, H. K.; et al., 1979, Wien. Tierarztl. Monatsschr., v. 66 (5), 181-184
    Sarcocystis spp., cattle, monthly occurrence, localization, diagnosis
    Rinder: Osterreich

Sarcocystis bovihominis, illus.
  Mehlhorn, H.; and Heydorn, A. O., 1978, Advances Parasitol., v. 16, 43-91
    Sarcosporidia, life cycle and fine structure, review

Sarcocystis capreolicanis n. spec., illus.
  Erber, M.; Boch, J.; and Barth, D., 1978, Berl. u. Munchen. Tierarztl. Wchnschr., v. 91 (24), 482-486
    Capreolus capreolus (Muskulatur) (nat. and exper.): Sudbayern
    Hunde (exper.)
    Füchse (exper.)

Sarcocystis cervi (von Hessling, 1854)
  Drost, S., 1977, Ang. Parasitol., v. 18 (4), 219-225
    Sarcocystis cervi in Cervus elaphus, incidence, localization, host age and sex, seasonal occurrence: Kreis-Wildsammelstelle Ludwigslust; Kreis-Wildsammelstelle Perleberg

Sarcocystis cruzi
  Barnett, D.; et al., 1977, Proc. 20. Ann. Meet., Am. Ass. Vet. Lab. Diagn. (Minneapolis, Minnesota, Oct. 16-18), 131-138
    Sarcocystis cruzi, pregnant cows (exper.), abortion, practical diagnosis using maternal caruncle

Sarcocystis cruzi
  Dissanaike, A. S.; and Kan, S. P., 1978, Ztschr. Parasitenk., v. 55 (2), 127-138
    Syn.: S. bovicanis

Sarcocystis cruzi
  Farmer, J. N.; et al., 1978, Vet. Rec., v. 102 (4), 78-80
    prevalence
    sheepdogs (feces): Gwyned, North Wales

Sarcocystis cruzi
  Fayer, R.; and Lynch, G. P., 1979, Parasitology, v. 79 (3), 325-336
    Sarcocystis cruzi-infected calves (exper.), pathophysiological changes in urine and blood, several specific effects beyond those induced by nutritional stress

Sarcocystis cruzi
  Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
    nomen dubium

Sarcocystis cruzi
  Kepka, O.; and Oesterreicher, H. D., 1979, Wien. Tierarztl. Monatsschr., v. 66 (5), 184-185
    Rinder: Steiermark, Osterreich

Sarcocystis cruzi, illus.
  Landsverk, T., 1979, Acta Vet. Scand., v. 20 (2), 238-244
    Sarcocystis cruzi, outbreak in cattle herd, histopathology: Telemark, Norway

Sarcocystis cruzi
  Leek, R. G.; and Fayer, R., 1978, Proc. Helminth. Soc. Washington, v. 45 (1), 135-136
    dogs fed beef and beef products purchased from retail food store in Maryland suburbs of Washington, D. C., presence of Sarcocystis cruzi sporocysts in dogs fed fresh beef and rare roast beef; no sporocysts found in feces of human volunteer, although results suggest that S. hominis, when present, could probably also survive in retail beef

Sarcocystis cruzi
  Leguia, G.; and Herbert, I. V., 1979, Research Vet. Sc., v. 27 (3), 390-391
    Sarcocystis in dogs, foxes, sheep, and cattle, Toxoplasma gondii in sheep, prevalence, indirect haemagglutination reaction for serodiagnosis
    dogs
    cattle
    all from New South Wales

Sarcocystis cruzi, illus.
  Pacheco, N. D.; Sheffield, H. G.; and Fayer, R., 1978, J. Parasitol., v. 64 (2), 320-325
    Sarcocystis cruzi, immature cyst, fine structure in relation to development and to multiplication of parasites within it

S[arcocystis] cuniculi Brumpt, 1913
  Meshkov, S., 1978, Vet. Med. Nauki, v. 15 (4), 72-78
    Lepus europeus (muscles): Bulgaria

Sarcocystis cuniculi, illus.
  Tadros, W.; and Laarman, J. J., 1978, Proc. K. Nederl. Akad. Wetensch., s. C, Biol. and Med. Sc., v. 81 (4), 469-491
    Sarcocystis spp., structure of muscle cyst walls, comparative study, light and electron microscopy, useful but not infallible tool for recognizing different species
    common European rabbit (abdominal wall muscles): Netherlands

Sarcocystis cymruensis n. sp., illus
  Ashford, R. W., 1978, Ann. Trop. Med. and Parasitol., v. 72 (1), 37-43
    muscle cysts not infective for Mustela putorius or dogs, sporocysts not infective for Mus musculus
    Rattus norvegicus (skeletal muscles) (nat. and exper.): area of Welshpool, mid Wales; border areas of England; Chobham and Petworth, Surrey
    Felis catus (intestine, feces) (exper.)

Sarcocystis debonei Vogelsang, 1929
  Duszynski, D. W.; and Box, E. D., 1978, J. Parasitol., v. 64 (2), 326-329
    Sarcocystis-infected muscles from Molothrus ater, Cassidix mexicanus, Quiscalus quiscula, and Anas acuta produced patent infections when fed to Didelphis virginiana but infected muscle from A. carolinensis and Spatula clypeata did not, sporocysts of cowbird and grackle origin designated as S. debonei, those of duck origin not given species designation, opossum likely to be important definitive host for Sarcocystis of icterid birds: Texas
    Syn.: Isospora boughtoni Volk, 1938

Sarcocystis dispersa
Cerna, Z.; and Senaud, J., 1977, Compt. Rend. Acad. Sc., Paris, v. 285, s. D, Sc. Nat., (4), 347-349
Sarcocystis dispersa in mice, asexual multiplication directly in cytoplasm of hepatic cells without formation of parasitophorous vacuole, new process of endogenesis (multiple synchronous endopolygenesis)

Sarcocystis equicanis Rommel and Geisel, 1975
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
definition, intermediate and final hosts, synonymy

Sarcocystis fusiformis (Railliet, 1897), illus.
Abdel Ghaffar, F.; Hilali, M.; and Scholtyseck, E., 1978, Tropenmed. u. Parasitol., v. 29 (3), 289-294
Sarcocystis fusiformis in Bubalus bubalis, fine structure morphology, large and small sarcocysts from muscular layer of oesophagus: Egypt

Sarcocystis fusiformis
Bordjochki, A.; et al., 1978, Bull. Acad. Vet. France, n.s., v. 51 (2), 189-195
Sarcocystis fusiformis, S. tenella, S. miescheriana, rabbits (exper.), diagnostic value of complement fixation and fluorescent antibody tests

Sarcocystis fusiformis (Railliet, 1897)
Drost, S.; and Brackmann, H., 1978, Monatsh. Vet.-Med., v. 33 (5), 175-178
Sarcocystis fusiformis, cattle, incidence tested by trichinoscopy, age of host, seasonal variation
Rinder (Muskel): Sanitatsschlachtbetriebes, Nordwesten DDR

Sarcocystis fusiformis (Railliet 1897)
Gill, H. S.; et al., 1978, J. Parasitol., v. 64 (3), 549-551
dogs fed Bubalus bubalis diaphragm muscles with visible cysts of Sarcocystis fusiformis shed unsporulated oocysts belonging either to Hammondia or Isospora

Sarcocystis fusiformis, illus.
Godoy, G. A.; Volcan G., G.; and Guevara, R., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (1), 68-72
bovinos (tejido miocardico): Estado Bolivar, Venezuela

S[arcocystis] fusiformis
Golubkov, V. I., 1978, Veterinariia, Moskva (5), 98-99
S[arcocystis] fusiformis, viability in meat and meat products in relation to temperature and salting

Sarcocystis fusiformis, illus.
Kan, S. P.; and Dissanaike, A. S., 1978, Ztschr. Parasitenk., v. 57 (2), 107-116
Sarcocystis levinei, S. fusiformis, comparative ultrastructure of cyst wall and zoites: Malaysia

Sarcocystis fusiformis
Proctor, S. J.; et al., 1976, Proc. 19. Ann. Meet., Am. Ass. Vet. Lab. Diagn., 329-336
Sarcocystis fusiformis, Jersey-cross cows (exper.), gross and histologic lesions

Sarcocystis fusiformis (Railliet, 1897), illus.
Scholtyseck, E.; and Hilali, M., 1978, Ztschr. Parasitenk., v. 56 (2), 205-209
Sarcocystis fusiformis, cats infected with cysts from Bubalus bubalis, ultrastructural study of sexual stages; first report describing sexual stages of Sarcocystis in final host

Sarcocystis fusiformis
Telles de Jesus Filho, M.; and Miraglia, T., 1977, Acta Histochem., v. 59 (1), 160-167
Sarcocystis fusiformis cysts in ox heart, histochemical (primarily histoenzymologic) observations, implications for parasite metabolism, no inflammatory reactions found around cysts and enzymatic reactivity of muscle fibers near to cysts was not different from normal fibers

Sarcocystis fusiformis
Thomas, V.; and Dissanaike, A. S., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (3), 303-306
Sarcocystis, human, indirect fluorescent antibody test using Sarcocystis fusiformis as antigen, antibody prevalence varies among 4 ethnic groups (Orang Asli, Malays, Indians, Chinese), some sera also reacted positively to Toxoplasma: West Malaysia

Sarcocystis gracilis (v. Ratz, 1909), illus.
Erber, M.; Boch, J.; and Barth, D., 1978, Berl. u. Munchen. Tierarztl. Wchnschr., v. 91 (24), 482-486
brief description
Capreolus capreolus (Muskulatur) (nat. and exper.): Sudbayern
Hunde (exper.)
Füchse (exper.)

S[arcocystis] gracilis v. Ratz, 1910
Meshkov, S., 1978, Vet. Med. Nauki, v. 15 (4), 72-78
Capreolus capreolus (muscles): Bulgaria

Sarcocystis hirsuta
Dissanaike, A. S.; and Kan, S. P., 1978, Ztschr. Parasitenk., v. 55 (2), 127-138
Syn.: S. bovifelis

Sarcocystis hirsuta Moule, 1888
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
nomen dubium

Sarcocystis hirsuta
Kepka, O.; and Oesterreicher, H. D., 1979, Wien. Tierarztl. Monatsschr., v. 66 (5), 184-185
Rinder: Steiermark, Osterreich

Sarcocystis hirsuta, illus.
Kisliakova, Z. I., 1973, Vet. Nauka--Proizvod., Trudy, Minsk, v. 11, 165-171
Sarcocystis hirsuta, cattle, morphology; review of prevalence and distribution

Sarcocystis hominis
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
nomen dubium

Sarcocystis hominis
Kepka, O.; and Oesterreicher, H. D., 1979, Wien. Tierarztl. Monatsschr., v. 66 (5), 184-185
Rinder: Steiermark, Osterreich

Sarcocystis hominis
Leek, R. G.; and Fayer, R., 1978, Proc. Helminth. Soc. Washington, v. 45 (1), 135-136
dogs fed beef and beef products purchased from retail food store in Maryland suburbs of Washington, D. C., presence of Sarcocystis cruzi sporocysts in dogs fed fresh beef and rare roast beef; no sporocysts found in feces of human volunteer, although results suggest that S. hominis, when present, could probably also survive in retail beef

Sarcocystis hominis, illus.
Tadros, W.; and Laarman, J. J., 1978, Proc. K. Nederl. Akad. Wetensch., s. C, Biol. and Med. Sc., v. 81 (4), 469-491
Sarcocystis spp., structure of muscle cyst walls, comparative study, light and electron microscopy, useful but not infallible tool for recognizing different species
calf (lower oesophageal muscles) (exper.)
Dutch child

Sarcocystis horvathi
Golubkov, V. I., 1979, Veterinariia, Moskva (1), 55-56
Sarcocystis horvathi and S. rileyi cysts from feces of carnivores fed muscle tissue from domestic birds

Sarcocystis kirmsei sp. nov., illus.
Garnham, P. C. C.; Duggan, A. J.; and Sinden, R. E., 1979, Ann. Parasitol., v. 54 (4), 393-400
Lophura diardi (brain): Thailand (London Zoological Gardens)
unidentified passerine bird (brain): Cerro Punta, Chiriqui Province, Panama

Sarcocystis kortei Castellani u. Chalmers, 1909
Prosl, H.; and Tamer, A., 1979, Zentralbl. Vet-Med., Reihe B, v. 26 (9), 696-709
Macaca mulatta (Skelettmuskulatur)

Sarcocystis levinei n. sp., illus.
Dissanaike, A. S.; and Kan, S. P., 1978, Ztschr. Parasitenk., v. 55 (2), 127-138
Sarcocystis levinei n. sp., life cycle
Bubalus bubalis (oesophageal muscles): abattoir, Shah Alam, Selangor, Malaysia
dog (exper.) (small intestine)

Sarcocystis levinei, illus.
Kan, S. P.; and Dissanaike, A. S., 1978, Ztschr. Parasitenk., v. 57 (2), 107-116
Sarcocystis levinei, S. fusiformis, comparative ultrastructure of cyst wall and zoites: Malaysia

Sarcocystis miescheriana, illus.
Bogush, A. A., 1973, Vet. Nauka--Proizvod., Trudy, Minsk, v. 11, 171-175
Trichinella spiralis, Sarcocystis miescheriana, swine, differential diagnosis

Sarcocystis miescheriana, illus.
Bogush, A. A., 1976, [Parasites in muscles of swine and measures for their prophylaxis (trichinosis and sarcocystosis)], 94 pp., illus.
trichinosis, sarcocystosis, swine muscles, general aspects, emphasis on diagnosis and prophylaxis, review: Belorussia

Sarcocystis miescheriana
Bordjochki, A.; et al., 1978, Bull. Acad. Vet. France, n.s., v. 51 (2), 189-195
Sarcocystis fusiformis, S. tenella, S. miescheriana, rabbits (exper.), diagnostic value of complement fixation and fluorescent antibody tests

Sarcocystis miescheriana
Farmer, J. N.; et al., 1978, Vet. Rec., v. 102 (4), 78-80
prevalence
sheepdogs (feces): Gwyned, North Wales
greyhounds (feces): London area

Sarcocystis miescheriana (Kuehn, 1865) Labbe, 1899
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
nomen dubium

Sarcocystis miescheriana
Kisliakova, Z. I., 1970, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 8, 124-126
Sarcocystis miescheriana, young pigs, diagnosis by compressor method

Sarcocystis miescheriana Kuehn, 1865
Kisliakova, Z. I., 1971, Nauch. Trudy, Nauchno-Issled. Vet. Inst., v. 9, 134-137
Sarcocystis miescheriana, cysts and trophozoites from swine used to infect white mice (muscles of heart, esophagus, diaphragm, and abdomen)

S[arcocystis] miescheriana
Meshkov, S., 1978, Vet. Med. Nauki, v. 15 (4), 72-78
Sus scrofa (muscles): Bulgaria

Sarcocystis miescheriana (Kuehn, 1865) Lankester, 1882
Tadros, W.; and Laarman, J. J., 1978, Proc. K. Nederl. Akad. Wetensch., s. C, Biol. and Med. Sc., v. 81 (4), 466-468
Syns.: Coccidium bigeminum Stiles, 1891; Isospora bigemina (Stiles, 1891) Luehe, 1906; C. bigeminum var. canis Railliet and Lucet, 1891, selected here as nominate subspecies of C. bigeminum and thus designated C. bigeminum bigeminum

Sarcocystis miescheriana, illus.
Tadros, W.; and Laarman, J. J., 1978, Proc. K. Nederl. Akad. Wetensch., s. C, Biol. and Med. Sc., v. 81 (4), 469-491
Sarcocystis spp., structure of muscle cyst walls, comparative study, light and electron microscopy, useful but not infallible tool for recognizing different species
Sus scrofa ferus (diaphragmal muscles): Netherlands
domestic pig and piglets (diaphragmal and oesophageal muscles) (nat. and exper.)
dogs (exper.)

Sarcocystis moulei
Rodriguez Osorio, M.; et al., 1978, Rev. Iber. Parasitol., v. 38 (3-4), 793-804
Sarcocystis moulei, caprine, comparative study of 3 diagnostic tests, peptic artificial digestion, immunodiffusion, and indirect immunofluorescence

Sarcocystis muris
Ashford, R. W., 1978, Ann. Trop. Med. and Parasitol., v. 72 (1), 95
"The authorship should, however, be corrected and the correct designation, hereby proposed, is Sarcocystis muris (Railliet, 1886)."

Sarcocystis muris
  Fayer, R.; and Frenkel, J. K., 1979, J. Parasitol., v. 65 (5), 756-762
    6 spp. of feline coccidia, oocysts fed to calves, calf tissue then fed to cats: neither Besnoitia, Hammondia, nor Sarcocystis were pathogenic for calves nor did they establish patent infections which could be transmitted back to cats; Cystoisospora spp. were not pathogenic for calves but could be transmitted back to cats; Toxoplasma strains were slightly to moderately pathogenic for calves and could be transmitted back to cats

Sarcocystis muris (Blanchard, 1885) Labbe, 1899 (type species)
  Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
    definition, intermediate and final hosts
    "Because the former type species, Sarcocystis miescheriana, is an indeterminate nomen dubium, we are proposing S. muris as the new type species."
    Syns.: Miescheria muris Blanchard, 1885; Coccidium bigeminum (Stiles, 1891) Luehe, 1906 pro parte; Coccidium bigeminum var. cati Railliet and Lucet, 1891 pro parte; Sarcocystis musculi (Blanchard, 1885) of Kalyakin and Zasukhin, 1975 (lapsus linguae)

Sarcocystis muris
  Lai, P. F., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (3), 417-419
    Rattus exulans
    R. jalorensis
    all from Malacca, Malaysia

Sarcocystis muris
  Last, M. J.; and Powell, E. C., 1978, J. Parasitol., v. 64 (1), 162-163
    mice used by Powell, E. C.; and McCarley, J. B., 1975, J. Parasitol., v. 61 (5), 928-931, were infected with both Sarcocystis muris and Isospora felis

S[arcocystis] muris R. Blanchard, 1885
  Meshkov, S., 1978, Vet. Med. Nauki, v. 15 (4), 72-78
    Rattus norvegicus
    Apodemus sylvaticus
    (muscles of all): all from Bulgaria

Sarcocystis muris
  Rommel, M., 1979, Ztschr. Parasitenk., v. 58 (2), 187-188
    Putorius putorius furo (exper.) (Kot)
    Mus musculus (exper.) (Muskulatur)

Sarcocystis muris
  Smith, D. D.; and Frenkel, J. K., 1978, J. Parasitol., v. 64 (2), 315-319
    Sarcocystis muris, Toxoplasma gondii, Isospora spp., experimental transmission to mice by ingestion of Blatella germanica and Periplaneta americana exposed to cat feces containing these coccidia or by direct ingestion of the feces, results indicate possible role of cockroaches as transport hosts and sources of cross-infection in laboratories, need for stringent control

Sarcocystis musculi (Blanchard, 1885) of Kalyakin and Zasukhin, 1975 (lapsus linguae)
  Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
    as syn. of Sarcocystis muris (Blanchard, 1885) Labbe, 1899

Sarcocystis nesbitti Mandour, 1969
  Prosl, H.; and Tamer, A., 1979, Zentralbl. Vet-Med., Reihe B, v. 26 (9), 696-709
    Macaca mulatta (Skelettmuskulatur)

Sarcocystis orientalis
  Lai, P. F., 1977, Southeast Asian J. Trop. M
  and Pub. Health, v. 8 (3), 417-419
    Rattus exulans
    R. annandeli
    R. jalorensis
    R. r. diardii
    all from Malacca, Malaysia

Sarcocystis ovicanis, illus.
  Boch, J.; et al., 1979, Berl. u. Munchen. Tierarztl. Wchnschr., v. 92 (7), 137-141
    Sarcocystis spp., Toxoplasma, sheep, parasitological and serological studies: slaughterhouses in Bavaria

Sarcocystis ovicanis
  Farmer, J. N.; et al., 1978, Vet. Rec., v. 10
  (4), 78-80
    prevalence
    Vulpes vulpes (feces): North Wales
    sheepdogs (feces): Gwyned, North Wales
    greyhounds (feces): London area

Sarcocystis ovicanis
  Fassi-Fehri, N.; et al., 1978, Ann. Recherche
  Vet., v. 9 (3), 409-417
    Sarcocystis spp., cattle, sheep, frequency
    infestation, host age and sex, localization
    in various muscles, pathology: Morocco

Sarcocystis ovicanis Heydorn et al., 1975
  Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
    definition, intermediate and final hosts, synonymy

Sarcocystis ovicanis, illus.
  Landsverk, T.; and Bratberg, B., 1979, Acta Vet. Scand., v. 20 (2), 206-308
    Sarcocystis ovicanis, possible cause of polyarteritis nodosa in sheep

Sarcocystis ovicanis
  Leguia, G.; and Herbert, I. V., 1979, Researc
  Vet. Sc., v. 27 (3), 390-391
    Sarcocystis in dogs, foxes, sheep, and cattle, Toxoplasma gondii in sheep, prevalence, indirect haemagglutination reaction for serodiagnosis
    dogs
    foxes
    sheep
    all from New South Wales

Sarcocystis ovicanis, illus.
  Mehlhorn, H.; and Heydorn, A. O., 1978, Advances Parasitol., v. 16, 43-91
    Sarcosporidia, life cycle and fine structure
    review

Sarcocystis ovicanis
  Munday, B. L., 1979, Vet. Parasitol., v. 5 (2-3), 129-135
    Sarcocystis ovicanis, deleterious effect on growth rate and haematocrit in lambs, presence of antibodies (presumably colostral)
    against Sarcocystis did not appear to provid
    significant protection

Sarcocystis ovifelis
Fassi-Fehri, N.; et al., 1978, Ann. Recherches Vet., v. 9 (3), 409-417
Sarcocystis spp., cattle, sheep, frequency of infestation, host age and sex, localization in various muscles, pathology: Morocco

Sarcocystis ovifelis Heydorn et al., 1975
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
definition, intermediate and final hosts, synonymy

Sarcocystis putorii (Railliet and Lucet, 1891) comb. nov.
Tadros, W.; and Laarman, J. J., 1978, Proc. K. Nederl. Akad. Wetensch., s. C, Biol. and Med. Sc., v. 81 (4), 466-468
Syn.: Coccidium bigeminum var. putorii Railliet and Lucet, 1891; Lucetina putorii (Railliet and Lucet, 1891) Henry and Leblois, 1926; Isospora putorii (Railliet and Lucet, 1891) Becker, 1934

Sarcocystis putorii, illus.
Tadros, W.; and Laarman, J. J., 1978, Proc. K. Nederl. Akad. Wetensch., s. C, Biol. and Med. Sc., v. 81 (4), 469-491
Sarcocystis spp., structure of muscle cyst walls, comparative study, light and electron microscopy, useful but not infallible tool for recognizing different species
Microtus arvalis (pectoral muscles) (exper.)
M. agrestis (pectoral muscles)
Mustela nivalis (exper.)

arcocystis putorii
Tadros, W.; and Laarman, J. J., 1979, Ztschr. Parasitenk., v. 58 (3), 195-200
Sarcocystis [sp.] in Mustela nivalis (abdominal, pectoral, and femoral muscles; subepithelium of small intestinal villi) and Strix aluco (droppings) (exper.), sarcocysts "strongly reminiscent of" and sporocysts "morphologically indistinguishable from" Sarcocystis sebeki, oocysts "morphologically identical to" S. putorii

Sarcocystis rileyi
Golubkov, V. I., 1979, Veterinariia, Moskva (1), 55-56
Sarcocystis horvathi and S. rileyi cysts from feces of carnivores fed muscle tissue from domestic birds

arcocystis sebeki, illus.
Tadros, W.; and Laarman, J. J., 1978, Proc. K. Nederl. Akad. Wetensch., s. C, Biol. and Med. Sc., v. 81 (4), 469-491
Sarcocystis spp., structure of muscle cyst walls, comparative study, light and electron microscopy, useful but not infallible tool for recognizing different species
Apodemus sylvaticus (pectoral muscles) (nat. and exper.)
Strix aluco (exper.)

arcocystis sebeki
Tadros, W.; and Laarman, J. J., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (3), 350 [Letter]
Sarcocystis sebeki from exper. infected Strix aluco, successful rodent to rodent (Apodemus sylvaticus) transmission by inoculation of precystic schizogonic stages

Sarcocystis sebeki
Tadros, W.; and Laarman, J. J., 1979, Ztschr. Parasitenk., v. 58 (3), 195-200
Sarcocystis [sp.] in Mustela nivalis (abdominal, pectoral, and femoral muscles; subepithelium of small intestinal villi) and Strix aluco (droppings) (exper.), sarcocysts "strongly reminiscent of" and sporocysts "morphologically indistinguishable from" Sarcocystis sebeki, oocysts "morphologically identical to" S. putorii

Sarcocystis suicanis n. sp., illus.
Erber, M., 1977, Berl. u. Munchen. Tierarztl. Wchnschr., v. 90 (24), 480-482
Sus scrofa domestica (Leber, Herzmuskel, quergestreifter Muskulatur)
Sus scrofa scrofa (Leber, Herzmuskel, quergestreifter Muskulatur)
Canis lupus familiaris (Dunndarm)
Vulpes vulpes (Dunndarm)
Canis lupus lupus (Dunndarm)

Sarcocystis suicanis, illus.
Boch, J.; Mannewitz, U.; and Erber, M., 1978, Berl. u. Munchen. Tierarztl. Wchnschr., v. 91 (6), 106-111
Sarcocystis suihominis, S. suicanis, occurrence in slaughter pigs of different ages; trypsin digestive method more reliable diagnosis than trichinoscopy, species differentiation: slaughterhouses in South Germany

Sarcocystis suicanis, illus.
Erber, M.; and Geisel, O., 1979, Berl. u. Munchen. Tierarztl. Wchnschr., v. 92 (10), 197-202
Sarcocystis suicanis, pigs (exper.), clinical findings, pathology, susceptibility to S. suihominis but not to reinfection with S. suicanis; S. suicanis cysts proved infective to dogs (exper.)

Sarcocystis suicanis
Erber, M.; Meyer, J.; and Boch, J., 1978, Berl. u. Munchen. Tierarztl. Wchnschr., v. 91 (20), 393-395
Sarcocystis suicanis, pregnant sows (exper.), abortion, clinical manifestations

Sarcocystis suicanis Erber, 1977
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
definition, intermediate and final hosts, synonymy

Sarcocystis suicanis, illus.
Goebel, E.; Katz, M.; and Erber, M., 1978, Zentralbl. Bakteriol., 1. Abt. Orig., Reihe A, v. 241 (3), 368-383
Sarcocystis suicanis, pigs (exper.), ultrastructure and development of sarcocysts in muscle cells, light and electron microscopy

Sarcocystis suicanis
Heydorn, A. O.; et al., 1978, Arch. Lebensmittel-Hyg., v. 29 (5), 184-185
Sarcocystis spp., pigs, abattoir survey, frequency, intensity of infection, trichinoscopy: Bundesrepublik Deutschland; DDR

Sarcocystis suihominis, illus.
Boch, J.; Mannewitz, U.; and Erber, M., 1978, Berl. u. Munchen. Tierarztl. Wchnschr., v. 91 (6), 106-111
Sarcocystis suihominis, S. suicanis, occurrence in slaughter pigs of different ages; trypsin digestive method more reliable diagnosis than trichinoscopy, species differentiation: slaughterhouses in South Germany

Sarcocystis suihominis, illus.
Erber, M., 1977, Berl. u. Munchen. Tierarztl. Wchnschr., v. 90 (24), 480-482
Sarcocystis suicanis n. sp., S. suihominis, pigs, two methods for detection of cystozoites described and compared, differential diagnosis

Sarcocystis suihominis
Erber, M.; and Geisel, O., 1979, Berl. u. Munchen. Tierarztl. Wchnschr., v. 92 (10), 197-202
Sarcocystis suicanis, pigs (exper.), clinical findings, pathology, susceptibility to S. suihominis but not to reinfection with S. suicanis; S. suicanis cysts proved infective to dogs (exper.)

Sarcocystis suihominis, illus.
Fayer, R.; et al., 1979, Ztschr. Parasitenk., v. 59 (1), 15-20
Sarcocystis suihominis, transmission from humans to swine to nonhuman primates, absence of clinical signs, prepatent period
swine (exper.) (skeletal muscle)
Pan troglodytes (exper.) (feces)
Macaca mulatta (exper.) (feces, small intestine)
M. irus (exper.) (feces)

Sarcocystis suihominis (Tadros and Laarman, 1976) Heydorn, 1977, illus.
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
definition, intermediate and final hosts, synonymy

Sarcocystis suihominis
Heydorn, A. O.; et al., 1978, Arch. Lebensmittel-Hyg., v. 29 (5), 184-185
Sarcocystis spp., pigs, abattoir survey, frequency, intensity of infection, trichinoscopy: Bundesrepublik Deutschland; DDR

Sarcocystis suihominis, illus.
Heydorn, A. O.; and Ipczynski, V., 1978, Berl. u. Munchen. Tierarztl. Wchnschr., v. 91 (8), 154-155
Sarcocystis suihominis, pigs (exper.), schizogony

Sarcocystis suihominis, illus.
Heydorn, A. O.; and Mehlhorn, H., 1978, Zentralbl. Bakteriol., 1. Abt. Orig., Reihe A, v. 240 (1), 123-134
Sarcocystis suihominis, pigs (exper.), fine structure of schizonts and formation of merozoites within various host organs

Sarcocystis suihominis, illus.
Mehlhorn, H.; and Heydorn, A. O., 1979, Ztschr. Parasitenk., v. 58 (2), 97-113
Sarcocystis suihominis, gamogony in human tissue cultures, electron microscopical study

Sarcocystis suihominis
Piekarski, G.; et al., 1978, Immun. u. Infekt., v. 6 (4), 153-159
Sarcocystis suihominis, medical students fed raw meat from experimentally infected pig, clinical, parasitological, and serological findings
Menschen (exper.)
Schweines (exper.)

Sarcocystis suihominis, illus.
Tadros, W.; and Laarman, J. J., 1978, Proc. K. Nederl. Akad. Wetensch., s. C, Biol. and Med. Sc., v. 81 (4), 469-491
Sarcocystis spp., structure of muscle cyst walls, comparative study, light and electron microscopy, useful but not infallible tool for recognizing different species
domestic pig (diaphragmal muscles)

Sarcocystis tenella
Bordjochki, A.; et al., 1978, Bull. Acad. Vet. France, n.s., v. 51 (2), 189-195
Sarcocystis fusiformis, S. tenella, S. miescheriana, rabbits (exper.), diagnostic value of complement fixation and fluorescent antibody tests

Sarcocystis tenella (Railliet, 1886) Moule, 1888
Frenkel, J. K.; et al., 1979, Ztschr. Parasitenk., v. 58 (2), 115-139
nomen dubium

Sarcocystis tenella, illus.
Mehlhorn, H.; and Heydorn, A. O., 1978, Advances Parasitol., v. 16, 43-91
Sarcosporidia, life cycle and fine structure, review

Sarcocystis tenella, illus.
Porchet, E.; and Torpier, G., 1977, Ztschr. Parasitenk., v. 54 (2), 101-124
Toxoplasma gondii, Sarcocystis tenella, freeze fracture studies of infective stages, outer and inner membranes, rhoptries membranes

Sarcocystis tenella, illus.
Sherkov, S. N.; Leitch, B.; and Kokash, L., [1977], Egypt. J. Vet. Sc., v. 13 (1), 1976, 37-43
Sarcocystis tenella, kittens (exper.), development in intestines, life cycle; attempted parasite suppression using statyl and pancoxin plus

Sarcomastigophora [sp.]
Keymer, I. F., 1978, Vet. Rec., v. 103 (25), 548-552
parasitic and other diseases of tortoises, necropsy survey
Testudo graeca
T. elegans
Geochelone pardalis
G. gigantea
Testudo hermanni
T. marginata
T. radiata
T. denticulata
T. nigriti

Sarcomastigophora [sp.]
Keymer, I. F., 1978, Vet. Rec., v. 103 (26-27), 577-582
parasitic and other diseases of terrapins and turtles, necropsy survey
Clemmys caspica leprosa
Pseudemys scripta elegans
Clemmys caspica rivulata
Geoemyda punctularia
Pseudemys ornata calirostris
Chrysemys picta picta
Pseudemys scripta scripta
Emys orbicularis
Macroclemys temmincki
Graptemys pseudogeographica ouachitensis
Pelomedusa subrufa
Kachuga smithi
Chelodina longicollis
Chelydra serpentina
Clemmys guttata
Staurotypus triporcatus
Chrysemys picta bellii
Podocnemis expansa
(alimentary tract of all)

Sarcosporidae [of] Henneguy and Thelohan, 1892
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Chapmanium macrocystis (Gurley, 1893) comb. n.

Sarcosporidi [of] Garbini, 1891
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Chapmanium macrocystis (Gurley, 1893) comb. n.

Sarcosporidia, illus.
Arru, E.; Cosseddu, A. M.; and Tarantini, S., 1978, Clin. Vet., Milano, v. 101 (4), 195-200
Sarcosporidia, ovines, swine, indirect immunofluorescence technique compared with post mortem microscopic studies, no constant correspondence of results, not practical diagnostic tool

Sarcosporidia, illus.
Sherkov, S. N.; Leitch, B.; and El Rabie, Y., [1977], Egypt. J. Vet. Sc., v. 13 (1), 1976, 45-51
Sarcosporidia, survey of incidence in domestic animals by examining for cystozoites rather than cysts, distribution by season, age of host, species of animal, and climate: Jordan

Sarcosporidia [sp.]
Drost, S., 1977, Ang. Parasitol., v. 18 (4), 219-225
Dama dama: Kreis-Wildsammelstelle Ludwigslust

Sarcosporidian cysts
Entzeroth, R.; Scholtyseck, E.; and Greuel, E., 1978, Naturwissenschaften, v. 65 (7), 395
"The application of different techniques led to the conclusion that, besides three ultramorphologically different sarcosporidian cysts, the roe-deer [Capreolus capreolus] muscles contain coccidia of the genera Toxoplasma and Hammondia."

Sarcosporidian cysts, illus.
Perrotin, Ch.; et al., 1978, Rev. Elevage et Med. Vet. Pays Trop., n. s., v. 31 (4), 423-426
description
Syncerus caffer: nord de la localite de Bangassou, Empire Centrafricain

Sarcosporidiosis "cisti di probabile natura sarcosporidica"
Cornaglia, E., 1976, Parassitologia, v. 18 (1-3), 19-20
bovina piemontese (cervello)

Sarcosporidiosis
Grossklaus, D., 1979, Zentralbl. Bakteriol., 1. Abt. Orig., Reihe B, v. 168 (1-2), 83-96
zoonoses transmitted via foodstuff, control measures, review

Sarcosporidiosis
Meshkov, S.; and Kotomindov, S., 1977, Vet. Sbirka, v. 75 (1), 34-36
sarcosporidiosis, sheep, meat inspection

Sarcosporidiosis
Sreemannarayana, O.; and Christopher, K. J., 1977, Indian J. Animal Health, v. 16 (2), 188
combined infection of Cysticercus bovis and sarcosporidiasis, bullock: slaughter house, Guntur, Andhra Pradesh, India

Saurocytozoon tupinambi, illus.
Telford, S. R., jr., 1978, Internat. J. Parasitol., v. 8 (2), 133-138
Saurocytozoon tupinambi, study of initial infection in juvenile Tupinambis teguixin provides evidence that schizogonic cycle in circulating cells may occur, but identity of intralymphocytic asexual stages with this species cannot be established due to presence of concurrent infection by small Plasmodium species, if confirmed data would justify removing Saurocytozoon from Leucocytozoidae: Venezuela

Schellackia sp.
Telford, S. R., jr., [1979], J. Parasitol., v. 64 (6), 1978, 1126-1127
Anolis carolinensis
Sceloporus undulatus
all from Florida

Schellackia brygooi n. sp., illus.
Landau, I., 1973, Mem. Mus. National Hist. Nat., Paris, n. s., s. A, Zool., v. 77, 1-62
Coccidiomorpha, life cycles, mechanisms assuring continuation of transmission, thesis
Oplurus sebae (nat. and exper.) (sang): Majunga
O. cyclurus (sang): Tulear; Ihotry
Chamaeleo brevicornis, probablement aussi (sang)
Culex pipiens fatigans (exper.)
C. p. pipiens (exper.)

Schizotrypanum Chagas, 1909 emend. Noeller, 1931
Barretto, M. P., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (4), 272-278
subgen. of Trypanosoma, includes: T. (S.) vespertilionis Battaglia, 1904; T. (S.) cruzi Chagas, 1909

Schizotrypanum, subgenus
Godfrey, D. G., 1977, Protozoology, v. 3, 33-49
identification of morphologically similar trypanosomes of mammals

Schizotrypanum [sp.]
Marinkelle, C. J., 1977, J. Wildlife Dis., v. 13 (3), 262-264
Pipistrellus kuhli: Iraq

Schizotrypanum cruzi Chagas, 1909
Pifano C., F.; and Ortiz, I., 1973, Rev. Inst. Nac. Hig., Caracas, v. 6 (1), 41-46
Schizotrypanum cruzi-infected Triatoma capitata found in wild areas close to beaches of northern Venezuela

Schizotrypanum cruzi
Velasquez-Antich, A., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (5), 347-353
Schizotrypanum cruzi, mice, ambilhar

Schizotrypanum hastatus, illus.
Barbosa, W.; Martins, S. P.; and de Oliveira, R. L., 1973, Rev. Patol. Trop., v. 2 (4), 367-376
Schizotrypanum hastatus, morphometric data
Phyllostomus hastatus: Caverna de Fercal, Brasilia, Brasil

Schneideria schneiderae (da Cunha et al., 1975), illus.
da Cunha, A. B.; et al., 1978, Rev. Brasil. Biol., v. 38 (4), 875-884
Schneideria schneiderae, effects of several metabolic inhibitors on penetration of sporozoites into host cells and intracellular development of trophozoites
Trichosia pubescens (ceco intestinal): Mogi das Cruzes

Schneideria schneiderae (da Cunha et al., 1975), illus.
da Cunha, A. B.; et al., 1978, Rev. Brasil. Biol., v. 38 (4), 885-893
Schneideria schneiderae, development in abnormal host Plastosciara sp. (exper.) compared to that in normal host Trichosia pubescens (exper.)

Schneideria schneiderae da Cunha et al., 1975, illus.
da Cunha, A. B.; and Jurand, A., 1978, Arch. Protistenk., v. 120 (3), 233-254
Schneideria schneiderae in Trichosia pubescens (exper.), entry into and development in cells of intestinal caecum, host cell-symbiont interrelations, metabolic exchanges, symbiotic bacteria in cytoplasm of Schneideria, ultrastructural study

Scyphidia sp.
Armas, G., 1979, J. Fish Dis., v. 2 (6), 543-547
Mugil cephalus (body surface): Rio Moche coastal lagoon, northern Peru

Scyphidia adunconucleata
MacKenzie, K.; and Gibson, D. I., 1970, Symposia Brit. Soc. Parasitol., v. 8, 1-42
Pleuronectes platessa (gills): Scotland

Sigmaperistomatus subgen. n.
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139
subgen. of Nyctotheroides
tod of subgen.: N. (S.) crossodactyli (Carini, 1945) comb. n.

Sinuolinea Davis 1917 emend, Laird 1953
Sankurathri, C. S., 1977, Canad. J. Zool., v. 55 (7), 1147-1150
differentiation from Conispora gen. nov. and Myxoproteus

? Solenophrya sp.
Armas, G., 1979, J. Fish Dis., v. 2 (6), 543-547
Mugil cephalus (body surface, gills): Rio Moche coastal lagoon, northern Peru

Sphaeromyxa sabrazesi, illus.
Grasse, P. P.; and Lavette, A., 1978, Compt. Rend. Acad. Sc., Paris, v. 286, s. D, Sc. Nat. (10), 757-759
Sphaeromyxa sabrazesi, 'germinal' cells are complex of three cellular types, observations support independence of Myxosporidia from Protozoa and Metazoa

Sphaerophrya insolita sp. n., illus.
Iankovskii, A. V., 1973, Parazitologiia, Leningrad, v. 7 (5), 401-407
Sphaerophrya insolita sp. n., morphology, life cycle, host specificity, detrimental effects on host
Bursaria truncatella: pond of Starogo Petergrofa; lake Mozhaiskogo near Krasnogo Sela

Sphaerospora carassii
Iskov, M. P., 1979, Gidrobiol. Zhurnal, v. 15 (4), 68-72
parasites causing intensive epizootic disease of food fishes: Kremenchug reservoir

Sphaerospora carassii Kudo, 1919, illus.
Molnar, K., 1979, Acta Vet., Budapest., v. 27 (1-2), 1977-1979, 99-113
Sphaerospora carassii, carp and grasscarp, incidence, pathology, moderately pathogenic, concurrent infection with Trichodina or other infections may depress host resistance and cause fatalities
Cyprinus carpio
Ctenopharyngodon idella
(gills of all): all from pond farms, Hungary

Sphaerospora caspialosae (Dogiel, 1939)
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
Syn.: Mitraspora caspialosae Dogiel, 1939

Sphaerospora caudata Parisi, 1910
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
Syn.: Mitraspora caudata (Parisi)

Sphaerospora cristata Shulman, 1962
Arthur, J. R.; Margolis, L.; and Arai, H. P., 1976, J. Fish. Research Bd. Canada, v. 33 (11), 2489-2499
Lota lota (urinary bladder): Aishihik Lake, Yukon Territory

Sphaerospora cristata Schulman, 1962, illus.
Chernysheva, N. B., 1973, Parazitologiia, Leningrad, v. 7 (6), 485-488
Sphaerospora cristata, plasmodia and spores found in Lota lota (kidneys), life cycle, host age, infection rate increases during winter: lake Vrevo, Leningradsk oblast

Sphaerospora cristata
Chernysheva, N. B., 1975, Izvest. Gosudarstv. Nauchno-Issled. Inst. Ozer. i Rech. Ryb. Khoziaistva, Leningrad, v. 93, 117-120
protozoans of young predatory fish, extent of infection correlated with some environmental factors
[Lota lota]: Vrevo Lake, Leningrad district

Sphaerospora elegans Thelohan
  Lester, R. J. G., 1974, Syesis, v. 7, 195-200
    Gasterosteus aculeatus (urinary tubules): near Vancouver, British Columbia

Sphaerospora irregularis
  MacKenzie, K.; and Gibson, D. I., 1970, Symposia Brit. Soc. Parasitol., v. 8, 1-42
    Pleuronectes platessa (urinary ducts): Scotland

Sphaerospora pectinacia sp. n., illus.
  Bocharova, T. A.; and Donets, Z. C., 1974, Parazitologiia, Leningrad, v. 8 (1), 74-76
    Perca fluviatilis (gall bladder, urinary bladder, kidney, liver, muscles): lake Durnoe (basin of middle Vasiugan river)

Sphaerospora tincae, illus.
  Koerting, W., 1977, Fisch u. Umwelt (4), 37-48
    fish parasites, histopathological changes

Sphenophrya [sp.]-like ciliate, illus.
  Newman, M. W., 1971, Proc. 1970 National Shellfish. Ass., v. 61, 59-63
    Crassostrea virginica (gills): New Haven Harbor, Connecticut

Spinifera subgen. n.
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 29-31
    subgen. of Cepedea
    type sp.: Cepedea (Spinifera) spinifera Metcalf, 1923

Spinifera
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
    subgen. of Cepedea, key

Spironucleus (Hexamita)
  Sebesteny, A., 1979, Lab. Animals, v. 13 (3), 189-191
    intestinal protozoa, successful transmission to and establishment in specific-pathogen-free mice exposed to intestinal contents of infected hamsters

Spironucleus elegans Lavier (1936), illus.
  Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
    description
    Syn.: Octomitus dujardini Dobell 1909 p. p.
    Bombina bombina
    Bombina variegata
    Bufo bufo
    B. viridis
    Pelobates fuscus
    Rana esculenta
    R. temporaria
    R. dalmatina
    Salamandra salamandra
    Triturus vulgaris
    T. alpestris
    T. montandoni
    all from CSSR

Spironucleus (Hexamita) muris, illus.
  Gruber, H. E.; and Osborne, J. W., 1979, Lab. Animals, v. 13 (3), 199-202
    Spironucleus muris, X-irradiated rats, ultrastructural changes in intestinal epithelium, no evidence of phagocytosis by Paneth cells

Spironucleus muris
  Kunstyr, I.; and Ammerpohl, E., 1978, Lab. Animals, v. 12 (2), 95-97
    Spironucleus muris, faecal cysts, resistance to physical and chemical factors tested, data may be useful for control of infection in rodents and for cryopreservation of parasite

Spiroperistomatus subgen. n.
  Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (4), 137-139
    subgen. of Nyctotheroides
    tod of subgen.: N. (S.) macropharyngeus (Bezzenberger, 1904) Amaro & Sena, 1967

Sporozoa
  Stunkard, H. W., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 725-739
    Sporozoa, review: early taxonomic concepts, developmental cycles and taxonomic revisions, host-parasite relations, microsporidian infections of fishes

'Sporozoaire' [of] Henneguy and Thelohan, 1892 [pro parte]
  Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
    as syn. of Thelohania contejeani Henneguy, 1892

'Sporozoaire' [of] Henneguy and Thelohan, 1892 [pro parte]
  Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
    as syn. of Thelohania giardi Henneguy, 1892

Sporozoan parasite, possibly Sarcocystis, illus.
  Clegg, F. G.; Beverley, J. K. A.; and Markson, L. M., 1978, J. Comp. Path., v. 88 (1), 105-114
    suspected Sarcocystis, cattle, clinical disease described, post-mortem findings, histopathology: Leicestershire, England

Steinhausia spraguei n. sp., illus.
  Kalavati, C.; and Narasimhamurti, C. C., [1978], Riv. Parassitol., Roma, v. 38 (2-3), 1977, 271-275
    Sepia elliptica (excretory cells in fluid from renal appendages): landings of offshore fishery station at Visakhapatnam; local fish market

Steinina coptotermi n. sp., illus.
  Kalavati, C.; and Narasimhamurti, C. C., 1978, Proc. Indian Acad. Sc., Sect. B, Animal Sc., v. 87 (12), 397-404
    Coptotermes heimi (midgut, faecal matter): Andhra University at Waltair

Stempellia polyspora (Leger et Hesse, 1921) Weiser, 1961, illus.
  Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135

Stempellia simulii Maurand et Manier, 1968, illus.
  Maurand, J.; and Loubes, C., 1978, Ztschr. Parasitenk., v. 56 (2), 131-146
    microsporidians from simuliid larvae, ultrastructural studies
    Tetisimulium bezzii: region languedocienne (Sud de la France)

Stenoductus carlogoni sp. n., illus.
  Janardanan, K. P.; and Ramachandran, P., 1979, Zool. Anz., Jena, v. 203 (5-6), 392-400
    Stenoductus carlogoni sp. n., systematics, biology and statistical analysis of morphogenesis
    Carlogonus palmatus (midgut, hindgut, faecal pellets): Chorakkulam, Thottada, and Panoor, Cannanore District, Kerala, India

Stenophora
  da Costa, S. C. G., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (5-6), 327-329
    Stenophora spp., types of motility, review

Stenophora juli (Frantzius, 1848) Labbe, 1899, illus.
  da Costa, S. C. G., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (5-6), 327-329
    Stenophora juli, motility

Sycia arachnoidea Devdhar & Gourishankar, 1971
  Devdhar, M. J.; and Amoji, S. D., 1978, Arch. Protistenk., v. 120 (1-2), 182-189
    Opalnia sp. (gut): Shomeshwar and Kalaghatagi (Dharwar District), Karnataka State, India
Sycia arachnoidea Devdhar & Gourishankar, 1971
  Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
    as syn. of Arachnocystis arachnoidea (Devdhar & Gourishankar, 1971) comb. n.

Symbionecta gen. n.
  Iankovskii, A. V., 1973, Parazitologiia, Leningrad, v. 7 (3), 214-219
    Hymenostomatida, Tetrahymenina, Entodiscidae fam. n.
    tod: S. cuenoti comb. n.

Symbionecta cuenoti (Florentin, 1898) comb. n. (tod)
  Iankovskii, A. V., 1973, Parazitologiia, Leningrad, v. 7 (3), 214-219
    Syn.: Cryptochilum cuenoti Florentin, 1898

Syndiomita neglecta Lavier (1936)
  Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
    as syn. of Octomitus neglecta Lavier, 1936

Systenostrema gen. n.
  Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
    Thelohaniidae fam. n.
    key, tod: S. tabani sp. n.

Systenostrema tabani sp. n. (tod), illus.
  Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
    Tabanus lineola (adipose tissue): Orange Lake, Florida, U.S.A.

Taeniocystis legeri Cognetti de Martiis, 1911
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
as syn. of Cognettia legeri (Cognetti de Martiis, 1911) comb. n.

Tetrahymena pro parte
Corliss, J. O.; and Coats, D. W., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 725-739
as syn. of Lambornella Keilin, 1921

Tetrahymena sp.
Corliss, J. O.; Berl, D.; and Laird, M., 1979, Tr. Am. Micr. Soc., v. 98 (4), 587-591
Tetrahymena sp. in Simulium damnosum (haemocoel), potential biocontrol agent: Ivory Coast

Tetrahymenidae
Corliss, J. O.; and Coats, D. W., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 725-739
familial diagnosis emended
ciliated protozoa found associated with culicine mosquitoes in nature, review (infectivity and host resistance, pathogenicity, mode of entry into host, facultative or obligate parasitism?, taxonomic and nomenclatural considerations, potential importance in biological control of mosquitoes)

Tetramastigamoeba n. g.
Singh, B. N.; and Hanumaiah, V., 1977, Protozoology, v. 3, 183-191
Schizopyrenidae
tod: T. hoarei n. sp.

Tetramastigamoeba hoarei n. g., n. sp. (tod), illus.
Singh, B. N.; and Hanumaiah, V., 1977, Protozoology, v. 3, 183-191
Tetramastigamoeba hoarei n. g., n. sp., mice (exper.), mildly pathogenic
garden soil, sewage sludge, Gomti river mud, and mud from Chinhat lake: Lucknow and vicinity of Lucknow

Tetramastix testudinis Grasse (1926)
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
as syn. of Karotomorpha bufonis Dobell (1909)

Theileria
Gutteridge, W. E.; and Coombs, G. H., 1977, Biochemistry of parasitic protozoa, 172 pp., illus.
biochemistry of parasitic protozoa, textbook: methodology; catabolism and generation of energy; nucleic acid metabolism; protein metabolism; lipid metabolism; biochemical mechanism of drug action; isolation of parasitic protozoa from infected animals; culture of parasitic protozoa

Theileria
Irvin, A. D.; Brocklesby, D. W.; and Purnell, R. E., 1979, Vet. Parasitol., v. 5 (1), 17-28
Babesia, Theileria, radiation and isotopic techniques in study and control of piroplasms of cattle, review

Theileria
Krylov, M. V., 1971, Parazitologiia, Leningrad, v. 5 (3), 201-208
Piroplasmida, distribution in different host groups and zoogeographic regions, speculations on phylogeny

Theileria
Krylov, M. V.; and Krylova, N. P., 1972, Parazitologiia, Leningrad, v. 6 (6), 493-505
Piroplasmida, analysis of host specificity

Theileria
Lumsden, W. H. R., 1978, Handb. Exper. Immunol. (Weir), 3. ed., 42.1-42.47
demonstration of antibodies to Protozoa, extensive review

Theileria
Mehlhorn, H.; et al., 1979, Ann. Biol., 4. s., v. 18 (3-4), 97-120
Sarcocystis, Theileria, life cycle, systematic position, pathology, review

Theileria
Smith, R. D., 1977, Interciencia, v. 2 (6), 335-344
current world research on ticks and tick-borne diseases of food producing animals, review

Theileria
van Vorstenbosch, C. J. A. H. V.; Uilenberg, G.; and van Dijk, J. E., 1978, Research Vet. Sc., v. 24 (2), 214-221
Syn.: Haematoxenus Uilenberg, 1964

Theileria, illus.
Young, A. S.; et al., 1978, Ann. Trop. Med. and Parasitol., v. 72 (5), 443-454
Theileria-infected Taurotragus oryx erythrocytes, bar and veil structures associated with parasite, light and electron microscopy, possible functions

Theileria [sp.]
Butabaeva, M., 1979, Uzbek. Biol. Zhurnal (2), 50-53
[Bos taurus]: Kashkadar'insk oblast

Theileria sp.
Han, T. W., 1978, Research Rep., Office Rural Develop., Min. Agric. and Fish., Korea, v. 20, 53-88
theileriosis, cattle, historical review, seasonal and host age incidence, duration of parasitaemia, relapse time; transmission of Theileria sp. to cattle using Boophilus microplus and Haemaphysalis longicornis; pathogenicity and immunogenicity of Korean and Japanese strains of T. sergenti compared: Korea

Theileria spp. (Korean strain)
Jeon, Y.; et al., 1977, Research Rep., Office Rural Develop., Min. Agric. and Fish., Korea (Vet. and Sericult.), v. 19, 27-32
Theileria spp., Korean strain, cattle, serological diagnosis, indirect fluorescent antibody technique

Theileria spp.
Jeon, Y.; et al., 1978, Research Rep., Office Rural Develop., Min. Agric. and Fish., Korea, v. 20, 1-3
cattle: abattoir in Seoul, Korea

Theileria [sp.]
Prabhakar, K. S.; and Hiregoudar, L. S., 1977, Indian J. Animal Sc., v. 47 (12), 848-850
Theileria [sp.], prevalence in sheep (blood): Karnataka

Theileria sp. (probably T. mutans)
Tustin, R. C.; and van Heerden, J., 1979, J. South African Vet. Ass., v. 50 (1), 49-51
Theileria sp., bovine, clinical signs, lesions in spinal cord and meninges, histopathology: Rietgat area of Pretoria district, South Africa

Theileria sp. (Githunguri)
Uilenberg, G.; et al., 1977, Tropenmed. u. Parasitol., v. 28 (4), 494-498
Theileria spp. (Idobogo and Mwanza), cattle (nat. and exper.); transmission by Rhipicephalus appendiculatus (exper.); clinical, parasitological, and serological observations; susceptibility of recovered animals to T. parva; similarity to Theileria sp. (Githunguri); may confuse studies on T. parva and T. mutans: Tanzania

Theileria sp. (Idobogo)
Uilenberg, G.; et al., 1977, Tropenmed. u. Parasitol., v. 28 (4), 494-498
Theileria spp. (Idobogo and Mwanza), cattle (nat. and exper.); transmission by Rhipicephalus appendiculatus (exper.); clinical, parasitological, and serological observations; susceptibility of recovered animals to T. parva; similarity to Theileria sp. (Githunguri); may confuse studies on T. parva and T. mutans: Tanzania

Theileria sp. (Mwanza)
Uilenberg, G.; et al., 1977, Tropenmed. u. Parasitol., v. 28 (4), 494-498
Theileria spp. (Idobogo and Mwanza), cattle (nat. and exper.); transmission by Rhipicephalus appendiculatus (exper.); clinical, parasitological, and serological observations; susceptibility of recovered animals to T. parva; similarity to Theileria sp. (Githunguri); may confuse studies on T. parva and T. mutans: Tanzania

Theileria-like macroschizonts
Young, A. S.; et al., 1978, Tropenmed. u. Parasitol., v. 29 (3), 281-288
theilerial parasites, incidence in Syncerus caffer, role as reservoir of cattle pathogenic theilerioses in East Africa

Theileria-like piroplasms
Young, A. S.; et al., 1978, Tropenmed. u. Parasitol., v. 29 (3), 281-288
theilerial parasites, incidence in Syncerus caffer, role as reservoir of cattle pathogenic theilerioses in East Africa

Theileria sp., illus.
Young, A. S.; Burridge, M. J.; and Payne, R. C., 1977, Trop. Animal Health and Prod., v. 9 (1), 37-34
Theileria sp. transmitted to Bos taurus (exper.) by Amblyomma cohaerens collected from Syncerus caffer, macroschizonts resemble T. barnetti but exper. host reacted serologically to T. mutans antigen only, description, blood transmission to splenectomized calf: Mara region, Kenya

Theileria annulata
Baharsefat, M.; et al., 1977, Arch. Inst. Ra
(29), 47-58
Theileria annulata, calves, unusual lesions
gross and histopathological changes

Theileria annulata
Bulman, G. M.; Arzo, G. M.; and Nassimi, M.
1979, Trop. Animal Health and Prod., v. 11 (1), 17-20
Theileria annulata, outbreak in Holstein bulls, history and clinical observations, diagnosis, treatment, infestation with Hyalomma anatolicum anatolicum: Kabul, Afghanistan

Theileria annulata
Deore, P. A.; Sabnis, M. G.; and Bendre, V. U
1979, Indian Vet. J., v. 56 (9), 794-795
Theileria annulata, bovine, atypical cutaneous case

Theileria annulata
Dhar, S.; and Gautam, O. P., 1978, Indian Vet.
J., v. 55 (9), 738-740
Theileria annulata, calves (exper.), hyperimmune serum inoculated, no therapeutic value, did not affect course of disease

Theileria annulata, illus.
Dhar, S.; and Gautam, O. P., 1979, Indian J. Animal Sc., v. 49 (2), 122-126
Theileria annulata, calves (exper.), anemia
produced by excessive removal of erythrocytes
from circulation by phagocytes, involvement
of auto-immune mechanism

Theileria annulata
Dhar, S.; and Gautam, O. P., 1979, Indian J. Animal Sc., v. 49 (7), 511-516
Theileria annulata, changes in serum proteins in cattle (exper.) with acute and chronic infections

Theileria annulata
El-Abdin, Y. Z.; Hamza, S. M.; and El Refaii, A
H., 1977, Egypt. J. Vet. Sc., v. 13 (2), 1976, 77-84
Theileria annulata, cattle, imidocarb dihydrochloride, serum enzyme activities and chemical constituents before and after treatment

Theileria annulata
El Refaii, A. H.; and Michael, S. A., 1979, Vet. Rec., v. 105 (3), 60
urine retention in Egyptian bulls possibly caused by or linked with subclinical Theileria annulata infection, imidocarb dipropionate treatment

Theileria annulata, illus.
Erturk, E.; Laiblin, C.; and Urman, H. K., 1976, Vet. Fak. Dergisi, Ankara Univ., v. 23 (3-4), 352-367
Theileria annulata, calves (exper.), clinicopathologic study

Theileria annulata
Gill, B. S.; et al., 1978, Internat. J. Parasitol., v. 8 (6), 467-469
Theileria annulata, calves, immunization by treating tick (Hyalomma anatolicum anatolicum) stabilate-induced infections with 1 or 2 doses of long-acting oxytetracycline vs. 8 doses of chlortetracycline

Theileria annulata
Goldman, M.; and Pipano, E., 1978, Tropenmed. u. Parasitol., v. 29 (1), 85-87
Theileria annulata, specific IgM and IgG antibodies detected in immunized or infected cattle

Theileria annulata
Halik, J., 1977, Veterinarstvi, v. 27 (10), 469-471
Babesia, Theileria, Anaplasma, diagnostic problems, review: North Africa

Theileria annulata
Hashemi-Fesharki, R., 1978, Arch. Inst. Razi (30), 91-99
Theileria annulata, 3 strains of varying virulence, calves (exper.), primary infections with different doses, parasitological findings, host temperature, resistance to challenge with homologous and heterologous strains

Theileria annulata
Jagdish, S.; et al., 1979, Vet. Rec., v. 104 (7), 140-142
Theileria annulata, immunising infection in calves by injecting ground up infected Hyalomma anatolicum anatolicum supernate, severity of reactions in rolitetracycline-treated vs. non-treated calves compared, adequate protection, durable immunity to subsequent severe homologous challenge

Theileria annulata Dschunkowsky et Luhs, 1904, illus.
Kolabskii, N. A., 1978, [Theileriasis of animals], 2. ed., 199 pp., illus.
Theileria spp. of domestic animals, economic importance, systematics, life cycle, pathology, diagnosis, therapy, prophylaxis, review

Theileria annulata
Laiblin, C., 1978, Berl. u. Munchen. Tierarztl. Wchnschr., v. 91 (3), 48-50
Theileria annulata, cattle (exper.), haematological study, toxic lesion of bone marrow caused aplastic anaemia

Theileria annulata
Laiblin, C.; Baysu, N.; and Mueller, M., 1978, Berl. u. Munchen. Tierarztl. Wchnschr., v. 91 (2), 25-27
Theileria annulata, cattle (exper.), extent of liver and kidney damage estimated by enzyme activity, bilirubin, and urea levels; there was no difference between cattle receiving liverprotecting medication and those that did not

Theileria annulata
Levchenko, F. F., 1978, Veterinariia, Moskva (7), 62-64
Theileria annulata, cattle, chinocide and bigumal treatment effective, treatment with hemostimulin and vitamins and microelements given in feed to counteract anemia and atonicity of digestive tract resulting from therapy: Gissarsk valley

Theileria annulata
McHardy, N., 1978, Ann. Trop. Med. and Parasitol., v. 72 (6), 501-511
Theileria parva- and T. annulata-infected bovine lymphoblastoid cell cultures used in in vitro screens to test wide range of compounds for chemotherapeutic activity

Theileria annulata
Melrose, T. R.; and Brown, C. G. D., 1979, Research Vet. Sc., v. 27 (3), 379-381
piroplasms isolated from bovine blood infected with Theileria annulata and T. parva, isoenzyme variation

Theileria annulata
Purnell, R. E., 1978, Vet. Sc. Commun., v. 2 (1), 3-10
Theileria annulata, distribution, tick vectors, suggested control methods, review: countries on northern Mediterranean littoral

Theileria annulata (Dchunkowsky and Luhs, 1904)
Samish, M.; and Pipano, E., 1978, Parasitology, v. 77 (3), 375-379
time-course for development of infectivity in Hyalomma detritum ticks fed as pre-imagoes on Theileria annulata-infected calves, unfed adults derived from infected nymphs were non-infectious whereas ticks of both sexes that had fed for 2-3 days or longer on calves or rabbits were always infectious

Theileria annulata (Dschunkowsky und Luhs, 1904), illus.
Schein, E.; and Friedhoff, K. T., 1978, Ztschr. Parasitenk., v. 56 (3), 287-303
Theileria annulata, development in haemolymph and salivary glands of Hyalomma anatolicum excavatum, light microscopy; hypothetical diagram of cycle in ticks

Theileria annulata
Singh, D. K.; et al., 1979, Trop. Animal Health and Prod., v. 11 (2), 87-90
infectivity of ground-up tick supernates prepared from Theileria annulata-infected Hyalomma a. anatolicum adults prefed for variable periods of time

Theileria annulata
Singh, D. K.; Jagadish, S.; and Gautam, O. P., 1979, Am. J. Vet. Research, v. 40 (6), 767-769
Theileria annulata, Bos indicus x Bos taurus, immunization using irradiated infective particles derived from Hyalomma a. anatolicum

Theileria annulata
Srivastava, P. S.; et al., 1976, Acta Vet. Brno, v. 45 (4), 245-250
Theileria annulata, cross-bred calves (exper.), histopathology

Theileria annulata
Srivastava, P. S.; and Sharma, N. N., 1978, Vet. Parasitol., v. 4 (1), 83-89
Theileria annulata-infected nymphs, adults, and ground tissues of Hyalomma anatolicum, infectivity for calves, all capable of inducing clinical theileriasis

Theileria annulata
Stepanova, N. I.; et al., 1977, Veterinariia, Moskva (3), 69-70
Theileria annulata, cattle, successful testing under farm conditions of live vaccine prepared from cell culture of parasite

Theileria annulata, illus.
Walker, A. R.; et al., 1979, Trop. Animal Health and Prod., v. 11 (1), 21-26
Theileria spp., rapid quantitative assessment of infection in ticks, simplified method for methyl green pyronin staining of salivary glands

Theileria annulata, illus.
Weber, G., 1978, Naturwissenschaften, v. 65 (11), 601-602
Theileria annulata, atypical mitochondria identified by ultracytochemical demonstration of mitochondrial marker enzymes, presence of both succinic dehydrogenase and cytochrome oxidase activity suggests that respiratory chain is operative in sporozoites

Theileria annulata, illus.
Weber, G.; and Friedhoff, K. T., 1977, Ztschr. Parasitenk., v. 53 (1), 83-92
Babesia bigemina, ultrastructure of supposed sexual stages from gut of Boophilus microplus, comparison with developmental stage of Theileria annulata

Theileria barnetti
Young, A. S.; Burridge, M. J.; and Payne, R. C., 1977, Trop. Animal Health and Prod., v. 9 (1), 37-34
Theileria sp. transmitted to Bos taurus (exper.) by Amblyomma cohaerens collected from Syncerus caffer, macroschizonts resemble T. barnetti but exper. host reacted serologically to T. mutans antigen only, description, blood transmission to splenectomized calf: Mara region, Kenya

Theileria dispar Sergent, et al., 1924, illus.
Kolabskii, N. A., 1978, [Theileriasis of animals], 2. ed., 199 pp., illus.
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Theileria hirci
Prabhakar, K. S.; and Hiregoudar, L. S., 1979, Gujarat Agric. Univ. Research J., v. 4 (2), 57-58
Theileria hirci, carrier sheep, parasite counts and haematological observations before and after splenectomy

Theileria lawrencei
Chambers, P. G., 1978, Rhodesian Vet. J., v. 9 (1-4), 40
protozoans, icteric cattle carcases, routine examination of blood and spleen smears: Sinoia abattoir, Rhodesia

Theileria lawrencei
Loehr, K. F., 1977, 3. Externe Veterinartagung, Ber. u. Arbeitsergebn. (40), 235-247
Theileria lawrencei, T. parva, cattle, epizootiology, diagnosis, immunity, prophylaxis, therapy: East Africa

Theileria lawrencei, illus.
Nyindo, M. B. A.; et al., 1978, Am. J. Vet. Research, v. 39 (1), 37-44
Theileria parva, T. lawrencei, cultivation of cell-free schizonts and merozoites in vitro, immunogenicity in cattle inoculated with T. parva merozoites and schizonts and later challenged

Theileria lawrencei
Radley, D. E.; et al., 1979, Vet. Parasitol., v. 5 (2-3), 117-128
Theileria lawrencei, immunization of Bos taurus by infection (with single and multiple Theileria spp. isolates) and chemoprophylaxis (long-acting oxytetracycline)

Theileria lawrencei, illus.
Young, A. S.; et al., 1978, Tropenmed. u. Parasitol., v. 29 (3), 281-288
theilerial parasites, incidence in Syncerus caffer, role as reservoir of cattle pathogenic theilerioses in East Africa

Theileria mutans, illus.
Guler, S., 1978, Vet. Fak. Dergisi, Ankara Univ., v. 25 (2), 323-338
Theileria mutans, cattle, occurrence, experimental infection of ticks successful only with Boophilus annulatus, possible vector: Turkey

Theileria mutans
Joyner, L. P.; et al., 1979, Research Vet. Sc., v. 26 (3), 387-388
British Theileria mutans, Japanese T. sergenti, serological comparison showed that the two parasites were indistinguishable, name of British parasite should be changed to T. sergenti

Theileria mutans Theiler, 1906, illus.
Kolabskii, N. A., 1978, [Theileriasis of animals], 2. ed., 199 pp., illus.
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Perie, N. M.; Uilenberg, G.; and Schreuder, B. E. C., 1979, Research Vet. Sc., v. 26 (3), 359-362
Theileria sp. (Zaria) and Theileria sp. (Katsina) shown to be T. mutans: Nigeria

Theileria mutans
Uilenberg, G.; et al., 1977, Tropenmed. u. Parasitol., v. 28 (4), 494-498
Theileria spp. (Idobogo and Mwanza), cattle (nat. and exper.); transmission by Rhipicephalus appendiculatus (exper.); clinical, parasitological, and serological observations; susceptibility of recovered animals to T. parva; similarity to Theileria sp. (Githunguri); may confuse studies on T. parva and T. mutans: Tanzania

Theileria mutans
Uilenberg, G.; et al., 1977, Tropenmed. u. Parasitol., v. 28 (4), 499-506
Theileria parva, T. mutans, cattle, immunization, large-scale field trial: Tanzania

Theileria mutans
Young, A. S., 1977, Tropenmed. u. Parasitol., v. 28 (4), 521-527
Theileria mutans, infectivity for cattle of parasites derived from prefed Amblyomma variegatum nymphs

Theileria mutans, illus.
Young, A. S.; et al., 1978, Parasitology, v. 76 (1), 99-115
Theileria mutans isolated from cattle exposed in Narok District of Kenya, transstadially transmissible by Amblyomma variegatum but not by Rhipicephalus appendiculatus, mechanically transmissible by blood containing piroplasms or lymphoid cells infected with schizonts, course of infection, pathogenicity, and morphology in cattle (exper.)

Theileria mutans
Young, A. S.; et al., 1978, Tropenmed. u. Parasitol., v. 29 (3), 281-288
theilerial parasites, incidence in Syncerus caffer, role as reservoir of cattle pathogenic theilerioses in East Africa

Theileria mutans
Young, A. S.; Burridge, M. J.; and Payne, R. C., 1977, Trop. Animal Health and Prod., v. 9 (1), 37-34
Theileria sp. transmitted to Bos taurus (exper.) by Amblyomma cohaerens collected from Syncerus caffer, macroschizonts resemble T. barnetti but exper. host reacted serologically to T. mutans antigen only, description, blood transmission to splenectomized calf: Mara region, Kenya

Theileria orientalis Yakimoff et Soudatschenkoff, 1931, illus.
Kolabskii, N. A., 1978, [Theileriasis of animals], 2. ed., 199 pp., illus.
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Theileria ovis Rodhain, 1916, illus.
Kolabskii, N. A., 1978, [Theileriasis of animals], 2. ed., 199 pp., illus.
Theileria spp. of domestic animals, economic importance, systematics, life cycle, pathology, diagnosis, therapy, prophylaxis, review

Theileria ovis Rodhain, 1916, illus.
Mehlhorn, H.; Schein, E.; and Warnecke, M., 1979, J. Protozool., v. 26 (3), 377-385
Theileria ovis, gamogony and sporogony in Rhipicephalus evertsi evertsi, electron-microscopic studies

Theileria parva
Aruo, S. K., 1977, Bull. Animal Health and Prod. Africa, v. 25 (3), 223-227
Theileria parva, European breeds of cattle, clinical and pathological picture of East Coast fever: Uganda

Theileria parva
Brown, C. G. D.; et al., 1977, Tropenmed. u. Parasitol., v. 28 (4), 513-520
Theileria parva, Bos taurus with patent East Coast fever induced by inoculation of cryopreserved stabilates of tick-derived infective particles, preliminary evaluation of n-pyrrolidinomethyl tetracycline and oxytetracycline

Theileria parva
Brown, C. G. D.; et al., 1978, Exper. Parasitol., v. 45 (1), 55-64
Theileria parva, experiments support hypothesis that the parasite (either as an infective particle derived from Rhipicephalus appendiculatus or as a macroschizont) may become closely associated with a member of the leukocyte series and that, on inoculation of such material into cattle, subsequent development to produce East Coast fever is dependent on acceptance of these cells by the recipient

Theileria parva
Duffus, W. P. H.; et al., 1978, Infect. and Immun., v. 22 (2), 492-501
Theileria parva, Trypanosoma rhodesiense, ^{51}Cr-labeled chicken erythrocytes coated with protozoal antigens form suitable targets for bovine antibody-dependent cell-mediated cytotoxicity assays

Theileria parva
Duffus, W. P. H.; Wagner, G. G.; and Preston, J. M., 1978, Clin. and Exper. Immunol., v. 34 (3), 347-353
Theileria parva-infected bovine lymphoid cell culture line (C2), no specific parasite antigen detected on C2 cell surface, no detectable surface immunoglobulin or secretion of immunoglobulins into tissue culture medium, C2 cells found to share membrane antigen with normal calf thymus cells and to possess strong transplantation antigen, C2 cells can act as weak stimulators in mixed leucocyte reaction

Theileria parva, probably
Giles, N.; et al., 1978, Vet. Rec., v. 102 (14), 313
Theileria parva, Masai cattle, clinical signs, serological tests: Loita Hills near Narosura, Kenya

Theileria parva, illus.
Irvin, A. D.; et al., 1977, Tropenmed. u. Parasitol., v. 28 (4), 507-512
Theileria parva, mice, piroplasm infection of erythrocytes following intraperitoneal inoculation with irradiated cultures of infected bovine lymphoid cells, infection could not be transmitted to cattle using Rhipicephalus appendiculatus

Theileria parva
Kimeto, B. A., 1978, Am. J. Vet. Research, v. 39 (7), 1117-1122
Theileria parva transmitted by Rhipicephalus appendiculatus to calves (exper.), histopathologic and electron microscopic studies of cutaneous lesions

Theileria parva Theiler, 1904, illus.
Kolabskii, N. A., 1978, [Theileriasis of animals], 2. ed., 199 pp., illus.
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Loehr, K. F., 1977, 3. Externe Veterinartagung, Ber. u. Arbeitsergebn. (40), 235-247
Theileria lawrencei, T. parva, cattle, epizootiology, diagnosis, immunity, prophylaxis, therapy: East Africa

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Maathai, W. M. J. M., 1977, Bull. Animal Health and Prod. Africa, v. 25 (4), 415-419
Rhipicephalus appendiculatus, adult unfed ticks fed as nymphs on animals suffering from Theileria parva, fine structure of uninfected and infected salivary cell types (c) and (d)

Theileria parva, illus.
McHardy, N., 1978, Ann. Trop. Med. and Parasitol., v. 72 (6), 501-511
Theileria parva- and T. annulata-infected bovine lymphoblastoid cell cultures used in in vitro screens to test wide range of compounds for chemotherapeutic activity

Theileria parva
Mchinja, S. J.; Young, A. S.; and Omwoyo, P., 1975, Bull. Animal Health and Prod. Africa, v. 25 (2), 158-161
Theileria parva, inoculation of oxytetracycline into rabbits on which infected Rhipicephalus appendiculatus are feeding did not inhibit development of parasites nor affect their subsequent infectivity for cattle, possible use of this system in screening prophylactic drugs against East Coast fever

Theileria parva
Melrose, T. R.; and Brown, C. G. D., 1979, Research Vet. Sc., v. 27 (3), 379-381
piroplasms isolated from bovine blood infected with Theileria annulata and T. parva, isoenzyme variation

Theileria parva, illus.
Nyindo, M. B. A.; et al., 1978, Am. J. Vet. Research, v. 39 (1), 37-44
Theileria parva, T. lawrencei, cultivation of cell-free schizonts and merozoites in vitro, immunogenicity in cattle inoculated with T. parva merozoites and schizonts and later challenged

Theileria parva
Oteng, A. K., 1977, Bull. Animal Health and Prod. Africa, v. 25 (2), 126-131
Theileria parva, cross-immunity trials in recovered cattle indicate that Ugandan isolates differ immunologically from T. parva (Muguga) and among themselves and that several strains and/or species are responsible for field cases of East Coast fever: Uganda

Theileria parva
Pearson, T. W.; et al., 1979, Nature, London (5733), v. 281, 678-680
cell-mediated immunity to Theileria parva-transformed cell lines

Theileria parva
Radley, D. E.; et al., 1979, Vet. Parasitol., v. 5 (2-3), 117-128
Theileria lawrencei, immunization of Bos taurus by infection (with single and multiple Theileria spp. isolates) and chemoprophylaxis (long-acting oxytetracycline)

Theileria parva (Muguga)
Roelants, G. E.; et al., 1977, Protides Biol. Fluids, v. 25, 743-745
Theileria parva, lymphocyte transformation

Theileria parva
Uilenberg, G.; et al., 1977, Tropenmed. u. Parasitol., v. 28 (4), 494-498
Theileria spp. (Idobogo and Mwanza), cattle (nat. and exper.); transmission by Rhipicephalus appendiculatus (exper.); clinical, parasitological, and serological observations; susceptibility of recovered animals to T. parva; similarity to Theileria sp. (Githunguri); may confuse studies on T. parva and T. mutans: Tanzania

Theileria parva
Uilenberg, G.; et al., 1977, Tropenmed. u. Parasitol., v. 28 (4), 499-506
Theileria parva, T. mutans, cattle, immunization, large-scale field trial: Tanzania

Theileria parva
Uilenberg, G.; and Zwart, D., 1979, Research Vet. Sc., v. 26 (2), 243-245
Theileria parva, calves (exper.) with advanced East Coast fever, occurrence of skin nodules with schizonts and/or skin nodules with Demodex bovis

Theileria parva
Walker, A. R.; et al., 1979, Research Vet. Sc., v. 26 (2), 264-265
methods for infecting Rhipicephalus appendiculatus ticks with Theileria parva by injection vs. by artificial feeding

Theileria parva, illus.
Walker, A. R.; et al., 1979, Trop. Animal Health and Prod., v. 11 (1), 21-26
Theileria spp., rapid quantitative assessment of infection in ticks, simplified method for methyl green pyronin staining of salivary glands

Theileria parva
Young, A. S.; Leitch, B. L.; and Omwoyo, P. L., 1979, Vet. Rec., v. 105 (23), 531-533
Theileria parva, induction of infective stages in salivary glands of infected unfed Rhipicephalus appendiculatus by exposure of ticks to high temperature, epidemiological significance

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Charyev, O. Ch., 1977, Izvest. Akad. Nauk Turkmen. SSR, s. Biol. Nauk (6), 70-71
piroplasmids, sheep, pure and mixed infections: Turkmenia

Theileria recondita Lestoquard, 1929
Kolabskii, N. A., 1978, [Theileriasis of animals], 2. ed., 199 pp., illus.
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van Vorstenbosch, C. J. A. H. V.; Uilenberg, G.; and van Dijk, J. E., 1978, Research Vet. Sc., v. 24 (2), 214-221
Syn.: Haematoxenus separatus Uilenberg and Andreasen, 1974

Theileria sergenti
Han, T. W., 1978, Research Rep., Office Rural Develop., Min. Agric. and Fish., Korea, v. 20, 53-88
theileriosis, cattle, historical review, seasonal and host age incidence, duration of parasitaemia, relapse time; transmission of Theileria sp. to cattle using Boophilus microplus and Haemaphysalis longicornis; pathogenicity and immunogenicity of Korean and Japanese strains of T. sergenti compared: Korea

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Hayashi, T.; et al., 1978, Bull. Fac. Agric. Tottori Univ., v. 30, 82-88
Theileria sergenti, grazing cattle, administration of pamaquine probably causes blood coagulation disorders

heileria sergenti
Joyner, L. P.; et al., 1979, Research Vet. Sc., v. 26 (3), 387-388
British Theileria mutans, Japanese T. sergenti, serological comparison showed that the two parasites were indistinguishable, name of British parasite should be changed to T. sergenti

neileria sergenti Yakimoff et Dekhtereff, 930, illus.
Kolabskii, N. A., 1978, [Theileriasis of animals], 2. ed., 199 pp., illus.
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neileria sergenti
Minami, T., 1977, Japan Agric. Research Quart., v. 11 (4), 234-238
Japanese Babesia sp., cattle (exper.), capillary-tube agglutination test, discrimination from B. bigemina and B. argentina was almost possible, no cross reactions with other protozoa

[heileria] sergenti
Rashidov, A. A.; and Elagin, V. I., 1978, Veterinariia, Moskva (6), 65-66
T[heileria] sergenti, splenectomized cattle, course of infection, clinical aspects; failure to transmit by Rhipicephalus bursa: Dagestan

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Kolabskii, N. A., 1978, [Theileriasis of animals], 2. ed., 199 pp., illus.
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Grootenhuis, J. G.; et al., 1979, Research Vet. Sc., v. 27 (1), 59-68
Theileria taurotragi stabilate from eland was infective for eland and for Bos taurus, course of infections, low pathogenicity, response of recovered animals to homologous and heterologous challenge, serology, tick transmission between cattle, establishment of infected lymphoblastoid cell lines; morphology of schizonts

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van Vorstenbosch, C. J. A. H. V.; Uilenberg, G.; and van Dijk, J. E., 1978, Research Vet. Sc., v. 24 (2), 214-221
Theileria velifera, ultrastructure of enigmatic veil using light and electron microscopy
Syn.: Haematoxenus veliferus Uilenberg, 1964
Amblyomma lepidum
A. hebraeum

eileria velifera
Berger, J., 1979, J. South African Vet. Ass., v. 50 (1), 45-46
case history, review
steers (nat. and exper.) (blood): farm Gulu near East London, Eastern Cape Province

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Avessalomov, I. S.; and Baenov, N., 1977, Veterinariia, Moskva (8), 76
theileriasis, cattle, delagil combined with vitamins and penicillin, effective and nontoxic

Theileriasis
Berggren, S. A., 1978, Vet. Parasitol., v. 4 (3), 289-297
cattle tick survey, 15 spp. found, concluded that establishment of Rhipicephalus appendiculatus makes situation potentially dangerous with regard to transmission of theileriasis: Malawi

Theileriasis
Gautam, O. P., 1977, Haryana Vet., v. 15 (2), 1976, 77-87
theileriasis, cattle, review: India

Theileriasis
Han, T. W., 1978, Research Rep., Office Rural Develop., Min. Agric. and Fish., Korea, v. 20, 53-88
theileriosis, cattle, historical review, seasonal and host age incidence, duration of parasitaemia, relapse time; transmission of Theileria sp. to cattle using Boophilus microplus and Haemaphysalis longicornis; pathogenicity and immunogenicity of Korean and Japanese strains of T. sergenti compared: Korea

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Kumar, B. J.; and Gowda, D. K. V., 1978, Indian Vet. J., v. 55 (9), 729-730
theileriasis, dairy cow treated with chloroquine and oxytetracycline, chloroquine toxicity causing corneal opacity and possibly abortion

Theileriasis
Norval, R. A. I., 1978, Rhodesian Vet. J., v. 9 (1-4), 9-16
ticks, cattle, significant increase in tick infestations and outbreaks of tick-borne diseases following collapse of dipping (1973-1978): African areas in Rhodesia

Theileriasis
Sharma, M. M.; et al., 1979, Indian Vet. Med. J., v. 3 (3), 187-191
theileriasis, Jersey cattle, incidence and treatment with berenil, babesan, nevaquine, and aureomycin: Exotic Nucleus Cattle Farm, Bassi, Jaipur

Theileriasis
Vecherkin, S. S.; et al., 1977, Veterinariia, Moskva (10), 77-78
theileriasis, cattle, Peganum harmala alkaloids effective, no harmful or cumulative effects

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Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
parasites of fishes: China

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parasites of fishes: China

Thelohanellus oculi-leucisci, illus.
Podlipaev, S. A., 1972, Parazitologiia, Leningrad, v. 6 (6), 506-508
Myxosporidia, 6 spp., lipid inclusions in spores and cysts

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Podlipaev, S. A., 1974, Parazitologiia, Leningrad, v. 8 (6), 535-542
Myxobolidae spp., polysaccharides in spores, nature of iodinophilous vacuole

Thelohanellus pyriformis
Iskov, M. P., 1979, Gidrobiol. Zhurnal, v. 15 (4), 68-72
parasites causing intensive epizootic disease of food fishes: Kremenchug reservoir

Thelohania
Grobov, O. F.; Karakuiumchian, M. K.; and Orlova-Sokol'skaia, I. A., 1975, Parazitologiia, Leningrad, v. 9 (2), 142-146
review of microsporidians reported from mammals

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Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Thelohaniidae fam. n., key

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Johnston, L. B.; Vernick, S. H.; and Sprague, V., 1978, J. Invert. Path., v. 32 (3), 278-290
"It is concluded that sporogony by means of three binary divisions and lack of a sporogonial plasmodium may be essential characters of the genus Thelohania Henneguy, 1892."

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Alikhanov, Sh. G., 1972, Parazitologiia, Leningrad, v. 6 (4), 381-384
epizootiology
Aedes caspius caspius: Azerbaidzhan

Thelohania sp. Hilsenhoff et Lowett, 1966, illus.
Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135

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Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora californica (Kellen and Lipa, 1960) comb. n.

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Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
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Thelohania sp. Chapman et al., 1966
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora canadensis (Wills and Beaudoin, 1965) comb. n.

Thelohania sp. Chapman et al., 1967
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora minuta (Kudo, 1924) comb. n.

Thelohania sp. Chapman et al., 1967
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora opacita (Kudo, 1922) comb. n.

Thelohania sp. Chapman et al., 1969
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Chapmanium cirritus sp. n.

Thelohania sp. Hazard and Weiser, 1968
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Parathelohania periculosa (Kellen and Wills, 1962) Hazard and Anthony, 1974

Thelohania sp. Noeller, 1920 [et auct.]
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora khaliulini sp. n.

Thelohania sp. Vey and Vago, 1972
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Thelohania contejeani Henneguy, 1892

Thelohania [sp.] or Plistophora [sp.], illus.
Herman, R. L.; and Putz, R. E., 1970, J. Wildlife Dis., v. 6 (3), 173
Ictalurus punctatus (ventricle of heart, intestinal sub-mucosa): Maryland

Thelohania sp. Vernick, Sprague, & Krause, 1977
Johnston, L. B.; Vernick, S. H.; and Sprague, V., 1978, J. Invert. Path., v. 32 (3), 278-290
as syn. of Thelohania butleri n. sp.

Thelohania [sp.]
Khaliulin, G. L., 1973, Parazitologiia, Leningrad, v. 7 (4), 370-373
Thelohania spp. in Aedes spp. larvae, infection rate under natural conditions and in tests with transovarial transmission did not exceed 2%, infection rate in tests with transmission per os reached 50%
Aedes diantaeus: Mariisk ASSR
A. cataphylla: Mariisk ASSR
A. communis: Chuvashsk ASSR
A. flavescens: Chuvashsk ASSR

Thelohania sp., illus.
Khaliulin, G. L., 1973, Parazitologiia, Leningrad, v. 7 (2), 180-182
description
Aedes excrucians (fat body, hemolymph): Mariisk ASSR

Thelohania sp.
Khaliulin, G. L., 1974, Parazitologiia, Leningrad, v. 8 (2), 183-184
Aedes annulipes: vicinity of Raifa, Volzhsk-Kamsk national forest, Tatar ASSR

Thelohania sp.
Kilochitskii, P. Ia., 1977, Vestnik Zool., Akad. Nauk Ukrainsk. SSR, Inst. Zool. (4), 71-75
Aedes caspius caspius: southern Ukraine

Thelohania sp., illus.
Loubes, C., 1979, J. Protozool., v. 26 (2), 200-208
Microsporida, synaptonemal complexes demonstrated in 6 genera but not in Nosema, implications for life cycles
Cricotopus sp.: environs de Montpellier

Thelohania sp. A
Tsai, Y. H.; Grundmann, A. W.; and Rees, D. M., 1969, Mosquito News, v. 29 (1), 102-110
parasites of mosquitoes, effect of mosquito control on incidence of larval parasitism
Culex tarsalis: northern Utah

helohania sp. B
Tsai, Y. H.; Grundmann, A. W.; and Rees, D. M., 1969, Mosquito News, v. 29 (1), 102-110
parasites of mosquitoes, effect of mosquito control on incidence of larval parasitism
Culiseta inornata: northern Utah

helohania sp. C
Tsai, Y. H.; Grundmann, A. W.; and Rees, D. M., 1969, Mosquito News, v. 29 (1), 102-110
parasites of mosquitoes, effect of mosquito control on incidence of larval parasitism
Aedes dorsalis: northern Utah

helohania sp. D
Tsai, Y. H.; Grundmann, A. W.; and Rees, D. M., 1969, Mosquito News, v. 29 (1), 102-110
parasites of mosquitoes, effect of mosquito control on incidence of larval parasitism
Culiseta impatiens: southwestern Wyoming

helohania acuta (Moniez, 1887) Schroeder, 1914
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

helohania apodemi Doby et al., 1963
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

helohania argyresthiae Issi and Lipa, 1968
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

helohania asterias Weiser, 1963, illus.
Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135

helohania asterias Weiser, 1963
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

helohania baetica Kudo, 1923
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania barbata Weiser, 1969 (nomen nudum)
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

elohania barra Pillai, 1968
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Parathelohania barra (Pillai, 1968) comb. n.

helohania baueri Voronin, 1974
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

helohania benigna Kellen and Wills, 1962
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora benigna (Kellen and Wills, 1962) comb. n.

Thelohania bicortex Baudoin, 1969
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora bicortex (Baudoin, 1969) comb. n.

Thelohania bicortex Baudoin (in part), 1969
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora trichostegiae (Baudoin, 1969) comb. n.

Thelohania bolinasae Kellen and Wills, 1962
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora bolinasae (Kellen and Wills, 1962) comb. n.

Thelohania bracteata: Debaisieux and Gastaldi (in part), 1919 [et auct.]
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora bracteata (Strickland, 1913) comb. n.

Thelohania bracteata (Stickland, 1913), illus.
Maurand, J.; and Loubes, C., 1978, Ztschr. Parasitenk., v. 56 (2), 131-146
microsporidians from simuliid larvae, ultrastructural studies
simuliid larvae: region languedocienne (Sud de la France)

Thelohania brasiliensis Kudo, 1924 (nomen nudum)
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania breindli Weiser, 1946, illus.
Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135

Thelohania breindli Weiser, 1946
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania butleri n. sp., illus.
Johnston, L. B.; Vernick, S. H.; and Sprague, V., 1978, J. Invert. Path., v. 32 (3), 278-290
Thelohania butleri n. sp., sporulation stages, light and electron microscopy
Syn.: Thelohania sp. Vernick, Sprague, & Krause, 1977
Pandalus jordani (cells of skeletal muscle): Queen Charlotte Sound, British Columbia, Canada

Thelohania californica Kellen and Lipa, 1960
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora californica (Kellen and Lipa, 1960) comb. n.

Thelohania cambari Sprague, 1950
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania campbelli Kellen and Wills, 1962
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora campbelli (Kellen and Wills, 1962) comb. n.

Thelohania cepede Hesse, 1905
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania chaetogastris Schroeder, 1909
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania cheimatobiae Kreig, 1956
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania chironomi Debaisieux, 1928
Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135
as syn. of Pleistophora chironomi (Debaisieux, 1928) comb. nov.

Thelohania chironomi Jirovec, 1940
Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135
as syn. of Thelohania debaisieuxi (Jirovec, 1940) nom. nov.

Thelohania chironomi Jirovec, 1940
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania cladocera (Pfeiffer, 1895) Jirovec, 1936
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania columbaczense Weiser, 1960
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania contejeani Henneguy, 1892
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
synonymy

Thelohania contejeani, illus.
Voronin, V. N., 1971, Parazitologiia, Leningrad, v. 5 (2), 186-191
Thelohania contejeani in Astacus astacus (skeletal and cardiac muscles, ovaries and eggs) (nat. and exper.), prevalence, pathogenesis, developmental cycle, possibility of transovarian as well as oral transmission: Leningrad oblast

Thelohania corethrae Schuberg and Rodriguez, 1915
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania corynoneurae sp. n., illus.
Voronin, V. N., 1979, Zool. Zhurnal, v. 58 (4), 592-594
Corynoneura sp. (fat body): lake Vrevo, Leningrad oblast and lake Uchenoe, Pskovsk oblast

Thelohania cyclopis Weiser, 1945
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania dasychirae Issi and Lipa, 1968
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania debaisieuxi (Jirovec, 1940) nom. nov., illus.
Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (2), 113-135
Syn.: Thelohania chironomi Jirovec, 1940

Thelohania diazoma Kramer, 1965
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania diazoma Kramer 1965
Pilley, B. M., 1976, J. Invert. Path., v. 28 (2), 177-183
as syn. of Vairimorpha necatrix (Kramer, 1965) [n. comb.]

Thelohania disparis Timofejeva, 1956
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania duorara Iversen and Manning, 1959
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
synonymy

Thelohania duorara, illus.
Kelly, J. F., 1979, J. Invert. Path., v. 33 (3), 331-339
Thelohania duorara, Agmasoma penaei, and Pleistophora sp. in Penaeus duorarum, pathology, tissue specificity: southern Biscayne Bay

Thelohania duorara Iversen and Manning, illus.
Thomas, M. M., 1978, Indian J. Fish., v. 23 (1-2), 1976, 282-284
description
Penaeus semisulcatus (musculature, gonads): Mandapam

Thelohania ephestiae Mattes, 1928
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania eriogastri Weiser, 1957
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania fibrata (Strickland, 1913) Debaisieux and Gastaldi, 1919
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania fibrata (Strickland), illus.
Loubes, C., 1979, J. Protozool., v. 26 (2), 200-208
Microsporida, synaptonemal complexes demonstrated in 6 genera but not in Nosema, implications for life cycles

Thelohania fibrata (Strickland, 1913), illus.
Maurand, J.; and Loubes, C., 1978, Ztschr. Parasitenk., v. 56 (2), 131-146
microsporidians from simuliid larvae, ultrastructural studies
simuliid larvae: region languedocienne (Sud de la France)

Thelohania giardi Henneguy, 1892
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
Syn.: 'sporozoaire' Henneguy and Thelohan, 1892

Thelohania gigantea Kellen and Wills, 1962
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora gigantea (Kellen and Wills, 1962) comb. n.

Thelohania grassii Missiroli, 1929
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania herediteria, illus.
Bulnheim, H. P., 1978, Helgoland. Wissensch. Meeresuntersuchg., v. 31 (1-2), 1-33
Octosporea effeminans and Thelohania herediteria in Gammarus duebeni duebeni, feminizing influence exerted on host's offspring by parasites, role of salinity and temperature on sex determination by parasites
Gammarus duebeni duebeni: Federal Republic of Germany; Denmark; France; United Kingdom
G. duebeni celticus: France; United Kingdom

Thelohania heredeteria Bulnheim, 1971
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania hessei (Leger et Hesse, 1921) Weiser, 1961, illus.
Coste-Mathiez, F.; and Tuzet, O., 1977, Ann. Sc. Nat., Zool. et Biol. Animalc, 12. s., v. 19 (2), 113-135

Thelohania hessei Weiser, 1961
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania hyphantriae Weiser, 1953
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania inimica: Bailey et al., 1967
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora canadensis (Wills and Beaudoin, 1965) comb. n.

Thelohania inimica Kellen and Wills, 1962
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora inimica (Kellen and Wills, 1962) comb. n.

Thelohania inimica canadensis Wills and Beaudoin, 1965
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora canadensis (Wills and Beaudoin, 1965) comb. n.

Thelohania janus Hesse, 1903
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania lairdi Weiser, 1965
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora lairdi (Weiser, 1965) comb. n.

Thelohania legeri: Fantham et al. (in part), 1941
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Parathelohania africana Hazard and Anthony, 1974

Thelohania legeri: Kudo (in part), 1929 [et auct.]
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Parathelohania anomala (Sen, 1941) Hazard and Anthony, 1974

Thelohania legeri: Sen (in part), 1941
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Parathelohania obscura (Kudo, 1929) Hazard and Anthony, 1974

Thelohania legeri
Khaliulin, G. L., 1973, Parazitologiia, Leningrad, v. 7 (4), 370-373
Thelohania spp. in Aedes spp. larvae, infection rate under natural conditions and in tests with transovarial transmission did not exceed 2%, infection rate in tests with transmission per os reached 50%
Aedes cinereus cinereus
A. cyprius
A. flavescens
all from Mariisk ASSR

Thelohania macrococcus sp. n., illus.
Voronin, V. N., 1979, Zool. Zhurnal, v. 58 (4), 592-594
Corynoneura sp. (fat body): lake Vrevo, Leningrad oblast

Thelohania macrocystis Gurley, 1893
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Chapmanium macrocystis (Gurley, 1893) comb. n.

Thelohania maenadis Perez, 1904
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104

Thelohania mesnili Paillot, 1924
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania minor Georgevitch, 1954
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania minuta
Khaliulin, G. L., 1973, Parazitologiia, Leningrad, v. 7 (4), 370-373
Thelohania spp. in Aedes spp. larvae, infection rate under natural conditions and in tests with transovarial transmission did not exceed 2%, infection rate in tests with transmission per os reached 50%
Aedes cinereus rossicus
A. caspius dorsalis
all from Mariisk ASSR

Thelohania minuta Kudo, 1924
Kilochitskii, P. Ia., 1977, Vestnik Zool., Akad. Nauk Ukrainsk. SSR, Inst. Zool. (4), 71-75
Aedes caspius caspius
A. caspius dorsalis
A. vexans
all from southern Ukraine

Thelohania mutabilis Kudo, 1923
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania nana Kellen and Lindegren, 1969
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania nepae Lipa, 1966
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Chapmanium nepae (Lipa, 1966) comb. n.

Thelohania noxia Kellen and Wills, 1962
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora noxia (Kellen and Wills, 1962) comb. n.

Thelohania obesa Kudo, 1924
Kilochitskii, P. Ia., 1977, Vestnik Zool., Akad. Nauk Ukrainsk. SSR, Inst. Zool. (4), 71-75
Aedes caspius caspius
A. caspius dorsalis
all from southern Ukraine

Thelohania ochridensis Georgevitch, 1952
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
"doubtful Thelohania species"

Thelohania opacita Kudo, 1922
Alikhanov, Sh. G., 1973, Parazitologiia, Leningrad, v. 7 (2), 175-179
Thelohania opacita in Aedes caspius caspius, changes in host sex ratio of populations as result of infection, more females than males

Thelohania opacita Kudo, 1922
Alikhanov, Sh. G., 1973, Parazitologiia, Leningrad, v. 7 (5), 389-391
Thelohania opacita, detrimental effect on growth and development of Aedes caspius caspius from natural populations

Thelohania opacita Kudo, 1922
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora opacita (Kudo, 1922) comb. n.

Thelohania opacita: Weiser (in part), 1947
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora khaliulini sp. n.

Thelohania opacita: Weiser (in part), 1961
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora californica (Kellen and Lipa, 1960) comb. n.

Thelohania nr. opacita Anderson, 1968
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora canadensis (Wills and Beaudoin, 1965) comb. n.

Thelohania opacitor Fulton et al., 1974
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora opacita (Kudo, 1922) comb. n.

Thelohania opacitor Fulton et al., 1974
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora canadensis (Wills and Beaudoin, 1965) comb. n.

Thelohania opacita
Khaliulin, G. L., 1973, Parazitologiia, Leningrad, v. 7 (4), 370-373
Thelohania spp. in Aedes spp. larvae, infection rate under natural conditions and in tests with transovarial transmission did not exceed 2%, infection rate in tests with transmission per os reached 50%
Aedes punctor: Mariisk and Tatarsk ASSR
A. intrudens: Mariisk ASSR
A. cantans: Mariisk ASSR
A. excrucians: Mariisk ASSR
A. communis (nat. and exper.): Mariisk ASSR

Thelohania opacita Kudo, 1922
Kilochitskii, P. Ia., 1977, Vestnik Zool., Akad. Nauk Ukrainsk. SSR, Inst. Zool. (4), 71-75
Aedes caspius caspius
A. caspius dorsalis
A. flavescens
A. vexans
Culex modestus
all from southern Ukraine

Thelohania opacita Kudo, illus.
Kukharchuk, L. P.; and Gulii, V. V., 1976, Izvest. Sibirsk. Otdel. Akad. Nauk SSSR (260) (5), s. Biol. Nauk (1), 75-79
Aedes caspius dorsalis
A. cantans
A. beklemishevi
A. communis
(fat body and haemolymph of all): all from central Priob'e

Thelohania opacita var. mariensis [n. var.], illus.
Khaliulin, G. L.; and Ivanov, S. L., 1971, Parazitologiia, Leningrad, v. 5 (1), 98-100 [For complete author reference see Supplement 20, Part 1]
Aedes communis (fat body, hemolymph): Mari ASSR

Thelohania opacita var. mariensis Khaliulin and Ivanov, 1971
Hazard, E. I.; and Oldacre, S. W., 1976, Tech. Bull. (1530), U. S. Dept. Agric., 1-104
as syn. of Amblyospora khaliulini sp. n.

Thelohania opacita var. mariensis
Khaliulin, G. L., 1973, Parazitologiia, Leningrad, v. 7 (4), 370-373
Thelohania spp. in Aedes spp. larvae, infection rate under natural conditions and in tests with transovarial transmission did not exceed 2%, infection rate in tests with transmission per os reached 50%
Aedes communis: Mariisk ASSR; Tatarsk ASSR

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Desportes, I.; Vivares, C. P.; and Theodorides, J., 1977, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 19 (3), 261-277
Porosporidae
mt: T. pisae (L. et D.) [n. comb.]

Thiriotia
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intestinal eugregarines of Brachyura, distribution among hosts confirms validity of new classification system for hosts

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Syn.: Porospora pisae Leger et Duboscq
Pisa tetraodon
P. nodipes
Lissa chiragra
Acanthonyx lunulatus
all from Villefranche-sur-Mer (Alpes-Maritimes)

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Corynoneura sp. (fat body): lake Vrevo

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Toxoplasma skin tests, mentally retarded children and their mothers, conclusion that toxoplasmosis does not play great role in mental retardation causation: Iraq

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coccidia, dogs (feces), coprological and serological survey: South Germany

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toxoplasmosis, Black Rhodesian youth, unusual presentation with cardiac arrhythmia and low cholinesterase levels, clindamycin

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toxoplasmosis, human, diagnosis, complement fixation reaction, evaluation of different antigens and methods, comparison with dye test and indirect immunofluorescence

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Camargo, M. E.; et al., 1978, Infect. and Immun., v. 21 (1), 55-58
toxoplasmosis, diagnostic possibilities of IgG and IgM micro-enzyme-linked immunosorbent assays, results compared to those of hemagglutination, complement fixation, and immunofluorescence tests in serum samples presenting serological characteristics of recent or acute infection, of transitional period following acute infection, and of ancient infection

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biochemistry of parasitic protozoa, textbook: methodology; catabolism and generation of energy; nucleic acid metabolism; protein metabolism; lipid metabolism; biochemical mechanism of drug action; isolation of parasitic protozoa from infected animals; culture of parasitic protozoa

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coccidia of cats, life cycle, epidemiology, review

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Plasmodium spp., rapid detection in blood smears by fluorescence microscopy with 4'6 diamidino-2-phenylindole; can also be used to stain Toxoplasma and microfilariae

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human ocular toxoplasmosis, long-term medical management, pathology, combined sulfamide and antibiotics as therapy

Toxoplasma
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Toxoplasma, humans (pig-breeders and their families), swine, stray cats, antibody prevalence: Hamaoka district, Shizuoka Prefecture, Japan

Toxoplasma
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Toxoplasma
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toxoplasmosis, human serological survey of 4 ethnic groups: Transvaal, South Africa

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Toxoplasma bradyzoites from Mastomys natalensis (cerebral tissue), not infectious or able to replicate after 24 hours of storage under freezing at -20°C, no intact bradyzoites demonstrable, changes in structure and substance of cells and cysts

Toxoplasma
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toxoplasmosis, children, survey for presence of antibodies using the direct agglutination and indirect immunofluorescence tests, gradual increase in infection rate with age (60% by age 14)

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toxoplasmosis, recurrent infection in woman treated with co-trimoxazole, normal clinical response to each course of therapy, no evidence of impaired immunity

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Toxoplasma, pregnant woman with past history of active toxoplasmic retinochoroiditis, monitoring of antibody titers throughout pregnancy by means of the hemagglutination test, case reports on individual pregnancies and deliveries

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Toxoplasma, possible cause of human mesenteric lymphadenitis, clinical report

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Strannegard, O.; et al., 1973, Scand. J. Infect. Dis., v. 5 (3), 181-186
serologic studies of infections in humans with hematologic malignancy, Toxoplasma observed in 3 patients: Sweden

Toxoplasma
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Sarcocystis, human, indirect fluorescent antibody test using Sarcocystis fusiformis as antigen, antibody prevalence varies among 4 ethnic groups (Orang Asli, Malays, Indians, Chinese), some sera also reacted positively to Toxoplasma: West Malaysia

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Tsubota, N.; et al., 1977, Kiseichugaku Zasshi (Japan. J. Parasitol.), v. 26 (4), 286-290
Toxoplasma, human, diagnosis, microtiter latex agglutination test

Toxoplasma
Tsubota, N.; et al., 1977, Kiseichugaku Zasshi (Japan. J. Parasitol.), v. 26 (4), 291-298
Toxoplasma, swine, rabbits, cats, mice, diagnosis, microtiter latex agglutination test

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human Toxoplasma, pathology, treatment, recent trends

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toxoplasmosis as cause of granulomatous hepatitis, reply to letter of Frenkel, J. K.; and Remington, J. S., 1980, N. England J. Med., v. 302 (3), 178

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Toxoplasma oocysts, No. 1 and Fukaya strains, effects of low temperature and dryness on viability

Toxoplasma
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Toxoplasma cyst in Macropus rufogriseus (retina, brain), ocular pathology, case report: Whipsnade Park, London Zoo

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Leptodactylus ocellatus (central nervous system): Brasil

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Toxoplasm[a sp.], puppies, radiculoneuritis, clinical and pathological findings
Junghunde (Nervenwurzeln)

Toxoplasma [sp.]
Hagiwara, T.; Katsube, Y.; and Kamiyama, T., 1978, Japan. J. Vet. Sc., v. 40 (4), 455-457
Toxoplasma [sp.], sheep (diaphragm, heart, brain, abdominal muscle, lung), goat (diaphragm), latent infection, parasitological and serological observations: Chiba Prefecture, Japan

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Pohl, R., 1977, Tierarztl. Umschau, v. 32 (12), 642-643
Toxoplasma-cysts, increased perinatal mortality in small flock of dairy sheep
Schafe (Lunge, Kotyledonen): Bodenseekreis

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Acosta, E.; and Finkelman, J., 1976, SPM Salud Pub. Mexico, v. 18 (2), 403-407
Toxoplasma gondii, humans, review of recent information on methods of transmission, possible transitory hosts, immunity

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Toxoplasma gondii, horses, dye test antibody titers, isolation in cats and mice: slaughtered in Ohio, obtained from Ohio, Indiana, Illinois, and Kentucky

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toxoplasmosis, human outbreak at industrial plant, epidemiological study: State of Sao Paulo, Brazil

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human toxoplasmosis, diagnostic problems encountered, discussion of need to consider patient's socio-economic status when toxoplasmosis has been confirmed

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human toxoplasmosis, etiology, epidemiology, clinical aspects, general review

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human toxoplasmosis, diagnosis, treatment, prophylaxis, conclusion of general review

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Toxoplasma gondii
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Toxoplasma gondii, human acquired lymphatic form, diagnosis by bone marrow examination

Toxoplasma gondii
Amato Neto, V.; et al., 1975, Rev. Inst. Med. Trop. S. Paulo, v. 17 (5), 277-282
Toxoplasma gondii, humans, agglutinotest compared with indirect immunofluorescence, agglutinotest not satisfactory

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test for anti-Toxoplasma gondii antibodies in persons with various non-parasitic diseases, Sabin-Feldman dye test and immunofluorescence compared using serum vs. ascitic fluid samples

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Toxoplasma gondii, members and visitors of small community, pigeons as source of infection: city of Sao Paulo, Brazil

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Toxoplasma gondii, differential diagnosis of human lymphatic form, finding that fever is not always a constant sign in the clinical diagnostic picture

Toxoplasma gondii
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Toxoplasma gondii, human acquired lymphatic infection, comparative study of serum and cerebro spinal fluid with Sabin-Feldman dye test, less antibodies in spinal fluid than serum, no evidence of nervous system involvement

Toxoplasma gondii
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T[oxoplasma] gondii, human, indirect hemagglutination test using whole antigen for serodiagnosis and for determining stage of disease, comparison with indirect immunofluorescence

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Toxoplasma gondii, survey of serum samples from blood donors, Sabin-Feldman dye test and indirect immunofluorescence tests compared; Sarcocystis sp. in local animals may have some bearing on test results: Belo Horizonte, Brazil

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onchocerciasis, malaria, humans, epidemiological and vector survey; no evidence of schistosomiasis but potential vectors are present; little evidence of Toxoplasma gondii: lac de retenue de Bamendjin, Cameroun

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Toxoplasma gondii
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cryopreservation of parasitic protozoa

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Toxoplasma gondii
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Toxoplasma gondii
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Toxoplasma gondii, survey of infection prevalence in mental patients as compared with a sampling from normal controls; slightly higher rate of positive serology in the mentally ill patients

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Toxoplasma gondii, diagnosis, immunofluorescence

Toxoplasma gondii
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Toxoplasma gondii, pregnant women, serodiagnosis, review

Toxoplasma gondii
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Toxoplasma gondii, human congenital, significance of different diagnostic tests in screening for inapparent infections, serologic tests and serologic patterns in mothers and children studied

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Camargo, M. E.; Leser, P. G.; and Rocca, A., 1972, Rev. Inst. Med. Trop. S. Paulo, v. 14 (5), 310-313
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Toxoplasma gondii, humans, review (life cycle, taxonomy, intermediate and definitive hosts, transmission, pathology, diagnosis, treatment)

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Trypanosoma cruzi, Toxoplasma gondii, antibody prevalence survey in pregnant women: Paraguay

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Toxoplasma gondii
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Entamoeba histolytica, Toxoplasma gondii, humans, serological survey using indirect hemagglutination test, prevalence of antibody titers by host age, sex, and village (altitude): Malili area, South Sulawesi, Indonesia

Toxoplasma gondii
de Castilho, E. A., 1976, Rev. Inst. Med. Trop. S. Paulo, v. 18 (3), 202-205
Toxoplasma gondii, human, estimation of incidence of congenital infections during 1970: Sao Paulo City, Brazil

Toxoplasma gondii
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Toxoplasma gondii, human, control reagent prepared from mouse peritoneal exudate used for toxoplasmin diagnostic skin test, comparison with the more commonly employed reagent of mouse spleen extract; study of 50 persons thought to have toxoplasmosis showed the new reagent to be equally reactive with some advantages over the spleen extract

Toxoplasma gondii
Chen, S. N.; et al., 1974, Taiwan i Hsueh Hui Tsa Chih (J. Formosan Med. Ass.), v. 73 (6), 324-327
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Toxoplasma gondii
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Toxoplasma gondii
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Toxoplasma gondii
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Toxoplasma gondii
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Toxoplasma gondii
Chhabra, M. B.; Mahajan, R. C.; and Ganguly, N. K., 1979, Haryana Agric. Univ. J. Research, v. 9 (1), 59-62
Toxoplasma gondii, mice (exper.), value of double vaccination by irradiated tachyzoites

Toxoplasma gondii
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Toxoplasma gondii
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Toxoplasma gondii, isolation from suspected human material by mouse inoculation, correlation between high serological titre and success in isolation, 4 of 6 isolations from lymph node biopsies were from patients with malignant disorders of lymphatics

Toxoplasma gondii, illus.
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Toxoplasma gondii, Besnoitia jellisoni, antigen-treated lymphocytes from immune hamsters can directly confer immunity not only to macrophages but also to fibroblasts and kidney cells, infection-specific mediators can be derived from immune lymphocytes in presence of antigen and also confer immunity to macrophages and somatic cells

Toxoplasma gondii
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Toxoplasma gondii, illus.
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Toxoplasma gondii oocysts, process of excystation, light and electron microscopy

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human parasites, differential diagnosis

Toxoplasma gondii
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Toxoplasma gondii, human, diagnosis, evaluation of direct agglutination test, less useful than the indirect fluorescent antibody test

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Toxoplasma gondii, illus.
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Toxoplasma gondii
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Toxoplasma gondii, congenital infection diagnosed in infant with microphthalmia and coloboma of the optic nerve, case report: Romania

Toxoplasma gondii
Copeland, D.; and Grove, D. I., 1979, Internat. J. Parasitol., v. 9 (3), 205-211
Toxoplasma gondii, Trichinella spiralis, concurrent infections in mice, intestinal worm burdens, muscle worm burdens, worm fecundity, resistance to newborn larvae, small bowel pathology, muscle inflammation, eosinophil levels, numbers of toxoplasma cysts in brain

Toxoplasma gondii
Correa, F. M. A.; Salata, E.; and Oliveira, M. R., 1978, Arq. Inst. Biol., S. Paulo, v. 45 (4), 209-212
Toxoplasma gondii, swine, indirect immunofluorescence diagnosis: State of Sao Paulo, Brazil

Toxoplasma gondii
Costa, A. J.; et al., 1978, Arq. Inst. Biol., S. Paulo, v. 45 (4), 299-301
anti-Toxoplasma gondii antibody, frequency in cows, indirect immunofluorescence: Jaboticabal, Sao Paulo, Brazil

Toxoplasma gondii
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sporozoans, staining for nucleic acids

Toxoplasma gondii
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Toxoplasma gondii, investigation of role of cat in disease transmission

Toxoplasma gondii
Couvreur, J.; and Desmonts, G., 1977, Rev. Prat., Paris, v. 27 (11), 647-648, 653-658
Toxoplasma gondii, trypanosomiasis, malaria, leishmaniasis, parasitic pathology of foetus, review

Toxoplasma gondii, illus.
Cravero, G. C., 1977, Ann. Fac. Med. Vet. Torino, v. 24, 144-153
Toxoplasma gondii, calf, congenital encephalomyelitis, perivascular mononuclear infiltrations, nodular gliosis and granulomatous lesions in spinal cord

Toxoplasma gondii
Cross, J. H., 1973, Taiwan i Hsueh Hui Tsa Chih (J. Formosan Med. Ass.), v. 72 (7), 407-412
parasitic diseases of human respiratory system, immunodiagnostic methods, review

Toxoplasma gondii
Cross, J. H.; and Santana, F. J., 1974, Taiwan i Hsueh Hui Tsa Chih (J. Formosan Med. Ass.), v. 73 (11), 676-678
Toxoplasma gondii, survey of local birds and mammals for possible infection
Apodemus agrarius (brain): Hsing-fu-liao, Taipei Hsien, Taiwan
kittens (exper.) (feces)

Toxoplasma gondii
Cruz, A. M. R.; et al., 1978, Rev. Portug. Farm., v. 18 (2), 123-126
Toxoplasma gondii, human, diagnosis, direct agglutination

Toxoplasma gondii, illus.
Deane, M. P.; et al., 1971, Rev. Inst. Med. Trop. S. Paulo, v. 13 (2), 110-113
Toxoplasma gondii, kittens (exper.), gametogonic cycle

T[oxoplasma] gondii
Delgado Garcia, G.; and Sanchez Torres, M., 1977, Rev. Cubana Med. Trop., v. 29 (3), 121-127
T[oxoplasma] gondii, immunoserological study of humans suspected of having infections resulting from their contacts with domestic animals

Toxoplasma gondii
Denmark, J. R.; and Chessum, B. S., 1978, Med. Lab. Sc., v. 35 (3), 227-232
Toxoplasma gondii, diagnosis, standardization of the enzyme-linked immunosorbent assay, discussion of apparatus available for mechanization of this process, importance of suitable carrier for the antigen

Toxoplasma gondii, illus.
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prenatal toxoplasmosis, human, clinical, diagnostic, and therapeutic aspects, symposium presentation

Toxoplasma gondii
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Toxoplasma gondii, affinity of 4 strains to mice brains over other organs

Toxoplasma gondii
Dotolo, J. R., 1974, Rhode Island Med. J., v. 57 (3), 104-105
Toxoplasma gondii causing symptoms resembling infectious mononucleosis, need for consideration in differential diagnosis of persons manifesting a glandular fever syndrome

Toxoplasma gondii
Dubey, J. P., 1978, J. Protozool., v. 25 (3, pt. 2), 380-382
Toxoplasma gondii, chronically infected cats re-excreted T. gondii oocyts after superinfection with Isospora felis, this re-excretion was prevented in cats infected with I. felis before T. gondii infection, administration of BCG before Toxoplasma infection had no apparent effect on outcome of infection

Toxoplasma gondii
Dubey, J. P., 1978, J. Protozool., v. 25 (3, pt. 2), 382-384
Hammondia hammondi, Besnoitia jellisoni, Toxoplasma gondii, BCG, comparison of cross-protection in hamsters

Toxoplasma gondii, illus.
Dubey, J. P., 1979, Am. J. Vet. Research, v. 40 (11), 1634-1637
Toxoplasma gondii, direct development of enteroepithelial stages in intestines of cats (exper.), hypothesis of extraintestinal pregametogonic stages not confirmed

Toxoplasma gondii, illus.
Dubey, J. P.; et al., 1979, J. Am. Vet. Med. Ass., v. 174 (6), 604-609
Toxoplasma gondii, pig (small intestine, lymph node, liver, spleen, lungs, brain), neonatal diarrhea, histopathology: Bloomfield, Indiana

Toxoplasma gondii
Dubey, J. P.; and Frenkel, J. K., 1974, Vet. Path., v. 11 (4), 350-379
Toxoplasma gondii, cats (exper.), immunity, effects of host age and corticosteroid administration; excretion of T. gondii, Isospora felis, and I. rivolta oocysts from cats previously infected and challenged with all three coccidia

Toxoplasma gondii
Dubey, J. P.; and Wong, M. M., 1978, J. Parasitol., v. 64 (3), 551-552
Hammondia hammondi, experimentally infectious to only 1 of 4 monkey species (Saguinus nigricollis), all 4 species developed low and transitory antibody titers to Toxoplasma gondii antigen after inoculation with Hammondia oocysts

Toxoplasma gondii
Durham, T. M.; and Colvin, H. M., 1978, J. Clin. Microbiol., v. 7 (3), 255-260
Toxoplasma gondii, premarket evaluation of commercial toxoplasmosis indirect fluorescent antibody reagents, only 49% of all of products tested met Center for Disease Control requirements

Toxoplasma gondii
Dvorak, J. A.; and Howe, C. L., 1979, J. Protozool., v. 26 (1), 114-117
Toxoplasma gondii-vertebrate cell interactions, kinetic study of reproductive phase utilizing controlled-environment culture system

Toxoplasma gondii
Dymon, M.; and Starzyk, J., 1977, Acta Biol. Cracov., s. Zool., v. 20 (1), 17-25
Toxoplasma gondii, wild hamster (Cricetus cricetus) highly susceptible to infection especially when parasite introduced intranasally, epidemiological implications

Toxoplasma gondii, illus.
Dymowska, Z.; and Zukowski, K., 1974, Przegl. Epidemiol., v. 28 (2), 181-184
Toxoplasma gondii, Dermestes lardarius fed on infected mice, possible role in natural transmission

Toxoplasma gondii, illus.
Entzeroth, R.; Scholtyseck, E.; and Greuel, E., 1978, Naturwissenschaften, v. 65 (7), 395
"The application of different techniques led to the conclusion that, besides three ultramorphologically different sarcosporidian cysts, the roe-deer [Capreolus capreolus] muscles contain coccidia of the genera Toxoplasma and Hammondia."

Toxoplasma gondii
Falade, S., 1978, Trop. Animal Health and Prod., v. 10 (3), 175-177
Toxoplasma gondii, goats, antibody prevalence, dye test, latex agglutinotest: Nigeria

Toxoplasma gondii, illus.
Faruqui, A. M. A.; et al., 1976, South. Med. J., v. 69 (9), 1234-1235
Toxoplasma gondii, man, presenting as prolonged remittent fever, diagnosed at abdominal laparotomy

Toxoplasma gondii
Fayer, R.; and Frenkel, J. K., 1979, J. Parasitol., v. 65 (5), 756-762
6 spp. of feline coccidia, oocysts fed to calves, calf tissue then fed to cats: neither Besnoitia, Hammondia, nor Sarcocystis were pathogenic for calves nor did they establish patent infections which could be transmitted back to cats; Cystoisospora spp. were not pathogenic for calves but could be transmitted back to cats; Toxoplasma strains were slightly to moderately pathogenic for calves and could be transmitted back to cats

Toxoplasma gondii
Felgner, P., 1978, Zentralbl. Bakteriol., 1. Abt. Orig., Reihe A, v. 240 (1), 118-122
new technique of heterogenous enzyme-linked immunosorbent assay (stick-ELISA), application to antigens of various infective agents (including Entameba histolytica, Toxoplasma gondii, Echinococcus spp., Schistosoma mansoni)

Toxoplasma gondii, illus.
Ferguson, D. J. P.; et al., 1978, Acta Path. et Microbiol. Scand., v. 86B (3), 165-167
Toxoplasma gondii, ultrastructure of sporocyst, initiation of sporozoite formation

Toxoplasma gondii, illus.
Ferguson, D. J. P.; et al., 1979, Acta Path. et Microbiol., Scand., v. 87B (3), 171-181
Toxoplasma gondii, oocyst sporulation, zygote development, sporoblast formation, ultrastructure, light and electron microscopy

Toxoplasma gondii, illus.
Ferguson, D. J. P.; et al., 1979, Acta Path. et Microbiol., Scand., v. 87B (3), 183-190
Toxoplasma gondii, oocyst sporulation, sporocyst formation, structure of sporocyst wall, ultrastructure

Toxoplasma gondii, illus.
Ferguson, H. W.; and Ellis, W. A., 1979, Vet. Rec., v. 104 (17), 392-393
Toxoplasma gondii, calf (lung, kidney), gross post mortem and histopathological changes, cat as suspected source of infection

Toxoplasma gondii
Fernandes, W. J.; and Barbosa, W., 1972, Rev. Patol. Trop., v. 1 (2), 259-265
Toxoplasma gondii, incidence in some domestic animals: Goiania

Toxoplasma gondii
Ferraroni, J. J.; and Marzochi, M. C. de A., 1978, Acta Amazonica, v. 8 (1), 83-89
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malaria, toxoplasmosis, screening of blood donors including those with possible parasitic infections

Toxoplasmosis
Wery-Paskoff, S.; et al., 1970, Ann. Soc. Belges Med. Trop. Parasitol., v. 50 (6), 703-710
toxoplasmosis, pregnant women, statistics of prevalence survey using the fluorescent antibody technique, incidence of infection in their new-born children: Kinshasa

Trichodina, illus.
Molnar, K., 1979, Acta Vet., Budapest., v. 27 (1-2), 1977-1979, 99-113
Sphaerospora carassii, carp and grasscarp, incidence, pathology, moderately pathogenic, concurrent infection with Trichodina or othe infections may depress host resistance and cause fatalities: pond farms, Hungary

Trichodina sp.
Armas, G., 1979, J. Fish Dis., v. 2 (6), 543-547
Mugil cephalus (body surface, gills): Rio Moche coastal lagoon, northern Peru

Trichodina sp.
Arthur, J. R.; Margolis, L.; and Arai, H. P., 1976, J. Fish. Research Bd. Canada, v. 33 (11), 2489-2499
Cottus cognatus (gills): Aishihik Lake, Yukon Territory

Trichodina [sp.]
Banerji, S. R.; Singh, U. N.; and Tiwari, S., 1978, Current Sc., Bangalore, v. 47 (8), 283-284 [Letter]
Trichodina [sp.] on exterior of Cyprinus carpio, severe infestation, copper sulphate effective treatment; quick lime, common salt, potassium permanganate, glacial acetic acid, and formalin were not effective: nursery ponds, Patna

Trichodina sp.
Cone, D. K.; and Anderson, R. C., 1977, Canad. J. Zool., v. 55 (9), 1410-1423
parasites of Lepomis gibbosus, prevalence and intensity in relation to host age and sex
Lepomis gibbosus (gills, body surface): Ryan Lake, Algonquin Park, Ontario

Trichodina sp.
Iziumova, N. A.; Mashtakov, A. V.; and Kashkovskii, V. V., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36) 64-68
[Lucioperca lucioperca] (nostril): Kamsk reservoir

Trichodina sp. (anguillae?)
Jackson, P. B. N., 1978, J. South African Vet. Ass., v. 49 (1), 57-59
parasites of Anguilla mossambica, importance of disease control in aquaculture
Anguilla mossambica (skin): South Africa

Trichodina sp., illus.
Kashkovskii, V. V., 1974, Parazitologiia, Leningrad, v. 8 (4), 368-378
Urceolariidae of fish, prevalence, intensity; parasite measurements
Perca fluviatilis (body surface, nasal cavity): Ural reservoirs

Trichodina spp.
Paperna, I.; and Baudin Laurencin, F., 1979, Aquaculture, v. 16 (2), 173-175
Dicentrarchus labrax
Sparus aurata
(gills of all): all from marine cultures in France

richodina sp., illus.
Shtein, G. A., 1973, Parazitologiia, Leningrad, v. 7 (6), 489-496
Zoarces viviparus (gills): Barents Sea

richodina sp.
Wierzbicki, K., 1970, Acta Parasitol. Polon., v. 18 (1-12), 45-55
Perca fluviatilis (gills): Lake Dargin, Mazurian Lakeland, Poland

richodina sp. 1, illus.
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
Ctenolabrus rupestris (gills): Black Sea

Trichodina sp. 2, illus.
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
Mullus barbatus ponticus (gills): Black Sea

Trichodina acuta
Chernysheva, N. B., 1975, Izvest. Gosudarstv. Nauchno-Issled. Inst. Ozer. i Rech. Ryb. Khoziaistva, Leningrad, v. 93, 117-120
protozoans of young predatory fish, extent of infection correlated with some environmental factors
[Lota lota]
[Perca fluviatilis]
all from Vrevo Lake, Leningrad district

Trichodina acuta Lom, 1961, illus.
Duncan, B. L., 1977, Tr. Am. Micr. Soc., v. 96 (1), 76-81
redescription
Tilapia zillii
T. mossambica
all from Freshwater Aquaculture Center, Central Luzon State University, Munoz, Nueva Ecija, Philippines

Trichodina ocuta [i. e. ?, acuta]
Iziumova, N. A.; Mashtakov, A. V.; and Kashkovskii, V. V., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36) 64-68
[Lucioperca lucioperca] (gills): Kamsk reservoir

Trichodina alburni Vojtek, 1957
Lom, J.; and Haldar, D. P., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 527-541
as syn. of Paratrichodina alburni (Vojtek, 1957) Lom, 1963

Trichodina borealis
MacKenzie, K.; and Gibson, D. I., 1970, Symposia Brit. Soc. Parasitol., v. 8, 1-42
Pleuronectes platessa (gills): Scotland

Trichodina bulbosa Davis
Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
parasites of fishes: China

Trichodina cottidarum Dogiel, 1948, illus.
Shtein, G. A., 1973, Parazitologiia, Leningrad, v. 7 (6), 489-496
measurements
Myoxocephalus scorpius
Pholis gunnelus
(gills of all): all from Barents Sea

Trichodina domerguei
Chernyshenko, A. S., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 105-113
ichthyoparasite fauna, extensity and intensity of invasion, species composition
[Pisces] bychok-zelenchak
[Pleuronectes flesus]
all from 4 estuaries, Black Sea (northern coastal region)

Trichodina domerguei
Dartnall, H. J. G., 1974, J. Zool., London, v. 172 (2), 207-214
Trichodina d. domerguei, T. tenuidens, salinity tolerance, activity of contractile vacuole of T. domerguei suggests mechanism for achieving euryhalinity

Trichodina domerguei (Wallengren)
Dartnall, H. J. G.; and Walkey, M., 1979, J. Fish Biol., v. 14 (5), 471-474
Gasterosteus aculeatus (skin, fins): Airds Bay, Loch Etive, Scotland

Trichodina domerguei (Wallengren)
Lester, R. J. G., 1974, Syesis, v. 7, 195-200
Gasterosteus aculeatus (fins, body surface): near Vancouver, British Columbia

Trichodina domerguei f. indet. 1, illus.
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
Gobius melanostomus (gills): Black Sea

Trichodina domerguei f. indet. 2
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
Crenilabrus quinquemaculatus
Symphodus scina
(gills of all): all from Black Sea

Trichodina domerguei f. indet. 3, illus.
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
Odontogadus merlangus euxinus (gills): Black Sea

Trichodina domerguei f. acuta
Astakhova, T. V.; and Stepanova, G. A., 1972, Parazitologiia, Leningrad, v. 6 (4), 364-368
Ctenopharyngodon idella (skin, gills): pond and spawning-nursery fisheries, Volga delta

Trichodina domerguei f. acuta Lom, 1961, illus.
Kashkovskii, V. V., 1974, Parazitologiia, Leningrad, v. 8 (4), 368-378
Urceolariidae of fish, prevalence, intensity; parasite measurements
Cyprinus carpio (body surface)
Ctenopharyngodon idella
[Hypophthalmichthys]
Perca fluviatilis
Acerina cernua
Carassius carassius
all from Ural reservoirs

Trichodina domerguei f. acuta Lom, 1961, illus.
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
description
Cyprinus carpio
Perca fluviatilis
Lucioperca lucioperca
Leucaspius delineatus
Rhodeus sericeus
Bombina bombina
Bufo bufo
Hyla arborea
Rana esculenta
R. ridibunda
R. temporaria
all from CSSR

Trichodina domerguei domerguei
Banina, N. N., 1973, Parazitologiia, Leningrad, v. 7 (3), 220-226
Infusoria living on Gasterosteus aculeatus and Pungitius pungitius, relationships in this biocoenosis between true parasites and other forms: Neva Delta

Trichodina domerguei domerguei
Dartnall, H. J. G., 1974, J. Zool., London, v. 172 (2), 207-214
Trichodina d. domerguei, T. tenuidens, salinity tolerance, activity of contractile vacuole of T. domerguei suggests mechanism for achieving euryhalinity
Gasterosteus aculeatus (fins): Great Britain

Trichodina domerguei f. esocis Lom, 1960, illus.
Kashkovskii, V. V., 1974, Parazitologiia, Leningrad, v. 8 (4), 368-378
Urceolariidae of fish, prevalence, intensity; parasite measurements
Esox lucius
Perca fluviatilis
all from Ural reservoirs

Trichodina domerguei f. gobii Raabe, 1959?, illus.
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
Gobius melanostomus (gills): Black Sea

Trichodina domerguei f. jadranica Raabe?, illus.
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
Mullus barbatus ponticus (gills): Black Sea

Trichodina domerguei subsp. saintjohnsi Lom et Laird, 1969, illus.
Shtein, G. A., 1973, Parazitologiia, Leningrad, v. 7 (6), 489-496
measurements
Myoxocephalus scorpius (gills)
Pholis gunnelus
all from Barents Sea

Trichodina esocis
Chernysheva, N. B., 1975, Izvest. Gosudarstv. Nauchno-Issled. Inst. Ozer. i Rech. Ryb. Khoziaistva, Leningrad, v. 93, 117-120
protozoans of young predatory fish, extent of infection correlated with some environmental factors
[Perca fluviatilis]
[Esox lucius]
all from Vrevo Lake, Leningrad district

Trichodina fariai da Cunha & Pinto, 1928
Lom, J.; and Haldar, D. P., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 527-541
trichodinids endoparasitic in fishes, survey of literature records together with some new observations

Trichodina heterodentata n. sp., Population A, illus.
Duncan, B. L., 1977, Tr. Am. Micr. Soc., v. 96 (1), 76-81
Tilapia mossambica (body and fins): Freshwater Aquaculture Center, Central Luzon State University, Munoz, Nueva Ecija, Philippines

Trichodina heterodentata n. sp., Population B, illus.
Duncan, B. L., 1977, Tr. Am. Micr. Soc., v. 96 (1), 76-81
Tilapia zillii
T. mossambica
all from Freshwater Aquaculture Center, Central Luzon State University, Munoz, Nueva Ecija, Philippines

Trichodina heterodentata n. sp., Population C, illus.
Duncan, B. L., 1977, Tr. Am. Micr. Soc., v. 96 (1), 76-81
Trichogaster trichopterus (body): Freshwater Aquaculture Center, Central Luzon State University, Munoz, Nueva Ecija, Philippines

Trichodina lieni Chen
Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
parasites of fishes: China

Trichodina liparisi Zhukov, 1964, illus.
Shtein, G. A., 1973, Parazitologiia, Leningrad, v. 7 (6), 489-496
measurements
Liparis liparis (gills): Barents Sea

Trichodina meridionalis (Dogiel, 1940), illus.
Kashkovskii, V. V., 1974, Parazitologiia, Leningrad, v. 8 (4), 368-378
Urceolariidae of fish, prevalence, intensity; parasite measurements
Cyprinus carpio
Carassius carassius
[Carassius auratus gibelio]
(nasal cavity of all): all from Ural reservoirs

Trichodina mira Kaschkowsky, illus.
Kashkovskii, V. V., 1974, Parazitologiia, Leningrad, v. 8 (4), 368-378
Urceolariidae of fish, prevalence, intensity; parasite measurements
Phoxinus percnurus (gills): Ural reservoirs

Trichodina mutabilis Kazubski et Migala, 1968, illus.
Kashkovskii, V. V., 1974, Parazitologiia, Leningrad, v. 8 (4), 368-378
Urceolariidae of fish, prevalence, intensity; parasite measurements
Cyprinus carpio (gills): Ural reservoirs

Trichodina (Vauchomia) nephritica Lom, 1958
Lom, J.; and Haldar, D. P., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 527-541
as syn. of Vauchomia nephritica Mueller, 1938

Trichodina nigra Lom, 1960, illus.
Kashkovskii, V. V., 1974, Parazitologiia, Leningrad, v. 8 (4), 368-378
Urceolariidae of fish, prevalence, intensity; parasite measurements
Cyprinus carpio
Ctenopharyngodon idella
[Hypophthalmichthys]
Rutilus rutilus
Abramis brama
Carassius carassius
[Carassius auratus gibelio]
[Coregonus peled]
all from Ural reservoirs

Trichodina nigra Lom, 1960
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
description
Cyprinus carpio
Scardinius erythrophthalmus
Rutilus rutilus
Abramis brama
Perca fluviatilis
Tinca tinca
Alburnus alburnus
Leuciscus cephalus
Bombina bombina
Bufo bufo
Hyla arborea
Rana esculenta
R. ridibunda
R. temporaria
all from CSSR

Trichodina nigra f. gobii Lom, 1960, illus.
Kashkovskii, V. V., 1974, Parazitologiia, Leningrad, v. 8 (4), 368-378
Urceolariidae of fish, prevalence, intensity; parasite measurements
Gobio gobio (body surface): Ural reservoirs

Trichodina nigra f. kamchatika G. Stein, 1967
Ashurova, M.; and Shtein, G. A., 1972, Parazitologiia, Leningrad, v. 6 (5), 476-480
as syn. of T. strelkovi Chan, 1961

Trichodina nigra kamchatika
Konovalov, S. M.; Shevliakov, A. G.; and Krasin, V. K., 1970, Parazitologiia, Leningrad, v. 4 (6), 547-556
parasite fauna of various groups of young Oncorhynchus nerka, comparative analysis reveals 3 ecological groups: Lake Azabach'e, Kamchatka river basin

Trichodina nigra kamchatica
Makhovenko, E. T., 1972, Parazitologiia, Leningrad, v. 6 (4), 369-375
Salvelinus alpinus inhabiting different ecological niches, possible use of differences in parasite fauna between groups as biological tags: Lake Azabach'e, Kamchatka

Trichodina nigra f. nemachili Lom, 1960, illus.
Kashkovskii, V. V., 1974, Parazitologiia, Leningrad, v. 8 (4), 368-378
Urceolariidae of fish, prevalence, intensity; parasite measurements
Phoxinus percnurus (gills): Ural reservoirs

Trichodina nobilis Chen, 1963, illus.
Kashkovskii, V. V., 1974, Parazitologiia, Leningrad, v. 8 (4), 368-378
Urceolariidae of fish, prevalence, intensity; parasite measurements
Ctenopharyngodon idella
[Hypophthalmichthys]
Cyprinus carpio
(body surface of all): all from Ural reservoirs

Trichodina nobilis Chen
Wu, P. H.; et al., 1975, Tung Wu Hsueh Pao (Acta Zool. Sinica), v. 21 (2), 190-198
parasites of fishes: China

Trichodina oviducti Poljansky, 1955, illus.
  Lom, J.; and Haldar, D. P., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 527-541
    synonymy
    trichodinids endoparasitic in fishes, survey of literature records together with some new observations
    Raja radiata: off Newfoundland

Trichodina ovonucleata Raabe, 1958 f. gracilis f. nova, illus.
  Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
    Ophidion rochei
    Crenilabrus griseus
    (gills of all): all from Black Sea (region near Sevastopol)

Trichodina ovonucleata f. subtilis f. nova, illus.
  Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
    Crenilabrus quinquemaculatus (gills): Black Sea (region near Sevastopol)

Trichodina pediculus Ehrenberg, 1838, illus.
  Hagargi, S. S.; and Amoji, S. D., 1979, Current Sc., Bangalore, v. 48 (17), 789-790 [Letter]
    description
    Barbus chola
    B. sarana
    B. stigma
    all from Gulbarga city, Karnataka (India)

Trichodina pediculus Ehrenberg
  Iziumova, N. A.; Mashtakov, A. V.; and Timoshechkina, L. G., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36), 71-76
    [Abramis brama]: Upper Kama

Trichodina pediculus Ehrenberg, 1838
  Kashkovskii, V. V., 1974, Parazitologiia, Leningrad, v. 8 (4), 368-378
    Urceolariidae of fish, prevalence, intensity; parasite measurements
    Cyprinus carpio (body surface)
    Acerina cernua
    Rutilus rutilus
    Perca fluviatilis
    all from Ural reservoirs

Trichodina pediculus, illus.
  Schneider, H., 1979, Mikrokosmos, v. 68 (11), 349-353
    Trichodina pediculus, morphology
    Hydra attenuata: Sudabschnitt des Landauer Stadtparkweihers

Trichodina polycirra Lom
  Iziumova, N. A.; Mashtakov, A. V.; and Timoshechkina, L. G., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36), 71-76
    [Rutilus rutilus]: Upper Kama

Trichodina polycirra Lom, 1960, illus.
  Kashkovskii, V. V., 1974, Parazitologiia, Leningrad, v. 8 (4), 368-378
    Urceolariidae of fish, prevalence, intensity; parasite measurements
    Rutilus rutilus (urinary bladder): Ural reservoirs

Trichodina polycirra Lom, 1960, illus.
  Lom, J.; and Haldar, D. P., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 527-541
    trichodinids endoparasitic in fishes, survey of literature records together with some new observations

Trichodina rajae (?)
  Lom, J.; and Haldar, D. P., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 527-541
    trichodinids endoparasitic in fishes, survey of literature records together with some new observations
    Raja radiata (urogenital tract): off Newfoundland

Trichodina rajae Evdokimova, Kuznetsova & Shtein, 1969, illus.
  Lom, J.; and Haldar, D. P., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 527-541
    trichodinids endoparasitic in fishes, survey of literature records together with some new observations

Trichodina rectuncinata Raabe, 1958, illus.
  Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
    Gaidropsarus mediterraneus
    Odontogadus merlangus euxinus
    Blennius sanguinolentus
    Crenilabrus griseus
    C. quinquemaculatus
    C. tinca
    Symphodus scina
    Ctenolabrus rupestris
    (gills of all): all from Black Sea

Trichodina (Vauchomia) renicola Lom, 1958
  Lom, J.; and Haldar, D. P., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 527-541
    as syn. of Vauchomia renicola (Mueller, 1931) Mueller, 1938

Trichodina reticulata Hirschmann et Partsch, 1955, illus.
  Kashkovskii, V. V., 1974, Parazitologiia, Leningrad, v. 8 (4), 368-378
    Urceolariidae of fish, prevalence, intensity; parasite measurements
    Carassius carassius (body surface, gills) [Carassius auratus gibelio]
    Cyprinus carpio
    all from Ural reservoirs

Trichodina rostrata Kulemina, 1968, illus.
  Kashkovskii, V. V., 1974, Parazitologiia, Leningrad, v. 8 (4), 368-378
    Urceolariidae of fish, prevalence, intensity; parasite measurements
    Rutilus rutilus (body surface, gills)
    Cyprinus carpio (body surface)
    all from Ural reservoirs

Trichodina schizothoraci sp. n., illus.
  Ashurova, M.; and Shtein, G. A., 1972, Parazitologiia, Leningrad, v. 6 (5), 476-480
    [lapsus p. 480 as T. schizothoracis]
    Schizothorax intermedius (urinary bladder): Bartang River, Western Pamir

Trichodina schizothoraci Ashurova & Shtein, 1972
Lom, J.; and Haldar, D. P., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 527-541
trichodinids endoparasitic in fishes, survey of literature records together with some new observations

Trichodina schizothoracis [lapsus p. 480 for T. schizothoraci sp. n.]
Ashurova, M.; and Shtein, G. A., 1972, Parazitologiia, Leningrad, v. 6 (5), 476-480

Trichodina spathulata Kulemina, 1968, illus.
Kashkovskii, V. V., 1974, Parazitologiia, Leningrad, v. 8 (4), 368-378
Urceolariidae of fish, prevalence, intensity; parasite measurements
Abramis brama (gills): Ural reservoirs

Trichodina strelkovi Chan, 1961
Ashurova, M.; and Shtein, G. A., 1972, Parazitologiia, Leningrad, v. 6 (5), 476-480
Syn.: T. nigra f. kamchatika G. Stein, 1967

Trichodina strelkovi
Paskeviciute, A., 1978, Lietuvos TSR Moks. Akad. Darbai, s. C (84), (4), 73-80
Dactylogyrus auriculatus
D. falcatus
all from Kursiu Marios Lagoon

Trichodina strelkovi f. badachschanica [lapsus pp. 476, 480 for T. strelkovi f. badachschanika f. n.]
Ashurova, M.; and Shtein, G. A., 1972, Parazitologiia, Leningrad, v. 6 (5), 476-480

Trichodina strelkovi f. badachschanika f. n., illus.
Ashurova, M.; and Shtein, G. A., 1972, Parazitologiia, Leningrad, v. 6 (5), 476-480
[lapsus pp. 476, 480 as T. strelkovi f. badachschanica]
Nemachilus stoliczkai (skin, gills): Murgab and Oksu rivers, Eastern Pamir
Schizopygopsis stoliczkai (gills): Sarezsk lake, Central Pamir

richodina strelkovi f. badachschanika Aschurova & G. Stein, 1972
Ashurova, M., 1973, Parazitologiia, Leningrad, v. 7 (2), 164-168
Schizopygopsis stoliczkai: Sarez Lake, central Pamir

richodina tenuidens
Dartnall, H. J. G., 1974, J. Zool., London, v. 172 (2), 207-214
Trichodina d. domerguei, T. tenuidens, salinity tolerance, activity of contractile vacuole of T. domerguei suggests mechanism for achieving euryhalinity
Gasterosteus aculeatus (gills): Great Britain

richodina tenuidens Faure-Fremiet
Dartnall, H. J. G.; and Walkey, M., 1979, J. Fish Biol., v. 14 (5), 471-474
Gasterosteus aculeatus (gills): Airds Bay, Loch Etive, Scotland

ichodina tenuidens (Faure-Fremiet)
Lester, R. J. G., 1974, Syesis, v. 7, 195-200
Gasterosteus aculeatus (gill chamber): near Vancouver, British Columbia

Trichodina tisae
Iziumova, N. A.; Mashtakov, A. V.; and Kashkovskii, V. V., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36) 64-68
[Lucioperca lucioperca] (gills): Kamsk reservoir

Trichodina urinaria Dogiel, 1940, illus.
Kashkovskii, V. V., 1974, Parazitologiia, Leningrad, v. 8 (4), 368-378
Urceolariidae of fish, prevalence, intensity; parasite measurements
Perca fluviatilis: Ural reservoirs

Trichodina urinaria Dogiel, 1940, illus.
Lom, J.; and Haldar, D. P., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 527-541
trichodinids endoparasitic in fishes, survey of literature records together with some new observations
Perca flavescens: Lake Opeongo, Canada

Trichodina urinaria Dogiel, 1940
Wierzbicki, K., 1970, Acta Parasitol. Polon., v. 18 (1-12), 45-55
Perca fluviatilis (urinary bladder, ureters, kidneys): Lake Dargin, Mazurian Lakeland, Poland

Trichodina urinicola Fulton, 1923
Frandsen, F., 1974, Acta Parasitol. Polon., v. 22 (1-11), 49-66
Triturus cristatus
T. vulgaris
(urinary bladder of all): all from Denmark

Trichodina urinicola Fulton, 1923
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
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Carassius carassius
[Carassius auratus gibelio]
[Tinca tinca]
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Ctenopharyngodon idella
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Kechker, V. I.; and Elizarov, V. V., 1978, Vestnik Dermat. i Venerol. (10), 81-82
recurrent gonorrhea in men complicated by concurrent infections including trichomoniasis

Trichomoniasis
Kholodovskaia, I. V.; Minasova, G. S.; and Khokhlov, A. P., 1979, Vestnik Dermat. i Venerol. (4), 58-60
trichomoniasis, human vaginal infections, clinical trials with tinidazole

Trichomoniasis
Kunii, K.; et al., 1976, Rinsho Fujinka Sanka (Clin. Gynec. and Obst.), v. 30 (3), 237-240
human vaginal trichomoniasis, single dose therapy with tinidazole, clinical studies

Trichomoniasis
Merchan Lopez, D.; et al., 1977, Rev. Obst. y Ginec. Venez., v. 37 (2), 267-273
human genital trichomoniasis, clinical trials testing efficacy of tinidazole, considered to be drug of choice for urogenital infections: Venezuela

Trichomoniasis
Merchan Lopez, D.; and Olmos, R., 1976, Rev. Obst. y Ginec. Venez., v. 36 (1), 181-185
human genital trichomoniasis, clinical trials with mimorazole, good results obtained: Venezuela

Trichomoniasis
Mettler, L., 1978, Therapiewoche, v. 28 (34), 6095-6098
trichomoniasis, women with vaginitis and their sexual partners, single dose therapy with ornidazole, cure rates, side effects

Trichomoniasis
Moore, J. R., 1979, J. Am. Coll. Health Ass., v. 28 (2), 128
trichomoniasis, women, vaginitis, metronidazole given as 1 gram, single dose: Columbia University, New York

Trichomoniasis
Ovchinnikov, N. M.; and Skuratovich, A. A., 1978, Vestnik Dermat. i Venerol. (6), 49-52
trichomoniasis and other human venereal diseases, experimental trials with chlorhexidine, humans, rabbits

Trichomoniasis
Rocha, A. M.; and Sarraff Neto, A., 1977, Rev. Brasil. Med., v. 34 (6), 365-366
trichomoniasis, human urogenital, successful therapy of sexual partners with secnidazole

Trichomoniasis
Shimotomai, K.; et al., 1976, Rinsho Fujinka Sanka (Clin. Gynec. and Obst.), v. 30 (9), 751-756
human vaginal trichomoniasis, oral use of tinidazole, clinical trials

Trichomoniasis
Shinohara, H.; and Sakai, H., 1975, Rinsho Fujinka Sanka (Clin. Gynec. and Obst.), v. 29 (8), 635-638
human trichomoniasis vaginitis, combined oral and vaginal tinidazole

Trichomoniasis
Shutskii, I. V.; et al., 1978, Vestnik Dermat. i Venerol. (9), 71-74
trichomoniasis, women, urogenital infections, clinical trials using various regimens of trichopol

Trichomoniasis
Takizawa, T.; and Ito, T., 1977, National Inst. Animal Health Quart., Tokyo, v. 17 (4), 179-183
secular trends of annual morbidities of animal infectious diseases (including bovine trichomoniasis), mathematical models, possible application in morbidity forecasting

Trichomoniasis
Tumasheva, N. I.; and Savchak, V. I., 1979, Vestnik Dermat. i Venerol. (8), 23-26
sexually transmitted mixed infections, including trichomoniasis, clinical review

Trichomoniasis
Yabur T., J. A., 1972, Rev. Obst. y Ginec. Venez., v. 32 (3), 425-432
urogenital trichomoniasis, women, statistics of case histories, clinical aspects, diagnosis, value of metronidazole therapy: Venezuela

Trichomoniasis
Zanella, D.; et al., 1976, Therapiewoche, v. 26 (44), 7244-7250
trichomoniasis, human vaginal, single or mixed candidiasis infections, methylpartricin therapy compared with metronidazole

Trichomoniasis
Zil'man, S. L., 1978, Vestnik Dermat. i Venerol. (5), 26-28
trichomoniasis, human urogenital, diagnosis, immunofluorescence using capillary blood

Trichophrya piscium
Konovalov, S. M.; Shevliakov, A. G.; and Krasin, V. K., 1970, Parazitologiia, Leningrad, v. 4 (6), 547-556
parasite fauna of various groups of young Oncorhynchus nerka, comparative analysis reveals 3 ecological groups: Lake Azabach'e, Kamchatka river basin

Trichophrya piscium, illus.
Lom, J., 1971, Folia Parasitol., v. 18 (3), 197-205
Trichophrya piscium, fine structure in relation to fish host, feeding, attachment, ultrastructural evidence is in favour of ectocommensal nature of this protozoon
Etheostoma flabellare (gills): Jolliet, Illinois

Trichophrya piscium
Makhovenko, E. T., 1972, Parazitologiia, Leningrad, v. 6 (4), 369-375
Salvelinus alpinus inhabiting different ecological niches, possible use of differences in parasite fauna between groups as biological tags: Lake Azabach'e, Kamchatka

Trichorhynchus pulcher Schneider, 1882, illus.
Marques, A.; Ormieres, R.; and Puissegur, C., 1978, Ann. Sc. Nat., Zool. et Biol. Animale, 12. s., v. 20 (1), 27-36
Trichorhynchus pulcher, description of life cycle stages, scanning electron microscopy
Scutigera coleoptrata (intestin): midi mediterraneen de la France

Tripartiella bulbosa
Astakhova, T. V.; and Stepanova, G. A., 1972, Parazitologiia, Leningrad, v. 6 (4), 364-368
Ctenopharyngodon idella (gills): pond and spawning-nursery fisheries, Volga delta

Tripartiella bulbosa (Davis, 1947), illus.
Kashkovskii, V. V., 1974, Parazitologiia, Leningrad, v. 8 (4), 368-378
Urceolariidae of fish, prevalence, intensity; parasite measurements
Ctenopharyngodon idella
[Hypophthalmichthys]
(gills of all): all from Ural reservoirs

Tripartiella copiosa Lom
Iziumova, N. A.; Mashtakov, A. V.; and Timoshechkina, L. G., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36), 71-76
[Rutilus rutilus]: Upper Kama

Tripartiella copiosa Lom, 1959, illus.
Kashkovskii, V. V., 1974, Parazitologiia, Leningrad, v. 8 (4), 368-378
Urceolariidae of fish, prevalence, intensity; parasite measurements
Rutilus rutilus (body surface, gills): Ural reservoirs

Tripartiella incisa Lom, 1959, illus.
Kashkovskii, V. V., 1974, Parazitologiia, Leningrad, v. 8 (4), 368-378
Urceolariidae of fish, prevalence, intensity; parasite measurements
Rutilus rutilus
Phoxinus percnurus
all from Ural reservoirs

Tritrichomonas, illus.
  De Carli, G. A.; et al., 1977, Rev. Microbiol., Sao Paulo, v. 8 (2), 53-54
    Tritrichomonas, fixation and staining, technique modification

Tritrichomonas angusta Alexeieff, 1911, illus.
  Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
    description
    Bufo bufo
    B. viridis
    B. calamita
    Rana esculenta
    R. ridibunda
    R. dalmatina
    R. temporaria
    Lacerta agilis
    L. vivipara
    (strevo of all): all from CSSR

Tritrichomonas batrachorum Perty (1852), illus.
  Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
    description
    Bombina bombina
    B. variegata
    Bufo bufo
    B. viridis
    B. calamita
    Pelobates fuscus
    Hyla arborea
    Rana esculenta
    R. ridibunda
    R. temporaria
    R. arvalis
    all from CSSR

Tritrichomonas fecalis
  Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
    parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Tritrichomonas foetus
  Brotherton, J., 1978, Arzneimittel-Forsch., v. 28 (10), 1665-1672
    trichomonads, in vitro testing of potential trichomonacides using Coulter Counter

Tritrichomonas foetus
  Cerkasov, J.; et al., 1978, J. Biol. Chem., v. 253 (4), 1207-1214
    Tritrichomonas foetus, respiration of hydrogenosomes, ADP-dependent oxidation of malate and pyruvate

Tritrichomonas foetus
  Christensen, H. R.; and Clark, B. L., 1979, Austral. Vet. J., v. 55 (4), 205 [Letter]
    Tritrichomonas foetus, prevalence in replacement bulls 4 years after introduction to infected herd, observations suggest routine culling of bulls at 4 years of age to minimize transmission: north-western Queensland

Tritrichomonas foetus
  De Carli, G. A.; and Guerrero, J., 1975, Rev. Microbiol., S. Paulo, v. 6 (3), 55-58
    Tritrichomonas suis, T. foetus, study of antigenic relationships using indirect immunofluorescence, results suggested presence of antigens that were common to both species

Tritrichomonas foetus
  De Carli, G. A.; and Guerrero, J., 1977, Rev. Microbiol., Sao Paulo, v. 8 (4), 107-109
    Tritrichomonas suis, T. foetus, antigenic comparison by immunoelectrophoresis

Tritrichomonas foetus
  Fuskova, A.; Fuska, J.; and Kettner, M., 1978, Folia Microbiol., v. 23 (5), 389-393
    Tritrichomonas foetus, in vitro inhibition by vermiculine, mode of action

Tritrichomonas foetus
  Habich, G. E.; et al., 1977, Gac. Vet., Buenos Aires (326), v. 39, 647-656
    Tritrichomonas foetus, dairy bulls, prevalence, diagnosis by preputial washings: Catamarca, Noroeste Argentino

Tr[itrichomonas] fetus
  Kolev, V.; et al., 1977, Vet. Med. Nauki, v. 14 (3), 54-60
    Protozoa as cause of abortions in cows: district of Rousse

Tritrichomonas foetus
  LaRusso, N. F.; Lindmark, D. G.; and Mueller, M., 1978, Biochem. Pharmacol., v. 27 (18), 2247-2254
    metronidazole, biliary and renal excretion, hepatic metabolism, and hepatic subcellular distribution in bile fistula rats, glucuronide conjugate of metronidazole was devoid of activity against Tritrichomonas foetus

Tritrichomonas foetus
  Lindmark, D. G.; and Mueller, M., 1976, Antimicrob. Agents and Chemotherapy, v. 10 (3), 476-482
    metronidazole and 11 other nitroimidazoles, antitrichomonad activity against Tritrichomonas foetus and Trichomonas vaginalis, mutagenic action in Salmonella test, reducibility of nitro group by T. foetus homogenates, results underscore role of reduction of nitro group in antitrichomonad and mutagenic activity of nitroimidazoles

Tritrichomonas foetus
  Lloyd, D.; Lindmark, D. G.; and Mueller, M., 1979, J. Parasitol., v. 65 (3), 466-469
    Tritrichomonas foetus, respiration: absence of detectable cytochromes

Tritrichomonas foetus
  Lloyd, D.; Lindmark, D. G.; and Mueller, M., 1979, J. Gen. Microbiol., v. 115 (2), 301-307
    Tritrichomonas foetus, adenosine triphosphatase activity

Tritrichomonas foetus
  Mack, S. R.; and Mueller, M., 1978, J. Parasitol., v. 64 (5), 927-929
    Trichomonas vaginalis, Tritrichomonas foetus, effect of oxygen and carbon dioxide on growth

Tritrichomonas foetus
  McLaughlin, J.; and Mueller, M., 1979, J. Biol. Chem., v. 254 (5), 1526-1533
    Tritrichomonas foetus, low molecular weight thiol proteinase, purification and characterization

Tritrichomonas foetus
  Meingassner, J. G.; et al., 1978, Antimicrob. Agents and Chemotherapy, v. 13 (1), 1-3
    Tritrichomonas foetus, metronidazole-resistant and susceptible strains, in vitro susceptibility testing, results suggest that the two strains differ in regulation of internal redox systems and underscore the role that testing methods may play in the in vitro detection of nitroimidazole-resistant protozoan parasites

ritrichomonas foetus
  Mueller, M.; et al., 1979, Comp. Biochem. and Physiol., v. 64B (1), 97-100
    Tritrichomonas foetus, Trichomonas vaginalis, Entamoeba invadens, effects of 2,4-dinitrophenol (including effect on accumulation of metronidazole)

ritrichomonas foetus
  Mueller, M.; and Lindmark, D. G., 1978, J. Biol. Chem., v. 253 (4), 1215-1218
    Tritrichomonas foetus, respiration of hydrogenosomes, effect of CoA on pyruvate oxidation

ritrichomonas foetus
  Nseka, K.; and Mueller, M., 1978, Compt. Rend. Soc. Biol., Paris, v. 172 (6), 1094-1098
    Tritrichomonas foetus, Entamoeba invadens, effect of glycolysis inhibitors on uptake of metronidazole

ritrichomonas foetus
  Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
    parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

itrichomonas foetus, illus.
  Sledge, W. E.; Larson, A. D.; and Hart, L. T., 1978, Science (4325), v. 199, 186-188
    Tritrichomonas foetus, costae, procedure for purification, chemical composition (95% carbohydrate, 5% protein), evidence of structural association among costae, kinetosomes, and flagella

itrichomonas foetus
  Tedesco, L. F.; Errico, F.; and Del Baglivi, I. P., 1979, Austral. Vet. J., v. 55 (7), 322-324
    Tritrichomonas foetus, bulls, diagnosis by direct microscopic examination and by cultural recovery, comparison of 2 methods for collection of preputial secretions, suitability of transport medium

itrichomonas foetus
  Tompel, Kh. Ia.; and Teras, Iu. Kh., 1978, Veterinariia, Moskva (9), 96-98
    Tritrichomonas foetus, strain differences in antigenic properties tested in rabbits

itrichomonas foetus (Riedmueller), illus.
  arton, A.; and Honigberg, B. M., 1979, J. Pro- ozool., v. 26 (1), 56-62
    Hypotrichomonas acosta, Trichomonas vaginalis, Pentatrichomonas hominis, Tritrichomonas foetus, scanning electron microscopy

Tritrichomonas suis
  De Carli, G. A.; and Guerrero, J., 1975, Rev. Microbiol., S. Paulo, v. 6 (3), 55-58
    Tritrichomonas suis, T. foetus, study of antigenic relationships using indirect immunofluorescence, results suggested presence of antigens that were common to both species

Tritrichomonas suis, illus.
  De Carli, G. A.; and Guerrero Ramirez, J., 1975, Rev. Fac. Med. Vet. e Zootec. Univ. S. Paulo, v. 12, 269-275
    Tritrichomonas suis in Sus scrofa (nasal cavity), incidence, morphology: Rio Grande do Sul

Tritrichomonas suis
  De Carli, G. A.; and Guerrero, J., 1977, Rev. Microbiol., Sao Paulo, v. 8 (4), 107-109
    Tritrichomonas suis, T. foetus, antigenic comparison by immunoelectrophoresis

Trypanoplasma beckeri sp. n., illus.
  Burreson, E. M., 1979, J. Protozool., v. 26 (3), 343-347
    Trypanoplasma beckeri sp. n., structure, life cycle, leech vector
    Scorpaenichthys marmoratus (blood and body fluids) (nat. and exper.): collected near entrance to Yaquina Bay, Oregon, U.S.A.; held in display tank at Oregon State University Marine Science Center Public Aquarium
    Malmiana diminuta (crop, proboscis sheath) (nat. and exper.): display tank at Oregon State University Marine Science Center Public Aquarium
    Trachelobdella oregonensis (exper.)

Trypanoplasma borelli Laveran & Mesnil, 1902
  Dykova, I.; and Lom, J., 1979, J. Fish Dis., v. 2 (5), 381-390
    Trypanosoma danilewskyi and Trypanoplasma borelli in Carassius auratus (exper.), histopathological changes

Trypanoplasma bullocki (Strout, 1965)
  Becker, C. D.; and Overstreet, R. M., 1979, J. Fish Dis., v. 2 (6), 469-479
    Paralichthys lethostigma (plasma)
    Micropogonias undulatus (plasma)
    Trinectes maculatus (spleen, liver)
    all from northern Gulf of Mexico

Trypanoplasma salmositica, illus.
  Woo, P. T. K., 1979, Exper. Parasitol., v. 47 (1), 36-48
    Trypanoplasma salmositica, successful in vitro culture and subpassage, course of infection in Salmo gairdneri (exper.), clinical signs (anemia, exophthalmia, abdominal distension with ascites, splenomegaly), diagnosis by wet mount examination more sensitive than hematocrit centrifuge technique, evidence of possible antigenic variation

Trypanosoma
  Baker, J. R.; et al., 1978, Bull. World Health Organ., v. 56 (3), 467-480
    salivarian trypanosomes, definitions and nomenclature, relationship of characterized populations to Linnaean taxa, maintenance of reference collections, documentation of reference collections, proposals

Trypanosoma
Barry, J. D., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (3), 348-349 [Letter]
trypanosome material, confusion in publishing pedigrees, need for standard system of nomenclature and recording of pedigrees

Trypanosoma
Brown, K. N., 1978, Colloque Immun. Parasit. Dis. (Thiverval-Grignon, Sept. 5-9, 1977), 59-70
Trypanosoma, Plasmodium, Babesia, antigenic variation (nature, consequences for protective immunity, possible implications for other protozoan infections), colloquium presentation

Trypanosoma
Cross, G. A. M., 1978, Trends Biochem. Sc., Ref. Ed., v. 3 (1), 49-51
Trypanosoma, molecular basis of antigenic variation, brief review

Trypanosoma
Cross, G. A. M., 1979, J. Gen. Microbiol., v. 113 (1), 1-11
trypanosomes, immunochemical aspects of antigenic variation, 3rd Fleming Lecture

Trypanosoma
Evans, D. A., 1978, Methods Cultiv. Parasites in Vitro, 55-88
Kinetoplastida, cultivation, review

Trypanosoma
Godfrey, D. G., 1978, Nature, London (5664), v. 273, 600-604
identification of economically important parasites (use of anatomical, biochemical, and behavioral tests), brief review

Trypanosoma
Gutteridge, W. E.; and Coombs, G. H., 1977, Biochemistry of parasitic protozoa, 172 pp., illus.
biochemistry of parasitic protozoa, textbook: methodology; catabolism and generation of energy; nucleic acid metabolism; protein metabolism; lipid metabolism; biochemical mechanism of drug action; isolation of parasitic protozoa from infected animals; culture of parasitic protozoa

Trypanosoma
Jaenson, T. G. T., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (3), 234-238
Glossina pallidipes, virus-like rods associated with salivary gland hyperplasia, exceedingly heavy trypanosome infections found in some hyperplastic glands, unlikely that virus-like particles can be used as biological control agents if they are favoring development of trypanosomes

Trypanosoma
Lumsden, W. H. R., 1972, Immun. Animal Parasites, 287-299
immune response to trypanosomes, review

Trypanosoma
Lumsden, W. H. R., 1974, Ciba Found. Symp., n.s. (20), 3-27
leishmaniasis and trypanosomiasis, causative organisms compared and contrasted, review

Trypanosoma
Lumsden, W. H. R., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 361-368
trypanosome populations, problems in characterization and nomenclature, review

Trypanosoma
Lumsden, W. H. R., 1978, Handb. Exper. Immunol. (Weir), 3. ed., 42.1-42.47
demonstration of antibodies to Protozoa, extensive review

Trypanosoma
Mansfield, J. M., 1978, Cellular Immunol., v. 39 (1), 204-210
African trypanosomiasis, lymphocyte dysfunction and immunosuppression (histopathological considerations; B cell function; T cell function; macrophage function), cellular bases of immunosuppression, review

Trypanosoma (Herpetosoma)
Molyneux, D. H., 1970, Ann. Soc. Belges Med. Trop. Parasitol., v. 50 (2), 229-237
Trypanosoma (Herpetosoma) spp., reproduction in vertebrate host, course of infection, transmission and development in fleas, review

Trypanosoma
Murray, M.; Clifford, D. J.; and McIntyre, W. I. M., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (1), 120-121 [Letter]
bovine African trypanosomiasis, usefulness of diagnosis by blood buffy coat examination by darkground microscopy, suggestions for planning of epidemiologic studies

Trypanosoma
Newton, B. A., 1974, Ciba Found. Symp., n.s. (20), 285-307
trypanosomiasis, leishmaniasis, chemotherapy, review

Trypanosoma
Peters, W., 1974, Ciba Found. Symp., n.s. (20), 309-334
trypanosomiasis, leishmaniasis, drug resistance, review

Trypanosoma
Snow, W. F.; and Boreham, P. F. L., 1979, Acta Trop., v. 36 (1), 47-51
Phacochoerus aethiopicus appears to be major maintenance host for Glossina morsitans submorsitans as well as potential reservoir of trypanosomiasis: The Gambia

Trypanosoma
Terry, R. J., 1976, Immunol. Parasit. Infect., 203-221
African trypanosomiasis, immunity, review

Trypanosoma
Trager, W., 1974, Ciba Found. Symp., n.s. (20), 225-254
trypanosomes, leishmanias, nutrition and biosynthetic capabilities, problems of in vitro cultivation and differentiation, review

Trypanosoma
Vickerman, K., 1974, Ciba Found. Symp., n.s. (20), 171-198
trypanosomatid flagellates, ultrastructure, review with emphasis on changes during life cycles

Trypanosoma
Williams, P.; and Coelho, M. de V., 1978, Advances Parasitol., v. 16, 1-42
Leishmania, taxonomy and nomenclature, Phlebotominae as hosts for Trypanosomatidae, review

Trypanosoma
Wilson, R. J. M., 1974, Ciba Found. Symp., n.s. (25), 185-203
soluable parasite antigens, possible modes of interference with immune response, review

Trypanosoma
Wright, I. G., 1979, Gen. Pharmacol., v. 10 (4), 319-325
Babesia, Plasmodium, Trypanosoma, kallikrein-kinin system, mechanisms of activation (parasite enzymes, immune complexes), role in hypotensive shock syndrome of infected animals, review

Trypanosoma [sp.]
Baker, J. R., 1974, J. Zool., London, v. 172 (2), 169-190
Corvus frugilegus
C. monedula
Delichon urbica
Emberiza citrinella
Fringilla coelebs
Garrulus glandarius
Linota rufescens
Muscicapa striata
Turdus ericetorum
T. merula
Athene noctua
Columba palumbus
Neomys fodiens
Talpa europaea
Nyctalus noctula
Pipistrellus pipistrellus
Oryctolagus cuniculus
Apodemus sylvaticus
Clethrionomys glareolus
Micromys minutus
Microtus agrestis
Mus musculus
Rattus norvegicus
R. rattus
all from Britain

Trypanosoma sp., illus.
Baker, J. R., 1977, Protozoology, v. 3, 75-82
Bufo regularis: Kinshasa, Zaire

Trypanosoma spp.
Chambers, P. G., 1978, Rhodesian Vet. J., v. 9 (1-4), 40
protozoans, icteric cattle carcases, routine examination of blood and spleen smears: Sinoia abattoir, Rhodesia

Trypanosoma (Herpetosoma) sp.
Edrissian, Gh. H.; Farhang-Azad, A.; and Neronov, V. M., 1976, J. Wildlife Dis., v. 12 (4), 497
Meriones persicus: Iran

Trypanosoma (Megatrypanum) spp.
Edrissian, Gh. H.; Farhang-Azad, A.; and Neronov, V. M., 1976, J. Wildlife Dis., v. 12 (4), 497
Minopterus schriebersi: Iran

Trypanosoma sp.
Fish, W. R.; Holz, G. G., jr.; and Beach, D. H., 1978, J. Parasitol., v. 64 (3), 546-547
26 trypanosomatid species, cultivation in new chemically-defined medium RE III

Trypanosoma (Megatrypanum) sp., illus.
Garnham, P. C. C., 1977, Protozoology, v. 3, 135-142
Miopithecus talapoin (blood): Rio Muni, West Africa, imported to United States of America

Trypanosoma [sp.], illus.
Guerrero, S.; and Ayala, S. C., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (5), 283-288
Dendrobates trivittatus (sangre): selva amazonica, Huanuco, Peru

Trypanosoma sp.
Hendricks, L. D.; Wood, D. E.; and Hajduk, M. E., 1978, Parasitology, v. 76 (3), 309-316
Leishmania spp., Trypanosoma spp., commercially available liquid media for rapid cultivation

Trypanosoma spp.
Lumsden, W. H. R., 1977, Protozoology, v. 3, 25-32
Trypanosoma spp., techniques, characterization, taxonomy, epidemiology, and pathogenesis, review of last 20 years

Trypanosoma (Trypanozoon) [sp.]
Mehlitz, D., 1979, Tropenmed. u. Parasitol., v. 30 (2), 212-219
Trypanosoma spp., infection rates in domestic animals; screening of dogs and pigs to indicate potential reservoirs of T. (Trypanozoon) brucei gambiense; comparison of sensitivity of 3 diagnostic techniques; comparison of number of primary isolations and derived stabilated Trypanosoma stocks
dogs: rain forest areas, Liberia

Trypanosoma, possibly T. vivax, illus.
Pereira, L. J.; and de Abreu, A. C. V. V., 1978, Pesquisa Agropec. Brasil., v. 13 (3), 17-21
Trypanosoma, possibly T. vivax, cattle and sheep, symptoms, laboratory tests, treatment: State of Para and Amapa Federal Territory, Brazil

Trypanosoma [sp.]
da Rocha e Silva, E. O.; et al., 1977, Rev. Saude Pub., S. Paulo, v. 11 (1), 87-96
Triatoma sordida: Estado de Sao Paulo, Brasil

Trypanosoma (Megatrypanum) sp., "designated tentatively as Trypanosoma magnusi" [? n. sp.], illus.
Shamsuddin, M.; and Mohammad, M. K., 1978, Bull. Nat. Hist. Research Cent., Univ. Baghdad, v. 7 (2), 35-47
Taphozous nudiventris magnus (blood): Al-Khasfa cave and Al-Kifal, Iraq
Pipistrellus kuhli (blood): Al-Akhaidhar, Iraq

Trypanosoma (Herpetosoma) sp., illus.
Shaw, J. J., 1977, Protozoology, v. 3, 119-128
Felis pardalis: Itaituba-Jacareacanga sector of Transamazon Highway, Para, Brazil

Trypanosoma sp. Indonesia 56
Weinman, D.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (2, pt. 1), 232-237
Macaca nemestrina: Indonesia
Rhodnius prolixus (gut) (exper.)
Triatoma rubrofasciata (gut) (exper.)
mice (exper.)

Trypanosoma sp. Indonesia 71
Weinman, D.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (2, pt. 1), 232-237
Macaca ira: Indonesia
Rhodnius prolixus (gut) (exper.)
Triatoma rubrofasciata (gut) (exper.)
mice (exper.)

Trypanosoma sp. Kashmir
Weinman, D.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (2, pt. 1), 232-237
experimental infection in Rhodnius prolixus "suggested survival and not multiplication"
Macaca mulatta: shipped from Kashmir-Jammu and bled in New Delhi, India

Trypanosoma sp. Lucknow
Weinman, D.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (2, pt. 1), 232-237
experimental infections in Rhodnius prolixus "suggested survival and not multiplication"
Macaca mulatta: Lucknow area, India

Trypanosoma [spp.]
Williams, N. A.; and Bennett, G. F., 1978, Canad. J. Zool., v. 56 (4, pt. 1), 596-607
Cyanocitta cristata: Maryland or New Jersey
Coccyzus erythrophthalmus: Maryland or New Jersey
Junco hyemalis: Maryland or New Jersey
Melospiza melodia: " " " "
Zonotrichia albicollis: Maryland, and/or New Jersey
Agelaius phoeniceus: Maryland, and/or New Jersey
Molothrus ater: Maryland, and/or New Jersey
Quiscalus quiscula: " " " "
Dumetella carolinensis: " " " "
Mimus polyglottus: Maryland or New Jersey
Geothlypis formosa: " " " "
Seiurus aurocapillus: " " " "
Calidris pusilla: Maryland
Sialia sialis: Maryland, and/or New Jersey
Turdus migratorius: Maryland or New Jersey

Trypanosoma sp., T. lewisi-like
Yap, L. F.; et al., 1977, Southeast Asian J. Trop. Med. and Pub. Health, v. 8 (3), 345-353
Rattus tiomanicus: Jenderak Utara, Malaysia

Trypanosoma (Trypanozoon) [sp.]
Zillmann, U.; and Mehlitz, D., 1979, Tropenmed. u. Parasitol., v. 30 (2), 244-248
Trypanosoma (Trypanozoon) [sp.] in Gallus gallus var. domesticus (nat. and exper.), diagnosed from morphology and infectivity in Mastomys natalensis (exper.), significance for epizootiology and epidemiology of trypanosomiasis: Ivory Coast

Trypanosoma akodoni Carini and Maciel, 1915
Barretto, M. P., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (4), 272-278
as syn. of Trypanosoma (Schizotrypanum) cruzi Chagas, 1909

Trypanosoma akodoni Carini & Maciel, 1915
Ribeiro, R. D.; and Barretto, M. P., 1972, Rev. Inst. Med. Trop. S. Paulo, v. 14 (3), 162-170
as syn. of T. cruzi

Trypanosoma (Megatrypanum) amilcari n. sp., illus.
Mello, D. A., 1979, Ann. Parasitol., v. 54 (5), 489-494
[lapsus p. 489 as T. amileari]
Oryzomys eliurus (blood): northern part of Formosa county, Goias State, Brazil (BR-020 road northeast direction, 156 km from Brasilia city)

Trypanosoma (Megatrypanum) amileari [lapsus p. 489 for T. amilcari n. sp.]
Mello, D. A., 1979, Ann. Parasitol., v. 54 (5), 489-494

Trypanosoma anabasi sp. nov., illus.
Mandal, A. K., 1978, Ang. Parasitol., v. 19 (3), 158-162
Anabas testudineus (blood): Canning, India

Trypanosoma armeti n. sp., illus.
Mandal, A. K., 1975, Ang. Parasitol., v. 16 (2), 87-93
Mastacembelus armatus (blood): India (Champahati; Parganas; West Bengal)

Trypanosoma (Schizotrypanum) assiutis sp. nov., illus.
Fahmy, M. A. M.; Abedl-Rahman, A. M.; and Khalifa, R., 1978, Parasitology, v. 77 (3), 249-254
Mus musculus (peripheral blood): Rifa village, Assiut, Upper Egypt

Trypanosoma ataevi sp. n., illus.
Khaibulaev, K. Kh., 1971, Parazitologiia, Leningrad, v. 5 (6), 551-555
Gobius ratan goebeli (blood): Bekdash, Caspian Sea
G. kessleri gorlap (blood): Bekdash, Caspian Sea
G. fluviatilis pallasi (blood): Tyuleni Island, Caspian Sea
Benthophilus macrocephalus (blood): Tyuleni Island, Caspian Sea

Trypanosoma avium
Bennett, G. F.; Cameron, M.; and White, E., 1975, Canad. J. Zool., v. 53 (10), 1432-1442
hematozoa of passeriforms, prevalence, effect of climate, application of insecticide, and large-scale environmental alteration: New Brunswick
Bombycilla cedrorum
Cyanocitta cristata
Coccyzus erythropthalmus
Carpodacus purpureus
Melospiza georgiana
M. melodia
Spizella passerina
Zonotrichia albicollis
Hirundo rustica
Agelaius phoeniceus
Dolichonyx oryzivoros
Dumetella carolinensis
Parus hudsonicua
Dendroica coronata
D. magnolia
D. pensylvanica
D. petechia
D. tigrina
Geothlypis trichas
Mniotilta varia

ypanosoma avium.-- Continued.
Bennett, G. F.; Cameron, M.; and White, E., 1975, Canad. J. Zool., v. 53 (10), 1432-1442
Setophaga ruticilla
Vermivora peregrina
Wilsonia canadensis
W. pusilla
Dendrocopos pubescens
Catharus guttatus
C. ustulatus
Turdus migratorius

Empidonax trailii
E. sp.
Nuttallornis borealis
Vireo olivaceous
Pheucticus ludovicianus
Colaptes auratus
Catharus fuscescens
all from New Brunswick

panosoma avium
Bennett, G. F.; White, E. M.; and Williams, N. A., 1977, J. Wildlife Dis., v. 13 (3), 251-257
Ispidina picta
Ploceus nigerrimus
Chlorocichla flaviventris
Pycnonotus virens
Camaroptera brevicaudata
(blood of all): all from Uganda

panosoma avium, illus.
esser, S. S., 1977, Canad. J. Zool., v. 55 8), 1359-1367
Trypanosoma avium in Simulium rugglesi (hindgut, rectum), ultrastructure of epimastigote stages

panosoma avium Danil. et Lav.
skina, G. V., 1971, Parazitologiia, Leninrad, v. 5 (5), 470-473
Trypanosoma avium, infections in blackflies experimentally fed on infected rock partridges, leptomonad and trypanosome forms
Obuchovia albella (exper.)
Tetisimulium coarctata (exper.)
T. alajensis (exper.)
Odagmia ornata (exper.)
O. ferganica (exper.)
[Alectoris graeca]

panosoma avium Danilewsky 1885
illiams, N. A., 1978, J. Parasitol., v. 64 3), 556-558
Junco hyemalis
Dendroica petechia
Oporornis tolmei
Catharus ustalatus
Turdus migratorius
Empidonax difficilis
all from 16 km west of Courtenay, British Columbia

anosoma avium
lliams, N. A.; Calverley, B. K.; and Mahrt, L., 1977, J. Wildlife Dis., v. 13 (3), 226-9
Anas platyrhynchos
Anas acuta
(blood of all): all from Mackenzie Delta, Northwest Territories

T[rypanosoma] avium complex
Wink, M.; and Bennett, G. F., 1976, J. Wildlife Dis., v. 12 (4), 587-590
Cyanomitra olivacea
Nectarinia seimundi
Francolinus ahantensis
Passer griseus
Ploceus cucullatus
Pycnonotus barbatus
all from Ghana

Trypanosoma (Herpetosoma) bandicotti
Edrissian, Gh. H.; Farhang-Azad, A.; and Neronov, V. M., 1976, J. Wildlife Dis., v. 12 (4), 497
Nesokia indica: Iran

Trypanosoma barbatulae Leger (1904)
Letch, C. A., 1979, Parasitology, v. 79 (1), 107-117
Trypanosoma cobitis should be regarded as single species of trypanosome from 6 spp. of British fish on basis of morphology, isoenzyme patterns, and cross-transmission (by syringe passage of culture forms and by leech vector Hemiclepsis marginata), specific names T. phoxini, T. elegans, T. barbatulae, T. occidentalis, and T. langeroni "should be disregarded"

Trypanosoma batai sp. nov., illus.
Joshi, B. D., [1979], J. Animal Morph. and Physiol., v. 25 (1-2), 1978, 1-7
Labeo bata: river Gomati [and/or] Chinhat lake of Lucknow district

Trypanosoma (Trypanozoon) brucei
Afchain, D.; et al., 1978, Bull. World Health Organ., v. 56 (3), 389-401
Trypanosoma brucei, antigenic variation, review with recommendations (WHO memorandum)

Trypanosoma brucei
Aiyedun, B. A., 1977, Bull. Animal Health and Prod. Africa, v. 25 (3), 245-250
Trypanosoma gambiense, T. brucei, rats, induction of high level of immunity by administration of Freund's complete adjuvant a week before infection followed by cure with berenil

Trypanosoma brucei
Alexander, J.; and Phillips, R. S., 1978, Exper. Parasitol., v. 44 (1), 136-142
Leishmania mexicana, L. tropica major, lesion growth in mice was markedly inhibited by concurrent Trypanosoma brucei infections, possible mechanisms, may or may not have immunological basis

Trypanosoma brucei
Bacchi, C. J.; et al., 1979, J. Protozool., v. 26 (3), 484-488
Trypanosoma brucei, bloodstream trypomastigotes, synthesis and content of polyamines

Trypanosoma brucei subspp.
Baker, J. R., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (1), 123 [Letter]
Trypanosoma brucei subspp., primary-isolate coding of stocks from Tanzania and Ethiopia, original and new designations given

Trypanosoma brucei
Barbet, A. F.; and McGuire, T. C., 1978, Proc. National Acad. Sc., v. 75 (4), 1989-1993
Trypanosoma brucei, T. congolense, crossreacting determinants in variant-specific surface antigens

Trypanosoma brucei
Barry, J. D., 1979, J. Cell Sc., v. 37, 287-302
Trypanosoma brucei, capping of variable antigen, immunological and biological significance

Trypanosoma brucei, illus.
Barry, J. D.; et al., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (2), 205-208
Trypanosoma brucei, detection of multiple variable antigen types in metacyclic parasite populations, possible use in diagnosis and vaccination

Trypanosoma brucei, illus.
Barry, J. D.; and Vickerman, K., 1979, Exper. Parasitol., v. 48 (2), 313-324
Trypanosoma brucei, loss of variable antigens during transformation from bloodstream to procyclic forms in vitro

T[rypanosoma] brucei
Bauer, F.; Raether, W.; and Seeger, K., 1978, Cahiers Bleus Vet. (27), 265-271
protozoal disease in exper. hosts, enhanced effect of berenil + reverin vs. berenil alone

T[rypanosoma] brucei
Boreham, P. F. L., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 249-255
T[rypanosoma] brucei, rabbits, possible pathologic effects of kallikrein and kinin release early in infection, consideration of similar mechanisms in human infection

Trypanosoma (Trypanozoon) brucei
oreham, P. F. L.; and Facer, C. A., 1977, tschr. Parasitenk., v. 52 (3), 257-265
Trypanosoma brucei in rabbits, fibrin degradation products in urine, possible mechanisms of renal damage

Trypanosoma brucei
Boreham, P. F. L.; and Parry, M. G., 1979, Advances Biosciences, v. 17, 85-92
Trypanosoma brucei, rabbits, role of urinary and plasma kallikreins in pathogenesis, immune complexes

T[rypanosoma] brucei
Bourn, D.; and Scott, M., 1978, Trop. Animal Health and Prod., v. 10 (4), 191-203
trypanosomiasis, successful use of Zebu work oxen in agricultural development of tsetse infested land, environmental conditions, epizootiology of trypanosomiasis in oxen and in Glossina morsitans, strategic drug use (alternation of diminazene aceturate and isometamidium to control trypanosomes; rafoxanide to control helminths): Wollega province, western Ethiopia

Trypanosoma brucei
owman, I. B. R., 1974, Ciba Found. Symp., n.s. 20), 255-284
trypanosomes, intermediary metabolism, review

Trypanosoma brucei
Breese, S. S., jr.; and Hsu, K. C., 1973, Immunol. Series, v. 2, 141-167
immuno-electron microscopic applications of ferritin-tagging, review with brief mention of Plasmodium berghei and Trypanosoma brucei

Trypanosoma brucei Palimmer et Bradford, 1899
Butabaeva, M., 1979, Uzbek. Biol. Zhurnal (2), 50-53
[Citellus fulvus]: Kashkadar'insk oblast

Trypanosoma brucei
Buyst, H., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 349-359
Trypanosoma brucei, T. rhodesiense, human, prehistorical and historical background of trypanosomiasis in Africa with special reference to the Luangwa Valley

Trypanosoma brucei
Chang, K. P.; et al., 1978, J. Protozool., v. 25 (1), 145-149
methylglyoxal bis(guanylhydrazone) (MGBG), little in vitro effect on Blastocrithidia culicis, Crithidia oncopelti, and Leishmania spp. but complete inhibition of growth of Trypanosoma brucei, reduced parasitemia of T. brucei and T. congolense in rats but infections relapsed, tracer studies with T. brucei showed that MGBG interfered with nucleoside incorporation

Trypanosoma brucei
Cook, R. M., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (5), 554-555 [Letter]
Trypanosoma brucei, different surfaces examined for ability to support phagocytic activity by mouse peritoneal exudate cells, possible significance of phenomenon of surface phagocytosis in disease process

Trypanosoma brucei
Cook, R. M., 1979, Vet. Parasitol., v. 5 (2-3), 107-115
Trypanosoma brucei-infected rabbits, quantitation of acute phase serum protein Cx-reactive, effect of anti-inflammatory drug treatment

T[rypanosoma] brucei
Cross, G. A. M., 1977, Ann. Soc. Belg Med. Trop., v. 57 (4-5), 389-402
T[rypanosoma] spp., identification, purification, and characterization of class of surface glycoproteins which appear to be primary mediators of antigenic variations

Trypanosoma brucei, illus.
Cross, G. A. M., 1978, Proc. Roy. Soc. London, s. B, Biol. Sc. (1146), v. 202, 55-72
Trypanosoma brucei, antigenic variation, characterization of major cell surface antigens, variations in amino acid sequence

Trypanosoma brucei
Cross, G. A. M., 1979, Nature, London (5694), v. 277, 310-312
Trypanosoma brucei, crossreacting determinants in C-terminal region of variant surface antigens

Trypanosoma brucei, illus.
Cunningham, I.; and Taylor, A. M., 1979, J. Protozool., v. 26 (3), 428-432
Trypanosoma brucei, restoration of infectivity of various stocks cultivated at 28 C with tsetse fly salivary glands

rypanosoma brucei
Dargie, J. D.; et al., 1979, Research Vet. Sc., v. 26 (2), 245-247
Trypanosoma brucei, Ndama and Zebu cattle (exper.), blood volumes and erythrokinetics, susceptibility differences between breeds

rypanosoma brucei
Donelson, J. E.; Majiwa, P. A. O.; and Williams, R. O., 1979, Plasmid, v. 2 (4), 572-588
Trypanosoma brucei, kinetoplast DNA minicircles share regions of sequence homology

rypanosoma (Trypanozoon) brucei
Draeger, N.; and Mehlitz, D., 1978, Tropenmed. u. Parasitol., v. 29 (2), 223-233
Trypanosoma spp., wildlife, prevalence determined by parasitological and/or serological techniques, correlations with high and low tsetse fly density areas (for buffalo and lechwe) and with host age (for buffalo)
Syncerus caffer
Kobus leche
all from Northern Botswana

rypanosoma brucei
Dube, D. K.; et al., 1979, Biochim. et Biophys. Acta, v. 561 (1), 10-16
Trypanosoma brucei, detection and characterization of DNA polymerase

rypanosoma brucei
Esuruoso, G. O., 1975, Bull. Animal Health and Prod. Africa, v. 23 (3), 323-332
trypanosomiasis, N'dama cattle, observations on outbreaks, association with various epizootiological factors: western Nigeria

rypanosoma brucei, illus.
Facer, C. A.; et al., 1978, Exper. Parasitol., v. 44 (2), 249-261
Trypanosoma brucei, rabbits, renal pathology, glomerular changes result from deposition of soluble trypanosome immune complexes, tubular changes are typical of tissue ischemia, trypanosomiasis in rabbit could be valuable model

rypanosoma brucei
Fruit, J.; et al., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 257-266
Trypanosoma gambiense, humans, T. brucei, rats, immune complexes, characterization by radioimmunoprecipitation

ypanosoma brucei
Galvao-Castro, B.; Hochmann, A.; and Lambert, P. H., 1978, Clin. and Exper. Immunol., v. 33 (1), 12-24
Trypanosoma brucei, comparative immunopathological studies in normal, athymic nude, irradiated, or newborn mice, results show that immunodeficiency suppresses development of characteristic muscle lesions of African trypanosomiasis and that passive transfer of normal spleen cells or normal T lymphocytes could induce characteristic lesions in infected athymic nude mice and that anti-trypanosome antibody could partly do so

ypanosoma brucei
Garlough, S.; Mason, M.; and Sanchez, G., 1978, Comp. Biochem. and Physiol., v. 61B (1), 33-36
Trypanosoma brucei-infected rats, host-parasite interaction in metabolism of tyrosine

Trypanosoma brucei
Geigy, R.; and Kauffmann, M., 1977, Protozoology, v. 3, 103-107
Trypanosoma brucei, experimental transmission by Auchmeromyia larvae feeding on rats

Trypanosoma brucei, illus.
Ghiotto, V.; et al., 1979, Exper. Parasitol., v. 48 (3), 447-456
Trypanosoma brucei, morphometric changes and loss of infectivity and of surface coat during transformation of bloodstream forms to procyclic culture forms in vitro

Trypanosoma brucei
Gobert, J. G.; Le Bras, J.; and Savel, J., 1977, Compt. Rend. Soc. Biol., Paris, v. 171 (5), 1008-1017
Trypanosoma brucei, survival in Blabera fusca, necessity for presence of red blood cells, subsequent infectivity for mice, toxicity of mouse hemoglobin to cockroach, original in vivo observation technique for studying trypanosomes in cockroach haemocoel

Trypanosoma brucei subgroup
Goodwin, L. G., 1974, Ciba Found. Symp., n.s. (20), 107-124
African trypanosomiasis, mechanisms of pathogenesis, review

Trypanosoma brucei
Gray, A. R., 1977, Ann. Soc. Belg Med. Trop., v. 57 (4-5), 403-408
trypanosomes, particularly Trypanosoma gambiense, syringe-passaged and cyclically transmitted isolates, antigenic variations

Trypanosoma brucei
Greer, C. A.; Cain, G. D.; and Schottelius, B. A., 1979, J. Parasitol., v. 65 (5), 825-827
Trypanosoma brucei-infected rats, changes in vascular smooth muscle contractility

Trypanosoma brucei
Gruenberg, J.; et al., 1979, Biochem. and Biophys. Research Commun., v. 88 (3), 1173-1179
Trypanosoma brucei, interactions of liposomes with plasma membrane, implications for use of liposome-encapsulated drugs in chemotherapy

Trypanosoma brucei
Gruenberg, J.; Sharma, P. R.; and Deshusses, J., 1978, European J. Biochem., v. 89 (2), 461-469
Trypanosoma brucei, D-glucose transport

Trypanosoma brucei
Gutteridge, W. E.; Dave, D.; and Richards, W. H. G., 1979, Biochim. et Biophys. Acta, v. 582 (3), 390-401
Kinetoplastida spp., Plasmodium spp., conversion of dihydroorotate to orotate, mechanism of reaction different in these 2 groups of protozoa, possible target of chemotherapeutic attack

Trypanosoma brucei
Hannon, R. H.; and Parr, C. W., 1978, Comp. Biochem. and Physiol., v. 60B (2), 177-181
Trypanosoma brucei, T. vivax, bloodstream forms, phosphoglucose isomerases, partial purification and characterization, comparison of the two species, many similarities in kinetic properties but differences in thermal stability and in isoelectric point

Trypanosoma brucei
Hawking, F., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (6), 592-595
Trypanosoma congolense, circadian rhythm in numbers of parasites in blood of laboratory rodents, indisputable rhythms not found in T. vivax, T. brucei, and T. lewisi

Trypanosoma (T[rypanozoon]) brucei
Hecker, H.; and Boehringer, S., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 465-471
Trypanosoma (T[rypanozoon]) brucei, quantitative ultrastructural composition of various forms in mammalian blood and organs of Glossina vectors investigated morphometrically

Trypanosoma brucei, illus.
Henley, G. L.; Lee, C. M.; and Takeuchi, A., 1978, Ztschr. Parasitenk., v. 55 (3), 181-187
Trypanosoma brucei, apical part of flagellar pocket, freeze-cleaving and thin-sectioning techniques, possible role of neck region in pinocytosis

Trypanosoma brucei
Holmes, P. H.; et al., 1979, Immunology, v. 36 (3), 415-420
Trypanosoma brucei, method of labelling with [^{75}Se]-methionine, suitability for in vivo studies of immunological clearance, liver found to be principal site of phagocytosis in immune mice; method equally applicable to T. congolense

Trypanosoma brucei
Hudson, K. M.; and Terry, R. J., 1979, Parasite Immunol., v. 1 (4), 317-326
Trypanosoma brucei, chronically infected mice, relationships between course of infection, antigenic variation, and immunodepression of antibody responses to heterologous antigens

Trypanosoma brucei, illus.
Ikede, B. O., 1979, Research Vet. Sc., v. 26 (2), 145-151
Trypanosoma brucei, rams (exper.), genital lesions, histopathology

Trypanosoma brucei
Jadin, J. M.; et al., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 525-531
Trypanosoma cruzi intra- and extracellular forms, T. brucei extracellular forms, mice, ethidium bromide vs. ethidium bromide-DNA complexes as therapy

Trypanosoma brucei, illus.
Jadin, J. M.; and Lumsden, W. H. R., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 459-463
Trypanosoma brucei, electron microscopic study of filopodium-like processes of trypomastigotes in infected mouse blood, thought to be herniation of cellular wall

T[rypanosoma] brucei
James, D. M., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (5), 471-476
T[rypanosoma] congolense, T. brucei, rats, mice, prophylactic activity of various trypanocides complexed with dextran, comparison with uncomplexed drugs and with suramin-complexed drugs

Trypanosoma brucei
Jayawardena, A. N.; Waksman, B. H.; and Eardley, D. D., 1978, J. Immunol., v. 121 (2), 622-628
Trypanosoma brucei, mice, activation of distinct helper and suppressor T cells, significance in relation to pathogenesis of trypanosomiasis

Trypanosoma brucei
Jennings, F. W.; et al., 1978, Research Vet. Sc., v. 25 (3), 399-400
Trypanosoma congolense, T. brucei, survival time of various strains of mice, C57 Bl mouse might provide laboratory model for study of trypanotolerance in cattle

Trypanosoma brucei
Jennings, F. W.; at al., 1979, Internat. J. Parasitol., v. 9 (4),381-384
Trypanosoma brucei, brain as source of relapsing infection in mice after berenil chemotherapy

Trypanosoma brucei
Johnson, J. G.; and Cross, G. A. M., 1979, Biochem. J., v. 178 (3), 689-697
Trypanosoma brucei, selective cleavage of variant surface glycoproteins

Trypanosoma (Trypanozoon) brucei
Le Ray, D., 1975, Ann. Soc. Belge Med. Trop., v. 55 (3), 129-311
Trypanosoma brucei, antigenic structures expressed in course of parasite life cycle, immunoelectrophoretic analysis and comparative study with various taxa within the family and phylum, extensive review

Trypanosoma brucei
Louis, J.; et al., 1978, Pharmacol. Immunoreg., 225-238
Trypanosoma brucei, mice, strong polyclonal B cell activation, appearance of autoantibodies with various specificites

Trypanosoma brucei
Luckins, A. G., 1977, Trop. Animal Health and Prod., v. 9 (1), 53-62
Trypanosoma spp., cattle, immunodiagnosis, micromodification of enzyme-linked immunosorbent assay, test did not distinguish between different trypanosome species but there were no cross-reactions with other protozoal antigens

Trypanosoma brucei
Luckins, A. G.; and Mehlitz, D., 1978, Trop. Animal Health and Prod., v. 10 (3), 149-159
trypanosomiasis, cattle, diagnosis, indirect fluorescent antibody test, enzyme-linked immunosorbent assay, and serum IgM levels compared: Liberia

Trypanosoma brucei
Lumsden, W. H. R.; et al., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (3), 312-317
Trypanosoma brucei, man, diagnosis using the miniature anion-exchange/centrifugation method, adaptation for field use: The Gambia

Trypanosoma (Trypanozoon) brucei
Lumsden, W. H. R.; Kimber, C. D.; and Doig, S., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (2), 214 [Letter]
Trypanosoma brucei, use of miniature anion-exchange/centrifugation technique to recover trypanosomes from tissue suspensions

rypanosoma brucei group
MacKenzie, P. K. I.; Boyt, W. P.; and Nesham, V. W., 1979, Brit. Vet. J., v. 135 (2), 178-184
trypanosomiasis, sheep (nat. and exper.), serum immunoglobulin levels during course of infection

rypanosoma (Trypanozoon) brucei
Martin, B. R.; Voorheis, H. P.; and Kennedy, E. L., 1978, Biochem. J., v. 175 (1), 207-212
Trypanosoma brucei, bloodstream forms, adenylate cyclase, location in plasma membrane, characterization

ypanosoma brucei
Mayor Withey, K. S.; et al., 1978, Clin. and Exper. Immunol., v. 34 (3), 359-363
Trypanosoma brucei, mice, extensive proliferation of B, T, and null cells in spleen and bone marrow, still unclear whether there is any primary target cell for immunosuppression

ypanosoma brucei, illus.
Meshnick, S. R.; Chang, K. P.; and Cerami, A., 1977, Biochem. Pharmacol., v. 26 (20), 1923-1928
Trypanosoma brucei, T. congolense, heme lysis of bloodstream forms, T. brucei, lytic effect of porphyrins, in vitro and in vivo (mice) studies, mechanism of action believed to be homolytic cleavage of intracellular H_2O_2 to form hydroxyl radicals which can react with vital cell components and kill the organism

ypanosoma brucei
Molyneux, D. H., 1973, Ann. Soc. Belge Med. Trop., v. 53 (6), 605-618
Gambian trypanosomiasis, evidence for existence of animal reservoirs of Trypanosoma brucei gambiense reviewed; use of modified blood incubation infectivity test for diagnosis of T. brucei subspp. suggested; recommendations for long-term study of man and various animal groups as sources of various strains

panosoma brucei
oors, A., 1972, Ann. Soc. Belge Med. Trop., v. 2 (1), 9-17
Trypanosoma brucei, changes in allantoic fluid composition during development of chick embryo that influence viability of parasite, increased phosphodiesterases main causal factor for disappearance of trypanosomes during embryonic development

panosoma brucei
rales, N. M.; and Roberts, J. F., 1978, omp. Biochem. and Physiol., v. 59B (1), 1-4
Crithidia fasciculata, Leishmania tropica, Trypanosoma brucei, comparison of ribosomal RNAs, possible evolutionary significance

panosoma brucei
oulton, J. E.; and Coleman, J. L., 1979, Am. . Vet. Research, v. 40 (8), 1131-1133
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pigs
all from rain forest areas, Liberia

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African trypanosomiasis, immunoenzymatic diagnosis comparing purified exoantigen and 5 crude extracts from strains of Trypanosoma b. brucei and T. b. gambiense; best sensitivity obtained with virulent strains of human origin

Trypanosoma brucei gambiense
Seed, J. R., 1978, J. Protozool., v. 25 (4), 526-529
Trypanosoma brucei gambiense, different antigenic variants compete with each other in vivo (mice) and one clone replaces another, this is a predictable phenomenon even under conditions analogous to chronic infection

Trypanosoma brucei gambiense
Seed, J. R.; et al., 1978, Am. Midland Naturalist, v. 100 (1), 126-134
Trypanosoma brucei gambiense-infected wild and laboratory Microtus montanus males, organ weights, parasite stress as cause of enlarged spleens and smaller gonads, splenomegaly can be used as survey marker to determine extent of parasitism in field populations, reduced reproductive potential suggests that parasitism plays role in limiting host population density: Jackson Hole, Wyoming

Trypanosoma brucei gambiense
Tizard, I.; et al., 1978, Microbiol. Rev., v. 42 (4), 661-681
African trypanosomes, biologically active products and pathogenesis

T[rypanosoma] (T[rypanozoon]) brucei gambiense
Van Meirvenne, N.; et al., 1975, Ann. Soc. Belge Med. Trop., v. 55 (5), 545-549
gambian and rhodesian sleeping sickness, humans, serological and parasitological diagnostic methods, review

Trypanosoma brucei gambiense
Wery, M.; et al., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 425-439
Trypanosoma brucei gambiense strains from Zaire, comparative infectivity in various laboratory animals

[Trypanosoma brucei gambiense]
van Wettere, P.; et al., 1978, PANS, v. 24 (4), 435-446
[Trypanosoma brucei gambiense], control of Glossina tachinoides, evaluation of insecticides applied as aerosols from helicopters: Komoe valley, Upper Volta, West Africa

rypanosoma brucei rhodesiense
Baker, J. R., 1974, Ciba Found. Symp., n.s. (20), 29-50
African sleeping sickness, epidemiology, review

rypanosoma (Trypanozoon) brucei rhodesiense
Barry, J. D.; Le Ray, D.; and Herbert, W. J., 1979, J. Comp. Path., v. 89 (4), 465-470
Trypanosoma brucei, white mice, dissociation of virulence and variable antigen type in relation to pleomorphism

rypanosoma brucei rhodesiense
Basson, W.; Page, M. L.; and Myburgh, D. P., 1977, South African Med. J., v. 51 (14), 453-457
trypanosomiasis, humans, problems in diagnosis and management emphasizing haematological and immunological aspects, clinical course in 5 cases, indications of continuation of immune complex process in spite of therapy that eradicates parasite: Voortrekkerhoogte, Tvl.

Trypanosoma brucei rhodesiense, illus.
Campbell, G. H.; et al., 1979, Am. J. Trop. Med. and Hyg., v. 28 (6), 974-983
Trypanosoma brucei rhodesiense, isolation and characterization of new serodeme, model for further study of immunopathology and antigenic variation in African trypanosomiasis

Trypanosoma brucei rhodesiense
D'hondt, J.; et al., 1979, Nature, London (5739), v. 282, 613-615
Trypanosoma brucei subspp., trypanocidal activity of normal human serum: Ca^{2+} is essential cofactor, α_2 macroglobulin might function as Ca^{2+} carrier, suppression by D-glucose, D-fructose, and D-mannose, glycerol has opposite effect

Trypanosoma brucei rhodesiense
Diffley, P., 1978, Infect. and Immun., v. 21 (2), 605-612
Trypanosoma brucei subspp., comparative immunological analysis of host plasma proteins bound to bloodstream forms (presence, location, host specificity, identity, and quantity)

Trypanosoma brucei rhodesiense
Evans, D. A., 1979, J. Protozool., v. 26 (3), 425-427
Trypanosoma brucei rhodesiense, T. congolense, cyclical transmission by Glossina morsitans morsitans infected with culture-form procyclic trypanosomes

Trypanosoma brucei rhodesiense, illus.
Evans, D. A.; and Ellis, D. S., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (6), 653-655
Trypanosoma brucei rhodesiense, evidence of active penetration and passage of trypanosomes across midgut cells of Glossina morsitans morsitans rather than passive uptake

Trypanosoma (Trypanozoon) brucei rhodesiense
Fife, E. H., jr., 1976, Immunol. Parasit. Infect., 77-93
African and American trypanosomiasis, serodiagnosis, review

Trypanosoma brucei rhodesiense, illus.
Fink, E.; and Schmidt, H., 1979, Tropenmed. u. Parasitol., v. 30 (2), 206-211
Trypanosoma brucei rhodesiense, EATRO 1989 strain in white mice induced chronic infection with meningoencephalitis similar to infection in humans, suitable model for studying human infection and screening drug compounds for activity during late stages of infections

Trypanosoma brucei rhodesiense
Luckins, A. G., 1977, Trop. Animal Health and Prod., v. 9 (1), 53-62
Trypanosoma spp., cattle, immunodiagnosis, micromodification of enzyme-linked immunosorbent assay, test did not distinguish between different trypanosome species but there were no cross-reactions with other protozoal antigens

Trypanosoma (Trypanozoon) brucei rhodesiense
Molyneux, D. H.; Baldry, D. A. T.; and Fairhurst, C., 1979, Acta Trop., v. 36 (1), 53-65
human West African trypanosomiasis, suggested role of wind-dispersed Glossina in distribution, entomological and epidemiological evidence, implications for control operations, significance of these ideas in relation to origin of Trypanosoma brucei rhodesiense in Ethiopia and of T. evansi outside the tsetse zone

Trypanosoma brucei rhodesiense, illus.
Rickman, W. J.; and Cox, H. W., 1979, J. Parasitol., v. 65 (1), 65-73
Trypanosoma brucei rhodesiense-infected rats, syndrome characterized by anemia, splenomegaly, and glomerulonephritis, accompanied by presence of 3 autoantibodies and by presence of fixed complement and fibrinogen on trypanosomes and erythrocytes

Trypanosoma brucei rhodesiense
Tizard, I.; et al., 1978, Microbiol. Rev., v. 42 (4), 661-681
African trypanosomes, biologically active products and pathogenesis

Trypanosoma brucei rhodesiense
Traub, N.; et al., 1978, East African Med. J., v. 55 (10), 477-481
Trypanosoma brucei rhodesiense, human congenital, fatal infection in mother, infant successfully treated with suramin and mel-B, immunoglobulin levels at diagnosis, during treatment, and post-treatment, case reports: Zambia

Trypanosoma brucei rhodesiense
Van Meirvenne, N.; Magnus, E.; and Janssens, P. G., 1976, Ann. Soc. Belge Med. Trop., v. 56 (1), 55-63
Trypanosoma brucei rhodesiense, serum-incubation-infectivity-tests on clone populations of distinct antigenic types

Trypanosoma cancili sp. nov., illus.
Mandal, A. K., 1978, Ang. Parasitol., v. 19 (3), 158-162
Xenentodon cancila (blood): Raidighi, India

Trypanosoma cecili n. sp., illus.
Lainson, R., 1977, Protozoology, v. 3, 87-93
Caiman crocodilus crocodilus (blood): Barcarena, north Para State, Brazil

Trypanosoma cervi Kingston and Morton, 1975, illus.
Kingston, N.; and Crum, J., 1977, Proc. Helminth. Soc. Washington, v. 44 (2), 179-184
Odocoileus virginianus (blood): Georgia; North Carolina; Alabama

Trypanosoma chattoni Mathis and Leger, 1911, illus.
Baker, J. R., 1977, Protozoology, v. 3, 75-82
Ptychadena mascareniensis
Phrynobatrachus natalensis
all from Illubabor Province, Ethiopia

Trypanosoma cobitis (Mitrophanow, 1883)
Letch, C. A., 1979, Parasitology, v. 79 (1), 107-117
Trypanosoma cobitis should be regarded as single species of trypanosome from 6 spp. of British fish on basis of morphology, isoenzyme patterns, and cross-transmission (by syringe passage of culture forms and by leech vector Hemiclepsis marginata), specific names T. phoxini, T. elegans, T. barbatulae, T. occidentalis, and T. langeroni "should be disregarded"
Nemacheilus barbatulus
Phoxinus phoxinus
Cottus gobio
Gobio gobio
Gasterosteus aculeatus
Pungitus pungitus
(blood of all): all from River Lee at Enfield Lock

Trypanosoma cobitis Mitrophanow, 1883
Letch, C. A.; and Ball, S. J., 1979, Parasitology, v. 79 (1), 119-124
Trypanosoma cobitis, prevalence in fish, seasonal fluctuation, host age
Nemacheilus barbatulus
Gobio gobio
Cottus gobio
Phoxinus phoxinus
Gasterosteus aculeatus
Pungitius pungitius
Carassius carassius
(blood of all): all from River Lee at Enfield Lock

Trypanosoma congolense
Assoku, R. K. G.; Hazlett, C. A.; and Tizard, I., 1979, Internat. Arch. Allergy and Applied Immunol., v. 59 (3), 298-307
Trypanosoma congolense, mice, significant depression of humoral immunity, simultaneous increase in background IgM plaque-forming cell levels, mitogenicity of trypanosome-derived saturated fatty acids

Trypanosoma congolense
Assoku, R. K. G.; and Tizard, I. R., 1978, Experientia, v. 34 (1), 127-129
Trypanosoma congolense, autolysates found to be highly mitogenic for spleen cells of normal and nude but not cyclophosphamide-treated mice, possible role of trypanosome-derived mitogen in immunosuppression associated with African trypanosomiasis

Trypanosoma congolense
Balber, A. E.; et al., 1979, Infect. and Immun., v. 24 (3), 617-627
Trypanosoma brucei brucei, T. congolense, inactivation or elimination of potentially trypanolytic complement-activating immune complexes containing antibodies to variant-specific antigens

Trypanosoma congolense, illus.
Banks, K. L., 1978, J. Protozool., v. 25 (2), 241-245
Trypanosoma congolense, rats, rabbits, localization of parasite in microvasculature is established by attachment of the organism to the vessel wall

Trypanosoma congolense, illus.
Banks, K. L., 1979, J. Protozool., v. 26 (1), 103-108
Trypanosoma congolense, in vitro binding to erythrocytes: method of studying trypanosome-host cell interaction, erythrocyte and trypanosome surface properties necessary for adhesion

Trypanosoma congolense
Barbet, A. F.; and McGuire, T. C., 1978, Proc. National Acad. Sc., v. 75 (4), 1989-1993
Trypanosoma brucei, T. congolense, crossreacting determinants in variant-specific surface antigens

T[rypanosoma] congolense
Bauer, F.; Raether, W.; and Seeger, K., 1978, Cahiers Bleus Vet. (27), 265-271
protozoal disease in exper. hosts, enhanced effect of berenil + reverin vs. berenil alone

T[rypanosoma] congolense
Bourn, D.; and Scott, M., 1978, Trop. Animal Health and Prod., v. 10 (4), 191-203
trypanosomiasis, successful use of Zebu work oxen in agricultural development of tsetse infested land, environmental conditions, epizootiology of trypanosomiasis in oxen and in Glossina morsitans, strategic drug use (alternation of diminazene aceturate and isometamidium to control trypanosomes; rafoxanide to control helminths): Wollega province, western Ethiopia

Trypanosoma congolense
Bowman, I. B. R., 1974, Ciba Found. Symp., n.s. (20), 255-284
trypanosomes, intermediary metabolism, review

Trypanosoma congolense
Chang, K. P.; et al., 1978, J. Protozool., v. 25 (1), 145-149
methylglyoxal bis(guanylhydrazone) (MGBG), little in vitro effect on Blastocrithidia culicis, Crithidia oncopelti, and Leishmania spp. but complete inhibition of growth of Trypanosoma brucei, reduced parasitemia of T. brucei and T. congolense in rats but infections relapsed, tracer studies with T. brucei showed that MGBG interfered with nucleoside incorporation

Trypanosoma congolense
Dargie, J. D.; et al., 1979, Parasitology, v. 78 (3), 271-286
Trypanosoma congolense-infected Ndama and Zebu cattle, red cell kinetics, concluded that anemia and its underlying processes are broadly in line with number of parasites in blood and that superior resistance of Ndama cattle lies in ability to control parasitemia rather than capacity to mount more efficient erythropoietic response

Trypanosoma congolense
Diffley, P.; and Honigberg, B. M., 1978, J. Parasitol., v. 64 (4), 674-681
Trypanosoma congolense, identification and quantitation of host albumin, nonspecific IgG, and complement (C3) bound to surface of bloodstream forms, possible functions for these surface-bound plasma proteins

Trypanosoma (Nannomonas) congolense
Draeger, N.; and Mehlitz, D., 1978, Tropenmed. u. Parasitol., v. 29 (2), 223-233
Trypanosoma spp., wildlife, prevalence determined by parasitological and/or serological techniques, correlations with high and low tsetse fly density areas (for buffalo and lechwe) and with host age (for buffalo)
Syncerus caffer
Kobus leche
Redunca arundium
all from Northern Botswana

Trypanosoma congolense
Esuruoso, G. O., 1975, Bull. Animal Health and Prod. Africa, v. 23 (3), 323-332
trypanosomiasis, N'dama cattle, observations on outbreaks, association with various epizootiological factors: western Nigeria

Trypanosoma congolense
Evans, D. A., 1979, J. Protozool., v. 26 (3), 425-427
Trypanosoma brucei rhodesiense, T. congolense, cyclical transmission by Glossina morsitans morsitans infected with culture-form procyclic trypanosomes

Trypanosoma congolense, illus.
Evans, D. A.; Ellis, D. S.; and Stamford, S., 1979, J. Protozool., v. 26 (4), 557-563
Trypanosoma congolense, development in Glossina morsitans morsitans, ultrastructure

Trypanosoma congolense
Forsberg, C. M.; et al., 1979, Vet. Path., v. 16 (2), 229-242
Trypanosoma congolense, calves, kinetics of blood coagulation

Trypanosoma congolense
Goodwin, L. G., 1974, Ciba Found. Symp., n.s. (20), 107-124
African trypanosomiasis, mechanisms of pathogenesis, review

Trypanosoma congolense
Gray, M. A.; et al., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (4), 406-408
Trypanosoma congolense, maintenance of infectivity in vitro with explants of infected skin incubated at 37°C

Trypanosoma congolense
Greig, W. A.; et al., 1979, Brit. Vet. J., v. 135 (2), 130-141
Trypanosoma congolense, T. vivax, cattle, factors affecting blood sampling for parasitemia and anemia (diurnal variation; ear-vein vs. jugular-vein blood): The Gambia

Trypanosoma congolense
Griffin, L.; and Allonby, E. W., 1979, Vet. Parasitol., v. 5 (2-3), 97-105
Trypanosoma congolense, sheep, goats, susceptibility of various breeds to experimental infection

Trypanosoma (Nannamonas) congolense
Griffin, L.; and Allonby, E. W., 1979, J. Comp. Path., v. 89 (4), 457-464
Trypanosoma congolense, sheep and goats, description of acute, sub-acute, and chronic disease syndromes: Kenya

Trypanosoma congolense
Gutteridge, W. E.; Dave, D.; and Richards, W. H. G., 1979, Biochim. et Biophys. Acta, v. 582 (3), 390-401
Kinetoplastida spp., Plasmodium spp., conversion of dihydroorotate to orotate, mechanism of reaction different in these 2 groups of protozoa, possible target of chemotherapeutic attack

Trypanosoma congolense
Hawking, F., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (4), 405-407
Trypanosoma evansi, sensitivity to human plasma; T. congolense, T. vivax, resistance to human plasma, suggests that latter 2 species might produce transient infection in man

Trypanosoma congolense
Hawking, F., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (6), 592-595
Trypanosoma congolense, circadian rhythm in numbers of parasites in blood of laboratory rodents, indisputable rhythms not found in T. vivax, T. brucei, and T. lewisi

Trypanosoma congolense
Holmes, P. H.; et al., 1979, Immunology, v. 36 (3), 415-420
Trypanosoma brucei, method of labelling with [^{75}Se]-methionine, suitability for in vivo studies of immunological clearance, liver found to be principal site of phagocytosis in immune mice; method equally applicable to T. congolense

Trypanosoma congolense, illus.
Jackson, P. R.; Honigberg, B. M.; and Holt, S. C., 1978, J. Protozool., v. 25 (4), 471-481
Trypanosoma congolense, bloodstream trypomastigotes and culture procyclics, lectin analysis of surface saccharides by agglutination and electron microscopic techniques

T[rypanosoma] congolense
James, D. M., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (5), 471-476
T[rypanosoma] congolense, T. brucei, rats, mice, prophylactic activity of various trypanocides complexed with dextran, comparison with uncomplexed drugs and with suramin-complexed drugs

Trypanosoma congolense
Jennings, F. W.; et al., 1978, Research Vet. Sc., v. 25 (3), 399-400
Trypanosoma congolense, T. brucei, survival time of various strains of mice, C57 Bl mouse might provide laboratory model for study of trypanotolerance in cattle

Trypanosoma congolense
Kaaya, G. P.; et al., 1979, Tropenmed. u. Parasitol., v. 30 (2), 230-235
Trypanosoma vivax, T. congolense, serum from infected cattle inhibited bovine granulocyte/macrophage colony formation in methyl cellulose culture, degree of inhibition appeared related to degree of parasitemia; no inhibitors of erythropoiesis were observed

Trypanosoma congolense
Kaaya, G. P.; Winqvist, G.; and Johnson, L. W., 1977, Bull. Animal Health and Prod. Africa, v. 25 (4), 397-408
Trypanosoma congolense, goats, clinico-pathological aspects with particular reference to pathogenesis of anemia, changes in peripheral blood and in bone marrow described in detail

Trypanosoma congolense
Kamara, J. A., 1975, Bull. Animal Health and Prod. Africa, v. 23 (3), 265-268
Tragelaphus s. scriptus: Sierra Leone

Trypanosoma congolense
Luckins, A. G., 1977, Trop. Animal Health and Prod., v. 9 (1), 53-62
Trypanosoma spp., cattle, immunodiagnosis, micromodification of enzyme-linked immunosorbent assay, test did not distinguish between different trypanosome species but there were no cross-reactions with other protozoal antigens

Trypanosoma congolense, illus.
Luckins, A. G.; and Gray, A. R., 1978, Nature, London (5654), v. 272, 613-614
Trypanosoma congolense, development in local skin reactions on rabbits at sites of infection transmitted by Glossina morsitans, evidence from light and electron microscopy that these trypanosomes develop in connective tissue and are not confined to the vascular system, implications

Trypanosoma congolense, illus.
Luckins, A. G.; and Gray, A. R., 1979, Parasitology, v. 79 (3), 337-347
Trypanosoma congolense, stocks from East and West Africa, antigenicity and serological relationships

Trypanosoma congolense, illus.
Luckins, A. G.; and Gray, A. R., 1979, Research Vet. Sc., v. 27 (1), 129-131
Trypanosoma congolense, sheep, Ayrshire calves (both exper.) (lymph nodes of both); should not be regarded as strict plasma parasite

Trypanosoma congolense
Luckins, A. G.; and Mehlitz, D., 1978, Trop. Animal Health and Prod., v. 10 (3), 149-159
trypanosomiasis, cattle, diagnosis, indirect fluorescent antibody test, enzyme-linked immunosorbent assay, and serum IgM levels compared: Liberia

Trypanosoma congolense (Broden, 1904)
Mackenzie, P. K. I.; et al., 1978, Research Vet. Sc., v. 24 (1), 4-7
Trypanosoma congolense, phagocytosis of erythrocytes and leucocytes in infected sheep is due to coating of blood cells with trypanosomal antigen, phagocytic activity is dependent on parasitemia and is significant in maintaining host anemia

Trypanosoma congolense
MacKenzie, P. K. I.; Boyt, W. P.; and Nesham, V. W., 1979, Brit. Vet. J., v. 135 (2), 178-184
trypanosomiasis, sheep (nat. and exper.), serum immunoglobulin levels during course of infection

Trypanosoma congolense
Maxie, M. G.; Tabel, H.; and Losos, G. J., 1978, Tropenmed. u. Parasitol., v. 29 (2), 234-238
Trypanosoma vivax, T. congolense, determination of volume of parasites separated from cattle blood during first growth phase of infections

Trypanosoma (Nannomonas) congolense
Mehlitz, D., 1979, Tropenmed. u. Parasitol., v. 30 (2), 212-219
Trypanosoma spp., infection rates in domestic animals; screening of dogs and pigs to indicate potential reservoirs of T. (Trypanozoon) brucei gambiense; comparison of sensitivity of 3 diagnostic techniques; comparison of number of primary isolations and derived stabilated Trypanosoma stocks
cattle
goats
sheep
pigs
dogs
all from rain forest areas, Liberia

Trypanosoma congolense
Meshnick, S. R.; Chang, K. P.; and Cerami, A., 1977, Biochem. Pharmacol., v. 26 (20), 1923-1928
Trypanosoma brucei, T. congolense, heme lysis of bloodstream forms, T. brucei, lytic effect of porphyrins, in vitro and in vivo (mice) studies, mechanism of action believed to be homolytic cleavage of intracellular H_2O_2 to form hydroxyl radicals which can react with vital cell components and kill the organism

T[rypanosoma] congolense
Mills, J. N.; and Valli, V. E. O., 1978, Tropenmed. u. Parasitol, v. 29 (1), 95-100
T[rypanosoma] congolense, rapid automated cytofluorometric method of counting trypanosomes

Trypanosoma (Nannomonas) congolense, illus.
Molyneux D. H.; Lavin, D. R.; and Elce, B. A., 1979, Ann. Trop. Med. and Parasitol., v. 73 (3), 287-290
Trypanosoma congolense-infected Glossina morsitans, study by stereoscan electron micrographs suggests close association between salivarian trypanosomes and host mechano-receptors, possible higher probing rate in infected flies

Trypanosoma congolense
Morrison, W. I.; et al., 1978, Clin. and Exper. Immunol., v. 32 (1), 25-40
Trypanosoma congolense, marked differences in susceptibility of inbred strains of mice to infection, correlation with changes in spleen lymphocyte populations

Trypanosoma congolense
Morrison, W. I.; and Murray, M., 1979, Exper. Parasitol., v. 48 (3), 364-374
Trypanosoma congolense, mouse strains, genetic basis of observed differences in susceptibility to infection examined with F1 hybrids and backcrosses, influence of H-2 haplotype on susceptibility

Trypanosoma congolense
Murray, M.; and Morrison, W. I., 1979, Parasitology, v. 79 (3), 349-366
Trypanosoma congolense, Trypanosoma brucei, non-specific induction of increased resistance in mice by immunostimulants

Trypanosoma congolense
Nielsen, K.; et al., 1978, Immunology, v. 35 (5), 811-816
Trypanosoma congolense-infected calves, changes in catabolism of serum immunoglobulins and complement components, possible relationship to pathological changes

Trypanosoma congolense
Nielsen, K.; et al., 1978, Immunology, v. 35 (5), 817-826
Trypanosoma congolense-infected calves, changes in serum immunoglobulins, complement, and complement components

Trypanosoma congolense
Nielsen, K.; et al., 1978, J. Parasitol., v. 64 (3), 544-546
Trypanosoma congolense, T. lewisi, direct activation of complement

T[rypanosoma] congolense
Pearson, T. W.; et al., 1978, European J. Immunol., v. 8 (10), 723-727
T[rypanosoma] congolense-infected mice, depressed T lymphocyte responses

Trypanosoma congolense
Pearson, T. W.; et al., 1979, European J. Immunol., v. 9 (3), 200-204
Trypanosoma congolense, suppression of both B and T lymphocyte responses by spleen cells from infected mice, mechanism of suppression is complex and probably involves more than 1 cell type

Trypanosoma congolense
Preston, J. M.; Wellde, B. T.; and Kovatch, R. M., 1979, Exper. Parasitol., v. 48 (1), 118-125
Trypanosoma congolense-infected calves, kinetics of anemia, important hemolytic component

Trypanosoma congolense
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Trypanosoma congolense, illus.
Reinwald, E.; Rautenberg, P.; and Risse, H. J., 1979, Exper. Parasitol., v. 48 (3), 384-397
Trypanosoma congolense, mechanical removal of surface coat in vitro

Trypanosoma congolense
Reinwald, E.; Risse, H. J.; and Saelker, R., 1978, Hoppe-Seyler's Ztschr. Physiol. Chem., v. 359 (8), 939-944
Trypanosoma congolense, radio-labelled diazoniobenzenesulfonate as marker for cell surface proteins, results indicate that surface coat is homogeneous layer composed of molecules of one type of protein

Trypanosoma congolense
Roelants, G. E.; et al., 1979, Clin. and Exper. Immunol., v. 37 (3), 457-469
Trypanosoma congolense-infected mice, numbers of parasites in peripheral blood, changes in spleen cell populations, immune depression, suppressor cell activity, changes after berenil treatment

Trypanosoma congolense
Roelants, G. E.; et al., 1979, European J. Immunol., v. 9 (3), 195-199
Trypanosoma congolense-infected mice, composition of spleen cell populations used for functional assays, drastic reduction in B and T lymphocyte function cannot be due simply to dilution of relevant cells by null cells

Trypanosoma (Nannomonas) congolense, illus.
Rosen, N. L.; et al., 1979, Exper. Parasitol., v. 47 (3), 378-383
Trypanosoma congolense, isolation and purification: methods for reducing peripheral distribution of trypanosomes, for increasing yield of infected blood from each rat, and for selectively lysing erythrocytes; latter 2 methods are equally applicable to T. brucei

Trypanosoma congolense
Rovis, L.; Barbet, A. F.; and Williams, R. O., 1978, Nature, London (5646), v. 271, 654-656
Trypanosoma congolense, characterization of surface coat, single specific glycoprotein as surface antigen, overall similarities with surface coat of T. brucei

Trypanosoma congolense
Rurangirwa, F. R.; et al., 1978, Research Vet. Sc., v. 25 (3), 395-397
Trypanosoma congolense, T. vivax, cattle, immunosuppressive effect on secondary humoral immune response to Mycoplasma m. mycoides

Trypanosoma congolense
Rurangirwa, F. R.; et al., 1979, Infect. and Immun., v. 26 (3), 822-826
Trypanosoma congolense- or T. vivax-infected Bos indicus, suppression of antibody response to Leptospira biflexa and Brucella abortus and recovery from immunosuppression after berenil treatment

Trypanosoma congolense
Scott, J. M.; et al., 1978, Research Vet. Sc., v. 25 (1), 115-117
Trypanosoma congolense, zebu cattle, attempted protection using multi-stabilate vaccine given either as live or dead organisms followed by trypanocidal therapy, disappointing results

Trypanosoma congolense
Staak, C.; and Lohding, A., 1979, Tropenmed. u. Parasitol., v. 30 (1), 13-18
Trypanosoma spp., evaluation of complement fixation antibody in African trypanosomiasis-infected cattle before and after treatment and followed by re-infection and treatment; applications for field studies

Trypanosoma congolense
Tizard, I.; et al., 1978, Microbiol. Rev., v. 42 (4), 661-681
African trypanosomes, biologically active products and pathogenesis

Trypanosoma congolense, illus.
Tizard, I. R.; et al., 1978, Tropenmed. u. Parasitol., v. 29 (1), 127-133
Trypanosoma congolense, concomitant generation of phospholipase A and hemolytic fatty acids by autolysing suspensions, autolysis of T. lewisi did not generate hemolytic activity unless exogenous phospholipase A was added

Trypanosoma congolense
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Trypanosoma congolense
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Trypanosoma spp., cattle, mice, suppressed antibody response to louping-ill vaccine, value of diminazene therapy in alleviating this effect

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Trypanosoma conorhini, illus.
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Trypanosoma conorhini
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Crocidura russula
Crocidura sp.
all from Iran

Trypanosoma cruzi
Afchain, D.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (3), 478-482
Trypanosoma cruzi, preparation of pure specific antigen and corresponding monospecific antiserum using immunoadsorption

Trypanosoma (Schizotrypanum) cruzi
Afchain, D.; et al., 1979, J. Parasitol., v. 65 (4), 507-514
Trypanosoma cruzi culture forms, antigenic make-up, comparison with salivarian and some other stercorarian trypanosomes and Leishmania using immunoprecipitation in gels and immunoelectrophoresis, identification of component specific to T. cruzi

Trypanosoma cruzi, illus.
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Dusicyon vetulus: Franca, State of Sao Paulo, Brazil
camundongos (exper.)
P[anstrongylus] megistus (exper.)
T[riatoma] infestans (exper.)
T. vitticeps (exper.)
R[hodnius] neglectus (exper.)

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Amato Neto, V.; et al., 1975, Rev. Goiana Med., v. 21 (1-2), 1-9
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Amato Neto, V.; et al., 1976, Rev. Med. IAMPSE, v. 7 (3-4), 3-6
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Trypanosoma cruzi, re-examination of person from whom extensively studied virulent Y strain had been isolated 23 years earlier, typical signs of chronic infection not evident

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Trypanosoma cruzi, characteristics of strain isolated in area of Salvador (virulence, tissue tropism, morphology, pathology), comparison with characteristics of strains from other areas, extensive bibliography: Bahia, Brazil

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Trypanosoma cruzi
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Trypanosoma cruzi, mice with chronic infection with Colombian strain, inoculation with virulent Y strain, results demonstrate that chronic infection produces partial immunity and that co-existence of parasite strains is possible

Trypanosoma cruzi
Andrade, S. G.; et al., 1972, Rev. Inst. Med. Trop. S. Paulo, v. 14 (3), 154-161
Trypanosoma cruzi, Colombian strain, mice with reticulo-endothelial blockade due to India ink injections, cortisone-treated mice, suckling mice, severe infection with high parasitemia occurred in these animals with lowered resistance but basic strain pattern was not changed

Trypanosoma cruzi
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Trypanosoma cruzi, mice, Peruvian or Colombian strain, Bay 2502, results varied with strain, suggestions for future research

Trypanosoma cruzi
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Trypanosoma cruzi
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    Trypanosoma cruzi, mice, activity of CL 71.366

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  Andrade, S. G.; and Macedo, V., 1973, Rev. Inst. Med. Trop. S. Paulo, v. 15 (6), 421-430
    Trypanosoma cruzi, mice, human, Bayer 2502, combined use of Bayer 2502 and corticoid (betamethasone) more effective than drug used alone

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    Chagas' cardiomyopathy, pathogenic mechanisms, review

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  Anthony, R. L.; Johnson, C. M.; and Sousa, O. E., 1979, Am. J. Trop. Med. and Hyg., v. 28 (6), 969-973
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  Araujo, F. G., 1976, Rev. Inst. Med. Trop. S. Paulo, v. 18 (6), 433-439
    Trypanosoma cruzi, acute infection in mice, circulating antigens capable of reacting against mouse serum in complement fixation and counterimmunoelectrophoresis tests but not in haemagglutination test

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Trypanosoma cruzi
  Araujo, F. G.; and Nunes. R. M. B., 1979, Rev. Inst. Med. Trop. S. Paulo, v. 20 (6), 315-317
    Trypanosoma cruzi, 3 strains with differences in antigenic structure, strain F1 always yielded lower titers in immunofluorescent test, use of pool of several strains recommended

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Trypanosoma cruzi
  Avila, J. L.; et al., 1979, J. Protozool., v. 26 (2), 304-311
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Trypanosoma cruzi
  Barbosa, W.; et al., 1974, Rev. Patol. Trop., v. 3 (3), 263-268
    Trypanosoma cruzi, diagnosis, sera from persons known to be infected, comparison of test results using counterimmunoelectrophoresis, hemagglutination, fluorescent antibody and complement fixation

T[rypanosoma] cruzi
  Barclay, C. A., 1977, Rev. Neurol. Argentina, v. 3 (3), 477-482
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Trypanosoma cruzi
  Barretto, A. C.; et al., 1978, Rev. Inst. Med. Trop. S. Paulo, v. 20 (4), 183-189
    Trypanosoma cruzi, man, xenodiagnosis, significantly more Dipetalogaster maximus showed infection than did Triatoma infestans in comparative trials

Trypanosoma (Schizotrypanum) cruzi Chagas, 1909
  Barretto, M. P., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (4), 272-278
    valid species, synonymy

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  Barretto, M. P.; and Albuquerque, R. D. R., 1971, Rev. Inst. Med. Trop. S. Paulo, v. 13 (5), 346-351
    Galictis vittata brasiliensis: Sao Simao, State of Sao Paulo, Brazil
    rats (exper.)
    white mice (exper.)
    P[anstrongylus] megistus (exper.)
    T[riatoma] infestans (exper.)
    T. sordida (exper.)
    T. vitticeps (exper.)
    R[hodnius] neglectus (exper.)

Trypanosoma cruzi, illus.
Barretto, M. P.; and Ribeiro, R. D., 1974, Rev. Inst. Med. Trop. S. Paulo, v. 16 (5), 270-275
Trypanosoma cruzi, strain isolated from Holochilus brasiliensis leucogaster, infective for baby mice and triatomines: Itapira, State of Sao Paulo, Brazil

Trypanosoma cruzi
Barretto, M. P.; Ribeiro, R. D.; and Belda Neto, F. M., 1978, Rev. Brasil. Biol., v. 38 (2), 455-459
Trypanosoma cruzi, infection of vertebrate hosts by feeding them infected triatomines or infected rats and mice demonstrates importance of oral route of transmission
mice
rats
dogs
Didelphis azarae
cats
guinea-pigs
(all exper.)

Trypanosoma cruzi
Barretto, M. P.; Ribeiro, R. D.; and Belda Neto, F. M., 1978, Rev. Brasil. Biol., v. 38 (4), 771-773
Trypanosoma cruzi, strains isolated from human cases of Chagas' disease and from infected triatomines and wild Brazilian mammals, behavior in Ricman and Robson's blood incubation infectivity test

Trypanosoma cruzi
Barretto, M. P.; Ribeiro, R. D.; and Rocha, G. de M., 1978, Rev. Brasil. Biol., v. 38 (3), 633-637
Trypanosoma cruzi in wild triatomines, preliminary survey
Rhodnius neglectus
Triatoma sordida
Panstrongylus megistus
P. geniculatus
all from Region of Triangulo Mineiro, State of Minas Gerais, Brazil

Trypanosoma cruzi
Barros, G. C.; et al., 1975, Rev. Inst. Med. Trop. S. Paulo, v. 17 (6), 319-329
human Chagas disease, extensive epidemiologic survey of school children for autochthonous cases, low incidence: State of Espirito Santo, Brazil

Trypanosoma cruzi
Barros, M. A. M.T.; et al., 1979, Clin. and Exper. Immunol., v. 38 (2), 376-380
Trypanosoma cruzi, patients with chronic myocardiopathy vs. indeterminate phase of Chagas' disease, leucocyte migration inhibition test using parasite vs. heart antigens

Trypanosoma cruzi
Baruffa, G.; and Alcantara Filho, A., 1973, Rev. Soc. Brasil. Med. Trop., v. 7 (5), 329-332
Trypanosoma cruzi, human acute and chronic forms, comparison of diagnosis by latex agglutination and by immunofluorescence

Trypanosoma cruzi
Baruffa, G.; and Alcantara Filho, A., 1974, Rev. Inst. Med. Trop. S. Paulo, v. 16 (3), 140-144
Trypanosoma cruzi, rural people, serological prevalence survey by complement fixation: South Region of Rio Grande do Sul State, Brazil

Trypanosoma cruzi
Baruffa, G.; and Alcantara Filho, A., 1975, Rev. Goiana Med., v. 21 (1-2), 11-21
Trypanosoma cruzi, humans, epidemiologic observations on multiple acute cases of Chagas disease, analysis of occupational and educational aspects, and standard of living of infected persons

Trypanosoma cruzi
Baruffa, G.; and Alcantara Filho, A., 1975, Rev. Soc. Brasil. Med. Trop., v. 9 (3), 121-12
Trypanosoma cruzi, humans, comparison of complement fixation and latex agglutination tests for immunodiagnosis of hospitalized patients and for epidemiologic surveys

Trypanosoma cruzi
Baruzzi, R. G.; et al., 1971, Ann. Soc. Belges Med. Trop. Parasitol., v. 51 (2), 205-213
high incidence of splenomegaly in Indians of Alto Xingu region, serological investigation for various parasitic diseases indicated only malaria as a possible cause: Brazil

Trypanosoma cruzi, illus.
Basso, B.; et al., 1977, Rev. Asoc. Argent. Microbiol., v. 9 (1), 11-16
Trypanosoma cruzi, isolation from wild rodent (Calomys musculinus) which is widely distributed in Argentina, possible implications for epidemiology of human Chagas disease: Las Higueras, Rio Cuarto, Cordoba

Trypanosoma cruzi
Batista, S. M.; et al., 1974, Rev. Inst. Med. Trop. S. Paulo, v. 16 (6), 317-323
Trypanosoma cruzi, hemagglutination test, serum from persons with chronic infections, antigens purified by alcohol-ether, acetone-butanol and with known protein content compared with those obtained from culture forms lysed in distilled water

T[rypanosoma] cruzi, illus.
Becker, P. F. L., 1975, Rev. Inst. Med. Trop. S. Paulo, v. 17 (3), 187-198
T[rypanosoma] cruzi, fatal infection in splenectomized woman with hemolytic anemia and under prolonged corticoid therapy, had received blood transfusion from Chagasic donor, clinical aspects, pathology: Sao Paulo State, Brazil

Trypanosoma cruzi
Bejarano, J. F. R.; and De Celis, M. R., 1973, Rev. San. Mil. Argent., v. 72 (1), 49-80
human Chagas disease, extensive epidemiologic and vector survey in the province of La Pampa, Argentina

Trypanosoma cruzi
Belda Neto, F. M., 1974, Rev. Soc. Brasil. Med. Trop., v. 8 (1), 27-36
Trypanosoma cruzi, morpho-biometric and biologic comparisons of 17 strains; results showed no relationships between morphology and pathogenicity of strains studied

Trypanosoma cruzi
Benard, J.; Riou, G.; and Saucier, J. M., 1979, Nucleic Acids Research, v. 6 (5), 1941-1952
Trypanosoma cruzi at different stages of culture and grown in presence of ethidium, kinetoplast DNA, characterization by sedimentation analysis

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Bertinetti, E. dos S., 1974, Radiol. Brasil., v. 7 (2), 91-94
Trypanosoma cruzi, human, dilatation and motion of esophagus as diagnostic feature in conjunction with serologic findings

Trypanosoma cruzi
Bittencourt, A. L., 1975, Rev. Inst. Med. Trop. S. Paulo, v. 17 (3), 135-139
Trypanosoma cruzi, human congenital infections, histopathology of skin infections

Trypanosoma cruzi, illus.
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Trypanosoma cruzi, congenital infections, autopsy pathology of abortus, stillborn, newborn, and infants

Trypanosoma cruzi
Bloch, M., 1976, Rev. Inst. Invest. Med., San Salvador, v. 5 (2), 198-205
Trypanosoma cruzi, evaluation of local epidemiologic information with that of countries known to be highly endemic areas in order to establish public health importance of local infection, suggestions for further research of this problem in El Salvador

Trypanosoma cruzi, illus.
de Boiso, J. F.; Docampo, R.; and Stoppani, A. O. M., 1974, Rev. Asoc. Argent. Microbiol., v. 6 (3), 119-126
Trypanosoma cruzi, scanning and transmission electronmicroscopy of epimastigotes grown in liquid medium

Trypanosoma cruzi
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Trypanosoma cruzi, evaluation of specificity and sensitivity of the direct agglutination test for diagnostic purposes in a non-endemic area, complement fixation and indirect hemagglutination tests used for comparison; when used with the addition of 2-mercaptoethanol in serum samples direct agglutination proved useful for screening in blood banks and for epidemiologic surveys

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Bomford, R.; and McHardy, N., 1979, Parasitology, v. 78 (1), 77-87
Trypanosoma cruzi, comparative efficacy of adjuvants for epimastigote vaccines (Corynebacterium parvum, saponin, Bordetella pertussis)

Trypanosoma cruzi
Bongertz, V.; and Hungerer, K. D., 1978, Exper. Parasitol., v. 45 (1), 8-18
Trypanosoma cruzi, protease, isolation and characterization

Trypanosoma cruzi
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cryopreservation of parasitic protozoa

Trypanosoma cruzi
Bouree, P., 1976, Rev. de Med., Paris, v. 17 (39), 2139-2150
Trypanosoma spp., geographic distribution, vectors, recommended therapeutic measures, review

Trypanosoma cruzi
Boveris, A.; et al., 1977, Rev. Asoc. Argent. Microbiol., v. 9 (2), 54-61
Trypanosoma cruzi, epimastigote forms in saline medium, action of β-lapachone and its metabolite form α-lapachone on parasite growth and production of hydrogen peroxide; β-lapachone inhibited growth but possible effect was mediated by the hydrogen peroxide and related free radicals

Trypanosoma cruzi
Boveris, A.; et al., 1978, Biochem. J., v. 175 (2), 431-439
Trypanosoma cruzi, differential effect of β-lapachone and α-lapachone on superoxide anion and hydrogen peroxide production and on growth of epimastigotes

Trypanosoma cruzi
Boveris, A.; et al., 1978, Comp. Biochem. and Physiol., v. 61C (2), 327-329
Trypanosoma cruzi, correlation between superoxide anion production and trypanocidal action of naphthoquinones

Trypanosoma cruzi
Bowman, I. B. R., 1974, Ciba Found. Symp., n.s. (20), 255-284
trypanosomes, intermediary metabolism, review

T[rypanosoma] cruzi
Braescher, H. M.; et al., 1977, Rev. Brasil. Pesqui. Med. e Biol., v. 10 (4), 255-260
T[rypanosoma] cruzi, leucocyte migration inhibition using soluble and particulate antigens and leucocytes from patients with chronic Chagas disease, results showed that stimulatory capacity of particulate antigens is greater than that of soluble antigens

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Cerisola, J. A.; et al., 1977, Bol. Chileno Parasitol., v. 32 (3-4), 51-62
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T[rypanosoma] cruzi, epidemiologic survey of autochthonous human infection, dispersion of infected Triatoma infestans in climatic environment thought to be unfavorable for development of this vector: Baixada fluminense, Municipio de Caxias, Estado do Rio de Janeiro

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Trypanosoma cruzi, epidemiologic survey of non-endemic area for evidence of Chagas disease, triatomes inhabiting housing, humans (mostly children) and 2 dogs found infected by serologic and xenodiagnostic methods: Caxias, Rio de Janeiro State, Brazil

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Trypanosoma cruzi, mice, transplacental transmission is dependent upon pathogenicity of parasite strain and phagocytic activity of placenta

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Trypanosoma cruzi, illus.
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Trypanosoma cruzi bloodstream forms, increase in respiration in presence of acetate, acetate oxidation took place via tricarboxylic acid cycle and involved antimycin A-sensitive respiratory pathway, immune sera had no effect on oxygen uptake

Trypanosoma cruzi
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Trypanosoma cruzi, cyanide-sensitive and insensitive respiration

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Trypanosoma cruzi, humans, 6 case reports, probable autochthonous origin, region contained many palm trees which harbored triatomes: Barcelos, State of Amazonas, Brazil

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Trypanosoma cruzi, laboratory-reared Triatoma infestans, cryopreservation of infective stages within intact vector may be useful for storing laboratory-adapted strains and naturally infected vectors collected in field studies

Trypanosoma cruzi
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Trypanosoma cruzi, mice, neuron lesions of juxtaprostatic pelvic ganglion, applications for human testicular lesions and genital misfunctions in human infections

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Trypanosoma cruzi, trypanocidal effect of various thiosemicarbazones compared with standard anti-trypanosomes, benzazon VII proved effective in vitro (cultured crithidial forms) and in exper. infected mice and compared favorably with nitrofurazone and lampit

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Forichon, E., 1975, Rev. Patol. Trop., v. 4 (1), 57-78
human Chagas disease, compilation and review of morbidity and mortality statistics gathered from various sources, extensive bibliography, thesis

Trypanosoma cruzi
Foxley, R., 1974, Rev. Chilena Obst. y Ginec., v. 39 (3), 83-86
Trypanosoma cruzi, survey of infertile women showed that Chagas disease was an uncommon cause of reproductive problems: Chile

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Trypanosoma cruzi, adenosine triphosphatase activities

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Trypanosoma cruzi, Mg^{2+}-activated adenosine triphosphatase, solubilization and some properties

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de Freitas, C. A., 1975, Rev. Brasil. Malariol., v. 26-27, 1974-1975, 61-92
Trypanosoma cruzi, humans, discussion of epidemiology and control measures in Brazil

Trypanosoma cruzi, illus.
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Trypanosoma cruzi, location of a specific antigen (antigen 5) on the surface of bloodstream trypomastigote and culture epimastigote forms

Trypanosoma cruzi
Fuentes, F. B.; Coura, J. R.; and Ferreira, L. F., 1971, Rev. Soc. Brasil. Med. Trop., v. 5 (1), 47-53
Trypanosoma cruzi, in survey of a "shanty town" for evidence of triatome vectors only Triatoma rubrofasciata was discovered and it was not infected: Sao Cristovao, Guanabara

Trypanosoma cruzi, illus.
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Trypanosoma cruzi, morphologic characteristics of strain isolated from bat reservoir hosts
Dasypterus ega argentinus
Molossus major crassicaudatus
Noctilio labialis albiventris
Lesiurus cinereus villosissimus
Glossophaga s. soricina
Eumops p. perotis
E. glaucinus
all from area of Sao Paulo

Trypanosoma cruzi, illus.
Funayama, G. K.; and Barretto, M. P., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (4), 257-265
Alouatta caraya: Franca, Sao Paulo, Brazil
camundongos (exper.)
P[anstrongylus] megistus (exper.)
R[hodnius] neglectus (exper.)
T[riatoma] sordida (exper.)
T. infestans (exper.)

Trypanosoma cruzi, illus.
Funayama, G. K.; and Barretto, M. P., 1973, Rev. Brasil. Biol., v. 33 (3), 439-444
Trypanosoma cruzi, new strain isolated from bat, infective for mice and triatomes in laboratory experiments
Epitesicus brasiliensis brasiliensis: Ribeirao Preto, Sao Paulo, Brazil

Trypanosoma cruzi, illus.
Funayama, G. K.; and Prado, J. C., jr., 1974, Rev. Soc. Brasil. Med. Trop., v. 8 (2), 75-81
Trypanosoma cruzi, strain isolated from Triatoma infestans captured in Vitichi, Bolivia, severe pathogenicity for mice, mice recovered from infection have high resistance against reinfection by the Y strain, Bolivia strain easily cultured and regularly infective for several triatomines

Trypanosoma cruzi
Galinari, S.; and Camargo, E. P., 1978, Exper. Parasitol., v. 46 (2), 277-282
trypanosomatid protozoa, 16 spp., survey for acetylornithinase and ornithine acetyltransferase, metabolic and nutritional implications

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Gallardo Zerpa, M. F., 1974, Rev. Fac. Farm., Univ. Central Venezuela (27), v. 14, 69-73
Trypanosoma cruzi, Leishmania donovani, L. braziliensis, cultured successfully in diphasic autoclaved medium of tryptose, glucose, mineral salts, vitamins, blood and agar; medium would not support growth of T. rangeli

Trypanosoma cruzi
Goncalves de Lima, V. M. Q.; Roitman, I.; and Kilgour, V., 1979, J. Protozool., v. 26 (4), 648-652
trypanosomatids, 7 species distinguished by electrophoretic mobilities of some isoenzymes

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de Gonzalez, G. R.; and Andres, A. S., 1977, Acta Bioquim. Clin. Latinoam., v. 11 (4), 353-356
Trypanosoma cruzi, diagnosis of Chagas disease in blood bank donors, direct agglutination and direct agglutination with 2-mercaptoethanol, comparison with complement fixation, immunofluorescence, and latex agglutination

rypanosoma cruzi, illus.
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Caluromys sp.
Metachirus sp.
Marmosa cinerea
Dasypus novemcinctus
Coendou spp.
Agouti paca
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Trypanosoma cruzi, 2 humans vaccinated with avirulent strain, 5-year follow-up

Trypanosoma cruzi
Mercado, T. I.; and Garbus, J., 1979, Comp. Biochem. and Physiol., v. 64B (1), 11-15
Trypanosoma cruzi, mice infected with myotropic vs. reticulotropic parasite strains, creatine phosphokinase isoenzymes in plasma and tissues

Trypanosoma cruzi
Milder, R. V.; Kloetzel, J.; and Deane, M. P., 1973, Rev. Inst. Med. Trop. S. Paulo, v. 15 (6), 386-392
Trypanosoma cruzi culture and blood stream forms, optical and electron microscopic observations on interactions with hamster peritoneal macrophages, blood forms observed to be wrapped by a conspicuous membrane and to multiply actively; culture forms were epimastigotes folded upon themselves with invagination of periplast and electron-dense material inside and outside invaginations

Trypanosoma cruzi, illus.
Milder, R.; Kloetzel, J.; and Deane, M. P., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (5), 313-322
Trypanosoma cruzi trypomastigotes, interaction with hamster peritoneal macrophages at optical and ultrastructural levels in vitro, possible mechanisms of parasite intracellular fate, strain differences

Trypanosoma cruzi
Miles, M. A.; et al., 1978, Nature, London (5656), v. 272, 819-821
Trypanosoma cruzi, enzyme electrophoresis reveals 3 discrete forms of parasite in Brazil including recently discovered type responsible for first autochthonous patients with Chagas' disease in Belem, Para state

Trypanosoma cruzi
Miles, M. A.; et al., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (5), 528-532
Trypanosoma cruzi, exper. infections in rhesus monkeys, electrocardiographic and histopathologic changes

rypanosoma (Schizotrypanum) cruzi
Minter, D. M.; Minter-Goedbloed, E.; and Marshall, T. F. de C., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (1), 84-91
Trypanosoma cruzi, comparative xenodiagnosis using 3 species of triatomes and materials from natural and experimental infections of man and animals; results show that interspecific differences of bloodmeal size and intrinsic differences in susceptibility to infection between species of triatomes are limiting factors for standardization and interpretation of results

rypanosoma (Schizotrypanum) cruzi, illus.
Minter-Goedbloed, E., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (1), 22-30
Trypanosoma cruzi, primary isolation by hemoculture of parasites from naturally infected Didelphis azarae, mice (exper.) and acute and chronic infections in humans; low proportion of successful isolations from human chronic infections due probably to lower parasitemia; hemoculture from chronic patients also differed markedly from other hosts in very slow growth-rate obtained which was probably due to continuing activities of humoral and cellular components in blood inoculum

rypanosoma (Schizotrypanum) cruzi
Minter-Goebloed, E.; Minter, D. M.; and Marshall, T. F. de C., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (3), 217-225
Trypanosoma cruzi, detection in mice, humans, and Didelphis azarae, acute and chronic infections, xenodiagnosis vs. haemoculture, quantitive comparison

rypanosoma cruzi
Mocelin, A. J.; et al., 1977, Transplantation, v. 23 (2), 163 [Letter]
Trypanosoma cruzi in 29-year-old woman recipient of kidney transplant, negative blood smears after lampit, kidney function preserved: Brazil

rypanosoma cruzi
Molyneux, D. H.; and Bafort, J. M., 1971, Ann. Soc. Belges Med. Trop. Parasitol., v. 51 (3), 335-348
Trypanosoma vespertilionis, morphology of bloodstream forms, sites and morphology of tissue stages (in cysts), morphology and ultrastructure of culture forms, difficulties in differentiating from T. cruzi

ypanosoma cruzi
Mott, K. E., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (3), 320 [Letter]
Trypanosoma cruzi, blood cultures using vacuum culture tubes, may be useful tool for parasite isolation in areas endemic for Chagas' disease

ypanosoma cruzi
Mott, K. E.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (6), 1116-1122
Trypanosoma cruzi, household distribution of seroreactivity analyzed in relation to style of house construction and distribution of Panstrongylus megistus: northeast Brazil

ypanosoma cruzi
Mott, K. E.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (6), 1123-1127
Trypanosoma cruzi, infection of domestic dogs and cats in relation to household seroreactivity and household Panstrongylus megistus infestation: northeast Brazil

Trypanosoma cruzi
Mourao, O. G.; and Milio, O. C., 1975, Rev. Soc. Brasil. Med. Trop., v. 9 (4), 183-188
Trypanosoma cruzi, method of processing blood cultures for use in diagnosis of chronic phase of Chagas disease

Trypanosoma cruzi
Nathan, C.; et al., 1979, J. Exper. Med., v. 149 (5), 1056-1068
Trypanosoma cruzi, activation of macrophages in vivo and in vitro, correlation between hydrogen peroxide release and trypanocidal activity

Trypanosoma cruzi
Neal, R. A.; and Miles, R. A., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (5), 325-332
Trypanosoma cruzi, indirect hemagglutination test, adapted for use with capillary blood dried on filter paper, useful for epidemiologic field surveys

Trypanosoma cruzi
Neal, R. A.; and Miles, R. A., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (3), 170-176
Trypanosoma cruzi, mice, detection of experimental infections, culture methods vs. xenodiagnosis

Trypanosoma cruzi
Neal, R. A.; and Miles, R. A., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (3), 177-181
Trypanosoma cruzi, xenodiagnosis, diagnostic parameters, numbers of trypomastigotes required to be ingested by triatomid bug to give rise to detectable infection

Trypanosoma cruzi, illus.
Neves, D. P., 1971, Rev. Inst. Med. Trop. S. Paulo, v. 13 (3), 155-161
Trypanosoma cruzi, effects of low and high temperatures on development in triatomines

Trypanosoma cruzi
Neves, D. P.; and Carvalho, A. L. M., 1972, Rev. Patol. Trop., v. 1 (2), 277-281
Trypanosoma cruzi-infected Triatoma infestans and T. vitticeps, use for xenodiagnostic purposes compared, T. vitticeps thought to be poor subject for actual use in diagnosis of human infections

Trypanosoma cruzi
Nogueira, N.; and Cohn, Z. A., 1978, J. Exper. Med., v. 148 (1), 288-300
Trypanosoma cruzi, conditions for inducing and maintaining trypanocidal activity in both resident and inflammatory mouse peritoneal macrophages maintained in vitro

Trypanosoma cruzi
O'Daly, J. A.; and Aso, P. M., 1979, Exper. Parasitol., v. 47 (2), 222-231
Trypanosoma cruzi, Leishmania spp., factor in cell-free extracts that induces lysis of mammalian red cells and Vero cells, postulated that this lytic factor is involved in penetration and damage produced by T. cruzi in vertebrate cells

Trypanosoma cruzi
Olabuenaga, S. E.; et al., 1979, Cellular Immunol., v. 45 (1), 85-93
Trypanosoma cruzi, antibody-dependent cytolysis of epimastigotes by human polymorphonuclear leukocytes

Trypanosoma cruzi
de Oliveira, A. R., 1973, Rev. Patol. Trop., v. 2 (1), 95-142
Trypanosoma cruzi, theories attempting to prove that Chagas disease does not cause megalogastria or achalasia of the pylorus

Trypanosoma cruzi
Ortega G., M.; Beltran H., F.; and Zavala V., J., 1976, SPM Salud Pub. Mexico, v. 18 (5), 837-843
Trypanosoma cruzi, human, epidemiological survey of 2 communities, infected Rhodnius prolixus found during vector search: northern Chiapas, Mexico

Trypanosoma cruzi, illus.
Pan, S. C., 1978, Exper. Parasitol., v. 45 (2), 215-224
Trypanosoma cruzi, highly enriched liquid medium that can support growth of amastigotes at 37 C

Trypanosoma cruzi, illus.
Pan, S. C., 1978, Exper. Parasitol., v. 45 (2), 274-286
Trypanosoma cruzi, amastigotes derived from culture in cell-free medium at 37 C are biologically and physiologically similar to intracellular amastigotes derived from natural infections in that both can multiply and complete the life cycle intracellularly

Trypanosoma cruzi, illus.
Pan, S. C. T., 1978, Exper. Parasitol., v. 46 (1), 92-107
Trypanosoma cruzi, ultrastructure of morphogenesis in vitro and in vivo (mice)

Trypanosoma cruzi
Pan, S. C. T., 1978, Exper. Parasitol., v. 46 (1), 108-112
Trypanosoma cruzi, cultivation in macromolecule-free semisynthetic and synthetic media

Trypanosoma cruzi
Patrucco, A.; et al., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (4), 425-426
Trypanosoma cruzi, human, leucocyte migration inhibition test with 3 antigenic fractions and whole homogenate, flagellar fraction proved to be antigen of choice, low incidence of positive response in patients with cardiomyopathy when compared with asymptomatic but serologically positive patients

T[rypanosoma] cruzi
Pereira, F. E. L., 1972, Rev. Inst. Med. Trop. S. Paulo, v. 14 (1), 30-32
T[rypanosoma] cruzi, human, chronic infection, increased presence of mast cells in esophageal musculature

Trypanosoma cruzi, illus.
Pereira, N. M.; et al., 1978, Exper. Parasitol., v. 46 (2), 225-234
Trypanosoma cruzi, membrane and flagellar fractions, isolation and characterization: electron microscopy; enzyme composition; gel immunodiffusion test

T[rypanosoma] cruzi
Perlowagora-Szumlewicz, A., 1973, Rev. Inst. Med. Trop. S. Paulo, v. 15 (3), 139-150
T[rypanosoma] cruzi vectors (Triatoma, Rhodnius, Panstrongylus), vector species and stage interaction in feeding behavior, application to standardization and efficacy of xenodiagnostic procedures

Trypanosoma cruzi
Perlowagora-Szumlewicz, A.; Petana, W. B.; and Figueiredo, M. J., 1975, Rev. Inst. Med. Trop. S. Paulo, v. 17 (2), 97-102
Trypanosoma cruzi, effects of juvenile hormone analogues on developmental changes in Panstrongylus megistus vector triatomines, juvenilized bugs less susceptible to natural gut infection, potential for biological control

Trypanosoma cruzi, illus.
Petana, W. B., 1978, Bol. Ofic. San. Panamer., v. 85 (4), 300-306
Trypanosoma cruzi, endemic in nearly all Central and South American countries facing Caribbean basin

Trypanosoma (Schizotrypanum) cruzi
Petana, W. B.; and Coura, J. R., 1974, Rev. Soc. Brasil. Med. Trop., v. 8 (6), 315-323
Trypanosoma cruzi, comparison of 9 strains isolated from man, animals, and triatomine bugs, host pathology, virulence, infectivity, importance of strain differentiation: Brazil

Trypanosoma cruzi, illus.
Peters, W., 1974, Ciba Found. Symp., n.s. (20), 309-334
trypanosomiasis, leishmaniasis, drug resistance, review

Trypanosoma cruzi
Pinchin, R.; et al., 1978, J. Econom. Entom., v. 71 (6), 950-955
Trypanosoma cruzi, 40 compounds bioassayed for juvenile hormone activity on Panstrongylus megistus

Trypanosoma cruzi
Pinzon Cantarell, J.; Quintal Aviles, R.; and Zavala Velazquez, J., 1976, SPM Salud Pub. Mexico, v. 18 (6), 999-1003
Trypanosoma cruzi, survey for infected triatomid vectors
Triatoma dimidiata maculipennis
T. phillosoma longipennis
all from state of Yucatan, Mexico

Trypanosoma cruzi
Polak, A.; and Richle, R., 1978, Ann. Trop. Med. and Parasitol., v. 72 (1), 45-54
Trypanosoma cruzi in vivo and in vitro, benznidazole, effect on growth and viability, aerobic and anaerobic respiration, and synthesis of protein, RNA, and DNA

Trypanosoma cruzi
Ponce, C.; Trochez, H.; and Zeledon, R., 1975, Rev. Biol. Trop., v. 22 (2), 1974, 289-303
Trypanosoma cruzi, T. rangeli, single and mixed infections, epidemiologic survey of area infested with Rhodnius prolixus: Departamento Francisco Morazan, Honduras

Trypanosoma cruzi
Prata, A.; et al., 1975, Rev. Patol. Trop., v. 4 (1), 35-38
Trypanosoma cruzi, diagnosis using complement fixation, results of tests from 3 different laboratories were compared and found to have substantial differences, study shows need for standardization of reagents and techniques

[rypanosoma] cruzi
de Queiroz, A. C.; and Ramos, E. A. G., 1979, Arq. Neuropsiquiat., v. 37 (4), 405-411
chronic Chagas infection with heart failure, cerebral pathology at autopsies compared with that of patients who died with idiopathic cardiomegaly

rypanosoma cruzi
Quintal Aviles, R.; and Navarrete Esquivel, R., 1975, Salud Pub. Mexico, v. 17 (3), 365-369
Trypanosoma cruzi; Toxoplasma gondii, humans, serological survey using the indirect hemagglutination test: Motul, Yucatan

rypanosoma cruzi
Quintal Aviles, R.; Zavala Velazquez, J.; and Rodriguez Lopez, M., 1975, Rev. Invest. Clin., v. 27 (4), 255-258
Trypanosoma cruzi, diagnostic review of acute and chronic human cases: Yucatan, Mexico

rypanosoma cruzi
Rabinovich, J. E.; Leal, J. A.; and Feliciangeli de Pinero, D., 1979, Tr. Roy. Soc. Trop. Med. and Hyg., v. 73 (3), 272-283
Rhodnius prolixus, domiciliary biting frequency and blood ingestion estimated by combining laboratory and field information: Venezuela

rypanosoma cruzi
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parásitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

rypanosoma cruzi
Ramos, C.; et al., 1978, Exper. Parasitol., v. 45 (2), 190-199
Trypanosoma cruzi, infection in mice induces immunosuppression to both T-dependent and T-independent antigens, depression observed is not due to alteration in macrophage function

ypanosoma cruzi
Ramos, C.; Schaedtler-Siwon, I.; and Ortiz-Ortiz, L., 1979, J. Immunol., v. 122 (4), 1243-1247
Trypanosoma cruzi, mice, infection elicits generation of T cells in spleen suppressive to T and B cell mitogenic responses

rypanosoma] cruzi
Raso, P.; and Tafuri, W. L., 1971, Rev. Soc. Brasil. Med. Trop., v. 5 (3), 135-153
T[rypanosoma] cruzi, humans, pathologic pericardial alterations compared with those seen in rheumatic and hypertensive heart disease, analysis of changes of acute and chronic infection, study of changes in exper. infections in mice

Trypanosoma cruzi
Rassi, A.; and Ferreira, H. de O., 1971, Rev. Soc. Brasil. Med. Trop., v. 5 (5), 235-262
Trypanosoma cruzi, extensive clinical trials testing efficacy of various nitrofuranes (singly, mixed nitrofuranes, or in association with primaquine), therapeutic response as based mainly on xenodiagnosis and the Guerreiro-Machado test showed nitrofurazone and especially Bayer 2502 to give best response

Trypanosoma cruzi
Reed, S. G.; Larson, C. L.; and Speer, C. A., 1977, Ztschr. Parasitenk., v. 52 (1), 11-17
Trypanosoma cruzi, mice immunized by Freund's adjuvant or oxazolone, acute infection suppressed cell-mediated immunity to these antigens; immunization with live T. cruzi before infection resulted in greater than normal oxazolone sensitivity, mice survived infection; inconclusive as to whether immunosuppression due to infection is directed toward induction or toward expression of cell-mediated response

Trypanosoma cruzi
Reed, S. G.; Larson, C. L.; and Speer, C. A., 1978, Infect. and Immun., v. 22 (2), 548-554
Trypanosoma cruzi, contact sensitivity responses in infected mice, results indicate that suppression of contact sensitivity during acute infection is directed toward efferent arm rather than afferent arm of response

Trypanosoma cruzi
Reis, A. P.; et al., 1976, Rev. Inst. Med. Trop. S. Paulo, v. 18 (6), 422-426
Trypanosoma cruzi, mice infected with different parasite strains, presence of delayed hypersensitivity demonstrated by leukocyte inhibition migration technique

Trypanosoma cruzi, illus.
Ribeiro, R. D., 1973, Rev. Brasil. Biol., v. 33 (3), 429-437
Trypanosoma cruzi, new rodent reservoir hosts, isolated strain infective for mice, rats and triatomes in laboratory experiments
Oryzomys subflavus
Calomys tener
Zygodontomys l. lasiurus
Cercomys c. cunicularius
Thomasomys dorsalis collinus
all from northeastern region, State of Sao Paulo, Brazil

Trypanosoma cruzi, illus.
Ribeiro, R. D.; and Barretto, M. P., 1972, Rev. Inst. Med. Trop. S. Paulo, v. 14 (6), 352-359
Trypanosoma cruzi from Sciurus aestuans ingrami, pathogenic for mice and rats, infective for several triatomines: Franca, State of Sao Paulo, Brazil

Trypanosoma cruzi, illus.
Ribeiro, R. D.; and Barretto, M. P., 1972, Rev. Inst. Med. Trop. S. Paulo, v. 14 (3), 162-170
Syn.: T. akodoni Carini & Maciel, 1915
T. cruzi strain isolated from Akodon nigritus nigritus pathogenic in baby rats and mice, infective for various species of triatomines: Ipanema, SP, Brazil

Trypanosoma cruzi, illus.
Ribeiro, R. D.; and Barretto, M. P., 1975, Rev. Inst. Med. Trop. S. Paulo, v. 17 (4), 247-252
Trypanosoma cruzi, strain isolated from Akodon lasiotis, infective for mice and triatomines: Ribeirao Preto, Sao Paulo, Brazil

Trypanosoma cruzi, illus.
Ribeiro, R. D.; Belda Neto, F. M.; and Barretto, M. P., 1977, Rev. Brasil. Biol., v. 37 (4), 907-912
Trypanosoma cruzi, life cycle in haemocoel of Panstrongylus megistus, development either extracellularly in haemolymphatic fluid or intracellularly in haemocytes

Trypanosoma cruzi, illus.
Ribeiro, R. D.; Belda Neto, F. M.; and Barretto, M. P., 1978, Rev. Brasil. Biol., v. 38 (2), 345-349
Trypanosoma cruzi, 4 strains with morphologically different blood trypomastigotes, differences in development in coelomic cavity of Panstrongylus megistus (exper.)

Trypanosoma cruzi, illus.
Ribeiro, R. D.; and Pereira Barretto, M., 1977, Rev. Brasil. Biol., v. 37 (2), 233-239
Trypanosoma cruzi, strain isolated from Dasyprocta a. aguti (blood), possible reservoir, infectivity to triatomines and mice, mice protected against subsequent infection by human strain: Colatina, E. S., Brasil

T[rypanosoma] cruzi
Ribeiro dos Santos, R.; et al., 1979, Tropenmed. u. Parasitol., v. 30 (1), 19-23
T[rypanosoma] cruzi-infected humans, presence of IgG and IgM antibodies to neurons demonstrated by immunofluorescence

Trypanosoma cruzi, illus.
Riou, G. F.; and Gutteridge, W. E., 1978, Biochimie, v. 60 (4), 365-379
Trypanosoma cruzi, comparative study of kinetoplast DNA in culture epimastigote, blood trypomastigote, and intracellular amastigote stages

T[rypanosoma] cruzi
Ritacco, V.; et al., 1977, Rev. Neurol. Argentina, v. 3 (3), 443-447
T[rypanosoma] cruzi, human, description of human serum gamma globulin factor associated with antibody against peripheral nerves

Trypanosoma cruzi, illus.
Rocha, G. de M.; and Peireira Barretto, M., 1977, Rev. Brasil. Biol., v. 37 (2), 419-424
Trypanosoma cruzi, isolated from Callithrix geoffroyi (blood), possible reservoir host, pathogenic for mice, infection by monkey strain gives good resistance in mice against reinoculation with Y-strain of T. cruzi: Governador Valadares, MG, Brazil

Trypanosoma cruzi, illus.
da Rocha e Silva, E. O.; et al., 1977, Rev. Saude Pub., S. Paulo, v. 11 (1), 87-96
[host uncertain]: insect colony, Moji Guacu

Trypanosoma cruzi
da Rocha e Silva, E. O.; et al., 1977, Rev. Saude Pub., S. Paulo, v. 11 (2), 258-269
Triatoma sordida, preference for avian versus human blood

Trypanosoma cruzi
Rogerson, G. W.; and Gutteridge, W. E., 1979, Internat. J. Biochem., v. 10 (12), 1019-1023
Trypanosoma cruzi, mammalian and culture forms, oxidative metabolism

Trypanosoma cruzi
Romanha, A. J.; et al., 1979, Comp. Biochem. and Physiol., v. 62B (2), 139-142
Trypanosoma cruzi, isoenzyme patterns of 4 soluble enzymes in seven parasite stocks determined by electrophoresis, changes after prolonged subculture

Trypanosoma cruzi
Rowland, E. C.; and Kuhn, R. E., 1978, Infect. and Immun., v. 20 (2), 393-397
Trypanosoma cruzi, mice (2 strains of differing resistance), suppression of cell-mediated responses during infection, significance unclear

Trypanosoma cruzi
Rowland, E. C.; and Kuhn, R. E., 1978, J. Parasitol., v. 64 (4), 741-742
Trypanosoma cruzi, mice, suppression of anamnestic cellular responses in immunized animals, suppression shows nonspecific character, could be speculated that T. cruzi acts as tolerogen during infection resulting in anergic condition similar to desensitization

Trypanosoma cruzi
Rubiolo, E. R.; and Vottero Cima, E., 1979, Acta Bioquim. Clin. Latinoam., v. 13 (3), 285-288
Chagas' disease, diagnosis, technique for preservation of red cells sensitized with Trypanosoma cruzi antigens to be used in indirect hemagglutination test

Trypanosoma cruzi
Salata, E.; Barbosa, M. A.; and Correa, F. M. A., 1976, Rev. Inst. Med. Trop. S. Paulo, v. 18 (4), 211-214
Trypanosoma cruzi, antibody production in mice inoculated with irradiated vs. non irradiated culture forms of parasite

Trypanosoma cruzi
Salata, E.; Wiendl, F. M.; and Correa, F. M. A., 1973, Rev. Inst. Med. Trop. S. Paulo, v. 15 (2), 66-71
Trypanosoma cruzi, effect of radiation on morphology, motility, reproduction, virulence and immunization potential

Trypanosoma cruzi
Sanderson, C. J.; Bunn Moreno, M. M.; and Lopez, A. F., 1978, Parasitology, v. 76 (3), 299-307
Trypanosoma cruzi, cytotoxicity of normal rat spleen cells to antibody-coated epimastigotes studied by assaying release of tritium-labelled RNA, DNA, and protein

Trypanosoma cruzi, illus.
Sanderson, C. J.; and de Souza, W., 1979, J. Cell Sc., v. 37, 275-286
Trypanosoma cruzi, interaction with rat eosinophils, neutrophils, and macrophages in vitro, light and electron microscopy

Trypanosoma cruzi
Santos, J. C. M., 1971, Rev. Inst. Med. Trop. S. Paulo, v. 13 (4), 279-284
Trypanosoma cruzi in Rhodnius prolixus, infectivity of avirulent PF strain compared with virulent Y strain

Trypanosoma cruzi
dos Santos, R. R., 1973, Rev. Patol. Trop., v. 2 (4), 433-463
Trypanosoma cruzi, lymphocytes of mice inoculated with avirulent PF strain conferred immunity in mice (treated with immunosuppressive drugs or untreated) against infections with the virulent Y strain; newborn mice treated with immunosuppressive drugs showed no protection against the virulent strain

Trypanosoma cruzi
dos Santos, R. R.; Amato Neto, V.; and Gioia, I., 1975, Rev. Goiana Med., v. 21 (1-2), 23-27
Leptomonas pessoai antigens and sera of patients infected with Trypanosoma cruzi gave frequent positive reactions with the passive hemagglutination, complement fixation and indirect immunofluorescence tests, possible implications for prophylactic vaccine for Chagas disease

T[rypanosoma] cruzi
dos Santos, R. R.; and Marquez, J. O., 1977, Rev. Neurol. Argentina, v. 3 (3), 439-442
T[rypanosoma] cruzi, human, physiopathology of neuronal destruction, presence of IgG and IgM antibodies to neurons discovered in human serum, sequence of pathologic events detailed in infected mice

Trypanosoma cruzi
dos Santos, R. R.; Menezes, H.; and Amato Neto, V., 1974, Rev. Inst. Med. Trop. S. Paulo, v. 16 (1), 47-49
Leptomonas pessoai unable to infect immunosuppressed mice, L. pessoai of possible use in immunization against Trypanosoma cruzi

T[rypanosoma] cruzi
dos Santos, R. R.; de Oliveira, J. C. R.; and Koeberle, F., 1976, Rev. Goiana Med., v. 22 (3-4), 235-243
T[rypanosoma] cruzi, human, immunopathology of nervous system lesions

Trypanosoma cruzi
Santos-Buch, C. A., 1977, Immunopathol., 7. Internat. Symp. (Bad Schachen, Germany, June 14-19, 1976), 205-220
Trypanosoma cruzi, role of immune reactions in pathogenesis of Chagas' disease, review

Trypanosoma cruzi
Sanz, O. P., 1977, Rev. Neurol. Argentina, v. 3 (3), 458-463
Trypanosoma cruzi, electrophysiological and histological study of skeletal muscles of chronically infected persons showed that a relevant proportion had motor unit involvement

Trypanosoma cruzi, illus.
Schaffer, G. D.; et al., 1978, J. Am. Vet. Med. Ass., v. 173 (9), 1148-1151
hematotropic parasites of Procyon lotor, carrier potential as related to translocation and release for hunting purposes, practice is considered biologically hazardous: southeastern United States
Procyon lotor: Glades County, Florida; Hillsborough County, Florida; Liberty County, Georgia; Brown County, Texas

Trypanosoma cruzi
Schenone, H.; et al., 1977, Bol. Chileno Parasitol., v. 32 (3-4), 63-66
Trypanosoma cruzi, patient with chronic infection and constant parasitemia, xenodiagnosis for study of diurnal and nocturnal periodicity

Trypanosoma cruzi
Schenone, H.; Villarroel, F.; and Alfaro, E., 1978, Bol. Chileno Parasitol., v. 33 (1-2), 2-7
Trypanosoma cruzi, epidemiologic survey in rural and suburban areas, positive xenodiagnosis found in both humans and their domestic animals, infected T. infestans collected from dwellings, study of housing conditions favoring the presence of triatomids: Chile

Trypanosoma cruzi
Schmunis, G. A.; et al., 1977, Rev. Hosp. Ninos, Buenos Aires (73), v. 19, 15-20
Trypanosoma cruzi, humans, diagnosis of acute infections by the direct agglutination test and by indirect immunofluorescence, techniques also useful in evaluating effects of therapy

Trypanosoma cruzi
Schmunis, G. A.; et al., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (5), 323-331
Trypanosoma cruzi-infected mice inoculated with sheep red blood cells, alterations in immune response and their possible mechanisms

Trypanosoma cruzi
Schmunis, G. A.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (3), 473-477
Trypanosoma cruzi, humans, immunoglobulin M, G, and A concentrations in treated acute Chagas' disease

Trypanosoma cruzi
Schmunis, G. A.; et al., 1978, J. Infect. Dis., v. 138 (3), 401-404
Trypanosoma cruzi, persistence of tissue-reacting (EVI) antibodies in nifurtimox-treated patients followed for several months to 2 years, implications for significance of EVI antibodies in pathogenesis of Chagas disease

Trypanosoma cruzi, illus.
Schmunis, G. A.; et al., 1978, Infect. and Immun., v. 20 (2), 567-569
Trypanosoma cruzi, induction of capping in blood-stage trypomastigotes by human specific antibody

Trypanosoma cruzi
Schofield, C. J., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (5), 449-455
Triatoma infestans, comparison of sampling techniques for domestic populations: Brazil

Trypanosoma cruzi
Schofield, C. J., 1979, Bull. Entom. Research, v. 69 (3), 363-379
Triatominae, behavior, review

Trypanosoma cruzi
Schottelius, J., 1977, Tropenmed. u. Parasitol., v. 28 (4), 533-538
Trypanosoma cruzi, course of infection in non-splenectomized SPF rats with and without Haemobartonella muris infection

Trypanosoma cruzi, illus.
Scorza, C.; and Scorza, J. V., 1972, Rev. Inst. Med. Trop. S. Paulo, v. 14 (3), 171-177
Trypanosoma cruzi, rats, pathology of myocardial lesions

Trypanosoma cruzi
Scott, M. T.; and Snary, D., 1979, Nature, London (5734), v. 282, 73-74
Trypanosoma cruzi, mice, protective immunization using cell surface glycoprotein

Trypanosoma cruzi
Sherlock, I. A.; and Almeida, S. P., 1973, Rev. Soc. Brasil. Med. Trop., v. 7 (2), 87-98
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de Siqueira, A. F.; Ribeiro, R. D.; and Fernandes, L. A. R., 1973, Rev. Inst. Med. Trop. S. Paulo, v. 15 (2), 76-80
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Sousa, O. E., 1972, Rev. Biol. Trop., v. 20 (2), 167-179
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Panstrongylus humeralis (intestinal tract):
Bayano River area, Panama Province
mice (exper.) (blood)

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Leptomonas pessoai, antigenic relationships with other trypanosomatids, cross-protection of mice against Trypanosoma cruzi

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antigens of Crithidia fasciculata, Trypanosoma cruzi and Leishmania brasiliensis showed cross-reacting precipitating bands with the antigen of Leptomonas pessoai as demonstrated by the agar gel diffusion technique

Trypanosoma cruzi
de Souza, W.; et al., 1978, Exper. Parasitol., v. 45 (1), 101-115
Trypanosoma cruzi incorporates exogenous proteins by pinocytosis, pinocytic vesicles can fuse forming multivesicular structures, cell membrane and membranes of pinocytic vesicles and large multivesicular structures contain carbohydrates, many intramembranous particles in cell membrane but few or none in membranes of vesicles and multivesicular structures

Trypanosoma cruzi, illus.
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Trypanosoma cruzi, fine structure morphology of epimastigotes maintained in acellular culture medium, cell division, observation of polysaccharide surface coat

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T[rypanosoma] cruzi, soldiers from 4 different provinces in 3 different years, prevalence of infection, correlation between different serodiagnostic methods: northeastern Argentina

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Sukhareva-Nemakova, N. N.; et al., 1977, Moscow Univ. Biol. Sc. Bull., v. 32 (2), 43-48
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Trypanosoma cruzi
Szarfman, A.; et al., 1977, An. Microbiol., Rio de Janeiro, v. 22, 1976-1977, 95-103
Trypanosoma cruzi, human, diagnosis, comparison between indirect immunofluorescent and indirect immunoperoxidase tests

Trypanosoma cruzi
Szarfman, A.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (4), 832-834
Trypanosoma cruzi, rhesus monkey infected for 29 years, detection of tissue-reacting antibodies similar to those described in human Chagas' disease

Trypanosoma cruzi
Tafuri, W. L.; et al., 1973, Rev. Inst. Med. Trop. S. Paulo, v. 15 (6), 347-370
Trypanosoma cruzi, human, chronic myocarditis, electron microscopic study of heart muscle cells and interstitial tissue, description of types of lesions

T[rypanosoma] cruzi
Tafuri, W. L.; Almeida Maria, T.; and Lopes, E. R., 1973, Rev. Ass. Med. Minas Gerais, v. 24 (1), 1-24
human trypanosomiasis, electron microscopic study of pathologic changes resulting in megaesophagus interstitium

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human Chagasic megaesophagus, role of neurosecretory vesicular component

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Triatoma infestans (exper.)
T. sordida (exper.)
Rhodnius prolixus (exper.)
R. neglectus (exper.)
mice (exper.)

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Tandon, A.; Zahner, H.; and Laemmler, G., 1979, Tropenmed. u. Parasitol., v. 30 (2), 189-193
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morphogenetic effects of precocene II on immature stages of Rhodnius prolixus

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Trypanosoma cruzi
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Trypanosoma cruzi, children with apparent vs. inapparent acute Chagas' disease, clinical and laboratory findings, humoral antibody response, delayed-type skin responses, inhibition of leukocyte migration, serum proteins and immunoglobulins; demonstration of cell-mediated immunodepression in inapparent acute disease

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infectious diseases including leishmaniasis and Chagas' disease, evidence for immunological basis of spectra of clinical manifestations, review

Trypanosoma cruzi
Urdaneta-Morales, S., 1973, Rev. Inst. Med. Trop. S. Paulo, v. 15 (4), 218-221
Rhodnius prolixus fed on Trypanosoma cruzi-infected mice, refeeding experiments with secondary hosts (avian, mammals, reptiles) showed that parasite development in gut is inherent in the insect and not dependent on type of blood supplied by refeeding

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Urrutia, L. E.; Ramirez, R.; and Ruiz, J., 1976, Rev. Inst. Invest. Med., San Salvador, v. 5 (2), 154-159
Trypanosoma cruzi, survey of 40 children with either acute Chagas disease or chagasic myocarditis, 5-year follow-up of relationship between therapy with lampit, electrocardiographic changes, and changes in body weight: San Salvador, El Salvador

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differentiation of vector Triatomidae by veins in wings, application to epidemiology of human trypanosomiasis: Mexico

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Vianna, L. G.; and Vianna, A. L., 1978, Rev. Inst. Med. Trop. S. Paulo, v. 20 (1), 64-66
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Trypanosoma cruzi
Villalta, F. V.; and Leon, W., 1979, J. Parasitol., v. 65 (1), 188-189
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Trypanosoma cruzi epimastigotes, evidence for NADH- and NADPH-linked glutamate dehydrogenases

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Walter, R. D.; and Ebert, F., 1979, Tropenmed. u. Parasitol., v. 30 (1), 9-12
Trypanosoma cruzi, inhibitory effect of polyamines on parasite protein kinases activity

Trypanosoma cruzi
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Trypanosoma cruzi, humans, clinical review, recommendations for using lampit as therapy

Trypanosoma cruzi-like strain, illus.
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Trypanosoma cruzi-like-strain, morphology, frequency and density of parasites in Sanguinus oedipus, infectivity to monkeys and rodents, no clinical or histopathological findings, parasitaemia, development in cell cultures, cyclical development in Rhodnius prolixus and Triatoma infestans
Sanguinus oedipus (nat. and exper.) (Blut): originally from Colombia
Ratten (exper.)
Maeuse (exper.)
Mastomys natalensis (exper.)
Meriones unguiculatus (exper.)
Rhodnius prolixus (exper.)
Triatoma infestans (exper.)

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Trypanosoma cruzi, ecological survey of triatomine vectors disclosed close association of Rhodnius pallescens and Triatoma dimidiata with widely distributed palm tree species; Didelphis marsupialis, Tamandua tetradactyla, and Proechimys semispinosus seem to be principal animal reservoirs: Panama

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Trypanosoma cruzi, leishmanial forms found more frequently in cardiac fibers of men than of women, autopsy survey, possible explanations

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Wilton, D. P.; and Cedillos, R. A., 1978, Bull. Pan Am. Health Organ., v. 12 (2), 116-123
house-dwelling triatomines, distribution and abundance, levels of infection with Trypanosoma cruzi and T. rangeli, impact of previous DDT and propoxur spraying inside houses for malaria control
Rhodnius prolixus (feces)
Triatoma dimidiata (feces)
all from El Salvador

ypanosoma cruzi
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activity of 16 novel 5-nitroimidazoles against protozoa in mice and golden hamsters, compared with metronidazole and tinidazole, structure-activity relationships

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Trypanosoma cruzi, saline extract of Rhodnius prolixus bugs added to insect-oriented culture medium, large increases in trypomastigote forms, useful for Chagas disease studies

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Trypanosoma cruzi in Triatoma protracta protracta, seasonal infection rates: Thousand Oaks, Ventura County, California

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Trypanosoma cruzi epimastigotes, epoxide hydrase, characterization, may be important in detoxication of drugs

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oneda, S., 1971, Rev. Brasil. Pesqui. Med. e iol., v. 4 (3), 205-218
Trypanosoma cruzi in tissue culture, purine metabolism in the intracellular phase of development; possible application to chemotherapeutic assays as during this period parasite exhibits greatest resistance to therapy

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Yoneda, S.; Carvalho, R. P. de S.; and Quiroga, M., 1974, Rev. Inst. Med. Trop. S. Paulo, v. 16 (6), 324-327
Trypanosoma cruzi, intra- and extracellular forms, pyrimidine metabolism

Trypanosoma cruzi
Yoshida, N.; and Camargo, E. P., 1978, J. Bacteriol., v. 136 (3), 1184-1186
trypanosomatids, excretion of urea or ammonia or both, varies according to genus, may be of taxonomic use

Trypanosoma cruzi, illus.
Zavala Velazquez, J.; Arjona Canto, D.; and Quintal Aviles, R., 1973, Rev. Invest. Clin., v. 25 (4), 367-371
Trypanosoma cruzi, elderly man, case report, diagnostic difficulties because of few specific initial symptoms: Hunucma, Yucatan

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Chagas' disease, epidemiology, modes of transmission, and reservoir hosts, review

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Zeledon, R.; de Trejos, M.; and Chinchilla, M., 1977, Protozoology, v. 3, 95-101
Trypanosoma cruzi trypomastigotes, mice, infectivity of blood, culture, and insect forms by 6 routes

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Zingales, B.; et al., 1979, Biochim. et Biophys. Acta, v. 550 (2), 233-244
Trypanosoma cruzi epimastigotes, procedure for purification of adenylyl cyclase-containing plasma membrane fraction, identity and purity of such membranes were assessed by biochemical and ultrastructural criteria

Trypanosoma (Schizotrypanum) cruzi cruzi Chagas 1909
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"T. cruzi sensu stricto thus becomes the nominate subspecies"

Trypanosoma (Schizotrypanum) cruzi marinkellei ssp. nov., illus.
Baker, J. R.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (3), 483-491
Trypanosoma (Schizotrypanum) spp. from Microchiroptera, characterization by DNA buoyant densities and by electrophoretic patterns of 6 isoenzymes
Phyllostomum discolor: Sao Felipe, Bahia, Brazil; Venezuela (Caracas?)
P. hastatus: Abaetetuba, Para, Brazil
Carollia perspicillata: Rio Virilla, San Jose, Costa Rica

Trypanosoma cyclops
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26 trypanosomatid species, cultivation in new chemically-defined medium RE III

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Heywood, P.; and Weinman, D., 1978, J. Protozool., v. 25 (3, pt. 2), 287-292
Trypanosoma cyclops, mitosis, ultrastructure

Trypanosoma cyclops, illus.
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Trypanosoma brucei, T. cyclops, application of scanning electron microscopic techniques to study of trypanosome biology

Trypanosoma danilewskyi Laveran & Mesnil, 1904
Dykova, I.; and Lom, J., 1979, J. Fish Dis., v. 2 (5), 381-390
Trypanosoma danilewskyi and Trypanoplasma borelli in Carassius auratus (exper.), histopathological changes

Trypanosoma deanei Coutinho and Pattolli
Barretto, M. P., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (4), 272-278
as syn. of Trypanosoma (Schizotrypanum) cruzi Chagas, 1909

Trypanosoma diemyctyli (Tobey)
Gill, D. E., 1978, J. Parasitol., v. 64 (5), 930-931
Notophthalmus viridescens eft stage and adults: George Washington National Forest, Rockingham County, Virginia

Trypanosoma dimorphon
Gill, B. S., 1977, Trypanosomes and trypanosomiases of Indian livestock, 137 pp., illus.
extensive review

Trypanosoma (Schizotrypanum) dionisii
Afchain, D.; et al., 1979, J. Parasitol., v. 65 (4), 507-514
Trypanosoma cruzi culture forms, antigenic make-up, comparison with salivarian and some other stercorarian trypanosomes and Leishmania using immunoprecipitation in gels and immunoelectrophoresis, identification of component specific to T. cruzi

Trypanosoma (Schizotrypanum) dionisii
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Trypanosoma (Schizotrypanum) spp. from Microchiroptera, characterization by DNA buoyant densities and by electrophoretic patterns of 6 isoenzymes
Pipistrellus pipistrellus: East Anglia, England; Antwerp, Belgium

Trypanosoma (Schizotrypanum) dionisii
Baker, J. R.; and Liston, A. J., 1978, J. Gen. Microbiol., v. 104 (1), 79-89
Trypanosoma dionisii, effect of various agents (including temperature, complement, trypsin, cytochalasin B and immune plasma) on attachment and entry to mouse peritoneal macrophages in vitro, and subsequent morphogenesis; attachment occurred to non-specific receptors, entry by phagocytosis

Trypanosoma (Schizotrypanum) dionisii Bettencourt & Franca, 1905
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Trypanosoma dionisii, extracellular growth in vitro at 37°C, influence of mouse peritoneal macrophages and calf sera

Trypanosoma dionisii
Ebert, F.; Schudnagis, R.; and Muehlpfordt, H., 1978, Tropenmed. u. Parasitol., v. 29 (1), 115-118
Trypanosoma cruzi and other Trypanosoma spp., protein typing by disc electrophoresis

Trypanosoma dionisii
Hart, P. A., 1979, Lysosomes Applied Biol. and Therap., v. 6, 409-423
phagosome-lysosome fusion in macrophages, possible role in intracellular fate of ingested microorganisms, review including some information on parasitic protozoa

Trypanosoma (Schizotrypanum) dionisii
Liston, A. J.; and Baker, J. R., 1978, J. Gen. Microbiol., v. 107 (2), 253-262
Trypanosoma dionisii, phagocytosis by mouse peritoneal macrophages in vitro and subsequent fate therein

Trypanosoma (Schizotrypanum) dionisii
Liston, A. J.; Baker, J. R.; and Selden, L. F., 1977, J. Gen. Microbiol., v. 98 (2), 615-617
Trypanosoma dionisii, T. vespertilionis, immunized mice, delayed-type hypersensitivity footpad response to homologous or heterologous species

Trypanosoma dionisii, illus.
Thorne, K. J. I.; et al., 1979, Parasitology, v. 79 (3), 367-379
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Trypanosoma dionisii
Thorne, K. J. I.; Svvennsen, R. J.; and Franks, D., 1978, Infect. and Immun., v. 21 (3), 798-805
Trypanosoma dionisii, cytotoxicity of granulocytes and lymphocytes to antibody-coated parasites, granulocytes (and probably also lymphocytes) kill the parasite with hydrogen peroxide by a peroxidase-mediated reaction

Trypanosoma duttoni
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Mus musculus (Blut, Herz, Lunge): Neusiedlerseegebiet, nordlichen Burgenland

Trypanosoma elegans Brumpt (1906)
Letch, C. A., 1979, Parasitology, v. 79 (1), 107-117
Trypanosoma cobitis should be regarded as single species of trypanosome from 6 spp. of British fish on basis of morphology, isoenzyme patterns, and cross-transmission (by syringe passage of culture forms and by leech vector Hemiclepsis marginata), specific names T. phoxini, T. elegans, T. barbatulae, T. occidentalis, and T. langeroni "should be disregarded"

Trypanosoma enhydris Sinha and Mandal
Mandal, A. K., 1978, Indian J. Animal Health, v. 17 (1), 97-98
Trypanosoma enhydris, virulence in Natrix stolata (exper.) and in Enhydris enhydris (nat. and exper.) after passage in Natrix stolata: Chakdah, Nadia Dist., West Bengal (India)

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Feldman, R., 1977, Acta Bioquim. Clin. Latinoam., v. 11 (2), 179-180
protozoa, techniques for microscopical diagnosis

Trypanosoma equinum
Costa, S. C. G.; Silva, A. M.; and Quintao, L. G., 1967, Atas Soc. Biol. Rio de Janeiro, v. 10 (6), 135-136
Trypanosoma equinum, T. lewisi, nucleic acids, cytochemical study

Trypanosoma equinum
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

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Bowman, I. B. R., 1974, Ciba Found. Symp., n.s. (20), 255-284
trypanosomes, intermediary metabolism, review

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Brooks, A. S., 1978, Tr. Am. Micr. Soc., v. 97 (3), 287-296
Crithidia fasciculata, Trypanosoma equiperdum, T. lewisi, ultrastructure of flagellar attachment site, thin section, freeze-etch, and cytochemical techniques

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Czesnel, Z., 1970, Roczniki Pomor. Akad. Med. im. Gen. K. Swierczewskiego Szczecinie, v. 16, 375-390
Trypanosoma equiperdum-infected guinea pigs (exper.), alterations in cardiac muscles, observations on ECG records, histological and histochemical estimations of glycogen content, pyruvic acid levels in blood, evidence of vitamin B_1 deficiency

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Trypanosoma lewisi, rats, Trypanosoma musculi, mice, precipitin responses of immune hosts repeatedly infected, antigenic relationships of these 2 trypanosome species, microimmunodiffusion, crossed immunoelectrophoresis (CIE), and tandem CIE analyses

Trypanosoma lewisi
Elpidina, E. N.; Zaitseva, G. N.; and Krasheninnikov, I. A., 1979, Biokhimiia, v. 44 (10), 1830-1841
Trypanosoma lewisi, histones from nuclei characterized in terms of chemical composition and RNA-polymerase activity

Trypanosoma lewisi
Ferrante, A.; Drew, P. A.; and Jenkin, C. R., 1978, Austral. J. Exper. Biol. and Med. Sc., v. 56 (6), 741-745
Trypanosoma lewisi, rats, method for assay of ablastin in serum, measurement of incorporation of ^{3}H-TdR into T. lewisi DNA

Trypanosoma lewisi
Ferrante, A.; and Jenkin, C. R., 1978, Austral. J. Exper. Biol. and Med. Sc., v. 56 (2), 201-209
Trypanosoma lewisi, rats, importance of monocytic phagocytic system in elimination of parasites during course of infection, relative importance of liver and spleen in removal of parasites, importance of specific antibody in uptake of parasites by liver, production of specific antibody during course of infection, effect of antibody and complement on parasites, fate of trypanosomes within chambers planted into peritoneal cavities of normal and immune rats

Trypanosoma lewisi, illus.
Ferrante, A.; and Jenkin, C. R., 1979, Cellular Immunol., v. 42 (2), 327-335
Trypanosoma lewisi, rat macrophages are able to ingest and then kill parasite in presence of specific antibody

Trypanosoma lewisi
Ferrante, A.; Jenkin, C. R.; and Reade, P. C., 1978, Austral. J. Exper. Biol. and Med. Sc., v. 56 (1), 47-59
Trypanosoma lewisi-infected rats, changes in activity of reticuloendothelial system

Trypanosoma lewisi
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Rattus norvegicus (Blut, Herz, Lunge, Leber): Neusiedlerseegebiet, nordlichen Burgenland

Trypanosoma lewisi, illus.
Giannini, M. S.; and D'Alesandro, P. A., 1978, Science (4359), v. 201, 916-918
Trypanosoma lewisi forms caps at 0°C when incubated with rabbit IgG directed against surface IgG from rat host, host IgG (which is specific for parasite antigens) probably does not cause capping of these antigens in vivo

Trypanosoma lewisi, illus.
Giannini, S. H.; and D'Alesandro, P. A., 1979, Exper. Parasitol., v. 47 (3), 342-355
Trypanosoma lewisi, accumulation of antigen-specific host IgG as component of surface coat during course of infection in rat

Trypanosoma lewisi
Gutteridge, W. E.; Dave, D.; and Richards, W. H. G., 1979, Biochim. et Biophys. Acta, v. 582 (3), 390-401
Kinetoplastida spp., Plasmodium spp., conversion of dihydroorotate to orotate, mechanism of reaction different in these 2 groups of protozoa, possible target of chemotherapeutic attack

Trypanosoma lewisi
Harris, L. M.; et al., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 599-606
Trypanosoma lewisi, blood and culture forms, T. conorhini, culture forms, isolation of DNA by 2 methods, characterization by GC base composition

Trypanosoma lewisi
Hawking, F., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (6), 592-595
Trypanosoma congolense, circadian rhythm in numbers of parasites in blood of laboratory rodents, indisputable rhythms not found in T. vivax, T. brucei, and T. lewisi

Trypanosoma lewisi
Hendricks, L. D.; Wood, D. E.; and Hajduk, M. E., 1978, Parasitology, v. 76 (3), 309-316
Leishmania spp., Trypanosoma spp., commercially available liquid media for rapid cultivation

Trypanosoma lewisi
Jacobson, H. A.; Kirkpatrick, R. L.; and McGinnes, B. S., 1978, Wildlife Monogr. (60), 53 pp.
disease and physiologic characteristics of cottontail rabbits in 2 study areas in relation to population density, includes data on seasonal and sex differences
Sylvilagus floridanus: Virginia

Trypanosoma lewisi, illus.
Kloetzel, J.; and Deane, M. P., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (6), 383-387
Trypanosoma lewisi, T. cruzi sensitized with specific antisera and complement, adherence to rat peritoneal cells; adherence is specific, without cross reactions; results suggest that phagocytosis as well as cytophilic antibodies plays a role in immunity

Trypanosoma lewisi
Kloetzel, J.; and Deane, M. P., 1971, Rev. Inst. Med. Trop. S. Paulo, v. 13 (1), 31-36
Trypanosoma cruzi, T. lewisi, cross-immunity studies in young mice and rats

Trypanosoma lewisi, illus.
Lage, H. A.; and Lage, S. A., 1971, Atas Soc. Biol. Rio de Janeiro, v. 14 (5-6), 125-127
Rattus norvegicus (pulmao): Rio de Janeiro, Guanabara

Trypanosoma lewisi
de Lara, W. G. F., 1972, Rev. Inst. Med. Trop. S. Paulo, v. 14 (5), 301-305
Trypanosoma cruzi, mice, T. lewisi antigen protected against infection but antigen fractions did not

Trypanosoma lewisi
Lee, C. M.; Glasgow George, Y.; and Aboko-Cole, G., 1977, Ztschr. Parasitenk., v. 53 (1), 1-6
Trypanosoma lewisi, rats, iron metabolism, serum iron and serum iron-binding capacity, diets with various levels of iron

Trypanosoma lewisi
Lee, C. M.; Harris, L. M.; and Aboko-Cole, G. F., 1978, Internat. J. Parasitol., v. 8 (3), 187-192
Trypanosoma lewisi, feral isolate, comparative activity in albino vs. black rats (parasitemia, trypanosome cell size and reproductive development, trypanosome respiration), data show that albino rats were significantly more susceptible to infection than black rats

Trypanosoma lewisi
Leong, T. S.; et al., 1979, Southeast Asian J. Trop. Med. and Pub. Health, v. 10 (1), 122-126
parasites of Rattus r. diardii, influence of human habitats on rat parasite fauna
Rattus rattus diardii (blood): Kuala Lumpur and nearby villages

Trypanosoma lewisi
Long, G. W.; and Dusanic, D. G., 1978, Exper. Parasitol., v. 44 (1), 56-65
Trypansosma lewisi, serological reactivities of exoantigens and cellular antigens of bloodstream parasites from immunosuppressed rats (precipitation and agglutination tests), results suggest that likely result of immunosuppressing host is trypansome antigen preparation that is more reactive serodiagnostic reagent

Trypanosoma lewisi
Mankau, S. K., 1975, Kiseichugaku Zasshi (Japan. J. Parasitol.), v. 24 (6), 379-384
Trypanosoma lewisi, white rats (exper.), sex difference in susceptibility to infection demonstrated, effect of gonadectomy and heterologus gonadal hormone treatment

Trypanosoma lewisi
Nielsen, K.; et al., 1978, Canad. J. Comp. Med., v. 42 (1), 74-79
Trypanosoma lewisi, complement activating factor(s), physiochemical characteristics of active components (carbohydrate-containing substance and lipid, possibly glyco lipid)

Trypanosoma lewisi
Nielsen, K.; et al., 1978, Experientia, v. 34 (1), 118-119
Trypanosoma lewisi-infected or decomplemented rats, increased susceptibility to Salmonella typhimurium infection; decomplemented rats subsequently infected with T. lewisi developed higher blood parasitemia than did normal T. lewisi-infected rats

Trypanosoma lewisi
Nielsen, K.; et al., 1978, J. Parasitol., v. 64 (3), 544-546
Trypanosoma congolense, T. lewisi, direct activation of complement

Trypanosoma lewisi
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Trypanosoma lewisi
Schraw, W. P.; and Vaughan, G. L., 1979, Exper. Parasitol., v. 48 (1), 15-26
Trypanosoma lewisi, membrane function (glucose, leucine, and potassium transport; 5'nucleotidase activity) in dividing and ablastin-inhibited trypanosomes

Trypanosoma lewisi
Shirshov, A. T.; Lunina, L. D.; and Zaitseva, G. N., 1978, Biokhimiia, v. 43 (5), 824-829
Trypanosoma lewisi, kinetoplast DNA associate, isolation and physico-chemical characterization

Trypanosoma lewisii
Sinniah, B., 1979, Southeast Asian J. Trop. Med. and Pub. Health, v. 10 (1), 115-121
distribution and prevalence
Rattus annandalei
R. argentiventer
R. rattus diardii
R. exulans
R. tiomanicus
R. norvegicus
all from Peninsular Malaysia

Trypanosoma lewisi, illus.
Thoongsuwan, S.; and Cox, H. W., 1978, J. Parasitol., v. 64 (4), 669-673
Trypanosoma lewisi, ATC strain in Sprague-Dawley rats, anemia, splenomegaly, and glomerulonephritis accompanied by presence of cold-active hemagglutinin for trypsinized rat erythrocytes

Trypanosoma lewisi
Thoongsuwan, S.; Cox, H. W.; and Patrick, R. A., [1979], J. Parasitol., v. 64 (6), 1978, 1050-1056
Trypanosoma lewisi, Babesia rodhaini, Plasmodium chabaudi, rats, acquired nonspecific resistance associated with recovery from various infectious anemias, association with immunoconglutinin activity, infections also stimulated production of cold-active hemagglutinin

Trypanosoma lewisi
Thoongsuwan, S.; Cox, H. W.; and Rickman, W. J., 1979, J. Parasitol., v. 65 (3), 426-429
Plasmodium chabaudi, Babesia rodhaini, Trypanosoma lewisi, rats, presence of antibody to fibrinogen/fibrin products, association with cold-activated hemagglutinin and immunoconglutinin, temporal relationships with anemia and parasitemia

Trypanosoma lewisi
Tizard, I. R.; et al., 1978, Tropenmed. u. Parasitol., v. 29 (1), 127-133
Trypanosoma congolense, concomitant generation of phospholipase A and hemolytic fatty acids by autolysing suspensions, autolysis of T. lewisi did not generate hemolytic activity unless exogenous phospholipase A was added

Trypanosoma lewisi
Uriniuk, V. M.; Sukhareva-Nemakova, N. N.; and Silaev, A. B., 1975, Izvest. Akad. Nauk SSSR, s. Biol. (4), 559-564
Trypanosoma lewisi, effect of sodium citrate on multiplication of parasites and on composition and biosynthesis of lipids, conditions of active aeration

Trypanosoma lewisi
Viyanant, V.; and Jones, A. W., 1978, Southeast Asian J. Trop. Med. and Pub. Health, v. 9 (1), 79-85
Trypanosoma lewisi, surface membrane antigen, isolation and characterization, protection of immunized rats against challenge

Trypanosoma lewisi
Warton, A.; and Modlinska, M., 1975, Acta Parasitol. Polon., v. 23 (1-11), 127-133
Trypanosoma spp., mice, rats, effect of acriflavine on dynamics of trypanosome population size and formation of dyskinetoplastic forms in host blood

Trypanosoma lineatus Iturbe and Gonzalez, 1916
Barretto, M. P., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (4), 272-278
species inquirendae

Trypanosoma (Megatrypanum) lizae n. sp., illus.
Miltgen, F.; and Landau, I., 1979, Ann. Parasitol., v. 54 (2), 163-169
Hipposideros cyclops (sang cardiaque): Gabon, environs de Makokou

Trypanosoma (Herpetosoma) longiflagellum sp. n., illus.
Marinkelle, C. J., 1977, J. Wildlife Dis., v. 13 (3), 262-264
Taphozous nudiventris (blood): Iraq (Numaniya, Wasit Province, 145 km southeast of Baghdad)

Trypanosoma magnusi [? n. sp.], illus.
Shamsuddin, M.; and Mohammad, M. K., 1978, Bull. Nat. Hist. Research Cent., Univ. Baghdad, v. 7 (2), 35-47
Taphozous nudiventris magnus (blood): Al-Khasfa cave and Al-Kifai, Iraq
Pipistrellus kuhli (blood): Al-Akhaidhar, Iraq

Trypanosoma (Herpetosoma) mariae n. sp., illus.
Mello, D. A., 1978, Ann. Parasitol., v. 53 (4), 345-350
Calomys callosus (nat. and exper.) (blood): Formosa county, state of Goias, Brazil
Oryzomys eliurus (exper.) "fleeting low parasitemias"
Xenopsylla cheopis (exper.) (intestine, rectum)
Polygenys jordani (exper.) (intestine, rectum)

Trypanosoma mega Dutton and Todd, 1903 (sensu Bone and Steinert, 1956), illus.
Baker, J. R., 1977, Protozoology, v. 3, 75-82
Bufo regularis: Illubabor Province, Ethiopia

Trypanosoma mega
Camargo, E. P.; et al., 1978, Exper. Parasitol., v. 46 (2), 141-144
Trypanosoma, Leishmania, and Leptomonas spp., enzymes of ornithine-arginine metabolism, existence of genus-specific enzyme patterns which may serve as biochemical markers in classification of Trypanosomatidae

Trypanosoma mega
Camargo, E. P.; Itow, S.; and Alfieri, S. C., [1979], J. Parasitol., v. 64 (6), 1978, 1120-1121
Trypanosomatidae, 18 spp. of 6 genera, proteolytic activities in cell extracts

Trypanosoma mega
Fish, W. R.; Holz, G. G., jr.; and Beach, D. H., 1978, J. Parasitol., v. 64 (3), 546-547
26 trypanosomatid species, cultivation in new chemically-defined medium RE III

Trypanosoma mega
Galinari, S.; and Camargo, E. P., 1978, Exper. Parasitol., v. 46 (2), 277-282
trypanosomatid protozoa, 16 spp., survey for acetylornithinase and ornithine acetyltransferase, metabolic and nutritional implications

Trypanosoma mega
Yoshida, N.; and Camargo, E. P., 1978, J. Bacteriol., v. 136 (3), 1184-1186
trypanosomatids, excretion of urea or ammonia or both, varies according to genus, may be of taxonomic use

Trypanosoma (Megatrypanum) megachiropterorum sp. n., illus.
Marinkelle, C. J., 1979, J. Protozool., v. 26 (3), 352-353
Pteropus tonganus (blood): Kolovai, Tongatapu Island, Kingdom of Tonga

Trypanosoma (Megatrypanum) megadermae
Edrissian, Gh. H.; Farhang-Azad, A.; and Neronov, V. M., 1976, J. Wildlife Dis., v. 12 (4), 497
Rhinolopus hipposiderus: Iran

Trypanosoma microti
Cox, F. E. G., 1977, Protozoology, v. 3, 129-134
Microtus agrestis

Trypanosoma microti
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Microtus oeconomus (Herz)
M. agrestis (Lunge)
all from Neusiedlerseegebiet, nordlichen Burgenland

Trypanosoma minasense
Deane, L. M.; da Silva, J. E.; and Loures, L. (filho), 1974, Rev. Inst. Med. Trop. S. Paulo, v. 16 (1), 1-6
Trypanosoma minasense, naturally infected marmosets, parasitemia has circadian rhythm with highest level at 16:00 hrs, natural vector may be hematophagous insect with preference for afternoon feeding
Callithrix penicillata jordani
C. geoffroyi
all from Brazil

Trypanosoma mrigali (sp. nov.), illus.
Joshi, B. D., 1978, Indian J. Zoot., v. 17 (1), 1976, 5-10
Cirrhina mrigala (plasma): River Gomati, Lucknow

Trypanosoma mugilicola n. sp., illus.
Becker, C. D.; and Overstreet, R. M., 1979, J. Fish Dis., v. 2 (6), 469-479
Mugil cephalus
Trinectes maculatus
(blood of all): all from Mississippi Sound, Gulf of Mexico

Trypanosoma murmanensis Nikitin, 1927
Khan, R. A., 1977, Canad. J. Zool., v. 55 (8), 1235-1241
Hippoglossoides platessoides (nat. and exper.)
Limanda ferruginea (nat. and exper.)
Glyptocephalus cynoglossus
Lycodes reticulatus (nat. and exper.)
Anarhichas minor
A. lupus (nat. and exper.)
Liparis cyclostigma
Gadus morhua (nat. and exper.)
Myoxocephalus octodecemspinosus (exper.)
M. scorpius (exper.)
Anguilla rostrata (exper.)
Pseudopleuronectes americanus (exper.)
Microgadus tomcod (exper.)
Tautogolabrus adspersus (exper.)
Lycodes vahlii (exper.)
Macrozoarces americanus (exper.)
Liparis atlanticus (exper.)
all from off coastal Newfoundland, Canada

Trypanosoma murmanensis
Khan, R. A., 1977, Canad. J. Zool., v. 55 (10), 1698-1700
Trypanosoma murmanensis, infectivity of different morphotypes in fish hosts to the leech vector (Johanssonia sp.)

Trypanosoma murmanensis
Khan, R. A., 1978, Canad. J. Zool., v. 56 (9), 2061-2063
Trypanosoma murmanensis, longevity in marine leech, Johanssonia sp., infection can persist through 5-6 host feedings, survival attributed to residual stages in proboscis

Trypanosoma musculi
Albright, J. W.; and Albright, J. F., 1978, Infect. and Immun., v. 22 (2), 343-349
Trypanosoma musculi, in vitro growth in cultures of murine spleen cells, analysis of requirement for supportive spleen cells, demonstration of utility of this culture system for analysis of host immune responses against the trypanosome

Trypanosoma musculi
Albright, J. W.; Albright, J. F.; and Dusanic, D. G., 1978, Proc. National Acad. Sc., v. 75 (8), 3923-3927
Trypanosoma musculi, mice, mechanisms of trypanosome-mediated suppression of humoral immunity, appears that soluble substances derived from parasites act directly on B lymphocytes or essential assistant cells rather than by activating suppressor T cells or macrophages

Trypanosoma musculi
Brooks, B. O.; and Reed, N. D., 1979, J. Reticuloendothel. Soc., v. 25 (3), 325-328
Trypanosoma musculi, nude and normal mice, trypan blue treatment, results suggest that macrophages may participate in early nonspecific control of parasitemia and in immune clearance of parasitemia

Trypanosoma musculi
Cox, F. E. G., 1977, Protozoology, v. 3, 129-134
Trypanosoma musculi, T. b. brucei, Babesia microti, mice, interactions between parasites

Trypanosoma musculi
Dusanic, D. G., 1978, Internat. J. Parasitol., v. 8 (4), 297-304
Trypanosoma musculi, precipitin responses of infected mice to exoantigens and cellular antigens

Trypanosoma musculi
Dusanic, D. G., 1979, Internat. J. Parasitol., v. 9 (6), 577-583
Trypanosoma lewisi, rats, Trypanosoma musculi, mice, precipitin responses of immune hosts repeatedly infected, antigenic relationships of these 2 trypanosome species, microimmunodiffusion, crossed immunoelectrophoresis (CIE), and tandem CIE analyses

Trypanosoma musculi
Hazlett, C. A.; and Tizard, I. R., 1978, Clin. and Exper. Immunol., v. 33 (2), 225-231
Trypanosoma musculi, immunosuppressive and mitogenic effects, possible relationship of mitogenesis to immunosuppression and non-specific antibody formation associated with infections

Trypanosoma musculi
Robinett, J. P.; and Rank, R. G., 1979, Infect. and Immun., v. 23 (2), 270-275
Trypanosoma musculi, mice, splenomegaly is T cell-dependent and is the result of proliferation of B and/or T lymphocytes

Trypanosoma musculi
Watson, L. P.; and Lee, C. M., 1979, Ztschr. Parasitenk., v. 59 (3), 299-301
Trypanosoma musculi, absense of ^{51}Cr binding to bloodstream trypomastigotes

Trypanosoma neinavana sp. n., illus.
Fattohy, Z. I., 1978, Current Sc., Bangalore, v. 47 (1), 33-35
Barbus grypus (blood): river Tigris, Mosul, Iraq

Trypanosoma ninae kohl-yakimov Yakimoff, 1918
Butabaeva, M., 1979, Uzbek. Biol. Zhurnal (2), 50-53
[Sus scrofa]: Kashkadar'insk oblast

Trypanosoma occidentalis Becker (1967)
Letch, C. A., 1979, Parasitology, v. 79 (1), 107-117
Trypanosoma cobitis should be regarded as single species of trypanosome from 6 spp. of British fish on basis of morphology, isoenzyme patterns, and cross-transmission (by syringe passage of culture forms and by leech vector Hemiclepsis marginata), specific names T. phoxini, T. elegans, T. barbatulae, T. occidentalis, and T. langeroni "should be disregarded"

Trypanosoma occidentalis
Ollenschlaeger, B., 1975, Tieraerztl. Prax., v. 3 (1), 99-107
blood and other parasites of commercial fish, pathology, transmission, therapy, clinical review

Trypanosoma pancali n. sp., illus.
Mandal, A. K., 1975, Ang. Parasitol., v. 16 (2), 87-93
Mastacembelus pancalus (blood): India (Champahati; Parganas, West Bengal)

Trypanosoma phoxini Brumpt (1906)
Letch, C. A., 1979, Parasitology, v. 79 (1), 107-117
Trypanosoma cobitis should be regarded as single species of trypanosome from 6 spp. of British fish on basis of morphology, isoenzyme patterns, and cross-transmission (by syringe passage of culture forms and by leech vector Hemiclepsis marginata), specific names T. phoxini, T. elegans, T. barbatulae, T. occidentalis, and T. langeroni "should be disregarded"

Trypanosoma phyllostomae Cartaya, 1910
Barretto, M. P., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (4), 272-278
as syn. of Trypanosoma (Schizotrypanum) cruzi Chagas, 1909

Trypanosoma (Schizotrypanum) pipistrelli or Trypanosoma (Schizotrypanum) vespertilionis
Edrissian, Gh. H.; Farhang-Azad, A.; and Neronov, V. M., 1976, J. Wildlife Dis., v. 12 (4), 497
Pipistrellus pipistrellus: Iran

Trypanosoma prowazeki Berenberg-Gossler, 1908
Barretto, M. P., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (4), 272-278
species inquirendae

Trypanosoma rabinowitschi
Frank, C., 1978, Ang. Parasitol., v. 19 (3), 137-154
Cricetus cricetus (Blut, Herz): Neusiedlerseegebiet, nordlichen Burgenland

Trypanosoma rajae
Daily, D. D., 1978, J. Parasitol., v. 64 (2), 361-362
Raja erinacea: Whiting Bay, Maine

Trypanosoma rajae Laveran and Mesnil, 1902, illus.
Laird, M.; and Bullock, W. L., 1969, J. Fish. Research Bd. Canada, v. 26 (4), 1075-1102
brief description
Raja erinacea: St. Andrews, New Brunswick; Woods Hole, Massachusetts
R. radiata: St. Andrews, New Brunswick
(blood of all)

Trypanosoma (H[erpetosoma]) rangeli
Afchain, D.; et al., 1979, J. Parasitol., v. 65 (4), 507-514
Trypanosoma cruzi culture forms, antigenic make-up, comparison with salivarian and some other stercorarian trypanosomes and Leishmania using immunoprecipitation in gels and immunoelectrophoresis, identification of component specific to T. cruzi

Trypanosoma rangeli
Anthony, R. L.; Johnson, C. M.; and Sousa, O. E., 1979, Am. J. Trop. Med. and Hyg., v. 28 (6), 969-973
Trypanosoma cruzi, T. rangeli, Panamanian villagers, diagnosis, micro-enzyme-linked immunosorbent assay, some serologic cross-reactivity between 2 species; comparison with complement xation, direct agglutination, and clinical diagnosis

ypanosoma rangeli
Balestrini, C.; and Rebollo, O. N., 1976, Rev. Inst. Invest. Med., San Salvador, v. 5 (2), 141-144
Trypanosoma cruzi, T. rangeli, human blood samples being tested for malaria found to be positive also for trypanosomiasis, epidemiologic survey, determination of possible vectors: El Salvador

ypanosoma rangeli
Brener, Z.; Filardi, L. S.; and Cuba, C., 1975, Rev. Inst. Med. Trop. S. Paulo, v. 17 (1), 16-19
Trypanosoma rangeli from salivary glands and haemolymph of Rhodnius ecuadoriensis, successful preservation in liquid nitrogen, infective for mice after 30 days preservation, with subsequent normal cyclical transmission

ypanosoma rangeli, illus.
Bretting, H.; and Schottelius, J., 1978, Ztschr. Parasitenk., v. 57 (3), 213-219
Trypanosoma cruzi and T. cruzi-like strains, differentiation from T. rangeli but not from T. conorhini by microimmunofluorescence using lectin of sponge (Aaptos papillata)

ypanosoma rangeli, illus.
Cuba, C. A. C., 1974, Rev. Inst. Med. Trop. S. Paulo, v. 16 (1), 10-18
Trypanosoma rangeli in Rhodnius ecuadoriensis, confirmation of natural infection by dissection and by transmission to white mice, parasite developmental stages
Rhodnius ecuadoriensis (estomago, duodeno, intestino posterior, hemolinfa, glandulas salivares): Distrito de Cascas, Provincia de Contumaza, Cajamarca, Peru
camundongos albinos (sangue) (exper.)

panosoma rangeli, illus.
uba, C. A. C., 1974, Rev. Inst. Med. Trop. S. Paulo, v. 16 (1), 19-27
Trypanosoma rangeli, Rhodnius ecuadoriensis experimentally infected by feeding on guinea pigs, capacity to transmit infection to mice by bite, but not by inoculation of forms eliminated in feces, parasite developmental periods
Rhodnius ecuadoriensis (fezes, tubo digestivo, hemolinfa, glandulas salivares) (exper.)
cobaias (sangue) (exper.)
camundongos albinos (sangue) (exper.)

ypanosoma rangeli, illus.
Cuba, C. A. C., 1975, Rev. Inst. Med. Trop. S. Paulo, v. 17 (4), 211-217
Trypanosoma rangeli in Panstrongylus herreri (exper.), intestinal and haemocoelic but not salivary gland infections, not considered natural vector

ypanosoma rangeli-like trypanosomes, illus.
uba, C.; et al., 1972, Rev. Inst. Med. Trop. S. Paulo, v. 14 (3), 191-202
Trypanosoma rangeli-like trypanosomes, natural infection in Rhodnius ecuadoriensis, potential as vector for humans: State of Cajamarca, Peru

ypanosoma rangeli, illus.
uba, C. A. C., 1975, Rev. Inst. Med. Trop. S. Paulo, v. 17 (5), 283-297
Trypanosoma rangeli, Peruvian strain, growth and development in Rhodnius ecuadoriensis (hemocele, glandulas salivares) (exper.)

Trypanosoma rangeli
Ebert, F.; Schudnagis, R.; and Muehlpfordt, H., 1978, Tropenmed. u. Parasitol., v. 29 (1), 115-118
Trypanosoma cruzi and other Trypanosoma spp., protein typing by disc electrophoresis

Trypanosoma rangeli
Gallardo Zerpa, M. F., 1974, Rev. Fac. Farm., Univ. Central Venezuela (27), v. 14, 69-73
Trypanosoma cruzi, Leishmania donovani, L. braziliensis, cultured successfully in diphasic autoclaved medium of tryptose, glucose, mineral salts, vitamins, blood and agar; medium would not support growth of T. rangeli

Trypanosoma rangeli
Hendricks, L. D.; Wood, D. E.; and Hajduk, M. E., 1978, Parasitology, v. 76 (3), 309-316
Leishmania spp., Trypanosoma spp., commercially available liquid media for rapid cultivation

Trypanosoma rangeli Tejera, 1920
Herrer, A.; Wygodzinsky, P.; and Napan, M., 1972, Rev. Biol. Trop., v. 20 (1), 141-149
Trypanosoma rangeli, geographic distribution survey of vector Rhodnius ecuadoriensis: Amazonian and Pacific watersheds, Peru

Trypanosoma rangeli (Tejera, 1920), illus.
de Lucena, D. T.; and Vergetti, J. G., 1973, Rev. Inst. Med. Trop. S. Paulo, v. 15 (4), 171-178
Panstrongylus megistus (conteudo intestinal): Ibateguara, Estado de Alagoas, Brasil

Trypanosoma rangeli
Marinkelle, C. J., 1967, Rev. Biol. Trop., v. 15 (1), 203-205
Triatoma dimidiata capitata (salivary glands, hemolymph, intestine): Colombia (Depts. of Boyaca, Cundinamarca, Huila, and Santander)
mice (blood) (exper.)
Rhodnius prolixus (salivary glands) (exper.)
people: San Joaquin, Dept. of Santander, Colombia

Trypanosoma rangeli Tejera, 1920
Panza Carpio, J. A., 1974, Rev. Fac. Farm., Univ. Central Venezuela (27), v. 14, 61-68
Trypanosoma rangeli, culture in diphasic autoclaved medium containing human albumin, implications for nutritional requirements

Trypanosoma rangeli
Ponce, C.; Trochez, H.; and Zeledon, R., 1975, Rev. Biol. Trop., v. 22 (2), 1974, 289-303
Trypanosoma cruzi, T. rangeli, single and mixed infections, epidemiologic survey of area infested with Rhodnius prolixus: Departamento Francisco Morazan, Honduras

Trypanosoma rangeli
Sousa, O. E., 1972, Rev. Biol. Trop., v. 20 (2), 167-179
Trypanosoma cruzi, T. rangeli, epidemiologic survey, vectors, reservoir hosts: Panama

Trypanosoma rangeli
Wilton, D. P.; and Cedillos, R. A., 1978, Bull. Pan Am. Health Organ., v. 12 (2), 116-123
house-dwelling triatomines, distribution and abundance, levels of infection with Trypanosoma cruzi and T. rangeli, impact of previous DDT and propoxur spraying inside houses for malaria control
Rhodnius prolixus (feces, hemolymph): El Salvador

Trypanosoma rangeli
Zeledon, R.; Ponce, C.; and Murillo, J., 1979, J. Parasitol., v. 65 (2), 275-279
Bradypus griseus
Choloepus hoffmanni (spleen)
all from Costa Rica

Trypanosoma remaki part. var. parva Laveran et Mesnil, 1901
Khaibulaev, K. Kh., 1971, Parazitologiia, Leningrad, v. 5 (6), 551-555
as syn. of T. schulmani sp. n.

Trypanosoma rhamdiae Splendore, 1910 nec Botelho, 1907
Froes, O. M.; et al., 1978, Rev. Brasil. Biol., v. 38 (2), 461-468
as syn. of Trypanosoma splendorei sp. n.

Trypanosoma rhamdiae Botelho, 1907, illus.
Froes, O. M.; et al., 1978, Rev. Brasil. Biol., v. 38 (2), 461-468
description
Rhamdia sapo (sangue): estuario de Guaiba, Rio Grande do Sul

Trypanosoma rhodesiense
Bouree, P., 1976, Rev. de Med., Paris, v. 17 (39), 2139-2150
Trypanosoma spp., geographic distribution, vectors, recommended therapeutic measures, review

Trypanosoma rhodesiense
Bowman, I. B. R., 1974, Ciba Found. Symp., n.s. (20), 255-284
trypanosomes, intermediary metabolism, review

Trypanosoma rhodesiense
Buyst, H., 1975, Ann. Soc. Belge Med. Trop., v. 55 (2), 95-104
Trypanosoma rhodesiense, human, specific treatment with suramin and mel B, adjuvant antimalarial treatment with chloroquine and proguanil; modifications of sleeping sickness therapy advocated on physio-pathological and epidemiological grounds: Luangwa Valley, Zambia

T[rypanosoma] rhodesiense
Buyst, H., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 201-212
sleeping sickness in children, epidemiologic data, age distribution, various therapeutic regimens, usefulness of antimalarial therapy administered simultaneously: Isoka, Zambia

Trypanosoma rhodesiense
Buyst, H., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 349-359
Trypanosoma brucei, T. rhodesiense, human, prehistorical and historical background of trypanosomiasis in Africa with special reference to the Luangwa Valley

Trypanosoma rhodesiense
Campbell, G. H.; Esser, K. M.; and Phillips, S. M., 1978, Infect. and Immun., v. 20 (3), 714-720
Trypanosoma rhodesiense infection in congenitally athymic (nude) mice, reduced parasitemia and prolonged survival, active immunity after infection and cure or after immunization with irradiated organisms, results indicate that resistance of mice to T. rhodesiense infection is relatively independent of thymic lymphocyte function

Trypanosoma rhodesiense
Cohen, C., 1973, South African Med. J., v. 47 (11), 485-491
trypanosomiasis, humans, clinical presentation and histopathological findings of 5 cases, need for inclusion in differential diagnosis and use of confirmatory tests discussed: Witwatersrand, South Africa

Trypanosoma rhodesiense
Duffus, W. P. H.; et al., 1978, Infect. and Immun., v. 22 (2), 492-501
Theileria parva, Trypanosoma rhodesiense, ^{51}Cr-labeled chicken erythrocytes coated with protozoal antigens form suitable targets for bovine antibody-dependent cell-mediated cytotoxicity assays

Trypanosoma rhodesiense
Finerty, J. F.; Krehl, E. P.; and McKelvin, R. L., 1978, Infect. and Immun., v. 20 (2), 464-467
Trypanosoma rhodesiense-immunized mice, delayed-type hypersensitivity elicited, results suggest that T-cell activation was necessary component in protective response

Trypanosoma rhodesiense, illus.
Fleischer, N. K. F., 1979, Med. Welt., v. 30 (44), 1625-1630
parasitic tropical diseases, humans, central nervous system involvement, clinical review

Trypanosoma rhodesiense
Flemmings, B.; and Diggs, C., 1978, Infect. and Immun., v. 19 (3), 928-933
Trypanosoma rhodesiense, antibody-dependent cytotoxicity against trypanosomes mediated through alternative complement pathway

Trypanosoma rhodesiense, illus.
Gear, J. H. S.; and Measroch, V., 1971, Recent Advances Stud. Cardiac Struct. and Metab., v. 2, 141-163
human filariasis, malaria and trypanosomiasis, etiology in endomyocardial fibrosis and other infective forms of myocarditis, review

Trypanosoma rhodesiense
Hill, G. C.; et al., 1978, Science (4369), v. 202, 763-765
Trypanosoma rhodesiense, infective cultures initiated and maintained in vitro on Chinese hamster lung cells, cultured trypomastigotes are infective for mice and rats and are morphologically similar to bloodstream trypomastigotes, in addition they possess L-α-glycerophosphate oxidase system characteristic of bloodstream forms

Trypanosoma rhodesiense
Kinnamon, K. E.; and Rane, D. S., 1978, Internat. J. Parasitol., v. 8 (6), 515-523
Trypanosoma rhodesiense, mice, greater than 1 year protection from lethal infections by prophylactic drugs and active immunity

Trypanosoma rhodesiense
Kinnamon, K. E.; Steck, E. A.; and Rane, D. S., 1979, Antimicrob. Agents and Chemotherapy, v. 15 (2), 157-160
Trypanosoma rhodesiense, mice, activity of antitumor drugs against infection

Trypanosoma rhodesiense
Kinnamon, K. E.; Steck, E. A.; and Rane, D. S., 1979, J. Med. Chem., v. 22 (4), 452-455
Trypanosoma rhodesiense, mice, activity of benzyltriphenylphosphonium salts, toxicity at higher dose levels

Trypanosoma rhodesiense
van Knapen, F.; Buys, J.; and Ruitenberg, E. J., 1977, Ann. Soc. Belg Med. Trop., v. 57 (4-5), 281-292
African trypanosomiasis, humans, diagnosis, evaluation of enzyme linked immunosorbent assay, comparison with other seroimmunologic tests

T[rypanosoma] rhodesiense
Limbos, P.; et al., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 495-500
T[rypanosoma] rhodesiense, 16-year-old girl, renal insufficiency after treatment with pentamidine: European visitor to Kagera park, Rwanda

Trypanosoma rhodesiense
Lindsley, H. B.; Nagle, R. B.; and Stechschulte, D. J., 1978, Am. J. Trop. Med. and Hyg., v. 27 (5), 864-872
Trypanosoma rhodesiense-infected rats, proliferative glomerulonephritis, hypocomplementemia, nucleic acid antibodies, feasibility of rat as model host

Trypanosoma rhodesiense
Mansfield, J. M.; and Bagasra, O., 1978, J. Immunol., v. 120 (3), 759-765
Trypanosoma rhodesiense, mice, B cell responses to helper T cell-independent and -dependent antigens, implications for mechanism of immune system dysfunction in chronic African trypanosomiasis

Trypanosoma rhodesiense
Morton, D. M.; Fuller, D. M.; and Green, J. N., 1973, Xenobiotica, v. 3 (4), 257-266
2-styryl-5-nitroimidazoles, metabolism and excretion in laboratory animals, activity against Trypanosoma rhodesiense

Trypanosoma (Trypanozoon) rhodesiense, illus.
Muehlpfordt, H., 1977, Protozoology, v. 3, 71-74
Trypanosoma spp., cell surface labelling with cationic ferritin

Trypanosoma rhodesiense
Powell, C. N., 1978, Experientia, v. 34 (11), 1450-1451
Trypanosoma rhodesiense, rats, inoculation with fraction 3, protection against challenge with T. brucei

Trypanosoma rhodesiense
de Raadt, P., 1974, Ciba Found. Symp., n.s. (20), 199-224
African trypanosomiasis, immunity and antigenic variation, clinical observations suggestive of immune phenomena, review

Trypanosoma rhodesiense
Raether, W.; and Seidenath, H., 1977, Ztschr. Parasitenk., v. 53 (1), 41-46
parasitic protozoans, survival following prolonged storage in liquid nitrogen, some species successfully recovered after preservation for over 10 years

Trypanosoma rhodesiense
Rifkin, M. R., 1978, Proc. National Acad. Sc., v. 75 (7), 3450-3454
trypanocidal factor in normal human serum is associated with high density lipoprotein (HDL), comparison of susceptibility of Trypanosoma brucei and T. rhodesiense to lysis by human serum and human HDL

Trypanosoma rhodesiense
Sadun, E. H., 1974, Parassitologia, v. 16 (1), 53-56
Trypanosoma rhodesiense in Macaca mulatta, clinical, parasitological, hematological, pathological, and immunological studies

Trypanosoma rhodesiense
Shetty, R. V.; and Blanton, C. D., jr., 1978, J. Med. Chem., v. 21 (9), 995-998
2-substituted primaquine analogues synthesized and evaluated in laboratory animals against Plasmodium berghei, P. cynomolgi, Trypanosoma rhodesiense, Leishmania donovani and Schistosoma mansoni; significant activity was observed against P. berghei and L. donovani

Trypanosoma rhodesiense
Van Meirvenne, N.; et al., 1973, Ann. Soc. Belge Med. Trop., v. 53 (2), 109-122
Trypanosoma gambiense, T. rhodesiense, detection in human blood using column separation and membrane filtration

Trypanosoma rhodesiense
Wellhausen, S. R.; and Mansfield, J. M., 1979, J. Immunol., v. 122 (3), 818-824
Trypanosoma rhodesiense-infected mice, spleen is major target organ for immunosuppression, suppression is attributable to emergence of suppressor macrophage population that is restricted to the spleen

Trypanosoma rhodesiense, illus.
Williamson, J.; and McLaren, D. J., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (6), 660-661
Trypanosoma rhodesiense, ultrastructural alterations induced by treatment with DAPI (new diamidine trypanocide)

Trypanosoma rhodesiense
Williamson, J.; and Scott-Finnigan, T. J., 1978, Antimicrob. Agents and Chemotherapy, v. 13 (5), 735-744
trypanocidal activity of antitumor antibiotics and other metabolic inhibitors, microtest for rapid preliminary assay in vitro, parasite motility and infectivity for mice are indexes respectively of respiration and glycolysis and of cell division, implications of results for combination chemotherapy and deposit prophylaxis (with polyanions)

Trypanosoma rotatorium (Mayer, 1843) Laveran and Mesnil, 1901, illus.
Baker, J. R., 1977, Protozoology, v. 3, 75-82
Bufo regularis: Illubabor Province, Ethiopia

Trypanosoma rotatorium Meyer, 1843
da Costa, S. C. G.; et al., 1970, Atas Soc. Biol. Rio de Janeiro, v. 12, Suppl., 7-13
Leptodactylus ocellatus: Manguinhos, Guanabara

Trypanosoma rotatorium Mayer, 1843, illus.
da Costa, S. C. G.; and da Silva, A. M., 1969, v. 12 (5-6), 245-246
Trypanosoma rotatorium, polymorphism, cytological aspects

Trypanosoma rotatorium, illus.
Desser, S. S., 1976, Canad. J. Zool., v. 54 (10), 1712-1723
Trypanosoma rotatorium in Batracobdella picta (gastric ceca) which had fed on infected Rana clamitans, ultrastructure of epimastigote stages

Trypanosoma rotatorium complex, illus.
Ramos, B.; and Urdaneta-Morales, S., 1977, Rev. Biol. Trop., v. 25 (2), 209-217
Trypanosoma rotatorium complex from frogs, experimental infection of hematophagous insects, course of development in Aedes aegypti gut, postulate transmission to frogs via ingestion of mosquitoes
Aedes aegypti (intestine, stomach, hind gut) (exper.)
Culex pipiens (exper.)
Colostethus trinitatis
Hyla crepitans (nat. and exper.)
Leptodactylus insularum (nat. and exper.)
L. macrosternum
L. sibilatrix
Pseudis paradoxus
all from Venezuela

Trypanosoma rotatorium (Mayer, 1843), illus.
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
description
Syn.: Amoeba rotatoria Mayer, 1893
Rana esculenta (krev): CSSR

Trypanosoma sanmartini Garnham and Gonzalez, 1962, pro parte
Barretto, M. P., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (4), 272-278
as syn. of Trypanosoma (Schizotrypanum) cruzi Chagas, 1909

Trypanosoma schulmani sp. n., illus.
Khaibulaev, K. Kh., 1971, Parazitologiia, Leningrad, v. 5 (6), 551-555
Syn.: T. remaki part. var. parva Laveran et Mesnil, 1901
Esox lucius (blood): estuary of Ural River, Sara Peninsula, and Agrakhan Gulf, Caspian Sea

Trypanosoma seenghali (sp. nov.), illus.
Joshi, B. D., 1978, Indian J. Zoot., v. 17 (1), 1976, 5-10
Mystus seenghala (plasma): River Gomati, Lucknow

Trypanosoma serveti, illus.
Guerrero, S.; and Ayala, S. C., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (5), 283-288
Anolis fuscoauratus (sangre): selva amazonica, Huanuco, Peru

Trypanosoma splendorei sp. n., illus.
Froes, O. M.; et al., 1978, Rev. Brasil. Biol., v. 38 (2), 461-468
Syn.: Trypanosoma rhamdiae Splendore, 1910 nec Botelho, 1907
Rhamdia quelen (sangue): rio Tiete, Sao Paulo

Trypanosoma stigmai sp. nov., illus.
Joshi, B. D., [1979], J. Animal Morph. and Physiol., v. 25 (1-2), 1978, 1-7
Puntius stigma: river Gomati [and/or] Chinhat lake of Lucknow district

Trypanosoma theileri
Esuruoso, G. O., 1975, Bull. Animal Health and Prod. Africa, v. 23 (3), 323-332
trypanosomiasis, N'dama cattle, observations on outbreaks, association with various epizootiological factors: western Nigeria

Trypanosoma theileri Laveran, 1902
Gill, B. S., 1977, Trypanosomes and trypanosomiases of Indian livestock, 137 pp., illus.
extensive review

Trypanosoma theileri
Grundboeck, M., 1977, Biul. Vsesoiuz. Ordena Lenin. Inst. Eksper. Vet., v. 30, 14-15
protozoa, cattle, as cofactors in leucotic process

Trypanosoma theileri
McHolland-Raymond, L. E.; Kingston, N.; and Trueblood, M. S., 1978, J. Protozool., v. 25 (3, pt. 2), 388-394
Trypanosoma theileri, continuous cultivation at 37 C in bovine cell culture

Trypanosoma theileri Laveran 1902, illus.
Mammerickx, M.; and Dekegel, D., 1976, Ann. Soc. Belge Med. Trop., v. 56 (1), 47-53
Trypanosoma theileri, cattle, frequent mixed infection with enzootic bovine leukosis, no correlation between diseases but common arthropod vector hypothesized: Belgium

Trypanosoma theileri
Marshall, J.; and Herbert, W. J., 1979, Brit. Vet. J., v. 135 (5), 416-419
Trypanosoma theileri, attempt to infect Stomoxys calcitrans with culture forms unsuccessful

Trypanosoma theileri
Niak, A., 1978, Trop. Animal Health and Prod., v. 10 (1), 26-27
Trypanosoma theileri, cattle (blood), incidence, seasonal distribution: Iran

Trypanosoma (Megatrypanum) theileri (Laveran 1902), illus.
Schlafer, D. H., 1979, Cornell Vet., v. 69 (4), 411-425
Trypanosoma theileri, cattle, incidence, comparison with reports throughout the world: New York

Trypanosoma theileri Laveran 1902, illus.
Warenycia, M. W.; et al., 1978, Experientia, v. 34 (4), 473-475
Trypanosoma theileri, identification as contaminant in primary cultures of bovine retina, successful subculture for 2 passages in sub-confluent cultures, some nutritional requirements, when cultures reached confluency no trypomastigotes or epimastigotes could be detected and attempts to recover trypanosomes from these cultures were unsuccessful

Trypanosoma (Megatrypanum) theileri, illus.
Wink, M., 1979, Internat. J. Parasitol., v. 9 (6), 585-589
Trypanosoma theileri, in vitro cultivation in semidefined insect and vertebrate cell culture media and in Glossina and vertebrate cell culture systems

Trypanosoma (Herpetosoma) ulanovi
Edrissian, Gh. H.; Farhang-Azad, A.; and Neronov, V. M., 1976, J. Wildlife Dis., v. 12 (4), 497
Allactaga williamsi: Iran

Trypanosoma venezuelense
Ercoli, N., 1978, Proc. Soc. Exper. Biol. and Med., v. 157 (3), 397-401
Trypanosoma venezuelense, rats, comparison of "true"and "false" prophylaxis using pentamidine, suramin and 98/202

Trypanosoma (Schizotrypanum) vespertilionis
Baker, J. R.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (3), 483-491
Trypanosoma (Schizotrypanum) spp. from Microchiroptera, characterization by DNA buoyant densities and by electrophoretic patterns of 6 isoenzymes
Nyctalus noctula: East Anglia, England
Pipistrellus pipistrellus: Rollesby, Norfolk, England; East Anglia, England

Trypanosoma (Schizotrypanum) vespertilionis Battaglia, 1904
Barretto, M. P., 1970, Rev. Inst. Med. Trop. S. Paulo, v. 12 (4), 272-278
valid species

Trypanosoma (Schizotrypanum) vespertilionis
Edrissian, Gh. H.; Farhang-Azad, A.; and Neronov, V. M., 1976, J. Wildlife Dis., v. 12 (4), 497
Eptesicus ognevi
Myotis blythi
all from Iran

Trypanosoma (Schizotrypanum) vespertilionis or Trypanosoma (Schizotrypanum) pipistrelli
Edrissian, Gh. H.; Farhang-Azad, A.; and Neronov, V. M., 1976, J. Wildlife Dis., v. 12 (4), 497
Pipistrellus pipistrellus: Iran

Trypanosoma (Schizotrypanum) vespertilionis Battaglia, 1904, illus.
Fahmy, M. A. M.; Abedl-Rahman, A. M.; and Khalifa, R., 1978, Parasitology, v. 77 (3), 249-254
Pipistrellus kuhli, syn. Vesperugo kuhli (blood): villages near Assiut city, Upper Egypt

Trypanosoma (Schizotrypanum) vespertilionis
Liston, A. J.; Baker, J. R.; and Selden, L. F., 1977, J. Gen. Microbiol., v. 98 (2), 615-617
Trypanosoma dionisii, T. vespertilionis, immunized mice, delayed-type hypersensitivity footpad response to homologous or heterologous species

Trypanosoma (Schizotrypanum) vespertilionis, illus.
Molyneux, D. H.; and Bafort, J. M., 1971, Ann. Soc. Belges Med. Trop. Parasitol., v. 51 (3), 335-348
Trypanosoma vespertilionis, morphology of bloodstream forms, sites and morphology of tissue stages (in cysts), morphology and ultrastructure of culture forms, difficulties in differentiating from T. cruzi
Pipistrellus pipistrellus (cardiac and skeletal muscle, gut wall, blood): Britain

T[rypanosoma] vivax
Bourn, D.; and Scott, M., 1978, Trop. Animal Health and Prod., v. 10 (4), 191-203
trypanosomiasis, successful use of Zebu work oxen in agricultural development of tsetse infested land, environmental conditions, epizootiology of trypanosomiasis in oxen and in Glossina morsitans, strategic drug use (alternation of diminazene aceturate and isometamidium to control trypanosomes; rafoxanide to control helminths): Wollega province, western Ethiopia

T[rypanosoma] vivax
Cross, G. A. M., 1977, Ann. Soc. Belg Med. Trop., v. 57 (4-5), 389-402
T[rypanosoma] spp., identification, purification, and characterization of class of surface glycoproteins which appear to be primary mediators of antigenic variations

Trypanosoma (Duttonella) vivax
Draeger, N.; and Mehlitz, D., 1978, Tropenmed. u. Parasitol., v. 29 (2), 223-233
Trypanosoma spp., wildlife, prevalence determined by parasitological and/or serological techniques, correlations with high and low tsetse fly density areas (for buffalo and lechwe) and with host age (for buffalo)
Syncerus caffer
Kobus leche
Redunca arundium
all from Northern Botswana

Trypanosoma vivax
Esuruoso, G. O., 1975, Bull. Animal Health and Prod. Africa, v. 23 (3), 323-332
trypanosomiasis, N'dama cattle, observations on outbreaks, association with various epizootiological factors: western Nigeria

Trypanosoma vivax
Esuruoso, G. O., 1977, Bull. Animal Health and Prod. Africa, v. 25 (3), 233-244
Trypanosoma vivax, Zebu vs. Muturu cattle (exper.), differences in innate resistance, comparison of haematological, clinical, and serological responses

Trypanosoma vivax
Evans, D. A.; and Holland, M. F., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (2), 203-204
Trypanosoma vivax, mice, effective treatment with salicylhydroxamic acid + glycerol

Trypanosoma vivax
de Gee, A. L. W.; Shah, S. D.; and Doyle, J. J., 1979, Exper. Parasitol., v. 48 (3), 352-358
Trypanosoma vivax, sequence of antigenic variants in mice and goats

Trypanosoma vivax
Goodwin, L. G., 1974, Ciba Found. Symp., n.s. (20), 107-124
African trypanosomiasis, mechanisms of pathogenesis, review

Trypanosoma vivax
Greig, W. A.; et al., 1979, Brit. Vet. J., v. 135 (2), 130-141
Trypanosoma congolense, T. vivax, cattle, factors affecting blood sampling for parasitemia and anemia (diurnal variation; ear-vein vs. jugular-vein blood): The Gambia

Trypanosoma vivax
Gutteridge, W. E.; Dave, D.; and Richards, W. H. G., 1979, Biochim. et Biophys. Acta, v. 582 (3), 390-401
Kinetoplastida spp., Plasmodium spp., conversion of dihydroorotate to orotate, mechanism of reaction different in these 2 groups of protozoa, possible target of chemotherapeutic attack

Trypanosoma vivax
Hannon, R. H.; and Parr, C. W., 1978, Comp. Biochem. and Physiol., v. 60B (2), 177-181
Trypanosoma brucei, T. vivax, bloodstream forms, phosphoglucose isomerases, partial purification and characterization, comparison of the two species, many similarities in kinetic properties but differences in thermal stability and in isoelectric point

Trypanosoma vivax
Hawking, F., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (4), 405-407
Trypanosoma evansi, sensitivity to human plasma; T. congolense, T. vivax, resistance to human plasma, suggests that latter 2 species might produce transient infection in man

Trypanosoma vivax
Hawking, F., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (6), 592-595
Trypanosoma congolense, circadian rhythm in numbers of parasites in blood of laboratory rodents, indisputable rhythms not found in T. vivax, T. brucei, and T. lewisi

Trypanosoma vivax
Ilemobade, A. A.; and Blotkamp, C., 1978, Tropenmed. u. Parasitol., v. 29 (4), 443-450
Eperythrozoon ovis, sheep (exper.), pathologic changes in blood picture, concurrent or superimposed infection with Trypanosoma vivax did not affect growth of either parasite

Trypanosoma vivax, illus.
van den Ingh, T. S. G. A. M.; and de Neijs-Bakker, M. H., 1979, Tropenmed. u. Parasitol., v. 30 (2), 239-243
Trypanosoma vivax-infected cattle, mononuclear pancarditis with extravascular trypanosomes, considered to be local immune response to these extravascular trypanosomes

Trypanosoma vivax
Isoun, T. T.; Isoun, M. J.; and Anosa, V. O., 1978, Tropenmed. u. Parasitol., v. 29 (3), 330-334
Trypanosoma vivax-infected sheep, free plasma amino acid profiles of infected sheep and the trypanosome organism, significance of amino acid changes in terms of host and parasite metabolism

Trypanosoma vivax
Kaaya, G. P.; et al., 1979, Tropenmed. u. Parasitol., v. 30 (2), 230-235
Trypanosoma vivax, T. congolense, serum from infected cattle inhibited bovine granulocyte/macrophage colony formation in methyl cellulose culture, degree of inhibition appeared related to degree of parasitemia; no inhibitors of erythropoiesis were observed

Trypanosoma vivax
Kamara, J. A., 1975, Bull. Animal Health and Prod. Africa, v. 23 (3), 265-268
Cephalophus sylvicultrix
Tragelaphus s. scriptus
all from Sierra Leone

Trypanosoma vivax
Kilgour, V.; and Godfrey, D. G., 1978, Trop. Animal Health and Prod., v. 10 (3), 145-148
Trypanosoma vivax, Nigerian trade cattle, use of lorries for cattle transport from northern grazing areas to south-western markets, important factor in reduced infection rate: Nigeria

Trypanosoma vivax
Leeflang, P.; Buys, J.; and Blotkamp, C., 1978, Internat. J. Parasitol., v. 8 (1), 15-18
Trypanosoma vivax, cattle, evaluation of parasitological diagnostic methods (thin and thick blood smear and lymph gland smear examination, haematocrit centrifuge technique, hypotonic lysis test, mouse inoculation)

Trypanosoma vivax
Luckins, A. G., 1977, Trop. Animal Health and Prod., v. 9 (1), 53-62
Trypanosoma spp., cattle, immunodiagnosis, micromodification of enzyme-linked immunosorbent assay, test did not distinguish between different trypanosome species but there were no cross-reactions with other protozoal antigens

Trypanosoma vivax
Luckins, A. G.; and Mehlitz, D., 1978, Trop. Animal Health and Prod., v. 10 (3), 149-159
trypanosomiasis, cattle, diagnosis, indirect fluorescent antibody test, enzyme-linked immunosorbent assay, and serum IgM levels compared: Liberia

Trypanosoma vivax
MacKenzie, P. K. I.; Boyt, W. P.; and Nesham, V. W., 1979, Brit. Vet. J., v. 135 (2), 178-184
trypanosomiasis, sheep (nat. and exper.), serum immunoglobulin levels during course of infection

Trypanosoma vivax
Maxie, M. G.; Tabel, H.; and Losos, G. J., 1978, Tropenmed. u. Parasitol., v. 29 (2), 234-238
Trypanosoma vivax, T. congolense, determination of volume of parasites separated from cattle blood during first growth phase of infections

Trypanosoma (Duttonella) vivax
Mehlitz, D., 1979, Tropenmed. u. Parasitol., v. 30 (2), 212-219
Trypanosoma spp., infection rates in domestic animals; screening of dogs and pigs to indicate potential reservoirs of T. (Trypanozoon) brucei gambiense; comparison of sensitivity of 3 diagnostic techniques; comparison of number of primary isolations and derived stabilated Trypanosoma stocks
cattle
goats
sheep
all from rain forest areas, Liberia

Trypanosoma vivax
Rurangirwa, F. R.; et al., 1978, Research Vet. Sc., v. 25 (3), 395-397
Trypanosoma congolense, T. vivax, cattle, immunosuppressive effect on secondary humoral immune response to Mycoplasma m. mycoides

Trypanosoma vivax
Rurangirwa, F. R.; et al., 1979, Infect. and Immun., v. 26 (3), 822-826
Trypanosoma congolense- or T. vivax-infected Bos indicus, suppression of antibody response to Leptospira biflexa and Brucella abortus and recovery from immunosuppression after berenil treatment

Trypanosoma vivax
Saror, D. I., 1979, Vet. Rec., v. 105 (5), 96-98
Trypanosoma vivax, zebu cattle (exper.), haematological changes, classification of anemia

Trypanosoma vivax
Staak, C.; and Lohding, A., 1979, Tropenmed. u. Parasitol., v. 30 (1), 13-18
Trypanosoma spp., evaluation of complement fixation antibody in African trypanosomiasis-infected cattle before and after treatment and followed by re-infection and treatment; applications for field studies

Trypanosoma vivax
Taze, Y.; and Gruvel, J., 1978, Rev. Elevage et Med. Vet. Pays Trop., n. s., v. 31 (1), 63-67
trypanosomiasis, sample surveys to evaluate effects of Glossina eradication and mass trypanocide treatment of animals after 2 year period, recommendations: Low-Chari Valley, Chad

Trypanosoma vivax
Tizard, I.; et al., 1978, Microbiol. Rev., v. 42 (4), 661-681
African trypanosomes, biologically active products and pathogenesis

Trypanosoma vivax
Toure, S. M.; et al., 1978, Rev. Elevage et Med. Vet. Pays Trop., n. s., v. 31 (3), 293-313
Trypanosoma vivax, T. congolense, zebu and N'Dama cattle, pathology compared, N'Dama not as susceptible as zebu and some displayed a remarkable immunity: Missira, Senegal

Trypanosoma vivax
Trager, W., 1974, Ciba Found. Symp., n.s. (20), 225-254
trypanosomes, leishmanias, nutrition and biosynthetic capabilities, problems of in vitro cultivation and differentiation, review

Trypanosoma vivax, illus.
Vickerman, K., 1974, Ciba Found. Symp., n.s. (20), 171-198
trypanosomatid flagellates, ultrastructure, review with emphasis on changes during life cycles

Trypanosoma vivax, illus.
Vickerman, K., 1974, Ciba Found. Symp., n.s. (25), 53-80
antigenic variation in African trypanosomes, review

Trypanosoma vivax
Whitelaw, D. D.; et al., 1979, Research Vet. Sc., v. 26 (1), 102-107
Trypanosoma spp., cattle, mice, suppressed antibody response to louping-ill vaccine, value of diminazene therapy in alleviating this effect

T[rypanosoma] vivax
Zwart, D., 1979, Vet. Sc. Comm., v. 3 (3), 187-206
T[rypanosoma] vivax, 3 mouse-infective strains, review: parasitology (history, morphology, surface coat, tsetse transmission, infectivity and virulence for rodents), clinical and pathological observations in ruminants (virulence, anaemia, free serum amino acids, bradykinin, serotonin), vascular leakage, thrombus formation, fever, myocarditis, drug susceptibility

Trypanosomatidae
Amato Neto, V.; da Silva, L. J.; and Camargo, M. E., 1977, Rev. Inst. Med. Trop. S. Paulo, v. 19 (2), 99-102
serodiagnostic tests for Chagas disease performed on sera of patients with visceral leishmaniasis gave positive results with several immunologic methods, indicates "group-reactions" within Trypanosomatidae

Trypanosomatidae
Evans, D. A., 1978, Methods Cultiv. Parasites in Vitro, 55-88
Kinetoplastida, cultivation, review

Trypanosomatidae
Wallace, F. G., 1977, Protozoology, v. 3, 51-56
trypanosomatid flagellates, developmental stages, terminology, historical review

Trypanosomes
Becker, C. D.; and Overstreet, R. M., 1979, J. Fish Dis., v. 2 (6), 469-479
Brevoortia patronus
Trachinotus carolinus
Micropogonias undulatus
Prionotus tribulus
Citharichthys spilopterus
Trinectes maculatus
Symphurus plagiusa
(blood of all): all from northern Gulf of Mexico

Trypanosomes, illus.
  Cohen, C., 1973, South African Med. J., v. 47 (11), 485-491
    trypanosomiasis, humans, clinical presentation and histopathological findings of 5 cases, need for inclusion in differential diagnosis and use of confirmatory tests discussed: Witwatersrand, South Africa

Trypanosomes
  Godfrey, D. G., 1977, Protozoology, v. 3, 33-49
    identification of morphologically similar trypanosomes of mammals

Trypanosomes
  Gray, A. R., 1977, Ann. Soc. Belg Med. Trop., v. 57 (4-5), 403-408
    trypanosomes, particularly Trypanosoma gambiense, syringe-passaged and cyclically transmitted isolates, antigenic variations

Trypanosome, illus.
  Hembree, S. C., 1979, Mosquito News, v. 39 (3), 575-582
    Toxorhynchites splendens: Thailand

Trypanosomes
  Joshi, B. D., 1979, Proc. Indian Acad. Sc., Sect. B, Animal Sc., v. 88 (1, pt. 1), 59-63
    Notopterus notopterus
    Puntius stigma
    Cirrhina mrigala
    Labeo bata
    Wallago attu
    Heteropneustes fossilis
    Clarias batrachus
    Mystus aor
    M. seenghala
    M. vittatus
    Channa punctatus
    C. gachua
    Mastocembelus armatus
    (blood of all): all from Lucknow (U.P.), India

Trypanosome
  Landau, I.; and Chabaud, A. G., 1978, Ann. Parasitol., v. 53 (3), 247-253
    Hipposideros cyclops: Makokou (Gabon)

Trypanosomes
  Letch, C. A., 1979, Parasitology, v. 79 (1), 107-117
    Carassius carassius: River Lee at Enfield Lock

Trypanosome (subgenus Megatrypanum)
  Marinkelle, C. J., 1977, J. Wildlife Dis., v. 13 (3), 262-264
    Taphozous nudiventris (blood): Iraq

Trypanosomes
  Mills, J. N.; and Valli, V. E. O., 1978, Tropenmed. u. Parasitol., v. 29 (1), 95-100
    T[rypanosoma] congolense, rapid automated cytofluorometric method of counting trypanosomes

Trypanosomes
  Moors, A.; Van Meirvenne, N.; and Janssens, P. G., 1973, Ann. Soc. Belge Med. Trop., v. 53 (2), 97-108
    techniques used to separate cells and viruses reviewed and summarized (centrifugation, filtration, gel-filtration, counter-current distribution, chromatography), application to the isolation of trypanosomes discussed

Trypanosomes
  Peirce, M. A.; and Mead, C. J., 1977, J. Nat. Hist., v. 11 (6), 597-600
    Pyrrhula pyrrhula: near Tring, Hertfordshire

Trypanosomes
  Sogandares-Bernal, F.; and Chandler, J., 1978, J. Parasitol., v. 64 (3), 547-548
    Haemobartonella muris infections of laboratory mice as problem in routine passage of trypanosomes, method devised to remove most of Haemobartonella from donor blood sample to be transferred to recipient

Trypanosomes
  Tandon, R. S.; and Chandra, S., 1977, Ztschr. Parasitenk., v. 52 (3), 195-198
    trypanosome-infected fish, reduced serum alkaline phosphatase levels and lowered metabolic activity

Trypanosomes
  Tandon, R. S.; and Chandra, S., 1977, Ztschr. Parasitenk., v. 52 (3), 199-202
    trypanosome-infected fishes, lowered serum cholesterol levels, possible causes

Trypanosomes
  Vickerman, K., 1978, Tr. Am. Micr. Soc., v. 97 (4), 485-502
    trypanoplasm-like flagellates, free-living, possibly mistakenly identified as trypanosomes and Trichomonas, potential cause of confusion in diagnosis of human and animal diseases

Trypanosomes
  Vickerman, K.; and Tetley, L., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 441-457
    Trypanosoma brucei, T. cyclops, application of scanning electron microscopic techniques to study of trypanosome biology

Trypanosomes, Malaysian isolates
  Weinman, D.; et al., 1978, Am. J. Trop. Med. and Hyg., v. 27 (2, pt. 1), 232-237
    experimental infections in Rhodnius prolixus "suggested survival and not multiplication"
    Macaca ira
    M. nemestrina
    all from Malaysia

Trypanosomes
  Wetzel, H.; and Luger, D., 1978, Tropenmed. u. Parasitol., v. 29 (2), 239-251
    in vitro feeding in rearing of tsetse flies

Trypanosomiasis
  Adekolu-John, E. O., 1978, Trop. and Geogr. Med., v. 30 (3), 285-294
    trypanosomiasis, transhumance pastoral mobility of Fulani nomads as possible important factor in any future outbreaks: Kainji Lake area, Nigeria

Trypanosomiasis
  Arandas, E. M. C. de A.; Loureiro, S.; and Azevedo, E. S., 1974, Rev. Inst. Med. Trop. S. Paulo, v. 16 (5), 283-285
    possible effect of human hemoglobin S on blood meal size of Rhodnius prolixus (vector of human South American trypanosomiasis)

Trypanosomiasis
  Bailenger, J.; et al., 1972, Therapeutique, v. 48 (2), 117-128
    dangers of cortisone therapy in the presence of human parasitic pathology, clinical review

Trypanosomiasis
Basson, W.; Page, M. L.; and Myburgh, D. P., 1977, South African Med. J., v. 51 (14), 453-457
trypanosomiasis, humans, problems in diagnosis and management emphasizing haematological and immunological aspects, clinical course in 5 cases, indications of continuation of immune complex process in spite of therapy that eradicates parasite: Voortrekkerhoogte, Tvl.

Trypanosomiasis
Bone, G. J.; and Charlier, J., 1975, Ann. Soc. Belge Med. Trop., v. 55 (5), 559-569
African trypanosomiasis, humans, capillary tube indirect hemagglutination test, useful diagnostic tool for field surveys

Trypanosomiasis
Bourn, D.; and Scott, M., 1978, Trop. Animal Health and Prod., v. 10 (4), 191-203
trypanosomiasis, successful use of Zebu work oxen in agricultural development of tsetse infested land, environmental conditions, epizootiology of trypanosomiasis in oxen and in Glossina morsitans, strategic drug use (alternation of diminazene aceturate and isometamidium to control trypanosomes; rafoxanide to control helminths): Wollega province, western Ethiopia

Trypanosomiasis
Busvine, J. R., 1977, Linn. Soc. Symposium Ser. (5), 145-150
human African and South American trypanosomiasis, currently used vector control measures

Trypanosomiasis
Buyst, H., 1975, Ann. Soc. Belge Med. Trop., v. 55 (5), 551-557
rhodesiense sleeping sickness, humans, diagnostic review of 400 cases (blood smear, cerebro-spinal fluid smear, gland aspirate): Zambia

Trypanosomiasis
Castagnino, H. E.; Cicco, J. A.; and Thompson, A., 1978, Semana Med. (5023), an. 85, v. 152 (6), 165-171, 178
trypanosomiasis, human, chronic myocarditis, clinical pathology, causes of death

Trypanosomiasis
Cohen, S. S., 1979, Science (4410), v. 205, 964-971
development of selective chemotherapeutic agents exploiting biochemical differences between disease agents and their hosts, review, includes discussion of several tropical diseases

Trypanosomiasis
Couvreur, J.; and Desmonts, G., 1977, Rev. Prat., Paris, v. 27 (11), 647-648, 653-658
Toxoplasma gondii, trypanosomiasis, malaria, leishmaniasis, parasitic pathology of foetus, review

Trypanosomiasis
Desfontaine, M.; et al., 1979, Med. Trop., v. 39 (5), 509-516
trypanosomiasis, humans, current epidemiologic situation in 7 O.C.C.G.E. nations (Benin, Ivory Coast, Upper Volta, Mali, Niger, Senegal, and Togo)

Trypanosomiasis
Dias, J. A. T. S.; Goncalves, A. C. B.; and Martins, J. N. O., 1971, Rev. Cien. Vet., Lourenco Marques, s. B, v. 14, 5-144
trypanosomiasis, bovine, extensive description of control program, results, and prospects: region of Muda (Sofala district)

Trypanosomiasis
Dunn, F. L., 1979, Bull. World Health Organ., v. 57 (4), 499-512
behavioral aspects of control of parasitic diseases, review

Trypanosomiasis
Duvallet, G.; et al., 1979, Med. Trop., v. 39 (5), 517-526
trypanosomiasis, humans, increased incidence in area of Vavoua, clinical, parasitological, and sero-immunological survey: Ivory Coast

Trypanosomiasis
Fernandes, W. J.; et al., 1972, Rev. Patol. Trop., v. 1 (1), 39-44
toxoplasmosis, serological testing of exceptional children, cross-reaction in children infected with American trypanosomiasis, no correlations found between mental diseases and these infections

Trypanosomiasis
Fernex, M., 1979, Therap. Umschau, v. 36 (3), 205-210
tropical protozoan diseases, humans travelling to endemic areas, preventive measures, review

Trypanosomiasis
Forattini, O. P.; et al., 1977, Rev. Saude Pub., S. Paulo, v. 11 (1), 73-86
Panstrongylus megistus, distribution patterns: areas in southern Brazil

Trypanosomiasis
Forattini, O. P.; et al., 1977, Rev. Saude Pub., S. Paulo, v. 11 (2), 199-213
Panstrongylus megistus, annual variations of nymph and adult populations and their mobility in artifical ecotopes: Brazil

Trypanosomiasis
Forattini, O. P.; et al., 1977, Rev. Saude Pub., S. Paulo, v. 11 (3), 362-374
population data on experimental colonies of Panstrongylus megistus and Triatoma sordida

Trypanosomiasis
Frezil, J. L.; and Coulm, J., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 315-322
African trypanosomiasis, human, current control measures: Republique populaire du Congo

Trypanosomiasis
Gill, B. S., 1977, Trypanosomes and trypanosomiases of Indian livestock, 137 pp., illus.
extensive review

Trypanosomiasis
Giordano, C.; et al., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 213-225
African trypanosomiasis, humans, diagnosis (radiology, electroencephalography, immunofluorescence, immunoelectrophoresis, electrophoresis)

Trypanosomiasis
Hedge, E. C.; Moody, A. H.; and Ridley, D. S., 1978, Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (4), 445 [Letter]
Leishmania donovani, human, screening of sera for kala-azar using Crithidia sp. as antigen in indirect fluorescent antibody test, some positive reactions with L. tropica, cross-reactions at low titer with Chagas' disease and African trypanosomiasis

Trypanosomiasis
Herrer, A., 1977, Rev. Peruana Entom., v. 20 (1), 19-24
trypanosomiasis, leishmaniasis, bartonellosis, human, transmission by insect vectors, control, review: Peru

Trypanosomiasis
Huhn, J., 1977, 3. Externe Veterinartagung, Ber. u. Arbeitsergebn. (40), 217-227
trypanosomiasis, cattle, possible mechanisms of tolerance, use of resistant breeds as control measure, economic importance: West Africa

Trypanosomiasis
Janssens, P. G., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 191-200
African sleeping sickness, historical review

Trypanosomiasis
Janssens, P. G.; and de Muynck, A., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 475-480
African trypanosomiasis, humans, clinical trials with nifurtimox

Trypanosomiasis
Jordan, A. M., 1978, Nature, London (5664), v. 273, 607-609
principles of eradication or control of tsetse flies, review

Trypanosomiasis
Kaschula, V. R.; Van Dellen, A. F.; and de Vos, V., 1978, J. South African Vet. Ass., v. 49 (3), 223-227
antibody levels against several parasitic infections in Cercopithecus aethiops pygerythrus: South Africa

Trypanosomiasis
Killick-Kendrick, R., 1977, Protozoology, v. 3, 109-117
early records of trypanosomiasis in Nigerian livestock

Trypanosomiasis
Lafaye, A.; and Saliou, P., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 335-347
trypanosomiasis, humans, statistical evaluation of immunological diagnostic findings

Trypanosomiasis
Laveissiere, C., 1976, Cahiers O.R.S.T.O.M., s. Entom. Med. et Parasitol., v. 14 (4), 359-367
trypanosomiasis, human, hypothesis on transmission without tsetse flies (antitubercular treatment stimulated virility of trypanosomes in carriers, transmitted mechanically by mosquitoes or bedbugs): Ouahigouya, Republique de Haute-Volta

Trypanosomiasis
Mangenot, M.; et al., 1979, Med. Trop., v. 39 (5), 527-530
African trypanosomiasis, humans, detection of foci, ELISA vs. immunofluorescence

Trypanosomiasis
Mangenot, M.; et al., 1979, Med. Trop., v. 39 (5), 531-535
African trypanosomiasis, humans, possible use of immunoglobulin assay in confirming diagnosis (increased IgG and IgM) in persons suspected to be infected after testing with immunofluorescence or ELISA

Trypanosomiasis
Mansfield, J. M., 1978, Cellular Immunol., v. 39 (1), 204-210
African trypanosomiasis, lymphocyte dysfunction and immunosuppression (histopathological considerations; B cell function; T cell function; macrophage function), cellular bases of immunosuppression, review

Trypanosomiasis
Mercenier, P., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 323-329
African trypanosomiasis, human, potential of multiple health services in detecting and controlling infections: Kasongo

Trypanosomiasis
Moulton, J. E.; and Stevens, D. R., 1978, Am. J. Path. (433), v. 91 (3), 693-696
trypanosomiasis, Peromyscus maniculatus as laboratory model for human disease, review

Trypanosomiasis
Ngandu-Kabeya, G., 1976, Ann. Soc. Belge Med. Trop., v. 56 (2), 85-93
African trypanosomiasis, children, classical features, frequency of central nervous system involvement: Zaire

Trypanosomiasis
Norval, R. A. I., 1978, Rhodesian Vet. J., v. 9 (1-4), 9-16
ticks, cattle, significant increase in tick infestations and outbreaks of tick-borne diseases following collapse of dipping (1973-1978): African areas in Rhodesia

Trypanosomiasis
Okiwelu, S. N., 1979, Ann. Trop. Med. and Parasitol., v. 73 (1), 73-77
observations on Glossina m. mortisans at a tsetse disinfection picket during rainy season (age composition, trypanosome infection rate, pregnancy rate, sex ratio, and settling behavior): Republic of Zambia

Trypanosomiasis
Palmero, H. A.; Caeiro, T. F.; and Iosa, D. J., 1977, Rev. Argent. Cardiol., v. 45 (5), 415-427
human trypanosomiasis, evaluation of age, sex and weight of Chagasic patients, possible effects on arterial blood pressure

Trypanosomiasis
Playfair, J. H. L., 1978, Current Topics Microbiol. and Immunol., v. 80, 37-64
effective and ineffective immune responses to parasites, evidence from experimental models, review with emphasis on malaria and trypanosomiasis

Trypanosomiasis
Poltera, A. A.; and Cox, J. N., 1977, Virchows Arch. A, Path. Anat. and Histol., v. 375 (1), 53-70
pancarditis with valvulitis in endomyocardial fibrosis and in human African trypanosomiasis, comparative study of 2 cases of each, possible relationship: Uganda

Trypanosomiasis
Ruitenberg, E. J.; et al., 1977, Biomedicine, v. 26 (5), 311-314
human parasitic infections, enzyme-linked immunosorbent assay in diagnosis, brief review

Trypanosomiasis
Ruppol, J. F.; and Libala, K., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 299-314
African sleeping sickness, humans, current control measures, geographic distribution in Zaire

Trypanosomiasis
Schalm, O. W., 1979, Canine Pract., Santa Barbara, v. 6 (6), 46-49
trypanosomiasis, Leishmania donovani, and other uncommon blood disorders, dogs, case reports

Trypanosomiasis
Staak, C., 1977, 3. Externe Veterinartagung, Ber. u. Arbeitsergebn. (40), 193-207
trypanosomiasis, several diagnostic methods evaluated under laboratory and field conditions

Trypanosomiasis
Sterner, G.; and Nåsander, L., 1977, Scand. J. Infect. Dis., v. 9 (2), 154-156
trypanosomiasis, probably Gambian type, Swedish woman who had vacationed in Gambia, case report, discussion of possible risk of Europeans acquiring infection while touring African nations

Trypanosomiasis
Tr. Roy. Soc. Trop. Med. and Hyg., v. 72 (2), 1978, 109-122
18. Seminar on trypanosomiasis, (discussion groups on: protozoology, entomology including insecticides, biochemistry including chemotherapy, immunology, pathology, epidemiology)

Trypanosomiasis
Tropenmed. u. Parasitol., 1976, v. 27 (special no. 1), 77 pp.
onchocerciasis, trypanosomiasis, paragonimiasis, humans, papers presented at the 8th Congress of the German Society of Tropical Medicine

Trypanosomiasis
Van Meirvenne, N.; et al., 1975, Ann. Soc. Belge Med. Trop., v. 55 (5), 545-549
gambian and rhodesian sleeping sickness, humans, serological and parasitological diagnostic methods, review

Trypanosomiasis
Voller, A., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 273-280
African trypanosomiasis, humans, various immunoserologic diagnostic tests, review

Trypanosomiasis
Voller, A., 1978, New Trends and Develop. Vaccines, 299-309
schistosomiasis, trypanosomiasis, malaria, potential and progress towards vaccines, review

Trypanosomiasis
Watanabe, Y., 1975, Rinsho Fujinka Sanka (Clin. Gynec. and Obst.), v. 29 (8), 625-628
pathological changes in human placenta due to infection, includes Toxoplasma gondii, Plasmodium, and trypanosomiasis

Trypanozoon
Evans, D. A., 1978, Methods Cultiv. Parasites in Vitro, 55-88
Kinetoplastida, cultivation, review

Trypanozoon, subgenus
Godfrey, D. G., 1977, Protozoology, v. 3, 33-4
identification of morphologically similar trypanosomes of mammals

Trypanozoon
Van Meirvenne, N.; Magnus, E.; and Vervoort, T., 1977, Ann. Soc. Belge Med. Trop., v. 57 (4-5), 409-423
trypanosome strains of subgenus Trypanozoon, comparison of variable antigenic types

Tuzetia debaisieuxi (Jirovec, 1943) [n. comb.], illus.
Maurand, J.; and Loubes, C., 1978, Ztschr. Parasitenk., v. 56 (2), 131-146
microsporidians from simuliid larvae, ultrastructural studies
simuliid larvae: region languedocienne (Sud de la France)

Tuzetia debaisieuxi, illus.
Loubes, C., 1979, J. Protozool., v. 26 (2), 200-208
Microsporida, synaptonemal complexes demonstrated in 6 genera but not in Nosema, implications for life cycles

Unicauda sp.
Joy, J. E.; Tarter, D. C.; and Franklin, H., 1978, Tr. Am. Micr. Soc., v. 97 (1), 100-104
Octomacrum spinum and Unicauda sp. in Campostoma anomalum (gills), prevalence and parasite load, sex of parasite, seasonal occurrence: Fourpole Creek, Cabell Co., West Virginia

Unicauda sp.
Markiw, M. E.; and Wolf, K., 1978, J. Fish. Research Bd. Canada, v. 35 (6), 828-832
Myxosoma cerebralis, rabbits immunized with antigens extracted from mature spores or prespore stages, antisera and globulins used in fluorescent antibody techniques, direct fluorescent antibody test showed higher specificity than indirect FAT in cross reactions with other species of myxosporidians

Unikaryon minutum Knell and Allen
Atkinson, T. H.; and Wilkinson, R. C., 1979, Florida Entom., v. 62 (3), 169-175
Unikaryon minutum and Contortylenchus brevicomi in Dendroctonus frontalis, incidence in live-trapped males vs. reared males and females; modified livetrap: Eglin AFB Forest, FL; near Athens, GA

Urophagus intestinalis Moroff (1903)
Vojtkova, L., 1976, Scripta Fac. Scient. Nat. Univ. Purkynianae Brun., Biol., v. 6 (5), 177-209
as syn. of Hexamita intestinalis Dujardin, 1841

Vahlkampfia sp.
De Jonckheere, J. F., 1979, Applied and Environment. Microbiol., v. 38 (4), 590-593
Naegleria and Acanthamoeba, incidence in aquaria, may be source of human infection: Belgium

Vahlkampfia sp.
Kadlec, V., 1978, J. Protozool., v. 25 (2), 235-237
15 strains of free-living amebae found in variety of domestic animals, 3 isolates tested were not pathogenic for laboratory animals, some features of isolates differed from those previously known for members of these genera
bull (prepucial cavity)

Vahlkampfia sp., illus.
Pernin, P.; and Riany, A., 1978, Ann. Parasitol., v. 53 (4), 333-344
survey of swimming pools for presence of free-living amebae, potential danger for swimmers: Lyon

Vahlkampfia avara
Kadlec, V., 1978, J. Protozool., v. 25 (2), 235-237
15 strains of free-living amebae found in variety of domestic animals, 3 isolates tested were not pathogenic for laboratory animals, some features of isolates differed from those previously known for members of these genera
swine (nasal cavity)

Vahlkampfia enterica
Kadlec, V., 1978, J. Protozool., v. 25 (2), 235-237
15 strains of free-living amebae found in variety of domestic animals, 3 isolates tested were not pathogenic for laboratory animals, some features of isolates differed from those previously known for members of these genera
turkey (intestine)

Vahlkampfia inornata
Kadlec, V., 1978, J. Protozool., v. 25 (2), 235-237
15 strains of free-living amebae found in variety of domestic animals, 3 isolates tested were not pathogenic for laboratory animals, some features of isolates differed from those previously known for members of these genera
sow (nasal cavity)

Vahlkampfiidae [sp.], illus.
Rinaldi, I.; et al., 1979, Virginia Med., v. 106 (9), 670-673
Vahlkampfiidae [sp.], woman (brain), chronic primary amebic meningoencephalitis, case report: Smithfield, Virginia

Vahlkampfiidae
Rivera, F.; et al., 1979, Tr. Am. Micr. Soc., v. 98 (3), 465-469
protozoa polluting tap water, concentration and identification in culture: Federal District, Mexico City

Vairimorpha new genus
Pilley, B. M., 1976, J. Invert. Path., v. 28 (2), 177-183
Microsporida
tod: V. necatrix (Kramer, 1965) [n. comb.]

Vairimorpha
Knell, J. D.; and Zam, S. G., 1978, J. Invert. Path., v. 31 (3), 280-288
Nosema spp., double immunodiffusion techniques used to investigate taxonomic relationships between 6 different microsporidian isolates
". . . Nosema necatrix isolates 942, 954, and 960 and Nosema plodiae . . . should be considered members of the genus Vairimorpha."

Vairimorpha necatrix (Kramer, 1965) [n. comb.] (tod), illus.
Pilley, B. M., 1976, J. Invert. Path., v. 28 (2), 177-183
Vairimorpha necatrix [n. comb.] in Spodoptera exempta, pathogenicity (occurrence of bacteriosis and cytoplasmic polyhedrosis virus), life cycle (disporoblastic life cycle at 25°C and both disporoblastic and octosporoblastic life cycle at 20°C), implications of polymorphism in relation to classification of Microsporida
Syns.: Thelohania diazoma Kramer 1965;
Nosema necatrix Kramer 1965
Spodoptera exempta (fat body) (exper.)

Vairimorpha necatrix (Kramer)
Chu, W. H.; and Jaques, R. P., 1979, Entomophaga, v. 24 (3), 229-235
Vairimorpha necatrix in Trichoplusia ni (exper.) (fat body, muscle tissue, midgut), symptoms, histopathology

Vairimorpha necatrix
Fuxa, J. R., 1979, J. Invert. Path., v. 33 (3), 316-323
Vairimorpha necatrix, interactions with bacterium, virus, and fungus in Heliothis zea (exper.)

Vairimorpha necatrix
Fuxa, J. R.; and Brooks, W. M., 1979, J. Invert. Path., v. 33 (1), 86-94
Vairimorpha necatrix, mass production and storage methods

Vairimorpha necatrix (Kramer)
Ignoffo, C. M.; and Garcia, C., 1979, J. Econom. Entom., v. 72 (5), 767-769
Agrotis ipsilon, susceptibility to Vairimorpha necatrix and other entomopathogens

Vairimorpha necatrix (Kramer)
Jordan, J. A.; and Noblet, R., 1978, Tech. Bull. (1069) South Carolina Agric. Exper. Station, 5 pp.
Nosema heliothidis and Vairimorpha necatrix, susceptibility of selected insect pests, spore dose levels necessary to obtain LD_{50} in selected host age levels
Heliothis zea
H. virescens
Spodoptera frugiperda
Galleria mellonella
(all exper.)

Vairimorpha necatrix
Kaya, H. K., 1977, J. Invert. Path., v. 30 (2), 192-198
Vairimorpha necatrix (potential biological control agent), survival (infectivity) of spores exposed to sunlight, ultraviolet radiation, and high temperature, laboratory and field tests

Vairimorpha (Nosema) necatrix (Kramer)
  Kaya, H. K., 1979, Proc. Hawaiian Entom. Soc. (1976), v. 13 (1), 91-94
    Pleistophora sp., Vairimorpha necatrix, retention of infectivity after passage through gut of Zelus exsanguis
    Spodoptera frugiperda
    Estigmene acrea (exper.)

Vairimorpha necatrix
  Pilley, B. M., 1978, J. Invert. Path., v. 31 (3), 341-344
    Vairimorpha necatrix, storage of infective spores in antibiotic solution at 4°C

Vauchomia nephritica Mueller, 1938, illus.
  Lom, J.; and Haldar, D. P., 1976, Tr. Am. Micr. Soc v. 95 (4), Finley Mem. Issue, pt. 2, 527-541
    Syn.: Trichodina (Vauchomia) nephritica Lom, 1958
    trichodinids endoparasitic in fishes, survey of literature records together with some new observations
    Esox masquinongy: Chautauqua Lake, New York

Vauchomia renicola (Mueller, 1931) Mueller, 1938, illus.
  Lom, J.; and Haldar, D. P., 1976, Tr. Am. Micr. Soc., v. 95 (4), Finley Mem. Issue, pt. 2, 527-541
    synonymy
    trichodinids endoparasitic in fishes, survey of literature records together with some new observations
    Esox niger: Oneida Lake, New York

Vexillifera bacillipedes Page, 1969, illus.
  Sawyer, T. K.; et al., 1978, Tr. Am. Micr. Soc., v. 97 (4), 596-600
    Vexillifera bacillipedes pathogenic to rainbow trout, morphology of culture specimens, free-living and endozoic forms compared
    rainbow trout (kidney lesions): hatchery, Italy

Wardia schulmani sp. n., illus.
  Bocharova, T. A.; and Donets, Z. C., 1974, Parazitologiia, Leningrad, v. 8 (1), 74-76
    Acerina cernua (kidney tubules): Tukhemitor lake (middle course of Vasiugan river)

Weiseria spinosa Golberg, 1971
  Kilochitskii, P. Ia., 1977, Vestnik Zool., Akad. Nauk Ukrainsk. SSR, Inst. Zool. (4), 71-75
    Culex theileri: southern Ukraine

Wenyonella baghdadensis sp. n., illus.
  Mirza, M. Y.; and al-Rawas, A. Y., 1978, J. Protozool., v. 25 (3, pt. 2), 285-286
    Nesokia indica (feces): Baghdad district, Iraq

Xysteroides subgen. n.
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 25-27
    subgen. of Zelleriella
    type sp.: Zelleriella (Xysteroides) telmatobii Metcalf, 1923

Xysteroides
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
    subgen. of Zelleriella, key

Zelleriella Metcalf, 1920
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 25-27
    Opalinidae
    includes: Xysteroides subgen. n.; Promitotica subgen. n.; Zelleriella Metcalf, 1920; Antunesia subgen. n.; Cornucopioides subgen. n.

Zelleriella Metcalf, 1920
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
    Opalininae, key
    key to subgenera

Zelleriella Metcalf, 1920
  Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (5-6), 237-243
    Zelleriella, subgenera Promitotica, Antunesia, and Cornucopioides, synopsis of Brazilian species

Zelleriella Metcalf, 1920
  Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (5-6), 307-314
    Zelleriella, subgen. Zelleriella, synopsis of Brazilian species

Zelleriella Metcalf, 1920
  Amaro, A., 1971, Atas Soc. Biol. Rio de Janeiro, v. 14 (5-6), 135-141
    synopsis of recent species

Zelleriella Metcalf, 1920
  Earl, P. R., 1979, Tr. Am. Micr. Soc., v. 98 (4), 549-557
    Hegneriellina [sic]

Zelleriella Metcalf, 1920
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 25-27
    subgen. of Zelleriella
    type sp.: Zelleriella (Zelleriella) antilliensis (Metcalf, 1914) Metcalf, 1923

Zelleriella
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 49-52
    subgen. of Zelleriella, key

Zelleriella Metcalf, 1920
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
    subgen. of Zelleriella
    key to species

Zelleriella sp. Metcalf, 1923
  Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (5-6), 307-314
    as syn. of Zelleriella (Z.) ovonucleata Metcalf, 1940

Zelleriella sp.
  Amaro, A.; Sena, S.; and dos Santos, E., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 123
    Atelopus moreirae: Veu da Noiva, Maromba, em Itatiaia, no Estado do Rio de Janeiro, Brasil

Zelleriella sp.
  Amaro, A.; Sena, S.; and dos Santos, E., 1968, Atas Soc. Biol. Rio de Janeiro, v. 12 (3), 123
    Hyla cuspidata: Recreio dos Bandeirantes, Lagoinha das Taxas, na cidade do Rio de Janeiro, Estado da Guanabara, Brasil

Zelleriella (Zelleriella) antilliensis (Metcalf, 1914) Metcalf, 1923 (tod of subgen.), illus.
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 25-27

Zelleriella (Zelleriella) antilliensis (Metcalf, 1914), Metcalf, 1923
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
    key

Zelleriella (Zelleriella) antilliensis (Metcalf, 1914) Metcalf, 1923, illus.
  Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (5-6), 307-314
    Syn.: Opalina antilliensis Metcalf, 1914

Zelleriella (Antunesia) antunesi Pessoa, 1934 (tod of subgen.), illus.
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 25-27

Zelleriella (Antunesia) antunesi, 1934
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
    key

Zelleriella (Antunesia) antunesi quadrata Metcalf, 1940
  Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (5-6), 237-243

Zelleriella (Antunesia) artigasi Unti, 1935
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
    key

Zelleriella (Zelleriella) boipevae Carini, 1938
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
    key

Zelleriella (Zelleriella) bolivari Beltran, 1941, illus.
  Amaro, A., 1971, Atas Soc. Biol. Rio de Janeiro, v. 14 (5-6), 135-141
    brief description

Zelleriella (Zelleriella) brasiliensis (Pinto, 1918) Metcalf, 1940
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
    key

Zelleriella (Zelleriella) brasiliensis (Pinto, 1918) Metcalf, 1923, illus.
  Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (5-6), 307-314
    Syn.: Opalina brasiliensis Pinto, 1918

Zelleriella (Zelleriella) carinii Otamendi, 1945, illus.
  Amaro, A., 1971, Atas Soc. Biol. Rio de Janeiro, v. 14 (5-6), 135-141
    brief description
    Bufo arenarum: La Plata, provincia de Buenos Aires, republica Argentina

Zelleriella (Promitotica) caryosoma Amaro, 1964 (tod of subgen.), illus.
  Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 25-27

Zelleriella (Zelleriella) casinae Boisson, 1966
Amaro, A., 1971, Atas Soc. Biol. Rio de Janeiro, v. 14 (5-6), 135-141
brief description

Zelleriella (Antunesia) caudata Carini, 1938
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Zelleriella (Cornucopioides) corniola Carini, 1938
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Zelleriella (Cornucopioides) cornucopia Carini, 1933 (tod of subgen.), illus.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 25-27

Zelleriella (Cornucopioides) cornucopia Carini, 1933
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Zelleriella (Cornucopioides) cuneata Carini, 1938
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Zelleriella (Zelleriella) dubia Metcalf, 1940
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Zelleriella (Zelleriella) elliptica Chen, 1948
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 25-27

Zelleriella (Antunesia) excavata Carini, 1938
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Zelleriella (Cornucopioides) falcata Carini, 1933
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Zelleriella (Zelleriella) foliacea Carini, 1938
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Zelleriella foliacea Carini, 1938, illus.
Amaro, A.; and Sena, S., 1968, Atas Soc. Biol. Rio de Janeiro, v. 11 (6), 205-206
redescription
Leptodactylus ocellatus (intestino grosso (regiao cecal)): Manguinhos, Rio de Janeiro, Estado da Guanabara, Brasil

Zelleriella (Zelleriella) freitasi Amaro, 1965
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 25-27

Zelleriella (Zelleriella) freitasi Amaro, 1964
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Zelleriella (Zelleriella) frolanoi Uttangi, 1951, illus.
Amaro, A., 1971, Atas Soc. Biol. Rio de Janeiro, v. 14 (5-6), 135-141
brief description

Zelleriella (Zelleriella) indica Uttangi, 1961, illus.
Amaro, A., 1971, Atas Soc. Biol. Rio de Janeiro, v. 14 (5-6), 135-141
brief description

Zelleriella (Zelleriella) jaegeri Carini, 1938 [i. e. 1933]
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Zelleriella kyii n. sp. (Fig. 3 of Delphin et al., 1972)
Earl, P. R., 1979, Tr. Am. Micr. Soc., v. 98 (4), 549-557
Kaloula pulchra: Burma

Zelleriella (Zelleriella) leptodeirae Beltran, 1941, illus.
Amaro, A., 1971, Atas Soc. Biol. Rio de Janeiro, v. 14 (5-6), 135-141
brief description

Zelleriella (Zelleriella) louisianensis Chen, 1948
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 25-27

Zelleriella magna Carini, 1938
Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (5-6), 307-314
as syn. of Zelleriella (Z.) proterocarya Amaro, 1964

Zelleriella mandalayensis n. sp. (Fig. 2 of Delphin et al., 1972)
Earl, P. R., 1979, Tr. Am. Micr. Soc., v. 98 (4), 549-557
Microhyla ornata: Burma

Zelleriella (Zelleriella) menendezi Schouten, 1934
Amaro, A., 1971, Atas Soc. Biol. Rio de Janeiro, v. 14 (5-6), 135-141
brief description

Zelleriella (Zelleriella) microhylae Uttangi, 1951, illus.
Amaro, A., 1971, Atas Soc. Biol. Rio de Janeiro, v. 14 (5-6), 135-141
brief description

Zelleriella nucleolata sp. n., illus.
Amaro, A.; and Sena, S., 1967, Atas Soc. Biol. Rio de Janeiro, v. 11 (3), 89-90
Leptodactylus ocellatus (intestino grosso (regiao cecal)): Rio Bonito, Estado do Rio de Janeiro, Brasil

Zelleriella (Antunesia) nucleolata Amaro & Sena, 1967, illus.
Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (5-6), 237-243

Zelleriella ooi n. sp. (Fig. 4 of Delphin et al., 1972)
Earl, P. R., 1979, Tr. Am. Micr. Soc., v. 98 (4), 549-557
Tylototriton verrucosus: Burma

Zelleriella (Zelleriella) ovonucleata Metcalf, 1940
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Zelleriella (Zelleriella) ovonucleata Metcalf, 1940, illus.
Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (5-6), 307-314
synonymy

Zelleriella ovonucleata bufonis Metcalf, 1940
Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (5-6), 307-314
as syn. of Zelleriella (Z.) ovonucleata Metcalf, 1940

Zelleriella paulista (Pinto, 1926) comb. n., illus.
Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (5-6), 237-243
Syn.: Opalina paulista Pinto, 1926

Zelleriella (Zelleriella) pfitzneri Chen, 1948
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 25-27

Zelleriella (Zelleriella) proterocarya Amaro, 1964
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 25-27

Zelleriella (Zelleriella) proterocarya Amaro, 1964
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Zelleriella (Zelleriella) proterocarya Amaro, 1964, illus.
Amaro, A., 1969, Atas Soc. Biol. Rio de Janeiro, v. 12 (5-6), 307-314
Syn.: Zelleriella magna Carini, 1938

Zelleriella (Zelleriella) siphonopsi Carini, 1942
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Zelleriella (Xysteroides) telmatobii Metcalf, 1923 (tod of subgen.), illus.
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 25-27

Zelleriella (Zelleriella) truncata Carini, 1938
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Zelleriella (Zelleriella) uruguaiensis Metcalf, 1940
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Zelleriella (Zelleriella) uruguaiensis quadrata Metcalf, 1940
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (3), 53-55
key

Zelleriella (Zelleriella) valliceps Chen, 1948
Amaro, A., 1966, Atas Soc. Biol. Rio de Janeiro, v. 10 (2), 25-27

Zschokkella sp.?, illus.
Zaika, V. E., 1966, Respublik. Mezhvedomstv. Sborn., Akad. Nauk Ukrain. SSR, s. Biol. Moria, 13-31
Gobius batrachocephalus
Scorpaena porcus
(gall bladder of all): all from Black Sea

Zschokkella embiotocidis sp. n., illus.
Moser, M.; and Haldorson, L., 1976, Canad. J. Zool., v. 54 (8), 1403-1405
Damalichthys vacca
Embiotoca lateralis
(gallbladders): all from Diablo Cove, central California

Zschokkella nova Klokacewa
Iziumova, N. A.; Mashtakov, A. V.; and Kashkovskii, V. V., 1977, Inform. Biul. Inst. Biol. Vnutren. Vod, Akad. Nauk SSSR (36) 64-68
[Pelecus cultratus] (gall bladder): Kamsk reservoir

Zschokkella nova
Otvodova, G. D., 1975, Izvest. Gosudarstv. Nauchno-Issled. Inst. Ozer. i Rech. Ryb. Koziaistva, Leningrad, v. 93, 112-116
[Abramis brama] (gills): Pskov-Chudskoe lake

Zschokkella orientalis
Makhovenko, E. T., 1972, Parazitologiia, Leningrad, v. 6 (4), 369-375
Salvelinus alpinus inhabiting different ecological niches, possible use of differences in parasite fauna between groups as biological tags: Lake Azabach'e, Kamchatka

Zygocystis pterotracheae Stuart, 1871
Levine, N. D., 1979, J. Protozool., v. 26 (4), 532-536
as syn. of Molluskocystis pterotracheae (Stuart, 1871) comb. n.

Zygocystis violacea sp. n., illus.
Segun, A. O., 1978, J. Protozool., v. 25 (2), 157-162
Libyodrilus violaceus (perivisceral coelom): Ibadan, Western Nigeria